RESUMO.- [Infeção por Vírus da Imunodeficiência Felina (FIV), vírus da leucemia felina (FeLV) e Leishmania sp. em gatos domésticos no Centro-Oeste do Brasil.] Esta pesquisa teve o objetivo de investigar a infeção em gatos domésticos por FIV e FeLV, analisando o perfil epidemiológico destas doenças, assim como a infeção por Leishmania sp. Oitenta e oito gatos domésticos foram avaliados pesquisando a infeção por FIV, FeLV e Leishmania sp. Onze (12,5%) gatos foram positivos para infeção por FIV, quatro (4,5%) foram positivos para FeLV, e dois gatos apresentaram co-infeção pelos dois vírus. Entretanto, nenhum gato doméstico apresentou infeção por Leishmania sp. A prevalência da infeção para FIV foi maior que a observada para FeLV, e que a observada em outras regiões, mas nenhum fator teve associação à infeção neste estudo.

TERMOS DE INDEXAÇÃO: Felino, FIV, FeLV, visceral leishmaniasis, PCR.

INTRODUCTION

Feline immunodeficiency virus (FIV) and feline leukaemia virus (FeLV) are retroviruses found within domestic cats around the world (Chhetri et al. 2015). Both viruses can be transmitted horizontally via saliva or other body fluids, and vertical transmission can also occur (Munro et al. 2014). The resultant infections are accompanied by dysfunction of the immune system, making the cats susceptible to other diseases (Hartmann 2012). Clinical manifestations due to the specific actions of the viruses, such as feline gingivitis-stomatitis complex in the case of FIV and lymphoma in certain FeLV-infected cats, have also been described (Beatty 2014).

In Brazil, related studies are still scarce, although a survey of data from several regions yielded prevalence ranging from 4.14 to 21.5% for FIV and from 10.8 to 32.5% for FeLV (Teixeira et al. 2007). There are no reports on the prevalence of these viruses in domestic cats in the Midwest of the country, where research has been performed only on wild felids kept in captivity (Schmitt et al. 2003).

Several serological tests detect the presence of antibodies against FIV proteins (p24 core protein or gp40 transmembrane protein) and the presence of soluble antigen of FeLV (p27 core protein). Among the serological tests, for FIV testing, Snap Combo Plus is recommended as the best...
performing in-clinic test, while for FeLV, the positive predictive value was 73.5% (Hartmann et al. 2007). However, molecular methods are more sensitive and specific to detect the pathogen during the first 2-4 weeks post-infection, as well as during the terminal stages of infection by FIV therefore the antibody response is generally low (Wilkes et al. 2015).

Infections with FIV and FeLV appear to increase the frequency of opportunistic infections, such as infection with *Leishmania* sp., for which felines are considered alternative hosts in endemic areas, such as Cuiabá, the capital of Mato Grosso (Dahroug et al. 2010, Sobrinho et al. 2012). In endemic areas for canine leishmaniosis, the subclinical feline infection by *Leishmania* are common, while clinical illness is rare. There is limited information on epidemiological and clinical manifestations of these infection (Pennisi et al. 2015).

Thus, the aim of the present study was to investigate FIV and FeLV infections in domestic cats and wild felids, analysing the epidemiological profile of the disease as well as additional infection with *Leishmania* sp.

MATERIALS AND METHODS

A transversal study was performed on 88 domestic cats healthy (17/19.3%) or sick (71/80.7%) at the University Veterinary Hospital in the city of Cuiabá, Mato Grosso State (15° 35’ 46” S 56° 05’ 48” W), in the Midwest of Brazil. The minimum sample size was defined as 71 cats, finite population of 310 cats, with an expected prevalence of 6.4% (Alves et al. 2011), an acceptable error rate of 5% and a 95% confidence interval, as defined by Epi Info 6.0 (Centers for Disease Control and Prevention [CDC], Atlanta, GA, USA). A questionnaire on the lifestyle and habits of the felines was given to each owner, followed by clinical evaluation of the felines and collection of blood samples by puncturing the jugular vein.

The blood samples were processed with EDTA for CBC automatic analysis and with phenol/chloroform/alcohol isooamyl (Sambrook et al. 1989) for DNA extraction, with later detection of FIV by nested PCR and of *Leishmania* by PCR.

The serum was used for the detection of FIV and FeLV using the IDEXX SNAP FIV/FeLV Combo Test diagnostic kit (IDEXX Laboratories, Markham, Ontario) in accordance with the manufacturer’s recommendations.

PCR for detection of FIV (Lara et al. 2008) and *Leishmania* sp. (Degrave et al. 1994) was performed using the primers listed in Table 1 in all samples. In all PCR reactions, used DNA reference strain of *L. infantum* (MHOM/BR/1974/PP75) and for FIV, we used one sequenced sample (unpublished data). For negative control we used the DNA-free reaction. For FIV and *Leishmania* sp., the amplification products were fractionated by 1.5 or 2.0% agarose gel electrophoresis, respectively, stained with ethidium bromide and analysed using a transilluminator (UV 300nm).

### Table 1. Primers used to detect FIV and *Leishmania* sp. and their expected amplicon sizes

<table>
<thead>
<tr>
<th>Primers name</th>
<th>Sequence 5’- 3’</th>
<th>Amplicons</th>
</tr>
</thead>
<tbody>
<tr>
<td>FIV PCR S2</td>
<td>AATATGACGCTATCTCTGTCG</td>
<td>658bp</td>
</tr>
<tr>
<td>FIV PCR A2</td>
<td>TTTTCCTTACGAGTCTTCTCGG</td>
<td>329bp</td>
</tr>
<tr>
<td>NESTED S FIV</td>
<td>TATTCAAAACGTAATGGGACG</td>
<td>120bp</td>
</tr>
<tr>
<td>NESTED A FIV</td>
<td>GGGG(G/T)AGGGGCGTTCT(C/G)CGAA</td>
<td>658bp</td>
</tr>
</tbody>
</table>

The data were analysed using the nonparametric chi-square test or Fisher’s exact test via Epi Info 6.0 (CDC, Atlanta, GA, USA). Agreement between the diagnostic techniques for FIV was determined using sensibility, specificity and Cohen’s kappa coefficient (κ), consider ELISA tests as the gold standard. A κ value of 0.2-0.6 represents fair to moderate agreement, a κ value of 0.6–0.8 represents substantial agreement, and a κ value of > 0.8 represents nearly complete agreement.

RESULTS

Eleven (12.5%; CI: 5.7-19.3) of the cats were positive for FIV infection, four (4.5%; CI: 1.1-9.1) were positive for FeLV, and two cats were co-infected (Table 2).

### Table 2. Clinical and haematological characteristics of cats with infection by FIV and / or FeLV

<table>
<thead>
<tr>
<th>Cat (year)</th>
<th>Gender</th>
<th>Street access</th>
<th>Origin</th>
<th>Clinical features</th>
<th>Haematological findings</th>
<th>ELISA FIV</th>
<th>PCR FIV</th>
<th>ELISA FeLV</th>
</tr>
</thead>
<tbody>
<tr>
<td>3</td>
<td>M</td>
<td>Cattery</td>
<td>Lymphadenomegaly</td>
<td>Anemia, thrombocytopenia</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>M</td>
<td>No</td>
<td>Lymphadenomegaly, weight loss, dehydration, otitis, FCG*•</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td></td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>F</td>
<td>Yes</td>
<td>Lymphadenomegaly, weight loss, anorexia, dehydration, FCG*•</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td></td>
<td></td>
</tr>
<tr>
<td>8</td>
<td>M</td>
<td>No</td>
<td>Lymphadenomegaly, FCG*•</td>
<td>Lymphopenia</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>9</td>
<td>M</td>
<td>No</td>
<td>FLUTD**</td>
<td>Leukopenia, lymphopenia, thrombocytopenia</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>10</td>
<td>M</td>
<td>Yes</td>
<td>Dermatitis</td>
<td>Hyperproteinemia</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td></td>
</tr>
<tr>
<td>11</td>
<td>M</td>
<td>Yes</td>
<td>Dermatitis</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td></td>
<td></td>
</tr>
<tr>
<td>12</td>
<td>M</td>
<td>No</td>
<td>Lymphadenomegaly, apathy, anorexia, dehydration weight loss, FRD**, diarrhea, dermatitis</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

* FCG = Feline chronic gingivostomatitis, ** FLUTD = Feline Lower Urinary Tract Disease, *** FRD = Feline Respiratory Disease, + positive, - negative.
Regarding the clinical characteristics and habits of 88 cats, there was no statistically significant association with FIV infection and gender (p=0.39), and outdoor access (p=0.57). For FeLV and gender (p=0.68) and outdoor access (p=0.66), as FIV/FeLV with unneutered cats (p=0.32) there was no statistically significant association, but many of the cats had disturbance in the oral cavity (Table 2). The cats had not received vaccine for FIV or FeLV. The haematological findings of infected cats (Table 2) were not statistically different from the uninfected.

Of the 11 cats with FIV, all were positive by ELISA, but only six were also positive by nested PCR, with substantial agreement between the tests (κ=0.678). The sensitivity was 54.5% (CI: 23.4-83.3) and the specificity was 100% (CI: 93.1-100) for detection the infection for FIV. Using the PCR, DNA was not detected Leishmania sp. in domestic cats.

**DISCUSSION**

The prevalence for FIV infection was higher than FeLV. It is contrasted by Munro et al. (2014) observed seroprevalence was 2.2% for FIV and 6.2% for FeLV in shelter cats in Canada, and Ortega-Pacheco et al. (2014) found 2.5% for FIV, 7.5% for FeLV in owned cats in Mexico, that used the same test. This difference may have related to the sampled population of cats, either by free access to the street, high population density or poor hygienic conditions (Teixeira et al. 2007, Lara et al. 2008, Munro et al. 2014).

We did not find significant correlation between genders and outdoor access with FIV or FeLV infections, despite the infections are more frequent in cats with free access to the street, adulthood and unneutered males (Teixeira et al. 2007, Lara et al. 2008, Hartmann 2012). According Chhetri et al. (2015) outdoor exposure is more important to acquire FIV infection than FeLV, due increased opportunity for transmission via fights between cats.

Although there was no statistical difference, the hematologic findings of the infected cats were consistent with findings in other studies on these infections, several of which were performed on cats with neoplastic or non-neoplastic manifestations induced by FIV or FeLV (Hartmann 2012).

The clinical evaluation showed that 84.6% of the FIV/FeLV-positive cats were sick, while 15.4% of cats has none clinical alterations. Disorders of the oral cavity, and particularly stomatitis, may be associated with FIV infection in many cases and may be due to the immune response to chronic antigenic stimulation (Hartmann 2012).

ELISA is a sensitive technique capable of detecting early antigens as well as viraemia due to only transient FeLV infection that can be overcome (Meinerzs et al. 2010). For FIV infection, ELISA may provide sufficient sensitivity for screening, while DNA PCR for FeLV may have low sensitivity for detecting lentiviral strains because of low proviral load in blood cells (Franklin et al. 2007), which explains five cats are negative in this test.

Though the region studied here is endemic for visceral leishmaniasis and *Leishmania* sp. infection in wild felids (Dahroug et al. 2010), DNA was not found in any of the samples of cat tested. However, co-infection with FIV and *Leishmania* has been described in an area endemic for visceral leishmaniasis in Brazil (Sobrinho et al. 2012).

**CONCLUSION**

The prevalence for FIV infection was higher than FeLV, and none factor was associated with the infection by FIV and FeLV in this study.

**Acknowledgements** - The authors are grateful to BioBrasil (IDEXX Laboratories) for partial supply of the kits. This research received no specific grant from any funding agency of the public sector.

**Conflict of interest** - The authors have no competing interests.

**REFERENCES**


