RESUMO. - Babesiose e anaplasmose em bovinos leiteiros no Nordeste do Brasil. O objetivo deste estudo foi caracterizar a situação epidemiológica e os fatores envolvidos na prevalência da babesiose e anaplasmose em bovinos da bacia leiteira de Parnaíba, Piauí, Brasil. O estudo foi realizado em 22 propriedades, sendo coletadas amostras de sangue de 202 bovinos para estudos sorológicos, molecular e determinação do volume globular (VG). Na PCR, Babesia spp. e A. marginale foram amplificados em 51,5% de amostras, mostrando co-infecção. A anaplasmose foi mais prevalente, com 76,2% de amostras positivas. O manejo semi-intensivo predominou em 68,0% das propriedades. O estudo indica que nesta região há instabilidade enzoótica para babesiose e estabilidade enzoótica para anaplasmose, reforçando o fato de que, no Brasil, existem áreas de instabilidade enzoótica, mesmo em regiões tropicais. O PCR technique foi um ferramenta valiosa para o diagnóstico de estas doenças e pode ser usado para caracterizar um regiões.

INDEX TERMS: Babesia bigemina, Babesia bovis, Anaplasma marginale, epidemiology, diagnosis.

Babesiosis and anaplasmosis in dairy cattle in Northeastern Brazil

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The goal of this study was to characterize the epidemiological situation and the factors involved in the prevalence of babesiosis and anaplasmosis in dairy cattle in the dairy basin of Parnaíba, Piauí, Brazil. The study was conducted in 22 farms, and collected blood samples from 202 cattle to study serological, molecular and determination of the packed cell volume (PCV). The farms were applied surveys involving epidemiological aspects. Sero-prevalence rates were: Babesia bigemina 52.5%, B. bovis 68.8%, and Anaplasma marginale 89.1%. Of the samples analyzed, 73.3% were reactive for Babesia spp. and A. marginale, showing co-infection. In PCR, B. bigemina and B. bovis were positive in 52.0% and 33.2% respectively, and A. marginale in 76.2%. Of these, 51.5% amplified DNA of Babesia spp. and A. marginale. The semi-intensive management predominated in 68.0% of the farms studied. The clinical history of babesiosis and anaplasmosis was reported from 73% of the farms. There was no significant difference (p>0.05) between age groups and for the PCV of positive compared with negative animals. The study indicates that in this region is enzootic instability for babesiosis and enzootic stability for anaplasmosis, reinforcing the fact that in Brazil there are areas of enzootic instability, even in tropical regions of the country. The PCR technique was a valuable tool for the diagnosis of these diseases and may be used to characterize a geographic region.
INTRODUCTION

Bovine babesiosis is caused by the hemoprotozoa *Babesia bovis* and *Babesia bigemina*, whereas anaplasmosis is caused by the intra-erythrocytic rickettsia *Anaplasma marginale*. These diseases are clinically and epidemiologically similar (Bock et al. 2004, Kocan et al. 2010), and they are responsible for significant economic losses for the livestock industry worldwide.

The most important biological vector for these three agents is the tick *Rhipicephalus (Boophilus) microplus*, which is distributed in tropical and subtropical regions (Estrada-Peña et al. 2006, OIE 2007). In addition to the biological transmission by ticks, *A. marginale* can be transmitted mechanically by bloodsucking flies or iatrogenically form by fomites contaminated with blood from infected cattle (Dreher et al. 2005, Kocan et al. 2010). The dynamics of infection of these parasites is dependent on factors such as vector population, transmission capability of the vector, and host susceptibility (Kocan et al. 2010).

Direct diagnosis of these diseases is based on detection of the agent in the blood smears (Bock et al. 2004) or by polymerase chain reaction (PCR) or *nested* PCR (nPCR) (Figueroa et al. 1993, Costa Júnior et al. 2006). Indirect diagnostic methods include serological methods such as enzyme-linked immunosorbent assays (ELISA) (Madruga et al. 2001), and indirect fluorescent antibody test (IFAT) (Santos et al. 2001).

Epidemiological assessment of babesiosis and anaplasmosis in a given area allows to predict the risk for occurrence of outbreaks. Therefore, the epidemiological situation can be classified as three types: enzootic stability, enzootic instability, and disease-free areas (Mahoney 1974). Most of the Brazilian territory is characterized as an area of enzootic stability for *Babesia* spp. and *A. marginale* (Barci et al. 1994, Araújo et al. 1997, Madruga et al. 2000, Santos et al. 2001). However, the Southern and Northern parts of Brazil include areas of enzootic instability (Artiles et al. 1995, Lima et al. 1999).

Studies on the epidemiological situation of babesiosis and anaplasmosis in Northeastern Brazil are extremely scarce. Considering that dairy cattle in this region play an significant social and economic role, epidemiological studies are very important in this region, particularly due to the fact that there exist peculiar climatic patterns.

The epidemiological situation of bovine babesiosis and anaplasmosis in Piauí (Northeast Brazil) is not known. Cattle mortality associated with clinical signs of these diseases have reported, but without laboratory confirmation. The goal of this study was to determine the prevalence of babesiosis and anaplasmosis in cattle in the dairy basin of Parnaíba, State of Piauí, with parasitological, serological, and molecular methods, in association with possible risk factors involved.

MATERIALS AND METHODS

**Study area.** This study was conducted in the dairy basin of Parnaíba, in the micro-region Litoral Piauiense, in the Northern part of the State of Piauí, Northeast Brazil. This area is located between longitudes 41°39'W e 42°05'W and latitudes 2°51'S e 3°55'S (Fig.1). This micro-region covers an area of 7,742 km², with a herd of 99,815 cattle (IBGE, 2010). The climate is classified as tropical with two well-defined seasons: the rainy season from December to April and the dry season from May to November. Temperature ranges from 23 to 33.6°C, with 40-80% relative air humidity, and an average rainfall of 1,200mm (Medeiros 2004).

**Blood samples.** Twenty two dairy farms were sampled. These farms have been randomly selected from the ADAPI (Agência de Defesa Agropecuária do Piauí) files, restricting the selection to farms that had no regular veterinary assistance and reasonably easy accessibility. Questionnaires regarding several epidemiological aspects such as type of breeding, sanitary conditions, clinical signs of diseases and control measures were applied to all herds. Samples were collected from all municipalities that are part of that particular dairy basin, proportionally to their cattle population. In total, 202 cattle over the age of one year were sampled. The sample size was determined based on a mathematical model developed by the Centro Pan Americano de Zoonosis (1979). An estimated prevalence of 67% was considered for calculation of the number of cattle to be sampled. This estimated prevalence...
was based on a preliminary experiment that included 78 cattle of this area, considering a degree of confidence of 95%, with an accepted 10% margin of error.

Blood samples with anticoagulant (EDTA) were used to determine the packed cell volume (PCV) by the microhematocrit technique, and for DNA extraction. Serum samples were obtained for IFAT.

**Serology.** Anti-*Babesia bigemina*, anti-*B. bovis* and anti-*Anaplasma marginale* IgG was detected by IFAT according to the Instituto Interamericano de Cooperación para la Agricultura (IICA) 1987. Samples that were reactive at a dilution ≥1:40 were considered positive.

**DNA extraction, PCR, nPCR and sequencing.** DNA was extracted from blood samples using a commercial kit (Illustra blood genomicPrep Mini Spin - GE Healthcare) following the manufacturer's instructions. PCR was used for amplification of DNA from *B. bovis* and *B. bigemina*, using previously described primers: GAU9/GAU10 and GAU7/GAU6 respectively (Linhares et al. 2002). Amplification of *A. marginale* was performed by nPCR using the primers Am9 and Am10, followed by an internal amplification in the case of nPCR was performed with a 1μl of a previous amplification with the primers Am11 and Am12 (Figueroa et al. 1993). PCR and nPCR were performed in a 25μl reaction, containing ultra-pure water; 1x Taq buffer; 0.2mM each dNTP (Promega); 1.0mM MgCl2; 0.4μM each primer; 1.5 U Taq DNA polymerase (Promega) and 200 ng of sample DNA. The second round of amplification in the case of PCR was performed with a 1μl of a previously amplified PCR product. Reactions were performed under the following conditions: 94°C for 2 min (*B. bovis* and *B. bigemina*) or 95°C for 7 min (*A. marginale*); 35 cycles of denaturation for 30s at 94°C (*B. bovis* and *B. bigemina*) or 95°C (*A. marginale*); annealing for 30s at 58°C (*B. bovis*) or 60°C (*B. bigemina* and *A. marginale*); extension for 1 min at 72°C, and a final extension at 72°C for 5 min in a Gene Pro thermocycler (Biomer).

Amplified products were subjected to electrophoresis in 1.5% agarose gel. PCR products had 541 bp for *B. bovis*, 690 bp for *B. bigemina*, and 170 bp for *A. marginale*.

Sequencing of PCR products amplified (*B. bovis*, *B. bigemina*, and *A. marginale*) from cattle blood samples were carried out in both directions using the ABI Prism BigDye Terminator Cycle Sequencing Ready Reaction Kit (Applied Biosystems). About 10ng of purified DNA, for each sequencing reaction, was combined with 3.2pmol of primer (forward and/or reverse) used in the amplification reaction. Nucleic acid sequence analyses were performed on an automated ABI PRISM 3500 Genetic Analyzer DNA sequencer.

Consensus sequences were obtained through the analysis of the sense and antisense sequences using the GAP version 1.4 - Staden Package (http://staden.sourceforge.net/). Comparisons were made with the sequences deposited in GenBank using the Basic Local Alignment Search Tool - BLAST (http://www.ncbi.nlm.nih.gov/BLAST/).

**Statistical analysis.** All statistical analyses were performed using a commercial statistical package GraphPad Prism, version 5.0. Data analysis was performed using Chi-square test, with a significance level of 0.05. The mean and standard deviations were estimated for PCV values. Mean PCV values of infected (as assessed by PCR) and seropositive cattle were compared with those of the uninfected and seronegative cattle using Student’s t-tests. Prevalence was estimated based on the total number of IFAT-positive cattle and total infected by PCR.

**Ethics and animal experimentation.** This study was conducted under the terms and conditions of the Ethics Committee for Animal Experimentation of the Universidade Federal do Piauí, approved under number 030/2010.

**RESULTS**

Prevalence of *Babesia bigemina*, *B. bovis*, and *Anaplasma marginale* as determined by IFAT and by PCR amplification of DNA, ranged from 33.2% to 89.1% depending on the species and method used for detection (Table 1). As shown in Figure 2, there was no statistically significant differences (p>0.05) in the prevalence of *B. bovis*, *B. bigemina*, and *A. marginale*, when various age groups were compared.

PCV ranged from 14% to 42%, by mean PCV of cattle positive for *B. bigemina*, *B. bovis*, *A. marginale* as well as co-infected cattle as demonstrated by PCR were not significantly different (p>0.05) from negative cattle (Table 1).

![Fig.2. Seroprevalence of Babesia bigemina, Babesia bovis and Anaplasma marginale antibodies by age group in cattle from the dairy basin of Parnaíba, state of Piauí, Northeastern Brazil.](http://example.com/fig2.png)

<table>
<thead>
<tr>
<th></th>
<th>B. bigemina</th>
<th>B. bovis</th>
<th>A. marginale</th>
<th>Co-infection (Babesia spp. and A. marginale)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Seroprevalence (IFA)</td>
<td>52.5% (106/202)</td>
<td>68.8% (139/202)</td>
<td>89.1% (180/202)</td>
<td>73.3% (148/202)</td>
</tr>
<tr>
<td>Prevalence of infection (PCR)</td>
<td>52.0% (105/202)</td>
<td>33.2% (67/202)</td>
<td>76.2% (154/202)</td>
<td>51.5% (104/202)</td>
</tr>
<tr>
<td>Mean PCV (%) of:</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Seropositive</td>
<td>29.38 ± 5.03*</td>
<td>29.27 ± 5.19*</td>
<td>29.07 ± 4.96*</td>
<td>29.17 ± 4.91*</td>
</tr>
<tr>
<td>Seronegative</td>
<td>28.84 ± 4.90*</td>
<td>28.76 ± 4.41*</td>
<td>29.48 ± 5.07*</td>
<td>27.20 ± 7.25*</td>
</tr>
</tbody>
</table>

*p* Means in the same column followed by the same letter do not differ (p>0.05) by Student’s *t*-test.
Amplification and sequencing of a 681 bp fragment of the 18S rRNA gene of \(B.\) \(b\)igemina (deposited in GenBank under accession number [X104106]) demonstrated 99% identity to sequences of \(B.\) \(b\)igemina from cattle in Spain (DQ788311 and FJ426361), Germany (EF458190), and China (AY603402); \(B.\) \(b\)igemina from water buffalo in China (HQ840960), and from white-tailed deer in the USA (HQ264115). Similar results were observed with \(B.\) \(b\)ovis 18S rRNA gene sequences generated in this study (accession number [X661267], a 543 bp fragment that had 100% identity with \(B.\) \(b\)ovis from cattle in the USA (L31922), and 99% identity from cattle in Germany (EF458218), and from white-tailed deer in USA (HQ264126). A 161 bp amplified fragment of the major surface protein gene of \(A.\) \(m\)arginale had 98% identity to sequences of complete genome \(A.\) \(m\)arginale Florida strain from cattle in USA (CP001079) and the Maries strain (CP000030).

A semi-intensive management was practiced in 68% of the 22 farms included in this study. Cattle in these farms were predominantly crossbreed of Gir (\(B.\) \(t\)aurus \(i\)ndicus) with Holstein (\(B.\) \(t\)aurus \(t\)aurus). Most farms (95%) had cultivated pastures, but only 59% of them adopted pasture rotation. All farms had cattle infested with ticks at the time of interview, whereas 64% also reported the presence of hematophagous diptera. All farms adopted procedures for controlling the population of ticks, based on acaricide treatments, but these procedures were perfomed without veterinarian supervision or a strategic control plan. Clinical history and/or signs of babesiosis and/or anaplasmosis including anorexia, apathy, pale mucous membranes, hemoglobinuria, fever, icteric mucous membranes, and abortion were reported in 73% of the farms, and only 68% of the farms treated cattle for these diseases. In 50% of the farms, reported deaths associated with clinical history of the disease. Most of the farms (95%) kept calves up to six months of age within a collective hut with food and water. These calves had contact with cows only during milking, but even at this stage calves were already infested with ticks.

**DISCUSSION**

Low seroprevalence rate of Babesia \(b\)igemina and \(B.\) \(b\)ovis observed in this study allow us to consider this region as an area of enzootic instability. In the case of Anaplasma \(m\)arginale, due to high seroprevalence, this same region is characterized by an area of enzootic stability. This classification is based on epidemiological concepts proposed by Mahoney (1974) that describes its application to control possible outbreaks.

Seroprevalence rates for \(B.\) \(b\)igemina and \(B.\) \(b\)ovis demonstrated in this study were lower than the seroprevalences previously reported in most states in the Northeast region of Brazil, where predominates areas of enzootic stability. In a study conducted in semi-arid region of Bahia, the prevalence of antibodies anti-\(B.\) \(b\)igemina and anti-\(B.\) \(b\)ovis was 77.7% and 75.5%, respectively (Barros et al. 2005). In the micro-regions of Feira de Santana, Jequié, Ilhéus, and Vitória da Conquista (Bahia State), the average percentages of cattle serologically positive for \(B.\) \(b\)ovis and \(B.\) \(b\)igemina were 97.2% and 95.0%, respectively (Araújo et al. 1997). In the region known as Araguáina (Tocantins State) seroprevalence determined for \(B.\) \(b\)ovis was 90.5% (Trindade et al. 2010). The unstable situation found in this study may high risk of clinical manifestation of infections in this region because there is the possibility of outbreaks of babesiosis in adult animals causing high mortality rate and economic losses. For \(A.\) \(m\)arginale the result seroprevalence found resembles the one recorded for other regions of Brazil (Souza et al. 2000, Souza et al. 2001), but differs in some Northeastern states, such as Sergipe, where a prevalence of 16.3% has been reported (Oliveira et al. 1992).

Regarding that age, seroprevalences of \(B.\) \(b\)ovis, \(B.\) \(b\)igemina, and \(A.\) \(m\)arginale did not differ statistically among groups, suggesting that these animals are infected mostly before they reach one year of age, when they tend to be more resistant to hemoparasites. Incidence of these diseases is highly variable among adult cattle as previously reported by Souza et al. (2000), Trindade et al. (2010), and Barros et al. (2005).

The frequency of infection with Babesia spp. and \(A.\) \(m\)arginale in cattle as determined by PCR and nPCR was similar to frequencies obtained by IFA serological test. These molecular methods allow detection of babesiosis and anaplasmosis during the initial phase of infection as well as asymptomatic carriers, and therefore they are powerful tools for epidemiological investigations, once these animals represent a major source of infection for female \(R.\) (\(B.\)) \(m\)icroplus (Oliveira-Sequeira et al. 2005). PCR and nPCR methods employed in this study have been previously used for research of these diseases in several Brazilian states (Figueroa et al. 1993, Madruga et al. 2002, Oliveira-Sequeira et al. 2005, Costa Júnior et al. 2006). This demonstrated that the Parnaiba dairy basin is an area of enzootic stability for \(A.\) \(m\)arginale and instability for \(B.\) \(b\)igemina and \(B.\) \(b\)ovis. In the geographic area of this study, semi-intensive farms are predominantly associated with high temperatures and low rainfall (close to zero during the dry season), what is likely to interfere with the life cycle of the tick, thus influencing the risk of infection with Babesia spp. This notion is in good agreement with a previous report by Ouhelli et al. (1987), who found that at high temperatures (25 to 35°C) there is a reduction in oviposition, hatching rate, and longevity of the larvae infected by \(B.\) \(b\)igemina and \(B.\) \(b\)ovis; this results in a decreased risk of transmission of Babesia spp., favoring enzootic instability, whereas enzootic stability in the case of Anaplasma may be due to a mechanical transmission.

As shown in this study, based on answers to the questionnaires, ticks and hematophagous flies were present in most of the farms. Improvement in health management, such as intensive use of insecticides and acaricides reduced the exposure of cattle to vectors of babesiosis and anaplasmosis, resulting in low infectious challenge for calves. This situation favors the emergence of areas of endemic instability (Guglielmone 1995, Bock et al. 2004). No characteristic clinical signs of babesiosis and/or anaplasmosis were observed while visiting the farms included in this study; what may be a consequence of widespread treatment of affected cattle adopted by farmers in this region (Kuttler & Johnson 1996, Kocan et al. 2010).
Our results indicated that cattle PCR and serologically positive for babesiosis and anaplasmosis had PCV values that were similar to negative cattle, and that anemia was occasionally observed in a few individual animals. Considering that anemia is proportional to parasitemia due to disseminated intravascular hemolysis as well as mechanisms sequestration and lysis of normal erythrocytes (Alfonso et al. 1996), it is likely that the levels of parasitaemia observed in this study were below the threshold required for triggering clinical disease.

CONCLUSION

Our results indicate that Babesia bigemina, B. bovis, and Anaplasma marginale infections occur in the dairy basin of Parnaíba in the state of Piauí, with features of enzootic instability in the case of babesiosis, and enzootic stability in the case of anaplasmosis.

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OIE. Rhipicephalus (Boophilus) microplus. <http://www.cfsph.iastate.edu/ Factsheets/pdfs/boophilus_microplus.pdf>


