ALLELOPATHIC EFFECT OF BLACK MUSTARD TISSUES AND ROOT EXUDATES ON SOME CROPS AND WEEDS

Efeito Alelopático de Tecidos de Mostarda-Preta e Exsudatos da Raiz de Algumas Culturas e Plantas Daninhas

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ABSTRACT - Laboratory and greenhouse experiments were conducted to evaluate the phytotoxic effect of black mustard extracts and root exudates on two crops: Trifolium alexandrinum and Triticum aestivum, and two weeds: Phalaris paradoxa and Sisymbrium irio. The seeds were treated with aqueous and ethanolic extracts and chloroform for eight days, or subjected to root exudates of just harvested mustard in a greenhouse for five weeks. High-performance liquid chromatography (HPLC) was used to quantify phytotoxins from plant tissues. Seed germination of P. paradoxa was reduced with the lowest concentration of the different extracts. However, the aqueous extract at 4% completely curtailed the germination of all the target species. In general, plant extracts had a concentration-dependent reduction of seedling growth of the target species. However, the ethanolic extract, at the lowest concentration, has stimulated the shoot length of both T. alexandrinum and T. aestivum, and the root length of the former. Mustard root exudates inhibited emergence and growth of the target species throughout the experiment. Ferulic and syringic acids were the dominant allelochemicals found when HPLC was used.

Keywords: allelopathy, Brassica nigra, plant extracts, isothiocyanates (ITCs), phenolic acids.

INTRODUCTION

Black mustard (Brassica nigra) is one of the common weeds worldwide. The allelopathic effect of Brassica spp. has been reported as the mechanism responsible for the inhibition of seed germination, for effects on coleoptile elongation and on radicle, for shoot and root development and for plant growth of many species (Tollsten & Bergström, 1988;
Choesin & Boerner, 1991; Vaughn & Boydston, 1997; Petersen et al., 2001; Turk & Tawaha, 2002; Turk et al., 2005). Sinigrin is one of the major secondary metabolites produced by *B. nigra* (Mohn et al., 2007). Under enzymatic hydrolysis this compound liberates mainly the mustard oil (allyl isothiocyanates (ITCs)), which is supposed to be an important allelochemical of *B. nigra* (Olivier et al., 1999).

Natural herbicides derived from allelochemicals could be used to control several weeds, and it is theorized that they can minimize the risks to the environment. In addition, allelochemicals released in the environment can affect other plant species. For instance, glucosinolates may be involved on weed control (Fenwick et al., 1983). The biological activity is not usually attributed to glucosinolates directly, but rather to other compounds produced under enzymatic hydrolysis such as organic cyanides (CN), oxazolidinethiones (OZT), ionic thiocyanate (SCN), and isothiocyanates, ITC, (Bangarwa et al., 2011). Myrosinase is responsible for such hydrolysis and remains stable within the dry plant tissues till hydrolysis, e.g. by soaking the dry plant material in water (Mohn et al., 2007). However, it may be difficult to determine the definite role of glucosinolate degradation products in biological activity. Moreover the assessment is complicated by the potential participation of other unidentified water soluble compounds (Choesin & Boerner, 1991; Brown & Morra, 1996). Therefore, it is necessary to know whether other phenolic compounds are participating in its allelopathic mechanism or not.

The objective of this study is to evaluate the allelopathic potential of *B. nigra* by using different extracts from the whole plant and from root exudates of this weed on seed germination and plant growth of two crops and two weeds.

**MATERIALS AND METHODS**

**Plant materials**

Black mustard plants, at the flowering stage (winter 2010), were collected from various farms located in Beni Suef Governorate, Egypt. They were left in open areas to be air dried and grinded into finely divided parts. The seeds of the most important cultivated crops, *Trifolium alexandrinum* (Egyptian clover) and *Triticum aestivum* (wheat), were obtained from the Agricultural Research Center (ARC) in Giza. Ripe seeds of both black mustard and two weeds; *Phalaris paradoxa* (hood canary grass) and *Sisymbrium irio* (London rocket), were collected from farms at the Beni Suef Governorate region.

**Preparation of extracts**

Aqueous extract was obtained by mixing different amounts of *Brassica* powder (10, 20, 30 and 40 gm) soaked in distilled water (1 L) for 24 hrs at 25 °C with mechanized shaking. The concentrations studied consisted of 1, 2, 3 and 4% (w/v). Filtration was carried out through a double layer filter paper (Whatman N.1). Distilled water was used as control.

Ethanol extract was obtained with the use of 150 g of finely divided residue of black mustard soaked in 2.5 L ethanol for 3 days. Afterwards, the solution filtrated three times in a Buchner funnel lined with a double layer of filter paper (Whatman N.1), and later centrifugation at 15,000 rpm for 15 minutes to remove fine plant debris. The extract was transferred into a rotary evaporator under vacuum at 50 °C to remove the solvent. The concentrated extract was then transferred into a 50 ml glass beaker to be completely dry. The removal of yield after solvent was 3.2 g of crude extract. The crude extract was dissolve din DMSO (Dimethyl sulfoxide) giving to the final concentration 60,000 ppm at controlled pH 7, using 10⁻² M 2-[N morpholino] of ethanesuphonic acid (MES) and adding 1 M NaOH of solution. From the stock solution, the test concentrations were prepared as follows: 300, 600, 900 and 1,200 ppm. Buffered 1% aqueous solution of DMSO was used as control.

Chloroform extract was obtained through the same procedures described in the ethanolic extract. But the recovery after solvent removal was 2.3 g of crude extract. The same concentrations mentioned above were prepared.
Bioassay

According to the different sizes for used seeds, twenty five seeds of Trifolium alexandrinum and Phalaris paradoxa, fifteen seeds of Triticum aestivum, and fifty seeds of Sisymbrium irio were sterilized with sodium hypochlorit (5.25 w/v) solution and placed on every Petri dish (9 cm diameter) lined with a filter paper. The treatments were arranged with four replicates receiving 10 mL of extract and placed in a slightly dark room with temperature ranging from 17 to 25 °C for 8 days. Germination percentage was monitored daily to obtain the germination rate. After the experiment period, plumule and radicle lengths as well as germination percentage were recorded. Radicle to plumule ratio (r/s) parameter was used to estimate the effect of extract on either radicle or plumule lengths.

Effect of root exudates

Five seeds of Brassica nigra were sown at 1 cm depth in each test pot containing well-drained silty clay loam soil with pH 7.5. Control pots were kept without Brassica seeds but watered regularly in the same time with test pots. After emergence, two healthy Brassica seedlings were left in each test pot. Twenty seeds of each test species were sown in each pot at 1 cm depth after thirty days from Brassica emergence. Four replicates were used for each treatment. Plants were harvested after 35 days. At harvest, whole pot was gently inverted and each individual plant was separated carefully from soil with the help of pressured tap water. Each individual plant was then divided into root, stem and leaves to measure the root size, shoot height, and leaf area. Leaf areas were indirectly measured by weighing their tracings on a high quality paper and comparing them with a paper of known area and weight. These organs were oven dried at 70 °C till obtaining constant weight. Dry mass was recorded for the total individual as a total dry mass.

Identification and quantification of allelopathic compounds

The residue of Brassica nigra was extracted with water in a glass soxlet for 8 hrs. Then, the crude materials were dissolved in methanol, acidified with dilute phosphoric acid to pH 2.5, and then partitioned two times with an equal volume of diethyl ether. The combined diethyl ether layers were evaporated and the resultant residue was dissolved in HPLC grade MeOH to give 1.000 ppm. 20 μl was injected into the HPLC system. Identification of the phenolic compounds in the sample was determined by comparing the retention times of known peaks with that of the following pure standards: p-hydroxybenzoic, p-coumaric, caffeic, protocatechuic, chlorogenic, salicylic, vanillic, ferulic, shikimic, gallic, syringic, sinapinic, pyrogallic, and trans-cinnamic acids as well as scopoletin, coumarin and vanillin.

Statistical analysis

The experimental design was completely randomized with four replication sets. The results of all experiments were analyzed with one-way of ANOVA and the means were compared at the level p<0.05. This analysis was carried out via the SPSS®/PC computer software package see. 11.1. (2001).

RESULTS AND DISCUSSION

Both the extracts and root exudates of black mustard had inhibitory effects on germination and growth of the target species.
Although the degree of inhibition in either radicle or plumule lengths of the target species increased gradually with the increase of the different extracts concentrations, significant stimulations for shoot lengths of Egyptian clover and wheat (14 and 20%, respectively) as well as root length of the former (14%) were detected at 300 ppm of ethanol extract. Meanwhile, there were recorded the most severely influenced radicle and plumule lengths by the 3% of water extract for Triticum aestivum (Figure 2A). The results showed that the radicle length was more sensitive to the different extracts compared to the plumule length; an obvious result due to the reduction in radicle/plumule ratio (Figure 3). Radicle/plumule ratio of Sisymbrium irio recorded the highest reduction at 3% of water extract, while that of Phalaris paradoxa was delayed at 1.200 ppm with either ethanolic or chloroform extracts.

**Effect of root exudates**

No seedling emergence was observed for Sisymbrium irio. However the emergence of Trifolium alexandrinum, Triticum aestivum and Phalaris paradoxa, was delayed (p<0.05) by 40, 25 and 27%, respectively, when compared with control (Figure 4).

Root exudates of Brassica nigra inhibited (p<0.05) both the root and the shoot lengths of all test species when compared with that of controls (Figure 5A, B). Plant mass of Trifolium alexandrinum, Triticum aestivum and Phalaris paradoxa decreased by about 74, 60 and 94% respectively (Figure 5C). Likewise, the leaf areas of the above test species were reduced by 69, 77 and 92%, respectively (Figure 5D).

**Identification of allelopathic compounds**

Six phenolic compounds were identified in water extract of plant tissues. Among the compounds identified in the whole plant extract, ferulic, syringic, and affeic acids were the major phenolics produced, with mass production of 124, 93, and 58 μg g⁻¹ of dry weight, respectively. P-coumaric and protocatechuic acids, and vanillin (aldehyde) recorded the minor amounts, 18, 39, and 12 μg g⁻¹ of dry mass, respectively (Table 1 and Figure 6).

The results showed that water extract of finely divided Brassica tissues greatly inhibited germination and seedling growth of the target species. Furthermore, the toxicity of the released compounds is species (target) dependant. This finding is in good agreement with the results of Mason-Sedun et al. (1986), where water extracts of B. nigra were mostly toxic to wheat comparing with the other test species. In addition, similar germination inhibition has been observed for lettuce, barnyardgrass and wheat when subjected to volatiles from B. nigra (Oleszek, 1987). Enzymatic hydrolysis of glucosinolates in Brassica tissues liberates various compounds (mainly ITC) that could inhibit seed germination (Brown & Morra, 1996). ITC were
Allelopathic effect of black mustard tissues and root exudates ... suggested as biocides that may interact with proteins and amino acids, particularly with the sulfhydryl groups (Fenwick et al., 1983; Kawakishi & Kaneko, 1985), amines (Kawakishi & Kaneko, 1987) and disulfide bonds (Kawakishi & Namiki, 1983) to form stable products. Thus, under higher concentrations of aqueous extracts of B. nigra, seed germination may be completely inhibited due to inactivation of the hydrolytic enzymes proceeding seed germination. This inhibition in both seed germination and seedling growth of the target species corroborates with germination (Turk et al., 2003) and growth (Turk et al., 2005) of alfalfa and radish. Furthermore, the results are in agreement with the literature in the fact that the inhibitory effect is dependent on the extract concentration (Turk et al. 2003, 2005; Ghareib et al., 2010; Hegab & Ghareib, 2010).

Figure 2 - Effect of (A) water extract, (B) ethanol extract and (C) chloroform extract of B. nigra on root length (cm) and shoot length (cm). Bars represent the standard deviation.
The data showed that the type of solvent used in the extract was important. Both ethanol and chloroform extracts were less toxic to seed germination when compared with water extract, even under the highest concentrations. This could be attributed to a partial inactivation (denaturation) of myrosinase, the enzyme responsible for hydrolysis of glucosinolates to the most toxic products (Bennett et al., 2007). The fact that at low concentration the allelochemicals (see the ethanol extract) are growth stimulators is in agreement with other results from the literature (Einhellig, 2004; Ghareib et al., 2010; Hegab & Ghareib, 2010).

It has been hypothesized that (Brown & Morra, 1996) the allelochemicals were not necessarily the hydrolysis products of glucosinolates, and there were other unidentified water soluble compounds participating in phytotoxicity. In our work, some phenolic compounds, recorded as allelochemicals, were detected in B. nigra throughout the HPLC analysis. Such identified phenolic compounds; ferulic, syringic, caffeic, p-coumaric, and protocatechuic acids, and vanillin (aldehyde) were recorded as allelopathic (Rice, 1984; Einhellig, 2004). The combined toxic action of such allelochemicals and ITCs is more effective. From literature reports, phenolic acids interfere with several enzymes and many physiological processes such as phytohormore activity, mineral uptake, photosynthesis, respiration and others (Einhellig, 1995). Thus, partial or complete germination inhibition may be attributed to inactivation of the enzymes responsible for germination by such allelochemicals. Also, complete germination inhibition at higher concentrations could result from death of embryos in seeds since such concentrations can be fatal. Reduction in growth may be attributed to water stress that reduces cell expansion or due to structural charges in membranes of the cells including alteration in membrane portions (Einhellig, 2004), or due to the suppression for cell division. Moreover, Einhellig (2004) stated that the early stage of seedling growth is very sensitive to phenolic acids. In this way, phytotoxicity of Brassica nigra may be strong due to the presence of some phenolic acids in addition to glucosinolates found in Brassica.

The results of this research may explain the field-observed long-term soil toxicity of B. nigra exudates (Tawaha & Turk, 2003).
Thus, in order for plant exudates to be persistent in the soil, it would be needed glucosinolate hydrolysis products to be released slowly and steadily from living plant parts (Haramoto & Gallandt, 2004).

One possible explanation for stronger inhibitory effect of B. nigra extracts on roots, compared to the shoots of reagent plant species, is that roots were in direct contact with the extract and subsequently with inhibitory chemicals released from the donor plant (Qasem, 1995; Turk et al., 2005; Hassan & Ghareib, 2009).

**Table 1** - Content of free phenolic compounds in *Brassica nigra* with their retention time (min.) and concentration (μg g⁻¹ dry weight)

<table>
<thead>
<tr>
<th>Standard phenolic compounds</th>
<th>Retention Time (min.)</th>
<th>Concentration (μg g⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ferulic acid</td>
<td>24.680</td>
<td>123.7</td>
</tr>
<tr>
<td>Protocatechuic acid</td>
<td>12.736</td>
<td>18.22</td>
</tr>
<tr>
<td>Caffeic acid</td>
<td>18.016</td>
<td>58.12</td>
</tr>
<tr>
<td>Syringic acid</td>
<td>18.379</td>
<td>93.5</td>
</tr>
<tr>
<td>p-coumaric acid</td>
<td>29.333</td>
<td>38.68</td>
</tr>
<tr>
<td>Vanillin</td>
<td>22.411</td>
<td>12.03</td>
</tr>
</tbody>
</table>

* Significant results at the 0.05 level.

**Figure 5** - Root exudates effect of *Brassica nigra* on (A) root length, (B) shoot length, (C) total dry weight (biomass), and (D) leaf area (cm²) of the study species. The bars represent the standard deviation with four replicates.
The fact that both weeds species, *Phalaris paradoxa* and *Sisymbrium trio*, were more sensitive to the extracts from *B. nigra* than crop species suggests that black mustard allelochemicals may be used as bioherbicides. Alternatively, the capacity of the target species to tolerate or detoxify the allelochemicals should be considered (Schultz & Wieland, 1999; Einhellig, 2004). The impact of allelochemicals on seed germination and seedling growth are acted from several aspects, which have been suggested to be on the hormone synthesis and usage, change cell division, elongation, microscopic structure, the membrane permeability and protein synthesis

In synthesis, this research has demonstrated that the toxicity of *B. nigra* extracts was caused by phenolic compounds. These chemicals are strong inhibitors of both seed germination and seedlings growth. At low concentration, the ethanol extract of black mustard had a stimulatory effect on the target species. The weed species *Phalaris paradoxa* and *Sisymbrium trio* are highly sensitive to the allelochemicals derived from *B. nigra*.

LITERATURE CITED


