Expression and Function of β1 Integrins on Human Eosinophils

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Eosinophils preferentially accumulate at sites of chronic allergic diseases such as bronchial asthma. The mechanisms by which selective eosinophil migration occurs are not fully understood. However, interactions of cell-surface adhesion molecules on the eosinophil with molecular counterligands on endothelial and epithelial cells, and on extracellular matrix proteins, are likely to be critical during the recruitment process. One possible mechanism for selective eosinophil recruitment involves the α4β1 (VLA-4) integrin which is not expressed on neutrophils. Correlations have been found between infiltration of eosinophils and endothelial expression of VCAM-1, the ligand for VLA-4, in the lungs of asthmatic individuals as well as in late phase reactions in the lungs, nose and skin. Epithelial and endothelial cells respond to the Th2-type cytokines IL-4 and IL-13 with selective de novo expression of VCAM-1, consistent with the possible role of VCAM-1/VLA-4 interactions in eosinophil influx during allergic inflammation. Both β1 and β2 integrins on eosinophils exist in a state of partial activation. For example, eosinophils can be maximally activated for adhesion to VCAM-1 or fibronectin after exposure to β1 integrin-activating antibodies or divalent cations, conditions that do not necessarily affect the total cell surface expression of β1 integrins. In contrast, cytokines like IL-5 prevent β1 integrin activation while promoting β2 integrin function. Furthermore, ligation of integrins can regulate the effector functions of the cell. For example, eosinophil adhesion via β1 and/or β2 integrins has been shown to alter a variety of functional responses including degranulation and apoptosis. Thus, integrins appear to be important in mediating eosinophil migration and activation in allergic inflammation. Strategies that interfere with these processes may prove to be useful for treatment of allergic diseases.

Key words: eosinophil - interleukin - β1 integrins - allergic diseases

Eosinophil accumulation is a distinctive feature of allergic airways inflammation (Bochner et al. 1994). Evidence for a role of eosinophils in the airway inflammation in asthma comes from a variety of studies. The presence of increased numbers of these cells has been demonstrated in bronchial biopsies, bronchoalveolar lavage (BAL) fluid and peripheral blood of patients with asthma. Furthermore, these cells appear to be in an activated state or in the process of degranulation and the levels of their granule proteins have been extensively correlated with clinical symptoms of asthma.

Recent studies on the role of eosinophils have focused on the mechanisms by which these cells infiltrate the airways (Resnick & Weller 1993, Bochner & Schleimer 1994). Although the exact mechanisms by which selective eosinophil recruitment occurs remain incompletely defined, leukocyte recruitment is known to result from the interaction of cell-surface adhesion molecules (e.g., selectins, integrins, and immunoglobulin superfamily members) with molecular counterligands on vascular endothelial cells, extracellular matrix (ECM) proteins, epithelial cells and other tissue structures (Carlos & Harlan 1994, Bochner & Schleimer 1997). While other factors determine the phenotype of infiltrating cells, such as cytokines and chemokines (Springer 1995), this chapter will focus on the role of β1 integrins in eosinophil trafficking and function.

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Exposure of Integrins on Eosinophils

Integrins are plasma membrane receptors composed of α and β heterodimeric transmembrane subunits generated from at least 16 α and 8 β subunits to produce over 20 different receptors (Bochner & Schleimer 1997). Both chains are required for normal receptor expression and for ligand binding. Members of the integrin family mediate cell-to-cell and cell-to-extracellular matrix interactions.

Table summarizes the expression of integrins on eosinophils. The predominant integrins on all leukocytes are in the β2 (CD18) subfamily (Bochner & Schleimer 1997). Granulocytes including eosinophils express the β2 integrins LFA-1, Mac-1, p150,95 and αdβ2 (Grayson et al. 1997). LFA-1 binds specifically to intercellular adhesion molecule-1 (ICAM-1), ICAM-2 and ICAM-3, Mac-1 binds to ICAM-1 and the iC3b product of activated complement, αdβ2 recognizes ICAM-3, while cellular ligands for p150,95 are as yet unknown. Because all granulocytes express β2 integrins, there appears to be no immediate explanation for how they might contribute to selective eosinophil recruitment. However, there are conditions under which eosinophil β2 integrins, especially Mac-1, may be selectively altered by stimuli such as cytokines [e.g., IL-5 (Walsh et al. 1990)] and chemokines (e.g., eotaxin) (Burke Gaffney & Hellewell 1996).

When other integrins are examined, more obvious differences in expression among granulocytes are observed (Georas et al. 1993, Ebisawa et al. 1995). Unlike neutrophils, eosinophils express the α4 integrins VLA-4 (α4β1) and α4β7 which mediate binding to VCAM-1, an immunoglobulin superfamily member induced by cytokines on endothelium and epithelial cell lines (Atsuta et al. 1997, Bochner & Schleimer 1997), and to an alternatively spliced domain in fibronectin, CS-1 (Anwar et al. 1994, Matsumoto et al. 1997). The α4β7 integrin binds to the mucosal addressin cell adhesion molecule-1 (MAdCAM-1) that has structural homology to ICAM-1 and VCAM-1 (Walsh et al. 1996, Briskin 1997). Eosinophils also express α6β1 (VLA-6), a ligand for the extracellular matrix protein laminin (Georas et al. 1993, Tourkin et al. 1993). Basophils resemble eosinophils in that they too express α4β1 and α4β7, but instead of α6β1, they express α5β1, another ligand for fibronectin (Saini et al. 1997).

### Eosinophil-Endothelial Interactions Through β1 Integrins

One mechanism of selective eosinophil recruitment involves the β1 integrin α4β1 (VLA-4), which is expressed on human eosinophils but not on neutrophils. This may be important for allergic inflammatory responses since it is a receptor for VCAM-1, and correlations have been found between infiltration of eosinophils and expression of VCAM-1 in the lungs of patients with asthma as well as in late phase reactions in the lungs, nose or skin (Kyan-Aung et al. 1991, Bentley et al. 1993, Lee et al. 1994, Gosset et al. 1994, Ohkawara et al. 1995, Fukuda et al. 1996). Expression of VCAM-1 also correlated with eosinophil numbers in nasal polyp tissues (Jahnsen et al. 1995, Beck et al. 1996).

Resting endothelial cells do not express VCAM-1. However, exposure of endothelial cells to IL-1, TNF, or bacterial endotoxin induces expression of endothelial adhesion molecules, including ICAM-1, E-selectin and VCAM-1. Specific antibodies to ICAM-1 and E-selectin have been shown to inhibit adherence of eosinophils to IL-1 stimulated endothelial monolayers by about 20-30% (Bochner et al. 1991). In contrast, VCAM-1 antibodies are extremely effective at inhibiting eosinophil but not neutrophil adherence (Bochner et al. 1991). Furthermore, anti-VLA-4 antibodies inhibit eosinophil, but not neutrophil, adherence to IL-1 stimulated endothelium (Dobrina et al. 1991, Walsh et al. 1991). These results indicate that specific induction of VCAM-1 on endothelial cells could selectively promote eosinophil adherence.

Eosinophils are predominant at inflammatory sites where Th2-type cytokines, such as IL-4 and IL-13, are prevalent (Hamilos et al. 1996, Rankin et al. 1996). In vitro, both of these cytokines selectively lead to the induction of VCAM-1 expression without any significant effect on the expression of E-selectin or ICAM-1 on endothelial cells (Schleimer et al. 1992, Kaiser et al. 1993, Bochner et al. 1995). Furthermore, incubation of endothelial cells with

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**Table**

<table>
<thead>
<tr>
<th>Integrin</th>
<th>CD designation</th>
<th>Expression</th>
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<tbody>
<tr>
<td>α1β1</td>
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</tr>
<tr>
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</tr>
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</tr>
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</tr>
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</tr>
<tr>
<td>β6</td>
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</tr>
<tr>
<td>α4β7</td>
<td>49d/103</td>
<td>Yes</td>
</tr>
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IL-4 or IL-13 has no effect on neutrophil adhesion but induces eosinophil adhesion in a dose-dependent manner (Schleimer et al. 1992, Bochner et al. 1995). These cytokines can be synergistic and selective in their ability to induce VCAM-1 on endothelial cells. The combination of IL-4 with either IL-1 or TNF results in at least 5-fold higher levels of VCAM-1 surface expression than either cytokine alone, with no induction of ICAM-1 or E-selectin (Iademarco et al. 1995, Ebisawa et al. 1997).

Further support for the potential importance of β1 integrins and their ligands is provided by in vivo studies where the function of these adhesion molecules, or the cytokines that induce their expression, has been blocked. Efforts to antagonize VLA-4, VCAM-1, and IL-4 have all been shown to reduce eosinophil recruitment and allergic airways.

Eosinophil-extracellular matrix (ECM) interactions through β1 integrins

After migration through the endothelium, eosinophils come into contact with the proteins of the basement membrane and ECM. The ECM is a complex web of large fibrillar proteins that underlies the endothelium and epithelium and surrounds connective tissue cells. Cellular interactions with ECM proteins can have profound consequences on leukocyte function (Hunt et al. 1997). Eosinophils interact with two ECM proteins, fibronectin and laminin, through two β1 integrins, namely VLA-4 and VLA-6 (α6β1), respectively.

Fibronectin - Fibronectin is encoded by a single gene, but alternative splicing of the primary RNA transcript gives rise to polypeptide diversity that appears to be regulated in a cell type-specific fashion (Walsh & Wardlaw 1997). The IIICS region of fibronectin contains a 25 amino acid site, named CS-1, that contains a sequence (LDV) recognized by VLA-4. Plasma fibronectin lacks the IIICS binding site in at least half of its subunits, whereas tissue fibronectin has it in both subunits. Despite expression of α4 integrins on eosinophils, whether they spontaneously attach to fibronectin remains controversial. Some studies have shown that resting eosinophils adhere to fibronectin in a VLA-4-dependent manner and exhibit prolonged survival via autocrine production of cytokines such as GM-CSF (Anwar et al. 1994, Neeley et al. 1994, Walsh et al. 1995). Other studies, however, have found little or no adhesion without prior activation with platelet-activating factor, Mn++ or a β1 integrin-activating antibody (Kuijpers et al. 1993, Kita et al. 1996, Matsumoto et al. 1997). A possible explanation for these discrepancies may arise from the fact that eosinophils express the β2 integrin Mac-1, and engagement through this receptor to a different site on fibronectin, or to the blocking protein (typically albumin), may be occurring.

Eosinophils also express α4β7, another ligand for fibronectin (Erle et al. 1994, Walsh et al. 1996). Levels of α4β7 on eosinophils are comparable to those for α4β1. In addition to functioning as a fibronectin ligand, it can also be a ligand for MAdCAM-1 and VCAM-1 (Erle et al. 1994, Walsh et al. 1996). However, α4β7 on eosinophils appears to be relatively inactive, because activation with Mn++ is required to demonstrate consistent adhesion (unpublished observations).

Laminin - Laminin consists of 3 distinct chains coded for by different but related genes. The mechanism by which laminin interacts with cells is complex (Walsh & Wardlaw 1997). It is recognised by different integrin receptors including α1β1, α2β1, α3β1, α6β1 and αvβ3, of which only α6β1 appears to be specific for laminin. Eosinophils can adhere to plate-bound laminin; this interaction requires divalent cations and is completely abolished by anti-α6 or anti-β1 antibodies. Indeed, eosinophils were shown by flow cytometry and immunoprecipitation to express α6β1 (Georas et al. 1993). As has been shown for fibronectin, eosinophils cultured on laminin exhibit prolonged survival (Tourkin et al. 1993).

Eosinophil–epithelial interactions through β1 integrins

Airway epithelium may also be an active participant in allergic inflammation. Epithelial cells are biologically active, express adhesion receptor proteins, and produce cytokines and chemokines (Polito & Proud 1997). Until recently, only ICAM-1, but not E-selectin or VCAM-1, had been identified in the respiratory epithelium in vitro and in vivo biopsies from patients with asthma (Bloemen et al. 1993, Fukuda et al. 1996, Stark et al. 1996). However, in the BEAS-2B bronchial epithelial cell line, culture with TNF or IL-1 was found to induce VCAM-1 mRNA and cell surface expression (as well as ICAM-1 expression), while culture with IL-4 induced VCAM-1 but not ICAM-1 expression (Atsuta et al. 1997). Maximal VCAM-1 expression resulted from the combination of TNF and IL-4. Furthermore, TNF treatment increased adhesion of eosinophils to BEAS-2B monolayers and this adhesion was blocked with VCAM-1 antibodies. These findings suggest that cytokine activation can induce expression of VCAM-1 on airway epithelium which can functionally interact with eosinophils through VLA-4.
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Levels of α4β1 and α6β1 on eosinophils are not altered after migration in vitro or in vivo or after cytokine activation, nor do levels differ among hypodense versus normodense eosinophils or cells from allergic versus nonallergic donors (Georas et al. 1992, 1993, Hansel & Walker 1992, Kroegel et al. 1994). However, in addition to the amount of expression of cell surface adhesion molecules, the functional state of integrins can be regulated, leading to changes in the affinity for counterligand binding without changing the level of cell surface expression (Diamond & Springer 1994). Recent studies have shown that the activation state of integrins can influence cell adhesion and function. The avidity of integrins, not just the total number of molecules expressed, influences cell adhesion and migration (Hunt et al. 1997). While a particular integrin may have more than one ligand, the avidity for each ligand may differ. It has recently been demonstrated that α4β1 integrins on eosinophils exist in a state of partial activation, and can be maximally activated for adhesion to ligands such as fibronectin and VCAM-1 after exposure to manganese or integrin-activating antibodies, conditions that do not affect the total cell surface expression of β1 integrins (Werfel et al. 1996, Matsumoto et al. 1997). Maintenance of basal levels of β1 integrin function on eosinophils appears to require tyrosine kinase activity, because reversible downregulation of VCAM-1 adhesion is seen in cells exposed to genistein or tyrphostins (Nagata et al. 1995, Matsumoto et al. 1997).

OUTSIDE-IN SIGNALING - Adhesion molecules are not only involved in adhesive interactions but also in transducing signals from the extracellular to the intracellular compartments and regulating effector functions of the cell (Ginsberg et al. 1992, Clark & Brugge 1995). For eosinophils, ligation of integrins has been shown to alter a variety of functional responses (Dri et al. 1991, Anwar et al. 1993, Tourkin et al. 1993, Neeley et al. 1994, Nagata et al. 1995, Kita et al. 1996). Signaling mechanisms via integrins are still poorly understood. The intracytoplasmic domains of integrins lack kinase or phosphatase activity of their own; they also lack sequence homology with known signaling proteins (Hemler et al. 1994). However, recent reports have shown that integrin engagement, either with ligand or with antibodies, is capable of transducing signals (Miyamoto et al. 1995) and induces the phosphorylation of the tyrosine kinase pp125FAK (FAK) (Schaller & Parsons 1994).

Outside-in signaling is initiated by the β subunit cytoplasmic-domain dependent rearrangement of cytoskeletal components and actin into focal adhesion complexes (FAC), found at areas of cell-ECM interaction (Clark & Brugge 1995). Formation of FAC’s in adherent cells is thought to be associated with cell spreading. A predominant FAC’s component, FAK, has been shown to physically interact with the cytoplasmic domain of β integrins, which in turn is thought to recruit several signaling molecules to FAC’s (Schaller & Parsons 1994). It is not clear whether the cytoskeletal and signaling components found in FAC’s associate with integrins in leukocytes. Besides FAK, β1 integrin interacts directly or indirectly with cytoskeletal proteins (McArthur Lewis & Schwartz 1995, Yamada & Miyamoto 1995, Wahl et al. 1996).

In addition to FAK, integrin receptor occupancy leads to the activation of the Src family of tyrosine kinases (Shattil et al. 1994) and the Ras/MAP kinase pathway (Schaller & Parsons 1994). Recently a novel serine/threonine kinase has been reported to associate with the β1 integrin cytoplasmic domain (Hannigan et al. 1996). The 59 kD protein, known as integrin-linked kinase (ILK), was found to phosphorylate a peptide representing the β1 integrin cytoplasmic domain and to co-localize with β1 in focal plaques.

Outside-in signaling is also regulated by the α subunit cytoplasmic tails. Those of α2 and α5 localize predominantly to FAC’s and show increased spreading on ECM. In contrast, the expression of the α4 cytoplasmic tail correlates with chemo and haptotactic migration, suggesting that α4 is responsible for weaker integrin-cytoskeletal interactions (Kassner et al. 1995). This is a potential mechanism by which α4β1, highly expressed in eosinophils, could increase cell motility.

Inside-out signaling - The rapidity of inside-out signaling insures that leukocytes can quickly modify their adhesiveness in response to stimuli. This is achieved by changes in integrin functional activity rather than integrin expression on the cell surface. The signaling pathways involved in inside-out signaling are still ill-defined (Hunt et al. 1997). Recent progress in this field has been mainly in T cells. Several activation stimuli have been shown to upregulate integrin function. Treatment of T cells with PMA or the Ca2+ ionophore A23187 has been shown to upregulate integrin-mediated T-cell adhesion, indicating that both protein kinase C and Ca2+ are involved in the intracellular signaling events (Shimizu et al. 1990), whereas treatment of Jurkat cells with the serine-threonine phosphatase inhibitor okadaic acid depresses fibronectin adhesion through β1 integrins (Seminario et al. 1997).


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