Study on the Elimination of *Angiostrongylus costaricensis* First Stage Larvae in the Experimental Infection of Swiss Mice

Cibele Canali, Aline H Goulart, Carlos Graeff-Teixeira+

Laboratório de Parasitologia, Curso de Pós-Graduação em Biociências, Instituto de Biociências, Faculdade de Medicina, PUCRS, Av. Ipiranga 6681, 90619-900 Porto Alegre, RS, Brasil

Abdominal angiostrongylosis is a nematode infection of wild rodents. Human infection may result in severe abdominal disease and has been reported from several countries in the Americas. The domestic mouse, *Mus musculus*, has not been found with natural infection and, like other urban rodents, should not be considered a natural host for *Angiostrongylus costaricensis*. Quantification of parasitic forms released for transmission may better express the coevolutionary status in parasite-host relationship. With this objective, five groups of experimentally infected Swiss mice were followed for up to 155 days post-infection (PI) days and the quantification of first stage larvae (L1) output revealed: an irregular elimination of L1 and a huge variation in the patency period (1 to 114 days) and in the number of L1 eliminated daily by individual animals (1 to 6340 L1/g). Overall mortality was 72% (range: 28% to 100%) at seven weeks PI. In conclusion, abdominal angiostrongylosis in *M. musculus* presents high mortality and a very variable and irregular elimination of L1 in feces.

Key words: *Angiostrongylus costaricensis* - abdominal angiostrongylosis - *Mus musculus* - coevolution

We describe some aspects of the kinetics of elimination of *A. costaricensis*' first stage larvae (L1) in feces of Swiss mice that further illustrate the lack of adaptation of this rodent to the etiological agent of abdominal angiostrongylosis.

**MATERIALS AND METHODS**

The *A. costaricensis* “Santa Rosa” strain has been maintained in the laboratory since 1992 with successive infections of Swiss *M. musculus* and the veronicelid slug *Phyllocaulis soleiformis* (D’Orbigny, 1835). Isolation of infective third stage larvae (L3) was performed as previously described (Graeff-Teixeira & Morera 1995). Five groups of Swiss mice were inoculated with eight L3 into the stomach with a metallic cannula. From the 21st day post-infection (PI), a sample of feces was individually collected every day up to 155 days post-infection (PI) and larvae were counted after their isolation with the Baermann method. Regularity of elimination was expressed by a ratio (PO, Patency/Observation) of the sum of days with larvae excretion (patency period) and total days of observation for individual animals. “A₀” is defined by the sum of daily larval output for individual animals, as proposed by Garnik (1992).

**RESULTS**

Mortality increased from the 4th week PI and varied from 28.5% to 100%, with a overall mor-
tality at seven weeks PI of 72% (Fig. 1). Twelve out of the 43 mice were alive after the 30th PI day: 3 died at 32, 25 and 27 PI day, while a number were sacrificed: 4 at 33rd PI and 5 at 155th PI day, without the identification of intra-arterial worms at necropsy. A large variability was seen with the patency period (1 to 114 days) and the PO ratio (0.03 to 1.00) (Fig. 2). Only 16 out of 43 animals (37.2%) presented with a PO > 0.5 and 11 (25.5%) had a PO > 0.8. Curves of frequencies for days of patency, PO ratio and survival days were deviated to the left, showing higher frequencies in categories of lower values (Fig. 2).

The daily elimination of L1 by individual mice varied from one L1/g feces to 6340 L1/g feces. Three patterns of curve were seen in graphics representing the daily individual larval output: early (5/29 - 17.2%), late (13/29 - 44.8%), continuous (11/29 - 37.9%), L1 elimination (Fig. 3). In six animals the larval output was very low and eight mice did not eliminate L1. The $A_o$ was individually calculated and varied from zero to 92982 (medium 3959.1; standard deviation 13727.12; variance: 188433714). The medium $A_o$ in each group varied from 17.2 to 13236.6. The huge variation in most of the data was similar among the five groups inoculated with different larvae isolates.

**DISCUSSION**

Both from epidemiological field work and from experimental studies there are indications that the main definitive hosts for *A. costaricensis* are wild rodents (Tesh et al. 1973, Morera 1985, Graeff-Teixeira et al. 1989, Santos et al. 1996). Sinantropic or urban rodents, like *M. musculus, Rattus norvergicus* (Linnaeus, 1758) and *Rattus rattus* (Linnaeus, 1758) probably do not play an

---

Fig. 1: mortality (%) along time post-infection in five groups of Swiss mice experimentally infected with *Angiostrongylus costaricensis* (Santa Rosa strain).

Fig. 2: distribution of frequencies for (a) PO ratio, (b) number of days with larvae elimination (patency) and (c) survival time in five groups of Swiss mice experimentally infected with *Angiostrongylus costaricensis*.
The classical wisdom about the characterization of adaptation in a host-parasite relationship put an emphasis in morbi-mortality of the host as descriptors for the coevolutionary status. An alternative and perhaps more universally applicable approach favors quantification of parasitic forms that are made available for transmission, like the L1 larvae eliminated in rodent feces in the *A. costaricensis* model. In graphics representing the daily larval output, like those of the Fig. 3, the area \( (Ao) \) beneath the curve would represent a quantitative assessment of coevolutionary status (Garnik 1992). This approach would have its best application in the comparative study of different host species and in the evaluation of their importance for the survival of a given parasite species. This report deals with a description of the kinetics of elimination of *A. costaricensis* larvae in a common strain of laboratory mice.

The high albeit variable mortality is a well-established feature of the experimental infection of mice strains, with the exception of C57BL/6 (Ishii & Sano 1989, Santos et al. 1996). Mice manifest disease (weight loss, diarrhea, hipoactivity) and most of the deaths occur from the 4th to the 6th week (Fig. 1) coincident with maturation of the worms. The incubation time (IT) in the human infection is unknown, but it may be similar to the observed IT in murine model.

Most animals survive less than 50 days, as can be seen in the distribution of frequencies of survival times (Fig. 2). Those with longer survival probably did not become infected or present an unisexual infection, as shown by Santos et al. (1996) and confirmed in the few animals that were

important role as hosts for this metastrongylid worm, as reviewed by Santos et al. (1996). Many confirmed human infections have been detected in small towns with a rural-like enviroment, where the sinanthropic rodents may have opportunities to contact the parasite in its wild cycle. Not a single case was been documented among residents at large urban areas, that are usually inadequate environments for the presence of molluscs and by consequence, to the transmission of the parasite (Rambo et al. 1997).

![Fig. 3: examples of graphics representing curves of first stage larvae fecal elimination (“y” axis) in individual Swiss mice with short (a, b, c) or long (d, e, f) survival time (days, “x” axis). Three main patterns can be depicted: early (a, d), late (b, e) or continuos and irregular (c, f) larval daily output.](image)
sacrificed and had a post-mortem examination performed, in the present experiments. These data on morbi-mortality indicate that *M. musculus* is not well-adapted to *A. costaricensis*.

Besides *Ao* quantification (Garnik 1992), other detailed arrangement of the data, focusing especially the regularity of patent days (PO), the length of patent period and the individual daily larval output may contribute to a better characterization of host-parasite coevolutionary status. In these experiments, patent period was frequently short, the elimination of larvae was not regular and individual daily larval elimination was very variable.

Without a comparative analysis of *Ao* among several host species this index may loose power as indicator of coevolutionary status. However, intra-species observation of the shape of the curves of daily larval elimination brings some information (Fig. 3). The irregularity of L1 elimination was expressed not only by low values of PO ratio, but also by the diversity of curve shapes. Animals presenting the “early” pattern of larval output (Fig. 3a, b) or the “late” pattern (Fig. 3 c, d), even if they survive for a long period, they offer a limited contribution for transmission of the parasite. The “continuous” pattern (quantitatively expressed by the PO ratio) associated with the amount of L1 (expressed by *Ao*) would describe animals presenting the most important contribution to the maintenance of the parasite. This kind of detailed analysis is required specially in non-comparative studies like the present one. The isolated *Ao* quantification probably will help to rank different species or strains, according to degree of host-parasite adaptation. Experiments are underway to compare *Ao* determinations of several rodent species.

These results give further support to the idea that *M. musculus* is not a well-adapted host for *A. costaricensis* and to the conception of abdominal angiostrongylosis as a zoonosis with a primordial wild cycle.

ACKNOWLEDGEMENTS
To Eva Medeiros, for technical help.

REFERENCES


