Lack of Correlation between Seminal and Plasma HIV-1 Viral Loads is Associated with CD4 T Cell Depletion in Therapy-naïve HIV-1\(^+\) Patients


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The present study was conducted to investigate a possible correlation between plasma (PVL) and seminal viral load (SVL) on treatment-naïve HIV-1-infected patients in Vitória, ES, Brazil. We also evaluated whether the progressive immunosuppression associated with HIV disease (as evidenced by declining CD4 T cell counts) has any impact on the correlation between PVL and SVL HIV-1. Viral load on paired blood and semen samples from 56 consecutive treatment-naïve patients were evaluated and compared to CD4 cell counts. Viral load and T cell counts (cells/µl) were determined by NASBA and by flow cytometry, respectively. Overall, a strong positive correlation between PVL and SVL was observed. However, when patients were grouped according to their CD4 counts, this correlation was only significant among patients with CD4 counts > 200 cells/µl. Results presented here demonstrate the existence of a strong correlation between PVL and SVL on patients with CD4 cell counts > 200 cells/µl, suggesting that this association may correlate with disease progression.

Key words: seminal viral load - plasmatic viral load - HIV - compartmentalization - CD4 T cell counts

Measures to prevent the continued propagation of the HIV pandemic not only require diagnosis of infected individuals, but also means for identifying subjects who are most likely to transmit HIV to others. Peripheral blood CD4 T-cell counts provide a good estimate of immune function (or disease stage), whereas HIV-1 viral load in plasma of infected individuals may be useful in assessing both the likelihood of disease progression and the likelihood of transmission of HIV-1 to previously uninfected persons (Quinn et al. 2000). Correlation between blood plasma viral load (PVL) and seminal viral load (SVL) has been a controversial issue (Hamed et al. 1993, Liu et al. 1996, Gupta et al. 1997, Vernazza et al. 1997). Ho et al. (1984) were the first authors to demonstrate the presence of HIV-1 in semen samples. Although, HIV pandemic is still driven by sexual transmission, little is known about the role of seminal viral load on HIV-1 infection. Data from other authors indicates that the efficiency of sexual transmission is directly associated with the inoculum’s size (Miller et al. 1992, Levy 1993, Dyer et al. 1998) and that SVL correlates strongly with HIV-1 shedding in seminal cell cultures (Vernazza et al. 1997). According to Dyer et al. (1998) high levels of HIV-1 replication probably contribute to both rapid disease progression and enhanced sexual transmission. Taken together these results suggest that SVL may correlate with HIV infectivity.

This cross-sectional study was designed to investigate the correlation between PVL and SVL in paired blood and seminal samples and their association with CD4 cell counts on anti-retroviral therapy naïve patients in Vitória, ES, Brazil.

MATERIALS AND METHODS

Patients - Fifty-six consecutive adult HIV-1\(^+\) men attending to the HIV clinic at Hospital da Santa Casa de Misericórdia, Vitória, from March 1998 to July 1999 were enrolled in the present study. All patients were treatment-naïve and did not display signs or symptoms of sexually transmitted diseases other than HIV and were asked to donate two blood and one semen sample at enrollment. Prior to its inception, this study was submitted and approved by the local ethical committee. A signed informed consent was obtained from all patients, preceding their inclusion in the study. Clinical staging was performed according to the revised Centers for Disease Control and Prevention classification system (CDC 1992).

Quantification of CD4\(^+\) lymphocytes in blood samples - Whole blood was collected in Vacutainer tubes (Becton Dickinson, Franklin Lakes, New Jersey, USA) using K\(_3\)-EDTA as anticoagulant. Blood CD4 T-cell counts (expressed as total numbers of lymphocytes per microliter of blood) were determined by flow cytometry (FACSCount, Becton & Dickinson, Mountain View, CA, USA) using the standard protocol recommended by the manufacturer. Blood samples were processed within 4 h of collection.

Quantification of HIV-1 RNA in blood and seminal plasma samples - Plasma was obtained from K\(_3\)-EDTA anti-coagulated blood (collected in a manner similar to
that described above) by centrifugation at 600 x g for 7 min. Cell free plasma samples were stored in aliquots at -70°C until assessment of viral load by NASBA. Prior to sample collection, patients were asked to adhere to a 72-hour sexual abstinence. Semen samples were processed within 4 h of ejaculation as described by Vernazza et al. (1996). HIV-1 RNA on both blood plasma and seminal plasma were quantified by the NASBA method (Nuclisens, Organon Teknika, Durham, North Carolina) (Dyer et al. 1996). This assay has a lower limit of detection of 400 HIV-1 RNA copies/ml.

Statistical analysis - The non-normal distribution of HIV-1 RNA titer in plasma and seminal samples was confirmed by Kolmogorov-Smirnov tests. Data were analyzed following log transformation and linear regression analysis by the Wilcoxon's test. Correlations between non-parametric variables (PVL and SVL, CD4 and CD8 counts) were evaluated using Spearman's rank correlation test. All statistical analyses were carried out using the SPSS 8.0 (SPSS Inc.) and SAS (SAS Institute Inc. SAS/STAT guide for personal computers, version 6, Cary, NC, USA) softwares.

RESULTS

Characteristics of the study population - Clinical staging, CD4 cell counts and age are summarized in the Table. The observed CD4 counts for the 56 patients ranged from 10 to 964 cells/µl (AVG ± SD = 359 ± 250 CD4+ cells/µl), and CD8 counts from 260 to 2292 CD8+ cells/µl (AVG ± SD = 1132 ± 500 CD8+ cells/µl). When patients were grouped according to their clinical stage, 15 patients (27%) were classified as Group 1 (CD4 > 500/µl), 24 (43%) as Group 2 (200/µl < CD4 < 500/µl) and 17 (30%) as Group 3 (CD4 < 200/µl). The observed CD4 counts for patients at Groups 1, 2, and 3 were 1308.3 ± 396, 1163.7 ± 676 and 930 ± 105 cells/µl, respectively. The average CD4/CD8 ratios were 0.53, 0.28, and 0.10 for Groups 1, 2, and 3, respectively.

The median PVL and SVL were 4.92 log_{10} copies/ml and 4.33 log_{10} copies/ml, respectively. When patients were grouped according to their CD4 cell counts/µl, the median PVL for Groups 1, 2, and 3 were 4.20, 5.04, and 5.11 log_{10} copies/ml, respectively, whereas the median SVL was 4.06 log_{10} copies/ml for Group 1, 4.15 log_{10} copies/ml for Group 2, and 4.51 log_{10} copies/ml for Group 3 (Table).

Correlation between viral load and T cell counts - When samples from all patients were considered, Spearman’s rank correlation test revealed a correlation (Spearman’s rank = 0.438, p < 0.001) between PVL and SVL, which was also confirmed by linear regression analysis (R^2 = 0.209, p < 0.001, Fig. 1A).

A negative correlation (Spearman’s rank = -0.502, p < 0.0001; R^2 = 0.239) was observed between PVL and CD4+ counts. Conversely, no correlation was observed between SVL and peripheral blood CD4+ counts (R^2 = 0.018, p = 0.126), and between CD8+ cell counts and HIV-1 viral load (either PVL or SVL, Figs 2A, 2B).

Relationship between viral load and immunological status - Correlation between PVL and SVL was only observed in patients with CD4 counts > 200 cells/µl, and most strikingly so (Spearman’s rank = 0.697, p < 0.004 and R^2 = 0.546) (Fig. 1B) on patients with the highest CD4 counts (Group 1). Although still significant, the correlation between PVL and SVL on patients in Group 2 was less pronounced (Spearman’s rank = 0.488, p < 0.016 and R^2 = 0.290) (Fig. 1C). No correlation (Spearman’s rank = 0.044, p > 0.866 and R^2 = 0.016) was observed on patients with CD4 counts < 200 cells/µl (Fig. 1D).

HIV-1 viral load on consecutive seminal or plasmatic samples - When paired samples were compared individually, 32 out of 56 patients (57%) had significantly discordant PVL and SVL (difference > 0.5 log_{10}). Among these 32 patients, 27 (84%) had PVL higher than SVL by more than > 0.5 log_{10}, whereas PVL was significantly lower than SVL only in 5 patients (16%) (Table). A decrease of the correlation between PVL and SVL in parallel to a decrease in the CD4 cell counts was also observed. Discordant results between PVL and SVL were most commonly observed among patients with more advanced HIV dis-

### TABLE

Characteristics of the study population

<table>
<thead>
<tr>
<th></th>
<th>Total (56)</th>
<th>Group 1 (15)</th>
<th>Group 2 (24)</th>
<th>Group 3 (17)</th>
</tr>
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<tbody>
<tr>
<td>N (%)</td>
<td>56 (100)</td>
<td>15 (27)</td>
<td>24 (43)</td>
<td>17 (30)</td>
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<tr>
<td>Age</td>
<td>34.4 ± 8.8</td>
<td>34.2 ± 7.0</td>
<td>35.1 ± 10.7</td>
<td>33.5 ± 11.1</td>
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<tr>
<td>CD4</td>
<td>359 ± 250</td>
<td>703 ± 114</td>
<td>328 ± 99</td>
<td>98 ± 46</td>
</tr>
<tr>
<td>CD8</td>
<td>1132 ± 500</td>
<td>1308 ± 396</td>
<td>1164 ± 676</td>
<td>930 ± 105</td>
</tr>
<tr>
<td>PVL</td>
<td>4.92</td>
<td>4.20</td>
<td>5.04</td>
<td>5.11</td>
</tr>
<tr>
<td>SVL</td>
<td>4.33</td>
<td>4.06</td>
<td>4.15</td>
<td>4.51</td>
</tr>
<tr>
<td>PVL = SVL (%)</td>
<td>24 (43)</td>
<td>8 (54)</td>
<td>10 (42)</td>
<td>6 (35)</td>
</tr>
<tr>
<td>PVL &gt; SVL (%)</td>
<td>27 (48)</td>
<td>5 (33)</td>
<td>13 (54)</td>
<td>9 (53)</td>
</tr>
<tr>
<td>PVL &lt; SVL (%)</td>
<td>5 (9)</td>
<td>2 (13)</td>
<td>1 (4)</td>
<td>2 (12)</td>
</tr>
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</table>

a: Groups 1, 2, 3: according to the CDC 1993 classification: (CDC 1992). Group 1: CD4 counts ≥ 500 cells/µl; Group 2: 200 ≥ CD4 counts > 500 cells/µl; Group 3: CD4 < 200 cells/µl; age: CD4 and CD8 T cell counts are expressed as mean average ± standard deviation. HIV-1 viral load, both plasmatic viral load (PVL) and seminal viral load (SVL) are expressed as medians; PVL = SVL: difference between PVL and SVL smaller than 0.5 log_{10}; PVL > SVL or PVL < SVL: differences between PVL and SVL greater than 0.5 log_{10}.
One may question if the lack of correlation between PVL and SVL was due to inter-assay variability or due to an intermittent HIV-1 shedding pattern (Krieger et al. 1995). We decided to investigate if a technical issue, as evidenced by lower CD4 T cell counts. PVL and SVL were discordant in 65%, 38%, and 46% of study subjects in Groups 3, 2, and 1, respectively (Table). To exclude the possibility of inter-assay variability, we collected two consecutive (collection time varying from 3-26 weeks) paired blood and semen samples from a subset of 17 therapy-naïve patients and compared their PVL and SVL. SVL on collections 1 and 2 were discordant in 35.3% of patients; whereas, 17.6% of these 17 patients had discordant PVL (Fig. 3).

**DISCUSSION**

A strong correlation between PVL and SVL was observed for all 56 HIV-1 patients, regardless of their clinical stage, confirming previous reports (Gupta et al. 1997, Vernazza et al. 1997, Dyer et al. 1998, Coombs et al. 1998, Fiscus et al. 1998). However, once patients were grouped according to their CD4 cell counts, correlation between PVL and SVL decreased in parallel to the decline of CD4+ T cell counts. Patients with CD4 cell counts > 500 cells/µl presented the strongest correlation between PVL and SVL; a significant but less strong correlation was also observed on patients with 500 < CD4 < 200 cells/µl; and no correlation among patients with CD4 cell counts < 200 cells/µl. To our knowledge this is the first report to show that the correlation between PVL and SVL is dependent on the peripheral blood CD4+ T cell concentration.

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Figure 3: variability of both plasmatic (PVL) and seminal viral loads (SVL) on two consecutive samples from 17 treatment-naive patients. Open circles: samples which varied less than 0.5 log_{10} copies/ml; closed circles: samples which varied more than 0.5 log_{10} copies/ml. Differences greater than 0.5 log_{10} copies/ml were considered significant.

Data presented here demonstrates for the first time the loss of correlation between PVL and SVL; by evaluating PVL and SVL on two consecutive samples from 17 drug naïve patients. In three patients (17.6%) PVL of two consecutive samples differed significantly, whereas SVL was discordant on six patients (35.3%). Although SVL varied more than PVL, HIV-1 RNA was detected in all semen samples on both collection points; suggesting, therefore, that these patients shed HIV-1 continuously but at different viral load levels.

When paired PVL with SVL from each one of the 56 patients were compared, PVL and SVL were significantly different in 32 patients. Among these discordant patients, a higher PVL was more frequent (84.4%) than a higher SVL (15.6%). Since leukocytes were not quantified in semen samples, we were not able to exclude the possibility of enrolled patients having asymptomatic urogenital infections, which could increase SVL. However, PVL was frequently higher than SVL in patients with discordant viral loads, suggesting that these patients were not harboring asymptomatic urogenital infections. Albeit, Coombs et al. (1998) reported a weak inverse correlation between blood CD4 cell counts and SVL, these findings could not be confirmed in our experiments. Recently, Ball et al. (1999) demonstrated that seminal HIV-1 proviral DNA, but not seminal HIV RNA was associated with elevated semen leukocyte counts.

Data presented here demonstrates for the first time that correlation between PVL and SVL decreases with the depletion of CD4+ T cells, suggesting an important role for CD4 cell counts in the maintenance of this correlation and support the hypothesis of HIV-1 compartmentalization (Gupta et al. 1997, Coombs et al. 1998, Ball et al. 1999).

A larger study aimed at the investigation of a possible correlation between seminal immune components and either PVL or SVL on paired semen and blood samples is currently been conducted at our laboratory, and may help to better understand the association between immune status and seminal viral load.

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