Systematics and biogeography of the genus *Phalloptychus* Eigenmann, 1907
(Cyprinodontiformes: Poeciliidae: Poeciliinae)

Paulo H. F. Lucinda

The genus *Phalloptychus* is revised. *Phalloptychus iheringii* is removed from the synonym of *P. januarius*. Three species are recognized: *P. eigenmanni*, *P. januarius*, and *P. iheringii*. *Phalloptychus eigenmanni* can be distinguished from its congeners by the number of pelvic-fin rays in females, the number of predorsal scales, and the predorsal distance in females. *Phalloptychus iheringii* and *P. januarius* can be distinguished by the number of epipleural ribs, number of gonopodial rays, and a significantly different number of vertical bars along body side in females. Lectotypes are designated for *Girardinus iheringii* and *G. januarius*. Redescriptions and known distribution ranges are provided for each species as well as an identification key. Derived features supporting the monophyly of the genus and infrageneric clades are presented and discussed. The biogeographic implications of the geographic distribution of *Phalloptychus* species are discussed.


**Key words:** Neotropical, ichthyofauna, taxonomy, phylogeny, live-bearing, killifishes.

**Introduction**

The genus *Phalloptychus* Eigenmann, 1907 contains poeciliids endemic to South America occurring along the coastal drainages from Bahia to Rio Grande do Sul states of Brazil (Fig. 1). Males are typically smaller than females; the gonopodium is long and asymmetrical and bears spines along its length. Papers concerning *Phalloptychus* are extremely rare in systematic literature. These are mostly confined to original descriptions. Even from a biological standpoint *Phalloptychus* species are poorly studied.

The taxonomic history of *Phalloptychus* began with the first described species currently in the genus: *Girardinus januarius* Hensel, 1868, based on specimens from Rio de Janeiro. A second species, *G. iheringii* Boulenger, 1889, was described from Rio Grande do Sul on the basis of specimens collected by Hermann von Ihering. Garman (1895) synonymized *G. iheringii* and *G. caudimaculatus* Hensel, 1868 [= *Phalloceros caudimaculatus*] with *G. januarius*, since the later had page priority over the second. Garman (1895) erected the genus *Glaridodon* on the basis of tooth and jaw morphology, with *G. uninotatus* Poey, 1854 as type species, and placed *G. januarius* in his new genus *Glaridodon*. The name *Glaridodon* is preoccupied in Therapsida, and Garman (1896) replaced it with *Glaridichthys* in the subsequent year. It is quite evident from Garman’s (1895) figure of a *Gl. januarius* male (Plate VIII, fig. 15), showing a distal appendix on gonopodium, that he had only *Phalloceros* Eigenmann, 1907 specimens at hand. That is the reason why he considered both genera to be synonyms. Schneider & Ribeiro’s (1903) examination of the fish collection in the Museu Nacional do Rio de Janeiro, identified and recognized *Gl. januarius* and described *G. zonatus* based on specimens with uncertain locality.

Steindachner (1907) did not treat *Glaridodon* as a preoccupied name and recognized *Glaridodon* as a subgenus of *Girardinus* Poey, 1854 based on his examination of material
Systematics and biogeography of the genus Phalloptychus

Lucinda & Reis (2005: fig. 2a) presented (but not discussed) a phylogenetic hypothesis of relationships among Phalloptychus species. This hypothesis (Fig. 2) is part of a more inclusive phylogenetic study on the relationships among poeciliine genera. Thus, the phylogenetic position of the genus Phalloptychus in the subfamily Poeciliinae was discussed by Lucinda & Reis (2005: fig. 1). Although the transformation series analysis was provided and intrageneric relationships of Phalloptychus were depicted by Lucinda & Reis (2005), these authors did not present the diagnoses of intrageneric clades. These clade diagnoses are provided herein.

Thus, this paper has the following aims: (1) to diagnose the species of Phallotorynus and their geographic distribution; (2) present the diagnoses of the genus and its intrageneric clades and (3) to propose a hypothesis of biogeographic relationships among those species.

Material and Methods

The entries under examined material for each species follow the sequence: country, state, museum acronym, catalogue number, total number of examined specimens in the lot (in parentheses; number of cleared and stained specimens is indicated by an asterisk and separated from total number by a bar), type status, collection locality, date, collector. Type localities are listed as in original descriptions. Museum acronyms are: BMNH - Natural History Museum, London; CM - Carnegie Museum, Pittsburgh; FMNH - Field Museum of Natural History, Chicago; FURG - Fundação Universidade de Rio Grande, Porto Alegre; MCP - Museu de Ciências e Tecnologia da Pontifícia Universidade Católica do Rio Grande do Sul, Porto Alegre; MHNCI - Museu de História Natural do Capão da Imbúia, Curitiba; MZUSP - Museu de Zoologia da Universidade de São Paulo, São Paulo; CAS-SU - Stanford University Collection at the California Academy of Sciences, San Francisco; UFPA - Universidade Federal da Paraíba, João Pessoa; UMMZ - University of Michigan Museum of Zoology, Ann Arbor; USNM - National Museum of Natural History, Smithsonian Institution, Washington, DC; and ZMB - Museum für Naturkunde, Humboldt-Universität, Berlin.

Counts of scales, fin rays and vertebrae were taken according to Lucinda (2005). The three numbers in parentheses separated by bars following the vertebrae counts indicate respectively: number of vertebrae anterior to first dorsal pterygiophore, number of vertebrae between first and last dorsal pterygiophore, and number of vertebrae posterior to last dorsal pterygiophore. Other counts include (a) number of teeth on outer premaxillary row; (b) number of teeth on outer dentary row; (c) number of branchiostegal rays; (l) number of caudal-fin rays attached to hyural plate; (d) number of upper accessory cartilages; and (e) number of lower accessory cartilages. All counts, except scales, were performed in cleared and stained specimens. The minute and incompletely ossified ray attached to the last normal gonopodial ray has been considered in gonopodial ray-counts. Rudimentary and procurent rays were included in pectoral and caudal fin.
counts, respectively. All counts were made on the left side of adult specimens whenever possible, except the number of teeth, which have been counted on both sides. In species descriptions, numbers in square brackets following the counts indicate number of specimens for each count.

Fourteen measurements were obtained as distances between 13 homologous landmarks on the lateral left surface of head and body, as described in Lucinda (2005). Tables of descriptive morphometrics were elaborated with “Datax, version 4.2” by Roberto Reis and Nelson Fontoura. Measurements, other than SL, are expressed as percents of SL, except those that are subunits of the head, which are expressed as percents of HL.

Nonparametric statistical tests of null hypotheses of character similarity were performed using the software “Sigma Stat for Windows (Jandel Scientific), in order to evaluate metric data that failed to pass tests for normality and equal variance. This methodology has been extensively described and commented by Weitzman & Malabarba (1999).

Number and disposition of cephalic pores followed the nomenclature of Rosen & Mendelson (1960), Gosline (1949), and Parenti (1981). Only adult individuals have been examined to avoid undesirable ontogenetic variation. Nomenclature of the gonopodium follows Rosen & Gordon (1953) and Lucinda & Reis (2005). Descriptions of gonopodium morphology are based on fully developed gonopodia of large adult males. Osteological nomenclature adopted followed Rosen & Bailey (1963) and Parenti (1981). Clearing and staining followed the method of Taylor & Van Dyke (1985). Cladistic procedures, character state assignments, transformation series and clade numbers follow the phylogenetic analysis performed by Lucinda & Reis (2005). Character states illustrations are provided or referred to by Lucinda & Reis (2005).

Results

Three *Phalloptychus* species are herein recognized: *P. eigenmanni*, *P. januarius*, and *P. iheringii*, whose descriptions are provided below. Autapomorphies and synapomorphies supporting the monophyly of the genus and the infrageneric clade are presented below. Two asterisks indicate uniquely derived and unreversed features.

**Phalloptychus Eigenmann**

Clade 100, Lucinda & Reis, 2005

*Phalloptychus* Eigenmann, 1907: 426 [diagnosis in key], 430 [formal diagnosis], fig. 6. Gender masculine. Type species: *Girardinus januarius* Hensel, 1868. Type by original designation and monotypy.

**Diagnosis.** *Phalloptychus* species share the following uniquely derived and unreversed features: (1) extremely large dorsolateral process of basipterygium in adult males [36-2**] (Lucinda & Reis, 2005: fig. 8c; 15); (2) first ray of left and right pelvic fins different from each other in adult males [40-1**] (Lucinda & Reis, 2005: fig. 10); (3) first pelvic-fin ray of adult males very wide, mainly right one [41-3**] (Lucinda & Reis, 2005: fig. 10); (4) lateral projection near the bifurcation of second right pelvic-fin ray of adult males [43-1**] (Lucinda & Reis, 2005: fig. 10a); (5) callosity at the distal portion of right pelvic fin of adult males [45-1**] (Lucinda & Reis, 2005: fig. 12); (6) third gonapophysis angled zero-10 degrees relative to vertebral column [55-3**]; (7) pleural rib 9 of adult males well-developed, longer than remaining pleural ribs, curved forward and expanded at distal tip [58-1**]; (8) gonactinost complex axis little inclined backwards, forming an angle between 45° and 90° relative to body longitudinal axis [68-3**]; (9) short and dorsal protuberance close to base of R4p [117-1**] (Lucinda & Reis, 2005: fig. 27); and (10) lower branch of R6 as long as upper branch [129-1**]. Additionally, *Phalloptychus* species can be diagnosed by the following not uniquely derived and/or reversed features: (1) parietals absent [2-2] (Ghedotti, 2000: fig. 3); (2) epistatic process absent [3-3] (Ghedotti, 2000: fig. 3); (3) posterior section of posterior remnant of infraorbital system open forming a groove [7-1] (Rosen & Mendelson, 1960: fig. 2A, B); (4) preorbital canal absent or open, forming a very shallow groove [9-2] (Rosen & Mendelson, 1960: fig. 2C, D); (5) four pelvic-fin rays in males [33-2]; (6) ligastyle tripartite (three axis) [46-3] (Lucinda & Reis, 2005: fig. 13c); (7) first gonapophysis approximately parallel to vertebral column [53-3]; (8) second gonapophysis approximately parallel to vertebral column [54-2]; (9) pleural ribs associated with haemal arches in males [60-1]; (10) pleural ribs associated with haemal arches in females [61-1]; (11) absence of a convex expansion on anterior border of second gonactinost [70-0]; (12) flanks with elongate vertical bars reaching dorsal and ventral profiles plus short bars on caudal peduncle [133-1] (Fig. 5; 7); and (13) orbital bones absent [143-0].

**Phalloptychus eigenmanni**

**Phalloptychus iheringii**
**Description.** Morphometric data in Tables 1-2. Range of SL: 21.8 to 22.8 mm (females), 15.7 mm (immature male). Body compressed; width in predorsal region uniform, and about half body depth. Postdorsal region compressed towards caudal peduncle. Dorsal profile of snout slightly concave. Posterior portion of supraorbital ramus represented by two neuromasts. Preorbital canal absent. Anterior portion of infra-orbital ramus represented by two neuromasts (pores 4b, 5, 6a) and by one canal open at both ends (pores 6b and 7). Ventral infra-orbital line neuromasts not visible due to poor condition of material. Preopercular canal along preopercular border and opened by two pores, posterior remnants of infra-orbital ramus represented by two neuromasts (pores 2b, 3b, 4a, 6b) and by one canal open at both ends (pores 6b and 7). Ventral infra-orbital line neuromasts not visible due to poor condition of material. Preopercular canal along preopercular border and opened by two pores, followed by two grooved neuromasts. Neuromasts on mandibular ramus not visible due to poor condition of material.

**Color in alcohol.** Description based on three female paratypes faintly colored. Eye light metallic-grey with white pupil. Ground color cream. Anus margin and genital papilla whitish yellow. Light yellow line along predorsal surface. Fins pale yellow. Belly and preanal area silvery yellow. Five to seven narrow brown bars along flanks.

**Common names.** Barrigudinho, guaru.

**Ecology.** *Phalloptychus eigenmanni* is included in the list of endangered fish species in Brazil (Rosa & Menezes, 1996; MMA, 2004). Various collecting efforts have revealed unfruitful, even in the type locality. *Phalloptychus eigenmanni* is seriously endangered, probably extinct (Rosa & Menezes, 1996).

**Distribution.** *Phalloptychus eigenmanni* is solely known from the type locality (Fig. 1).
Catarina, Brazil. (a) male, 18.3 mm SL; (b) female, 25.7 mm SL.

Tubarão and lateral channels near Campo Verde, Santa Catarina, Brazil. Furthermore, diagnosed by the possession of eight anal-fin rays in males and five pelvic-fin rays in females.

Diagnosis: Phalloptychus iheringii + Phalloptychus januarius

Phalloptychus iheringii + Phalloptychus januarius

Phalloptychus iheringii (Boulenger)

Fig. 5; Tables 1-2

Girardinus iheringii Boulenger, 1889: 266. Type locality: Rio Grande do Sul. Restricted by Ihering (1893: 29) to “an der Mündung des Rio Camaquam” [= in the mouth of rio Camaquã].

Type material. Lectotype [present designation]: BMNH 1886.1.21.73. Paralectotypes: BMNH 1886.1.21.74-82 (9), CAS 62348 (33).

Diagnosis. Phalloptychus iheringii can be autapomorphically diagnosed by the possession of eight anal-fin rays in males (8 vs. 6, respectively), the number of predorsal scales (10-12 vs. 13, respectively), and the shorter predorsal distance in females (55.6-61.3 vs. 64.0-68.6 mm SL, respectively). P. iheringii is distinguished from P. januarius by the number of epipleural ribs (10-11 vs. 13-12 in the other, respectively), by the number of gonopodial rays (8 vs. 9, respectively), and by a significantly greater number of vertical bars along body sides of females (range = 8-18, median = 12 vs. range = 5-12, median = 7, respectively). Although some overlap occurs, Mann-Whitney rank sum tests (Fig. 6) indicate significant differences (P < 0.001).


Preorbital ramus of cephalic sensory system represented by two to five grooved neuromasts. Preorbital canal absent. Anterior portion of supraorbital ramus (pores 1 and 2a) parallel to upper lip with three inconspicuous neuromasts on each side. Posterior portion of supraorbital ramus (pores 2b, 3, 4a) composed of two grooved neuromasts. Posterior remnants of infra-orbital ramus represented by three neuromasts (pores 4b, 5, 6a) and by one short canal (pores 6b and 7). Preopercular ramus represented by large canal (sometimes completely open, forming groove) along preopercular posterolateral border and by prolonged canal along preopercular ventral border opened by four pores. Opercular canal absent. Mandibular ramus composed of two or three superficial neuromasts (pores Z, Ya, and Yb) on anterior border of ventral surface of mandible and by one superficial neuromast near maxillary distal end (pore W).

Gonopodial complex composed of three functional gonopophyses and nine gonactinosts. Gonactinosts 2, 3, 4 fused. Gonactinost 4 with wing-like expansions. Ligastyle present. Gonopodium sinistrally asymmetrical. Eight gonopodial rays. R1 and R2 unbranched and short, with 8 segments. R3 with dorsal convexity located near base ranging from second to tenth or eleventh segments 2 to 10 or 11. Tip of R3, R4a, and R4p ventrally bent and joint. Dorsal con-
vexity located between segments 12 to 20 of R4p. Fourteen to fifteen spines on distal segments of R4p. Spines retrace except two or three last spines directed forwards or upwards. R6, R7, and R8 branched. Five or six segments before bifurcation of R6. Anterior and posterior branches of R6 almost or fully ankylosed. Distal end of R6 modified in arrow-shaped expansion. R7 with six to eight segments anterior to bifurcation. Anterior and posterior branches of R7 moderately expanded and partially to completely ankylosed. Anterior branch of R7 greater than posterior. R8 with six or eight segments anterior to bifurcation. Anterior and posterior branches of R8 normal. Anterior branch of R8 greater than posterior. Six to eight segments on anterior branch of R8. R9 very minute not attached to pterygiophore.


Common names. Barrigudinho.

Distribution. Phalloptychus iheringii occurs in coastal drainages in Santa Catarina and Rio Grande do Sul states of Brazil (Fig. 1).

Remarks. Boulenger (1889) described Girardinus iheringii in honor of Hermann von Ihering who collected and sent him the specimens on which the description has been based (Ihering, 1893). This name has subsequently been used by Eigenmann & Eigenmann (1891) referring to Rio Grande do Sul specimens. Boulenger originally cited the type locality as “Rio Grande do Sul”, probably because it was the only collection information he had. Ihering (1893) states that type specimens have been collected “an der Mündung des Rio Camaquã” [= in the mouth of rio Camaqua], thus restricting the type locality. In the same paper, however, Ihering claimed that G. iheringii should be regarded as a junior synonym of G. januarius, based on information provided in letter by Hingeldorf. Presumably, for this reason, subsequent authors have considered both names synonyms. However, the study of color pattern has revealed that the number of vertical bars along body sides of females is significantly greater in southern populations of Phalloptychus from Santa Catarina and Rio Grande do Sul (range = 8-18, median = 12) than for northern populations from Rio de Janeiro, São Paulo and Paraná (range = 5-12, median = 7). See Fig. 6 for a nonparametric statistical expression of differences and similarities among Phalloptychus populations. These differences along with differences in number of epipleural ribs and gonopodial rays give support to the resurrection of P. iheringii as a valid species.


Phalloptychus januarius (Hensel)
Fig. 7; Tables 1-2

Girardinus januarius Hensel, 1868: 360. Type locality: “aus den Pfützen und Gräben um Rio de Janeiro” [= from the puddles and ditches around Rio de Janeiro].

Type material. Lectotype [present designation]: ZMB 7422. Paratelectyopes: ZMB 33219 [originally ZMB 7422], ZMB 7424, and ZMB 31497.

Diagnosis. Phalloptychus januarius can be autapomorphically diagnosed by the possession of nine anal-fin rays in males [86-4]. Furthermore, Phalloptychus januarius can be distinguished from P. eigenmanni by the number of pelvic-fin rays in females (5 vs. 6, respectively), by the number of predorsal scales (10-12 vs. 13, respectively), and the shorter predorsal distance in females (55.4-62.2 vs. 64.0-68.6 % SL, respectively). Phalloptychus januarius is distinguished from P. iheringii by the number of epipleural ribs (12-13 [one specimen had 11 in one side and 12 in the other] vs. 10-11, respectively), by the number of gonopodial rays (9 vs. 8, respectively), and by a significantly lower number of vertical bars along body sides of females (range = 5-12, median = 7, vs. range = 8-18, median = 12, respectively). Although some overlap occurs, Mann-Whitney rank sum tests (Fig. 6) indicate significant differences (P < 0.001).
Significant differences were found between the distributions of Phalloptychus januarius and P. iheringii (Rio de Janeiro, São Paulo and Paraná) and P. iheringii populations (Santa Catarina and Rio Grande do Sul). * Type locality of P. januarius, ** type locality of P. iheringii.


**Fig. 6.** Tukey box plot of number of bars along flanks in female specimens of *Phalloptychus januarius* and *P. iheringii*. Significant differences were found between *P. januarius* (Rio de Janeiro, São Paulo and Paraná) and *P. iheringii* populations (Santa Catarina and Rio Grande do Sul). * Type locality of *P. januarius*, ** type locality of *P. iheringii*.

**Fig. 7.** *Phalloptychus januarius*. UMMZ 231550, Farol de São Tomé, Rio de Janeiro, Brazil. (a) male, 16.1 mm SL; (b) female, 21.9 mm SL.

Preorbital ramus of cephalic sensory system represented by one to three grooved neuromasts. Preorbital canal absent. Anterior portion of supraorbital ramus (pores 1 and 2a) parallel to upper lip with three inconspicuous neuromasts on each side. Posterior portion of supraorbital ramus (pores 2b, 3, 4a) composed of two or three grooved neuromasts. Posterior remnants of infra-orbital ramus represented by three neuromasts (pores 4b, 5, 6a) and by one canal opened in both ends (pores 6b and 7). About 15 inconspicuous neuromasts on ventral infra-orbital line. Preopercular ramus represented by large canal (sometimes completely open, forming groove) along preopercular posterolateral border and by prolonged canal along preopercle ventral border opened by four pores. Opercular canal absent. Mandibular ramus composed of two or three superficial neuromasts (pores Z, Ya, and Yb) on anterior border of ventral surface of mandible and by one superficial neuromast near maxillary distal end (pore W).

Gonopodal complex composed of three functional gonopophyses and nine gonactinosts. Gonactinosts 2, 3, 4 fused. Gonactinost 4 with wing-like expansions. Ligastyle present. Gonopodium sinistrally asymmetrical. Eight gonopodial rays. R1 and R2 unbranched and short, with 8 segments. R3 with dorsal convexity located near base ranging from second to tenth or eleventh segments. Tip of R3, R4a, and R4p ventrally bent and joint. Dorsal convexity located between segments 14 to 21 (rarely 12 to 18) of R4p. Twelve to 17 spines on distal segments of R4p. Spines retrorse except two or three last spines, directed forwards or upwards. R6, R7, R8 branched. Five to 8 segments before bifurcation of ray 6. Anterior and posterior branches of R6 almost or fully ankylosed. Distal end of R6 modified in arrow-shaped expansion. R7 with six or seven segments anterior to bifurcation. Anterior and posterior branches of R7 moderately expanded and partially to completely ankylosed. Anterior branch of R7 greater than posterior. R8 with seven or eight segments anterior to bifurcation. Anterior and posterior branches of R8 normal. Anterior branch of R8 greater than posterior. Four to six segments on anterior branch of R8. Four segments on posterior branch of R8. R9 minute not attached to pterygiophore.
Distribution. *Phalloptychus januarius* is known from coastal drainages in Rio de Janeiro, São Paulo and Paraná States of Brazil (Fig. 1).

**Remarks.** Besides the types examined, the type series includes the lots ZMB 7423 (4) and CAS-SU 1132 (1). It is not absolutely sure, whether the lot ZMB 31497 belongs to the type series, since this number is based secondarily on an old label from the anatomical collection: “No. 25204, R. Janeir [sic]’” - the remainder of the label is illegible. This is the collection of the former “Institute of Anatomy of the Humboldt-University” in the 19th century. And this belonged to the Medical Faculty. This is due to the fact that human-anatomists were traditionally doing much research work on comparative anatomy of vertebrates. Thus, collected material was often divided between Zoological (Philosophical) Faculty with the Zoological Collection and the Medical Faculty. Only from approximately 1890 onwards the collections were reunited. But, apparently also R. Virchow, the famous pathologist working at the Charity Hospital at Humboldt-University took over some of the original anatomical collection for study (Paepeke & Seegers 1986; P. Bartsch *in litt.*, 2000). As there is no apparent evidence that Hensel has not examined specimens from lot ZMB 31497, it is advisable to label the lot paralectotypes. If future evidence demonstrates that these specimens are not syntypes, they will lose their paralectotype status.


**Key to the species of Phalloptychus.**

1. Females with six pelvic-fin rays; 13 predorsal scales; predorsal distance 64.0-68.6 % SL (rio Catu, Bahia) ……………………………………………………………………………………………………………… **Phalloptychus eigenmanni**

1’. Females with five pelvic-fin rays; 10-12 predorsal scales; predorsal distance 54.4-62.2 % SL ………………………………………………………………………………………………………………… 2

2. Five to 12 (median = 7) vertical bars along body side in females; 12 or 13 epipleural ribs in adult specimens; 9 gonopodial rays; ninth minute, incompletely ossified gonopodial ray present (Coastal drainages from Rio de Janeiro to Paraná) ………………………………………………………………………………………………………………………………………………………………………………………………………… **Phalloptychus januarius**

2’. Eight to 18 (median = 12) vertical bars along body sides in females; 9 to 11 epipleural ribs on adult specimens; 8 gonopodial rays (Coastal drainages of Santa Catarina and Rio Grande do Sul) ……………………………………………………………………………………………………………………………………………………………………………………………………………………………… **Phalloptychus iheringii**

**Discussion**

The genus *Phalloptychus* is a well-diagnosed, monophyletic group (Lucinda & Reis, 2005). The genus is traditionally allocated in the tribe Cenesterodontini Hubbs. This tribe, as

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Table 1. Descriptive morphometrics of males of *Phalloptychus* species. Measurements 1-10 are percents of standard length and measurements 11-13 are percents of head length.

<table>
<thead>
<tr>
<th>Character</th>
<th><em>P. eigenmanni</em> n = 1</th>
<th><em>P. iheringii</em> n = 19</th>
<th><em>P. januarius</em> n = 25</th>
</tr>
</thead>
<tbody>
<tr>
<td>Standard length (mm)</td>
<td>15.7</td>
<td>14.4 - 20.0</td>
<td>13.6 - 19.8</td>
</tr>
<tr>
<td>1. Head length</td>
<td>22.0</td>
<td>21.0 - 23.9</td>
<td>20.7 - 26.2</td>
</tr>
<tr>
<td>2. Snout-occipital distance</td>
<td>16.5</td>
<td>17.1 - 20.4</td>
<td>16.8 - 21.9</td>
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<tr>
<td>3. Predorsal distance</td>
<td>9.0</td>
<td>49.8 - 55.0</td>
<td>51.6 - 56.0</td>
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<tr>
<td>4. Dorsal-fin base length</td>
<td>7.0</td>
<td>9.7 - 14.1</td>
<td>8.4 - 14.2</td>
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<tr>
<td>5. Anal-fin base length</td>
<td>11.1</td>
<td>8.5 - 12.2</td>
<td>6.6 - 13.4</td>
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<tr>
<td>6. Body depth</td>
<td>32.9</td>
<td>31.1 - 34.6</td>
<td>30.8 - 36.3</td>
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<td>7. Pre-pelvic length</td>
<td>30.2</td>
<td>27.6 - 30.7</td>
<td>28.7 - 33.5</td>
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<td>8. Pre-anal length</td>
<td>35.6</td>
<td>31.2 - 37.1</td>
<td>31.7 - 37.9</td>
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<td>9. Post-anal length</td>
<td>56.5</td>
<td>58.0 - 63.6</td>
<td>57.5 - 62.6</td>
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<td>10. Caudal peduncle depth</td>
<td>16.5</td>
<td>14.9 - 16.6</td>
<td>13.9 - 18.0</td>
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<tr>
<td>11. Snout length</td>
<td>19.6</td>
<td>16.6 - 25.1</td>
<td>20.2 - 25.0</td>
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<tr>
<td>12. Orbital diameter</td>
<td>43.5</td>
<td>35.4 - 43.1</td>
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<td>13. Postorbital length</td>
<td>42.4</td>
<td>40.6 - 47.6</td>
<td>37.7 - 50.1</td>
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Table 2. Descriptive morphometrics of females of *Phalloptychus* species. Measurements 1-10 are percents of standard length and measurements 11-13 are percents of head length.

<table>
<thead>
<tr>
<th>Character</th>
<th>P. eigenmanni n = 2</th>
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<th>P. iheringii n = 48</th>
<th></th>
<th>P. januarius n = 34</th>
<th></th>
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<tbody>
<tr>
<td>Standard length (mm)</td>
<td>21.8-22.8</td>
<td>22.3</td>
<td>22.4-34.1</td>
<td>26.4</td>
<td>18.4-32.6</td>
<td>23.7</td>
</tr>
<tr>
<td>1. Head length</td>
<td>23.2-23.8</td>
<td>23.5</td>
<td>20.9-26.2</td>
<td>23.1</td>
<td>21.7-26.6</td>
<td>24.1</td>
</tr>
<tr>
<td>2. Snout-occipital distance</td>
<td>18.6-21.4</td>
<td>20.0</td>
<td>16.4-21.7</td>
<td>18.6</td>
<td>17.9-21.8</td>
<td>20.0</td>
</tr>
<tr>
<td>3. Predorsal distance</td>
<td>64.0-68.6</td>
<td>66.3</td>
<td>55.6-61.3</td>
<td>58.4</td>
<td>55.4-62.2</td>
<td>58.4</td>
</tr>
<tr>
<td>4. Dorsal-fin base length</td>
<td>7.3-8.8</td>
<td>8.1</td>
<td>8.8-13.8</td>
<td>11.2</td>
<td>8.1-13.5</td>
<td>10.8</td>
</tr>
<tr>
<td>5. Anal-fin base length</td>
<td>7.6-8.9</td>
<td>8.2</td>
<td>8.6-11.9</td>
<td>10.2</td>
<td>7.7-12.2</td>
<td>10.1</td>
</tr>
<tr>
<td>6. Body depth</td>
<td>26.9-29.8</td>
<td>28.4</td>
<td>27.1-33.3</td>
<td>29.6</td>
<td>25.4-32.2</td>
<td>27.7</td>
</tr>
<tr>
<td>7. Pre-pelvic length</td>
<td>43.7-45.9</td>
<td>44.8</td>
<td>43.5-49.6</td>
<td>46.6</td>
<td>42.3-49.8</td>
<td>45.6</td>
</tr>
<tr>
<td>8. Pre-anal length</td>
<td>56.5-58.6</td>
<td>57.6</td>
<td>55.8-62.8</td>
<td>59.0</td>
<td>54.4-63.0</td>
<td>57.8</td>
</tr>
<tr>
<td>9. Post-anal length</td>
<td>36.4-38.9</td>
<td>37.6</td>
<td>32.9-37.8</td>
<td>35.8</td>
<td>33.0-39.1</td>
<td>36.2</td>
</tr>
<tr>
<td>10. Caudal peduncle depth</td>
<td>15.7-16.4</td>
<td>16.1</td>
<td>13.5-16.8</td>
<td>15.1</td>
<td>13.3-16.8</td>
<td>15.2</td>
</tr>
<tr>
<td>11. Snout length</td>
<td>15.8-21.4</td>
<td>18.6</td>
<td>16.5-27.0</td>
<td>21.2</td>
<td>19.7-27.7</td>
<td>23.4</td>
</tr>
<tr>
<td>12. Orbital diameter</td>
<td>40.5-40.6</td>
<td>40.6</td>
<td>31.9-41.0</td>
<td>36.7</td>
<td>29.0-39.4</td>
<td>34.7</td>
</tr>
<tr>
<td>13. Postorbital length</td>
<td>41.4-45.3</td>
<td>43.3</td>
<td>38.8-50.7</td>
<td>46.7</td>
<td>38.6-49.6</td>
<td>45.3</td>
</tr>
</tbody>
</table>

Originally defined by Hubbs (1924), was composed of genera *Phalloceros*, *Cnesterodon* Garman, 1895, *Phallotorynus* Henn, 1916, and *Diphyacantha* Henn, 1916. Hubbs (1926) added *Darienichthys* Hubbs, 1926 to the Cnesterodontini, which was excluded together with *Diphyacantha* by Rosen & Bailey (1963). These authors also included *Phalloptychus* in the tribe. Later, Ghedotti (2000) included *Tomeurus* Eigenmann, 1907. Lucinda & Reis (2005) put forward a proposal of a new classification for the subfamily Poeciliinae in the basis of a phylogenetic analysis of 144 primarily osteological characters, in which Cnesterodontini is redefined and restricted to the genera *Cnesterodon*, *Phalloceros*, and *Phallotorynus*. According to Lucinda & Reis (2005) the tribe Cnesterodontini *sensu* Rosen & Bailey is paraphyletic, given that *Phalloptychus* is more closely related to the genera *Poeciliopsis* Regan, 1913; *Xenophallus* Hubbs, 1924; *Phallichthys* Hubbs, 1924; and *Girardinus* than to *Cnesterodon*, *Phalloceros* and *Phallotorynus*. The genera *Poeciliopsis*, *Xenophallus*, *Phallichthys*, and *Girardinus* together with *Phalloptychus* constitute a monophyletic assemblage, namely the tribe *Girardinini* Hubbs, 1924 *sensu* Lucinda & Reis (2005).

At present, the distribution pattern of *Phalloptychus* is decipherable on the basis of current evidence; as a result few assumptions can be attempted in order to enlighten such a pattern. The three species inhabit lowland drainages along the Brazilian coast. *Phalloptychus januarius* and *P. iheringii* are sister species isolated from each other by the Serra Geral formation, which prevents freshwater exchange during inundation events. Similar patterns have been identified for characiform and cyprinodontiform groups by different authors, e.g. monophyletic group of *Spintherobolus* Eigenmann, 1911; some species of *Hyphessobrycon* Durbin, 1908 (Weitzman & Malabarba, 1999), *Hollandichthys* Eigenmann, 1909 (V. Bertaco, pers. comm.), *Glandulocauda* Eigenmann, 1911; *Minnowiataes* Regan, 1907; *Rachoviscus* Myers, 1926; *Deuterodon* Eigenmann, 1907 (Weitzman et al., 1988), *Jenynsia* Günther, 1866 (Ghedotti, 1998). Weitzman & Malabarba (1999) suggested that sea level fluctuations in the Pleistocene are related to the isolation of ancient widespread species and to speciation events concerning this biota.

A striking feature of *Phalloptychus* distribution is the large gap between the *P. eigenmanni* and the remaining species. There is no apparent geological evidence that could explain this gap. Past sea level fluctuations may also be involved in the isolating of *P. eigenmanni*, but they are not sufficient to explain the gap. Extinction of intermediate ancient populations may also have occurred, which leads to the assumption that *Phalloptychus* may have been more widespread in ancient times than nowadays.

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