Estimated features from surface EMG of the lower limb correlate with the subjective sensation of pain

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Abstract
Pain assessment is very important in establishing the efficacy of analgesics and therapies, but because pain is a subjective experience, using methods that represent pain objectively is necessary. A number of biopotentials have been employed in studies of the objective assessment of pain. However, few investigations have considered the peripheral nervous system response to electrical stimulation. The present study evaluated a method for pain quantification based on the analysis of biopotentials. We assessed electromyographic activity that resulted from evoked movements from the nociceptive flexion reflex (NFR). We investigated correlations between stimulus intensity, features extracted from surface electromyography (EMG), and subjective pain reported by subjects using a Visual Analog Scale (VAS). A total of 10 healthy male subjects without any pain disorder, aged 20-27 years, participated in the study. A high correlation ($r^2 > .87$) was found between stimulus intensity and the following features extracted from the EMG: area, root mean square (RMS), and entropy. A high correlation ($r^2 > .99$) was also found between stimulus intensity and subjective pain reported on the VAS. We conclude that estimating features from electromyographic signals that are correlated with subjective pain sensations and the intensity of the electrical stimulus is possible. Entropy, RMS, and the area of the electromyographic signal appear to be relevant parameters in correlations with subjective pain. Keywords: pain, electromyography, nociceptive flexion reflex, electrical stimulation.

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Introduction
The International Association for the Study of Pain reports that pain is an unpleasant sensory and emotional experience associated with actual or potential tissue damage (IASP), and the definition of pain is clearly made in terms of human experience (Ong & Seymour, 2004). Knowing the differences between pain and nociception is important. Pain is mediated by the nervous system, and nociception is the neural process that involves the transduction and transmission of a noxious stimulus to the brain through pain pathways as a response to tissue damage. Awareness of the stimulus is not implicated in or required for these definitions (Kandel, Schwartz, & Jessell, 2003; Steeds, 2009). With regard to pain, tissue injury is not necessary—only sensory perception is sufficient (Holdcroft & Jaggar, 2005). This perception is the result of abstraction of the brain and the development of sensory information. Pain is the result of a complex interaction between signaling and modulation from higher centers and unique perception by the individual (Kandel et al., 2003; Steeds, 2009).

Pain measurement occurs only using subjective scales, such as self-reports, with potential susceptibility to contamination by several factors that are external to the immediate pain sensation. Some of these factors are anxiety, expectations, and past experiences, which may contribute to errors and make subjective pain ratings confusing (Chan & Dallaire, 1989). Although self-reported pain provides useful clinical information and has proven to be an effective approach to pain assessment in most situations, it can fail when applied to certain vulnerable populations. Subjects with major cognitive or communicative impairments, such as intensive care unit patients or elderly individuals with dementia, may not be able to provide valid self-reports of pain (M. Averbuch, 2000). For these individuals, few methods are available for determining the presence or absence of pain.

Because of various physiological and psychosocial factors, subjects may report different levels of pain in response to the same painful stimulus (Kane, Bershadsky, Rockwood, Saleh, & Islam, 2005). Pain evaluation is important and necessary to establish the efficacy of analgesics and other therapies (Noble et al., 2005). This is generally done using a Visual Analog Scale (VAS), which is a subjective scale for the quantification of pain (M. Averbuch, 2000).
Several studies reported the importance of identifying pain intensity, but most of them used subjective methods for such measurements (Ahlers et al., 2008; Lee, 2001; Machado, Oliveira, Alves, & Andrade, 2011). Thus, according to Chan and Dallaire (1989), validating the measurement of pain using psychophysical and physiological instruments simultaneously is desirable, such as the nociceptive flexion reflex (NFR) evoked from electrical stimulation that has been proposed as a physiological nociceptive indicator (Chan & Dallaire, 1989; Rhudy & France, 2007; Willer, 1977).

The NFR is typically evaluated by monitoring the electromyographic activity of the biceps femoris when the sural nerve is electrically stimulated (France, Rhudy, & McGlone, 2009; Willer, 1977). The stimulation intensity required to evoke the NFR is used as an objective indicator of nociception threshold and applied in clinical and experimental studies of nociception and pain modulation (Dincklage, Olbrich, Baars, & Rehberg, 2013; France et al., 2009; Rhudy & France, 2007, 2011).

Chan and Dallaire (1989) demonstrated that volunteers reported the highest score for pain, based on the VAS, with increasing intensity of the electrical stimulus to yield the NFR. An increase in pain was also associated with an increase in the area of the electromyogram.

The main focus of the present study was to determine the set of characteristics extracted from the electromyographic signal that better represent the correlation with pain sensation. One of the features we evaluated was approximate entropy (S. Pincus, 1995; S. M. Pincus, 1991), which has become an additional tool in electromyographic studies related pain.

Methods

Subjects

A total of 10 healthy male subjects, aged 20-27 years, participated in the study. None of the subjects were athletes. The number of subjects was based on the studies by (Chan & Dallaire, 1989; Willer, 1977). Data collection was performed at the Biomedical Engineering Laboratory, Federal University of Uberlândia, Brazil. The data were collected after issuance of the opinion of the Ethics Committee in Research of the institution.

The subjects underwent a prior physical evaluation to evaluate physical and functional status. For all of the subjects, the inclusion criteria were the following: no history of surgery or injury or chronic pain in the right lower limb and age between 18 and 30 years. The exclusion criteria were the following: central or peripheral neurological disorders and rheumatic affections, use of a pacemaker or any heart problems, obesity (body mass index > 30 kg/m²), use of medications that cause changes in motor control and peripheral sensitivity (e.g., benzodiazepines, opioid narcotics, antihistamines, anticonvulsants, and antidepressants), lower limb amputation, and diabetes mellitus that causes impairment in peripheral sensitivity.

Apparatus

A commercial Myosystem-Br1 amplifier (DataHominis Technology, Brazil) was used for conditioning, digitalization, and recording of surface electromyographic activity. Neuropack S1 MEB-9400 equipment (Nihon Kohden, Japan) was used for electrical stimulation.

Procedure

The subjects were initially informed of the objectives of the research and data collection and subsequently signed a consent form. These participants were subjected to training to familiarize them with the data collection procedures. Data collection occurred in a calm and comfortable environment with controlled temperature (21-23°C). During the test, the subjects were asked to maintain all of their muscles as relaxed as possible in the prone position.

An active Ag/AgCl electromyography (EMG) parallel bar electrode (gain, 20; length, 25 mm; inter-electrode distance, 10 mm) was fixed with adhesive tape to the skin at the right biceps femoris, 10 cm above the popliteal fossa, according to previous reports (France et al., 2009; Rhudy & France, 2011). The EMG reference electrode was placed on the head of the right fibula over electrically conductive gel. For electrical stimulation, the cathode was positioned on the external retromalleolar pathway of the right sural nerve, and the reference electrode (anode) was positioned proximally to the right medial malleolus (Oliveira et al., 2012). These were disposable electrodes (Meditrace Ag/AgCl; 1.5 cm diameter).

An additional EMG electrode was positioned over the right extensor digitorum brevis muscle on the right lower limb with the aim of aiding the detection of the exact time when the electrical stimulus occurred. This is possible because of the short distance between the extensor digitorum brevis muscle and stimulation site (Oliveira, et al., 2012). Before positioning the sensors on the skin it was properly cleaned with alcohol and shaved whenever necessary. This procedure may help reduce skin impedance and thus increase the signal-to-noise ratio of the collected electromyographic activity.

The electromyographic signals were amplified with a gain of 2000, filtered by means of a band-pass filter with cutoff frequencies set at 20 Hz and 5000 Hz and digitized at 10 kHz by means of a 16-bit analog-to-digital signal converter.

Rectangular pulse trains (pulse width, .2 ms; inter-pulse interval, 10 ms) were used for stimulation. This current was used because of the fact that it is more uncomfortable for the subject. Electromyographic data collection began 100 ms before the electrical stimulation and finished 200 ms after it (Figure 1). Data from these two periods were collected to compare both electromyographic activities (Chan & Dallaire, 1989).
The RIII component of the flexion reflex that was evaluated in the present study has a long latency and normally appears between 85 and 120 ms after the stimulus. However, individual differences exist at the beginning and end of the flexion reflex (Chan & Dallaire, 1989; Sandrini et al., 2005; Skljarevski & Ramadan, 2002). Thus, we analyzed the full 200 ms period after the electrical stimulus. To prevent anticipatory reactions and habituation, the inter-stimulus interval varied randomly between 10 and 20 s (Chan & Dallaire, 1989).

The threshold of perception by the subjects was determined according to the method of limits, which consisted of the presentation of 10 series of ascending and descending stimuli during which the subject should report the perceived pain (Sidowski, 1966). The subject’s pain tolerance level was determined by gradually increasing the intensity of the stimuli to the maximum tolerable limit using a VAS (Figure 2). The VAS allows the subject to quantify the perceived pain from 0 (no pain) to 10 (maximum tolerable pain). For purposes of analysis, the stimulus intensity was normalized between 0% and 100%, with 0% corresponding to the pain threshold and 100% corresponding to the maximum tolerable pain for each subject. The VAS value reported by the subjects was multiplied by 10.

The change between these two intensities was then divided into 10% increments, yielding a total of 11 stimulus intensities. Each stimulus intensity was presented 10 times to the subject in random order. The data were collected on different days for each subject and saved in a text file offline for analysis using MatLab software (MathWorks).

Approximate entropy (S. Pincus, 1995), the area of the electromyogram, and root mean square (RMS) were estimated from the electromyographic signal (evoked response). These features were correlated with the pain reported on the VAS and intensity of the electrical stimulus. The ApEn and RMS were estimated 100 ms before and 200 ms after electrical stimulation as suggested by Chan and Dallaire (1989).

Approximate entropy is a tool used to quantify the regularity of a signal, returning a value between 0 and 2. ApEn = 0 represents a deterministic signal as well as a sinusoid. ApEn = 2 represents a random signal, such as white noise.

The procedure for calculating the ApEn was performed by considering the electromyographic signal sequence (demg). Choosing values for the parameters m (standard length) and s (tolerance or similarity criterion of comparison) was necessary to calculate the ApEn of the sequence. If an m signal sample window, starting from sample i, is denoted by pm (i), then two windows, pm (i) and pm (j), are similar if the difference between any pair of corresponding samples of the windows are smaller than r \[ demg (i + k) - demg (j + k) \] < r, for 0 ≤ k < m. pm is the set of all windows of length m of demg, the number of windows of length m that resemble the window of the same length, starting from i is Cim (r). Cim (r) is the number of windows in pm that are similar to pm (i). Thus, Cim (r) can be calculated for each pm window, estimating Cm (r) as an average of these values. Cm (r) measures the regularity or frequency of similar windows in a given set of windows, demg that contained an m length, complying with tolerance r. Then, d ApEn can be defined as in Equation 1.

\[
\text{ApEn}(m, r, \text{demg}) = \ln \left[ \frac{Cm(r)}{Cm+1(r)} \right]
\]

Equation 1

where demg is the electromyographic signal, m is the window length, and r is the tolerance.

Approximate entropy, ApEn, measures the similarity between the windows of lengths m and m+1. The technique was applied to the electromyographic signal with a value of m = 2 and r = .2 SD (demg), where .2 SD (demg) is the standard deviation of demg as suggested by Pincus (S. M. Pincus, 1991). For data processing, we developed specific software tools using MatLab.

Statistical analysis

The study analyzed the ApEn before and after stimulation, the area of the electromyographic signal, and VAS and RMS before and after stimulation as a function of the intensity of electrical stimuli applied to correlate these variables with the sensation of pain.

We calculated the Pearson correlation coefficient (r) and coefficient of determination (r²) and performed linear regression analysis. The alpha level of the statistical tests was 5%. The data followed a normal distribution, based on the Shapiro-Wilk test, with a confidence level of 95%. These data are shown in Table 1. The results are presented as the average for the subjects.

Results

Table 1 presents the estimated linear models, describing the correlation among the stimulus intensity (%) and investigated features (i.e., area, entropy before and after the stimulus, and VAS and RMS before and after the stimulus). Table 2 presents the mean values of the VAS and standard deviation in 10 presentations...
of current intensities for the subjects at each 10% increment. Figures 3-6 show the regression lines and estimated features. In the figures, the mean estimates for all subjects are given.

A high positive linear correlation was found between VAS score and stimulus intensity (Figure 3). A weak correlation was found between entropy estimated before the stimulus and stimulus intensity. Entropy after the stimulus had a high negative correlation with stimulus intensity (Figure 4). The area had a high correlation with stimulus intensity (Figure 5). A weak correlation was found between the RMS before the stimulus and stimulus intensity. The RMS after the stimulus had a high correlation with stimulus intensity (Figure 6). Figure 7 presents an example of raw data of the NFR.

### Table 1. Correlations between the stimulus intensity (%) and investigated features.

<table>
<thead>
<tr>
<th>Feature</th>
<th>Pearson’s correlation coefficient ($r$)</th>
<th>Estimated linear model</th>
<th>Coefficient of determination ($r^2$)</th>
<th>$p$</th>
</tr>
</thead>
<tbody>
<tr>
<td>VAS</td>
<td>.998307</td>
<td>$y = .5502x + 18.011$</td>
<td>.9966</td>
<td>$p&lt;.05$</td>
</tr>
<tr>
<td>Entropy (before stimulus)</td>
<td>-.677000</td>
<td>$y = -.0573x + 52.935$</td>
<td>.4589</td>
<td>$p&gt;.05$</td>
</tr>
<tr>
<td>Entropy (after stimulus)</td>
<td>-.933150</td>
<td>$y = -2.433x + 60.415$</td>
<td>.8708</td>
<td>$p&lt;.05$</td>
</tr>
<tr>
<td>Area</td>
<td>.9740940</td>
<td>$y = .3559x + 16.971$</td>
<td>.9489</td>
<td>$p&lt;.05$</td>
</tr>
<tr>
<td>RMS (before stimulus)</td>
<td>.09000000</td>
<td>$y = .0114x + 40.141$</td>
<td>.0082</td>
<td>$p&gt;.05$</td>
</tr>
<tr>
<td>RMS (after stimulus)</td>
<td>.97000000</td>
<td>$y = .321x + 20.333$</td>
<td>.9460</td>
<td>$p&lt;.05$</td>
</tr>
</tbody>
</table>

### Table 2. Mean VAS score and standard deviation (SD) of 10 presentations of current intensities for each subject at each 10% increment.

<table>
<thead>
<tr>
<th>10% Increments</th>
<th>V1 Mean (SD)</th>
<th>V2 Mean (SD)</th>
<th>V3 Mean (SD)</th>
<th>V4 Mean (SD)</th>
<th>V5 Mean (SD)</th>
<th>V6 Mean (SD)</th>
<th>V7 Mean (SD)</th>
<th>V8 Mean (SD)</th>
<th>V9 Mean (SD)</th>
<th>V10 Mean (SD)</th>
<th>Mean of the Mean (SD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0%</td>
<td>.00</td>
<td>.00</td>
<td>42.5 (12.08)</td>
<td>1 (3.2)</td>
<td>29.3 (12.6)</td>
<td>14 (5.2)</td>
<td>45 (22.7)</td>
<td>8.5 (3.4)</td>
<td>8.5 (4.1)</td>
<td>14.9 (6.6)</td>
<td>16.37 (6.19)</td>
</tr>
<tr>
<td>10%</td>
<td>0.0 (0.0)</td>
<td>0.0 (0.0)</td>
<td>44 (10.49)</td>
<td>13 (9.5)</td>
<td>39.5 (14.8)</td>
<td>22 (6.3)</td>
<td>49 (19.1)</td>
<td>13 (4.2)</td>
<td>19.5 (6.7)</td>
<td>21.1 (7.04)</td>
<td>22.21 (7.04)</td>
</tr>
<tr>
<td>20%</td>
<td>2.8 (4.3)</td>
<td>1.0 (0.0)</td>
<td>51 (8.43)</td>
<td>20 (12.5)</td>
<td>46 (12.0)</td>
<td>22 (6.3)</td>
<td>51 (16.0)</td>
<td>18.5 (10.0)</td>
<td>30 (18.6)</td>
<td>26.2 (7.5)</td>
<td>26.85 (18.04)</td>
</tr>
<tr>
<td>30%</td>
<td>8.1 (3.8)</td>
<td>3.5 (0.0)</td>
<td>56 (6.99)</td>
<td>30 (12.5)</td>
<td>55.5 (14.1)</td>
<td>26 (7.0)</td>
<td>52 (19.3)</td>
<td>25 (9.1)</td>
<td>32 (17.4)</td>
<td>31.8 (6.3)</td>
<td>31.99 (18.26)</td>
</tr>
<tr>
<td>40%</td>
<td>16 (6.6)</td>
<td>10 (5.0)</td>
<td>60 (7.07)</td>
<td>46 (24.1)</td>
<td>58.1 (10.2)</td>
<td>38 (7.9)</td>
<td>53 (20.6)</td>
<td>35 (16.0)</td>
<td>38 (6.7)</td>
<td>39.4 (16.46)</td>
<td>39.35 (16.46)</td>
</tr>
<tr>
<td>50%</td>
<td>29.3 (8.9)</td>
<td>14 (0.0)</td>
<td>60.5 (5.50)</td>
<td>41 (21.3)</td>
<td>67.7 (11.4)</td>
<td>44 (12.6)</td>
<td>57 (22.1)</td>
<td>36.5 (13.1)</td>
<td>54 (26.1)</td>
<td>45.4 (7.2)</td>
<td>44.44 (15.91)</td>
</tr>
<tr>
<td>60%</td>
<td>29.5 (8.5)</td>
<td>21 (5.0)</td>
<td>62.5 (9.20)</td>
<td>55 (21.2)</td>
<td>68 (10.3)</td>
<td>46 (14.3)</td>
<td>61 (19.7)</td>
<td>44 (10.7)</td>
<td>54 (25.4)</td>
<td>50.2 (6.4)</td>
<td>49.19 (14.74)</td>
</tr>
<tr>
<td>70%</td>
<td>44.5 (7.2)</td>
<td>30 (11.0)</td>
<td>67 (6.32)</td>
<td>71 (21.3)</td>
<td>67.3 (11.7)</td>
<td>49 (19.7)</td>
<td>65 (21.2)</td>
<td>51 (11.5)</td>
<td>58 (17.4)</td>
<td>57.3 (3.7)</td>
<td>56.01 (12.66)</td>
</tr>
<tr>
<td>80%</td>
<td>53.2 (12.5)</td>
<td>23.5 (11.4)</td>
<td>71 (8.76)</td>
<td>88 (4.2)</td>
<td>84.7 (7.4)</td>
<td>59 (13.7)</td>
<td>68 (16.9)</td>
<td>56 (17.3)</td>
<td>59 (21.3)</td>
<td>64.2 (6.7)</td>
<td>62.66 (18.02)</td>
</tr>
<tr>
<td>90%</td>
<td>68.1 (5.6)</td>
<td>45 (13.6)</td>
<td>75 (5.27)</td>
<td>95.5 (5.0)</td>
<td>85 (7.9)</td>
<td>61 (19.1)</td>
<td>69 (17.9)</td>
<td>61 (14.9)</td>
<td>75 (14.9)</td>
<td>72.5 (3.7)</td>
<td>70.75 (13.86)</td>
</tr>
<tr>
<td>100%</td>
<td>80.2 (5.7)</td>
<td>51 (15.8)</td>
<td>80.5 (4.97)</td>
<td>95 (8.5)</td>
<td>88.8 (6.0)</td>
<td>65 (15.8)</td>
<td>69 (20.8)</td>
<td>71 (13.5)</td>
<td>75 (15.6)</td>
<td>77.6 (4.0)</td>
<td>75.31 (12.39)</td>
</tr>
</tbody>
</table>

| Estimated linear model | $y = .8245x - 515x - .3623x^2 + .9955x + 5734x + .5236x + .2591x + .6164x + .6456x + .1686x + .5947x$ | $r = .97$ | .95 | .99 | .98 | .98 | .98 | .99 | .98 | .95 | .99 |

Note: The last column shows the mean of the mean and standard deviation. The last two rows show the estimated linear model and Pearson’s correlation coefficient ($r$). Notice that the subjects presented a linear relationship with pain. V = voluntary/subject.

### Discussion
To find objective EMG parameters that can be correlated with pain, several features were analyzed. This protocol showed a strong linear correlation between VAS score and stimulus intensity. Chan and Dallaire (Chan & Dallaire, 1989) also reported a linear correlation between VAS score and stimulus intensity. As the stimulus intensity increased, the subjects reported a higher VAS score. Entropy estimated after the stimulus was strongly correlated with pain sensation (Oliveira et al., 2012). The area of the electromyographic signal was positively correlated with the intensity of the electrical stimulation that caused the pain. The RMS showed a strong positive linear correlation with the stimulus. This
Correlation between electromyography and pain

Figure 3. Linear regression and correlation between VAS and the painful stimulus. On the x-axis, 0% and 100% correspond to the intensity threshold and pain tolerance, respectively ($p < .05$).

Figure 4. Linear regression and correlation between entropy (after stimulus) and the painful stimulus. On the x-axis, 0% and 100% correspond to the intensity threshold and pain tolerance, respectively ($p < .05$).

Figure 5. Linear regression and correlation between area and stimulus. On the x-axis, 0% and 100% correspond to the intensity threshold and pain tolerance, respectively ($p < .05$).

Figure 6. Linear regression and correlation between RMS (200 ms after the stimulus) and the stimulus. On the x-axis, 0% and 100% correspond to the intensity threshold and pain tolerance, respectively ($p < .05$).

Figure 7. Electromyographic response elicited in the biceps femoris muscle of subject V1. The electromyographic signal was rectified and filtered at 75 Hz. The dashed line corresponds to stimulation intensity of 20%, and the solid line corresponds to 100%, noting that the variation among the pain threshold and maximum tolerable stimulation was divided into 10% increments, producing 11 stimulus intensities.
correlation was high during the period 200 ms after electrical stimulation and was not observed during the 100 ms period before stimulation.

According to S Pincus (1995) and S.M. Pincus (1991), ApEn was introduced for use in time series by quantifying the predictability, regularity, or complexity of experimental data. This decrease in entropy according to the increase in pain may be attributable to the fact that some muscle fibers are already pre-recruited to implement the reflex response. As the pain increases, the proportion of recruited fibers increases. These observations suggest that the nociceptive response is a highly organized mechanism that allows painful stimuli to activate appropriate muscles that initiate the most adequate withdrawal response (Skljarevski & Ramadan, 2002). Therefore, the ApEn has a lower value in regular time series and a greater value in irregular and complex temporal series.

Skljarevski and Ramadan (2002) reported that Sherrington, in the early twentieth century, observed that pain caused by electrical stimulation in a limb in animal experiments caused a withdrawal reflex or the hip, knee, and ankle of the same limb, a phenomenon called the NFR. Because the reflex does not appear without the activation of nociceptive fibers, the method has become a useful tool for clinical pain research. Furthermore, digital technology advances have allowed greater reproducibility (Skljarevski & Ramadan, 2002).

In addition to the NFR, different studies utilized other methods, such as the M wave, algometry, pain scales and questionnaires, hypertonic saline solution, electroencephalography, thermosensitivity, functional magnetic resonance imaging, and lasers (Bottega & Fontana, 2010; Buchgreitz, Egsgaard, Jensen, Arendt-Nielsen, & Bendtsen, 2008; Ervilha, Farina, Arendt-Nielsen, & Graven-Nielsen, 2005; Graven-Nielsen, Arendt-Nielsen, & Mense, 2002; Hogeweg, Langereis, Bernards, Faber, & Holders, 1998; Iannetti, Hughes, Lee, & Mouraux, 2008; Iannilli, Gratta, Gerber, Romani, & Hummel, 2009; Kane et al., 2005; Machado et al., 2011; Ong & Seymour, 2004).

The NFR can also be studied in subjects with headaches, fibromyalgia, back pain, and knee pain. In such disorders, central hyperexcitability occurs because of central facilitation sustained by continuous peripheral nociceptive afferents from an unrecognized source, which leads to signal amplification by hyperexcited nociceptive spinal neurons (Lim, Sterling, Stone, & Vicenzino, 2011).

The present study evaluated male subjects because many women experience hormonal changes during their menstrual cycle (Sheffield, Biles, Orom, Maixner, & Sheps, 2000; Sherman & LeReche, 2006), which may interfere with pain assessment.

Some reports have indicated that the very slow loss of muscle mass occurs at 25-50 years of age (Matsudo, Matsudo, & Neto, 2000). Because this sarcopenia is rather insignificant at 25 to 28 years of age (i.e., accounting for < 1% loss (Powers & Howley, 2005), we chose 20- to 27-year-old subjects for this study, for whom no significant sarcopenia would be expected.

An important contribution of the present study was the verification that entropy after stimulation, RMS, and area of the flexion reflex are relevant parameters for quantifying pain. The identification of these features may be used in more complex algorithms and tools for automatic pain quantification procedures. The methodology used for data collection in the present EMG-based protocol was based on investigations of correlations performed by several research groups (Chan & Dallaire, 1989; France, et al., 2009; Rhudy & France, 2007, 2011; Willer 1977) that used similar techniques in studies of pain assessment. Electromyography does not measure pain itself but provides an inference to pain with parameters that can be analyzed, including the area of the electromyographic signal RMS, and entropy. The discovery of the relationship between the above-cited features and pain is very important for the development of new protocols that can objectively contribute to solving problems associated with pain and its quantification. A relevant factor is that this research utilized an easily reproducible protocol that can be used for the development of future studies.

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