In vitro effect of intracanal medicaments on strict anaerobes by means of the broth dilution method

Efeito in vitro de medicações intracanal sobre anaeróbios estritos pelo método de diluição em caldo

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INTRODUCTION

Irrigation and mechanical cleaning of the root canal are of the utmost importance in reducing the amount of bacteria during the endodontic treatment. However, without the utilization of intracanal medication, half of the endodontically treated root canals would still present with bacteria in the second session. Intracanal dressing, thus, helps eliminating bacteria that are not affected by the chemical-mechanical preparation and prevents, or at least delays, the reinfection of canals between sessions. Therefore, any medication chosen for intracanal application must be evaluated as to its antimicrobial activity by means of in vitro susceptibility tests. Usually, the evaluation is carried out on facultative and aerobic bacteria, and the agar diffusion technique is employed. However, the infection of root canals is polybacterial, with the prevalence of anaerobic bacteria, whose susceptibility is ideally determined by means of dilution tests in agar or broth.

In the endodontic literature, evaluations of the susceptibility of anaerobes to antimicrobial agents are relatively scarce; besides, studies carried out with dilution quantitative techniques practically do not exist. Hence, the aim of this study was to standardize a broth dilution test, utilizing traditional intracanal medicaments and reference anaerobic strains, in order to guarantee the reproducibility of the results and correlate the in vitro...
results with the clinical performance of these substances.

MATERIAL AND METHODS

Microorganisms and drugs

Reference strains of *Prevotella nigrescens* (ATCC 33563) and *Fusobacterium nucleatum* (ATCC 25586), representative of the most frequently isolated anaerobes in endodontic infections, as well as of *Clostridium perfringens* (ATCC 13124), commonly employed in tests with antibiotics, were utilized. Bacteria were maintained at −20°C in cryotubes containing skim milk (0.5 ml of active growth added to 0.5 ml of a sterile 20% solution of powdered skim milk). Solutions of 10% calcium hydroxide (Biodinâmica, Brazil); 2% chlorhexidine digluconate (FGM, Brazil); 35% camphorated paramonochlorophenol (PMCC) (Inodon, Brazil) and Formocresol (Probem, Brazil) were evaluated.

Microdilution test

In order to select the dilutions that would be utilized in the broth macrodilution test, microdilution tests were carried out in microplates. Initially, two screw-capped tubes, measuring 16 x 160 mm and containing 10 ml of the reinforced clostridial medium (RCM, Oxoid, USA) or Brucella broth (Difco, USA) supplemented with 5 µg/ml of hemin (Sigma, USA) and 10 µg/ml of vitamin K 1 (Merck, Germany) were inoculated with 0.1 ml of the skim milk stock and incubated in anaerobiosis (jar and envelopes Gaspak-BBL, USA), at 37°C, for 24 hours. As soon as growth was observed, two additional subcultures were made, in the same way, with incubation in anaerobiosis at 37°C, for 24 hours, in order to obtain bacteria in an exponential phase of growth. After the last incubation, the turbidity of the tubes was read in an Ultrospec 1000 spectrophotometer (Pharmacia, USA), at 540 nm. Having the 0.5 McFarland standard as a reference, the dilution of the cultures in RCM and Brucella broths was carried out up to the concentration of 5 x 10⁵ colony forming units/ml (CFU/ml)²³. This standardized inoculum was always used within 30 minutes after the adjustment²⁵.

The microdilution test was carried out in a sterile flat-bottomed microplate, with 96 wells (Corning, USA). Initially, each well received 100 µl of the standardized inoculum. The tests were carried out in duplicate. Six controls were utilized for all tests. Incubation with PMCC and formocresol was carried out separately, due to the activity of the vapors of those medicaments. Microplates were incubated in anaerobiosis, at 37°C for 48 hours.

In the face of the impossibility of visual reading of growth in the wells, and considering the impossibility of determining values of minimal inhibitory concentration (MIC), the determination of minimal bactericidal concentration (MBC) was carried out¹³. For that, 25 µl from each well were dispensed on Petri plates containing RCM agar with 5% sheep blood (RCM-blood), each plate holding 25 drops. These plates were incubated in anaerobiosis, at 37°C for 48 hours.

Macrodilution test²¹

The sequence was the same previously described, but it was carried out in tubes measuring 13 x 100 mm with screw caps, which initially received 2.5 ml of RCM or supplemented Brucella broth. After obtaining the desired concentrations of the medicaments, according to serial two-fold dilutions, all tubes received 2.5 ml of the standardized inoculum. Positive and negative controls were included. Before and after incubation at 37°C in anaerobiosis, for 48 hours for RCM broth and for 96 hours for Brucella broth, the reading of the optical density (OD) of all tubes was carried out in the spectrophotometer and, whenever possible, also by means of visual reading. Once MIC had been established, MBC was determined by means of spreading, with a Drigalsky loop, 25 µl of the material from each tube without growth on Petri plates measuring 52 x 12 mm, which contained 5 ml of RCM-blood agar. The samples were incubated in anaerobiosis, at 37°C for 48 hours.

RESULTS

The values of MBC obtained with the microdilution tests are shown in Tables 1 and 2, for RCM and Brucella broth, respectively. MICs and MBCs obtained through the macrodilution test in RCM broth are described in Table 3, and those obtained in Brucella broth, in Table 4.

DISCUSSION

The method recommended by the National Committee for Clinical Laboratory Standards (NCCLS, USA) to test the susceptibility of anaero-
bes is the agar dilution method, with the alternative of macrodilution test in broth when there are few isolated bacteria and agents to be tested. The agar diffusion method is counterindicated. Since both methods are expensive and very time-consuming, the microdilution test has been developed.

With antibiotics, the results obtained by means of this method are pretty similar to those obtained with the agar dilution method. In general, the utilized dilution is smaller than that employed in macrodilution in broth, which makes the reading easier, since the transparency of drugs does not complicate the naked-eye visualization of the changes that take place in the wells. However, the solutions utilized in this study significantly change the employed media, which makes impossible to visually determine MICs. For that reason, MBCs were determined (Tables 1 and 2), even though microdilution is not adequate for that procedure since with the small amount of inoculum utilized there is the risk of not detecting the elimination of 99.9% of the viable CFU in the broth (MBC).

The discrepancy between MBCs obtained through the micromethod and by means of the macromethod (Tables 1 to 4) – the values obtained with the micromethod were much lower or higher than those obtained through the macromethod – confirms that the first method is inadequate to this determination. Nevertheless, this fact does not restrain the future utilization of the micromethod to determine the MIC of medications utilized in endodontics, with readings of growth carried out with the photometer employed in the technique of enzyme immunoassay (ELISA). SHAPIRO et al. (1994), who utilized the microdilution test for thymol, did not observe any difference between the

### TABLE 1 - Minimal bactericidal concentrations (µg/ml) of substances employed against anaerobic bacteria – microdilution test in RCM broth.

<table>
<thead>
<tr>
<th>Bacteria</th>
<th>Ca(OH)₂</th>
<th>Chlorhexidine</th>
<th>PMCC*</th>
<th>Formocresol</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>F. nucleatum</em></td>
<td>1,562.50</td>
<td>0.30</td>
<td>0.0025</td>
<td>92.75</td>
</tr>
<tr>
<td><em>P. nigrescens</em></td>
<td>3,125.00</td>
<td>0.0044</td>
<td>0.33</td>
<td>740.00</td>
</tr>
<tr>
<td><em>C. perfringens</em></td>
<td>&gt; 50,000</td>
<td>156.25</td>
<td>340.00</td>
<td>92.75</td>
</tr>
</tbody>
</table>

* 35% camphorated paramonochlorophenol.

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<th>Formocresol</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>F. nucleatum</em></td>
<td>6,250.00</td>
<td>0.00014</td>
<td>10.62</td>
<td>185.00</td>
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<tr>
<td><em>P. nigrescens</em></td>
<td>3,125.00</td>
<td>0.00050</td>
<td>0.33</td>
<td>0.36</td>
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<tr>
<td><em>C. perfringens</em></td>
<td>&gt; 50,000</td>
<td>39.06</td>
<td>170.00</td>
<td>742.00</td>
</tr>
</tbody>
</table>

* 35% camphorated paramonochlorophenol.

### TABLE 3 - Minimal inhibitory (MIC) and bactericidal concentrations (MBC) (µg/ml) of substances employed against anaerobic bacteria – macrodilution test in RCM broth.

<table>
<thead>
<tr>
<th>Bacteria</th>
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</tr>
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<tr>
<td><em>F. nucleatum</em></td>
<td>781.25</td>
<td>1,562.50</td>
<td>0.30</td>
<td>1.22</td>
</tr>
<tr>
<td><em>P. nigrescens</em></td>
<td>781.25</td>
<td>1,562.50</td>
<td>0.30</td>
<td>1.22</td>
</tr>
<tr>
<td><em>C. perfringens</em></td>
<td>6,250.00</td>
<td>6,250.00</td>
<td>2.44</td>
<td>2.44</td>
</tr>
</tbody>
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<th>Bacteria</th>
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</tr>
</thead>
<tbody>
<tr>
<td><em>F. nucleatum</em></td>
<td>781.25</td>
<td>781.25</td>
<td>0.15</td>
<td>0.15</td>
</tr>
<tr>
<td><em>P. nigrescens</em></td>
<td>781.25</td>
<td>781.25</td>
<td>0.15</td>
<td>0.15</td>
</tr>
<tr>
<td><em>C. perfringens</em></td>
<td>1,562.50</td>
<td>1,562.50</td>
<td>2.44</td>
<td>2.44</td>
</tr>
</tbody>
</table>

* 35% camphorated paramonochlorophenol.

### TABLE 4 - Minimal inhibitory (MIC) and bactericidal concentrations (MBC) (µg/ml) of substances employed against anaerobic bacteria – macrodilution test in Brucella broth.

<table>
<thead>
<tr>
<th>Bacteria</th>
<th>Ca(OH)₂</th>
<th>Chlorhexidine</th>
<th>PMCC*</th>
<th>Formocresol</th>
</tr>
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<tr>
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</tr>
<tr>
<td><em>P. nigrescens</em></td>
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<td>0.15</td>
<td>0.15</td>
</tr>
<tr>
<td><em>C. perfringens</em></td>
<td>1,562.50</td>
<td>1,562.50</td>
<td>2.44</td>
<td>2.44</td>
</tr>
</tbody>
</table>

* 35% camphorated paramonochlorophenol.
MICs obtain by means of the micro and macromethods for the anaerobic bacterium Porphyromonas gingivalis.

The results displayed in Tables 3 and 4 indicate chlorhexidine as the drug that was efficient in the lowest concentrations, followed by PMCC, formocresol and calcium hydroxide, for all bacteria, which confirms that the first drug shows good results in eliminating anaerobes. Despite the need for higher concentrations of calcium hydroxide for the obtainment of an antimicrobial effect, the results do contest the absence or low intensity of the activity of that drug, which has been shown by means of different in vitro methods. However, the present results are in agreement with those reported by SUZUKI et al. (1999), who found MICs ranging from 1,560 to 25,000 µg/ml, for twelve distinct anaerobes, demonstrating that the minimal inhibitory concentrations depend on the studied bacterium.

The good performance of PMCC against anaerobic bacteria, which has been previously observed by other authors, was confirmed. The liquid form, which presents a higher antimicrobial effect than the clinically utilized vapor form, was employed. Therefore, the minimal bactericidal and inhibitory concentrations of the vapor form must be higher than the those presented here. According to SPANGBERG (1994), when utilizing the vapor form, the concentration of the medicament should be 100 to 1,000 times higher than that employed when utilizing the liquid form.

OHARA et al. (1993) tested the in vitro effect of vapors of volatile drugs and found out that formocresol was more efficient than PMCC. In their study, except for P. nigrescens, MICs for PMCC and formocresol were similar; MBCs were similar or lower (with the variation of one dilution) for PMCC, depending on the utilized medium.

Comparisons between the three most commonly utilized substances can be found in the literature, with divergences as to the antimicrobial activity against species of anaerobic bacteria, mainly due to differences in the employed methodologies and in the interpretation of data. Nevertheless, in a clinical evaluation, BARBOSA et al. (1997) did not observe any significant difference between the percentages of cases rendered negative for bacteria after receiving intracanal dressings of PMCC, calcium hydroxide and chlorhexidine (69%, 73% and 78%, respectively). This fact is especially important because it refers the clinician to other criteria, besides antibacterial efficacy, for choosing an intracanal agent, such as toxicity, inflammatory potential, diffusibility, ability to neutralize endotoxins and stimulate tissue repair in the periapical area.

It is worth to mention that the bacterium Clostridium perfringens, which is not a member of the oral cavity normal flora and is commonly used as a test control, was the microorganism with the most resistant profile to all drugs.

In relation to the utilized culture media, employing a standardized inoculum, the growth of the positive control (without any drug) of all bacteria in supplemented Brucella broth after incubation for 96 hours (OD between 1,000 and 1,400) was smaller than that observed in RCM broth after 48 hours (OD ≥ 2,000). RCM broth always presented the best performance, which was also verified in the activation of strains. Yet, the averages of MICs and MBCs were not expressively different for the three species of bacteria, in both media, with final values quite similar or showing differences of one or two lower dilutions in Brucella broth. In the future, other broths should be tested.

It is known that pre-reduced anaerobically sterilized media (PRAS) allow for a better growth of anaerobes, but its preparation is impossible in most clinical laboratories. That justifies the attempt to establish a practicable technique for average laboratories that have anaerobiosis jars, which allows for the preparation of the rich media one day before the experiment. The methodology has shown a good performance with the tested microorganisms. It may present problems when employed with more fastidious microorganisms, for which more intensive subcultures do not afford the growth proper to constitute the inoculum for susceptibility tests – that growth must preferably occur within 18 to 24 hours.

In vitro studies, such as the present research, are carried out with isolated bacteria while endodontic infections are usually mixed – with complex microbial interactions and an average concentration of viable bacteria of 10³⁷ CFU/g of sample, which may interfere with the action of different medicaments. In spite of that, the importance of presenting quantitative data (µg/ml) on the antimicrobial effect of drugs must be pointed out, since those studies are based on standardized methodologies that can be reproduced by microbiology laboratories that do not have the same resources as great research centers.
CONCLUSIONS

Spectrophotometric reading renders good reliability to the interpretation of MIC. MICs and MBCs obtained by means of the macrodilution method are consistent with the clinical performance of the obtained by means of the macrodilution method.

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BIBLIOGRAPHIC REFERENCES


