Conduction velocity of the rabbit facial nerve: a noninvasive functional evaluation

Velocidade de condução no nervo facial do coelho: uma avaliação funcional não invasiva

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ABSTRACT: The aim of this study was to evaluate standardized conduction velocity data for uninjured facial nerve and facial nerve repaired with autologous graft nerves and synthetic materials. An evaluation was made measuring the preoperative differences in the facial nerve conduction velocities on either side, and ascertaining the existence of a positive correlation between facial nerve conduction velocity and the number of axons regenerated postoperatively. In 17 rabbits, bilateral facial nerve motor action potentials were recorded pre- and postoperatively. The stimulation surface electrodes were placed on the auricular pavilion (facial nerve trunk) and the recording surface electrodes were placed on the quadratus labii inferior muscle. The facial nerves were isolated, transected and separated 10 mm apart. The gap between the two nerve ends was repaired with autologous nerve grafts and PTFE-e (polytetrafluoroethylene) or collagen tubes. The mean of maximal conduction velocity of the facial nerve was 41.10 m/s. After 15 days no nerve conduction was evoked in the evaluated group. For the period of 2 and 4 months the mean conduction velocity was approximately 50% of the normal value in the subgroups assessed. A significant correlation was observed between the conduction velocity and the number of regenerated axons. Noninvasive functional evaluation with surface electrodes can be useful for stimulating and recording muscle action potentials and for assessing the functional state of the facial nerve.

DESCRIPTORS: Nerve regeneration; Electrophysiology; Neural conduction.

INTRODUCTION

Nerve conduction data are used routinely in the diagnosis of peripheral neuropathies and other conditions.1,14,26 Surface electrodes have been used for stimulating and recording the muscle action potentials. Normally three different types of electrodes are used in electromyography: surface, needle and implanted electrodes15,16,30. The recording of peripheral nerves conduction velocity by directly stimulating and recording the surgically exposed nerves is one of the most precise approaches to nerve regeneration and muscle reinnervation. However, such single brief recordings makes it difficult to evaluate the nerve regeneration process

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over time\textsuperscript{15,30}. On the other hand, needle electrodes and electrodes implanted around peripheral nerves for sustained periods of time may generate artifacts that interfere with the interpretation of the effects of nerve injury\textsuperscript{15,16,19,20,30}. Furthermore, infection also may occur as a result of invasive puncture with needles or the surgical implantation of devices, in particular when percutaneous contacts are used\textsuperscript{15,30}.

Contrasting with needle and implanted electrodes, surface electrodes require no surgery, and evaluations can be repeated over sustained periods of time thereby affording an acceptable measurement approach\textsuperscript{15,30}. Nerve conduction velocity is calculated by dividing the interelectrode distance by the latency of the first positive deflection in the waveform\textsuperscript{13}.

The present experimental model studied normal conduction velocity in the rabbit facial nerve before surgery, and across facial nerve repaired with autologous nerve grafts and synthetic materials, with the aim of establishing the effectiveness of noninvasive functional testing.

**MATERIALS AND METHODS**

The stimulation and recording of antidromic evoked motor action potentials were carried out in 17 male New Zealand white rabbits (2.5 to 3.0 kg). The animals were anesthetized with an intramuscular injection of a mixture of ketamine 50 mg/kg (Ketolar\textsuperscript{®}, Parke-Davis, El Prat Llobregat, Barcelona, Spain) and acepromazine maleate 5 mg/kg (Calmo neosan\textsuperscript{®}, SmithKline-Beechan, Madrid, Spain), repeated as necessary. In addition, local anesthesia with 3\% mepivacaine (Isogaine\textsuperscript{®}, Clarben, Madrid, Spain) was applied via subcutaneous injection in the surgical area. A mandibular incision towards the preauricular area identified the facial nerve. All animals were subjected to noninvasive functional study before and after surgery.

**Nerve lesions**

The buccal division of the facial nerve on each side and distally to the parotid gland was identified and isolated. A 10 mm segment was removed from the buccal division of the facial nerve in all cases (n = 34, nerves) (Figure 1). The defects, in turn, were repaired with autologous nerve grafts, collagen and polytetrafluoroethylene (PTFE-e) conduits (Figures 2, 3 and 4). The autologous nerve grafts were obtained from the left facial nerve, of the same animal.

The animals recovered from the surgery on heated pads and were housed in individual cages, with free access to food and water. All animals survived the surgery without problems. At the completion of the study, all rabbits were sacrificed with overdose of sodium pentobarbital via an intracardiac injection.

After sacrifice, the anastomotic sites, graft or conduits plus a segment of approximately 5 mm from the proximal and distal nerve stumps were removed and marked. All specimens were fixed in glutaraldehyde and stained with toluidine blue. Cross-sectional biopsy specimens were examined under light microscopy (100 X) to assess intra-neural morphology. The axon counts were obtained in the sections from the midpoint of the primary anastomosis.

**Electrophysiological studies**

Electrophysiological analysis was performed following anesthesia. The stimulation surface electrodes were placed on the auricular pavilion (facial nerve trunk) and the recording surface electrodes were placed on the quadratus labii inferior muscle. Evoked muscle action potential was recorded by electromyography (Medelec MS25 Mistro, Surrey, England). Normal conduction velocity for the right and left facial nerve was recorded pre- and postoperatively. Conduction velocities were evoked across the reconstructed defect in the postoperative period, after peripheral nerve repair the rabbits were evaluated at 15 days and at 1, 2 and 4 months, and correlated to the number of axons regenerated.

**Statistical Analysis**

Data analysis of the results was performed using the SPSS 7.5 (Statistical Package for Social Science - Microsoft, Illinois, Chicago, USA). Conduction velocity comparisons between the two sides (left and right) were made with the Student t-test for paired data. Correlations between conduction velocity and regenerated axon count were in turn evaluated by Pearson correlation analysis.

**RESULTS**

**Normal conduction velocity**

Bilateral facial nerve conduction tracings were obtained for each of the 17 rabbits studied. There were no untoward events or complications. The
mean normal right conduction velocity evoked preoperatively was 41.27 ± 3.27 m/s, versus 40.92 ± 2.71 m/s on the left side. Nerve conduction velocity at the peak of the signal was 47.05 m/s. There were no statistically significant differences between sides at the 95% confidence level (Student t-test, p = 0.67). The mean global normal conduction velocity evoked before surgery was 41.10 ± 2.97 m/s.

**Postoperative nerve conduction velocity**

No nerve conduction was evoked in the animals evaluated after 15 days, while the mean conduction velocity recorded after 2 and 4 months was approximately 50% of the normal value. The conduction velocity evoked at 4 months was 21.21 ± 8.46 m/s (repair with nerve graft) and 18.26 ± 5.69 m/s (repair with polytetrafluoroethylene tube).

The conduction velocity at 2 months for the polytetrafluoroethylene tube and collagen groups were 22.95 ± 6.52 m/s and 24.11 ± 6.12 m/s and the results at 4 months were 21.68 ± 3.7 m/s (polytetrafluoroethylene tube) and 23.29 ± 2.47 m/s (collagen).

**Normal axonal nerve**

The total number of myelinated axons ranged from 156.37 to 176.21. The mean myelinated nerve count (± standard deviation) was 166.29 ± 9.45 myelinated axons (oculoreticular area) (Figure 5).

**Postoperative axonal regeneration**

15 days after PTFE-e, autologous nerve grafts and collagen repair, an absence of axonal regeneration was observed (Figure 6). However, within 4 months, a good axonal regeneration was observed.
for all groups (Figure 7). In the postoperative period, a significant correlation between conduction velocity and axonal regeneration was observed for all groups and techniques (n = 34; 0.002 – Pearson test).

**DISCUSSION**

One of the challenges of peripheral nerve regeneration is the early and quantitative assessment of the regeneration process in terms of the number of active axons found in the bundle. Reinnervation dynamics can be studied using different techniques. In the present study the efficacy of a noninvasive method was evaluated in an application in a rabbit model.

The use of single-fiber electromyography for evaluating the microphysiology of an individual motor unit has been described in the literature.

This technique can be used to study the denervation and reinnervation occurring as a result of peripheral nerve injury.

A comparative study between magnetic nerve stimulation with conventional electric nerve stimulation in normal subjects and in patients with disorders of the peripheral nervous system has been reported. In general, both nerve stimulation modalities yielded similar results in patients with disorders and healthy subjects.

The literature reports an evaluation of the inferior alveolar nerve function using a repeated nerve conduction test during mandibular sagittal split osteotomy. The intraoperative sensory nerve conduction velocity recorded and the mental nerve blink reflex tests conducted two weeks after the operation were correlated, whereas comparison of clinical neurosensory testing with the intraoperative events and sensory conduction velocities was less consistent.

Another research studied conduction velocity of the deep temporal nerve and the mylohyoid nerve motor branches of the trigeminal nerve. These nerves were stimulated intraorally with a pediatric surface stimulator. Compound muscle action potentials were recorded over the temporalis and mylohyoid muscles using surface electrodes. The mean latency of the deep temporal nerve was 2.1 m/s ± 0.3 m/s, versus 1.9 m/s ± 0.2 m/s for the mylohyoid nerve. The maximal side-to-side latency difference was 0.4 m/s. These authors concluded that such techniques should prove useful in patients with trigeminal nerve disorders.

A new technique for recording sensory conduction velocity of the inferior alveolar nerve was re-
ported by Jaaskelainen. The sensory responses of this nerve were obtained in all 42 nerves examined with silver wire or monopolar needle electrodes inserted beneath the zygomatic arch, anteriorly to the temporomandibular joint and to a depth of 4 to 4.5 cm, near the oval foramen. A small bipolar surface electrode was used for stimulation at the mental foramen. The mean sensory conduction velocity of the inferior alveolar nerve was 62.8 m/s ± 6.7 m/s on the right side, and 63.0 m/s ± 5.4 m/s on the left side.

The influence of temperature, age, and height upon conduction velocity has been described by a number of authors. Frassen et al. reported differences in evoked potentials at different temperature in humans. Our own observations show mean conduction velocity of the facial nerve (41.10 m/s) at 25°C to be similar to the results reported by Frassen et al. (45.0 m/s). In this case, environmental temperature and age of the animals were controlled. In addition, the mean conduction velocity of the facial nerve was consistent with values previously reported in the literature (range 35.0-64.6 m/s).

There were no significant serial differences between the results of conduction velocity between the left and right facial nerves. Ours results agree with those reported by Colin et al.

In this study a significant correlation between the conduction velocity and the number of regenerated axons involving different techniques was observed (p < 0.01; r = 0.002). This could explain why the study variables exhibited an increasing tendency during nerve regeneration.

CONCLUSION

In all groups and techniques, a significant correlation between conduction velocity and axonal regeneration was observed.

Noninvasive functional evaluation with surface electrodes can be useful for stimulating and recording muscle action potentials and for assessing the functional state of the facial nerve following injury or surgery.

REFERENCES