A NEW SECO-LIMONOID FROM THE INFLORESCENCES OF Dictyoloma vandellianum

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Phytochemical investigation of the inflorescences of *Dictyoloma vandellianum* A. Juss. (Rutaceae) has led to the isolation of a new A-*seco*-limonoid, named dictyolomol (1), along with a known quinolone alkaloid, casimiroine (2), previously isolated from the same species. The structures were established by spectroscopic analysis such as one- and two-dimensional NMR, IR, high-resolution mass spectrometry and chemical transformations. The isolated compounds (1 and 2) and *O*-acetyl derivative (1a) were evaluated for their cytotoxicity against MCF-7 and HeLa cancer cell lines. The results indicated that 1, 1a and 2 showed no significant cytotoxicity.

Keywords: A-seco-limonoid; alkaloid; Dictyoloma vandellianum; Rutaceae; cytotoxicity.

INTRODUCTION

The genus *Dictyoloma* contains two species, *Dictyoloma peruvianum*, which occurs in Peru and Bolivia, and *Dictyoloma vandellianum* A. JUSS., in Brazil.¹ Previous phytochemical investigations of *D. vandellianum* have demonstrated the presence of limonoids, quinolone alkaloids, and pyranochromones.²⁻⁵ Although hundreds of limonoids have been isolated from various plants, their occurrence in the plant kingdom is limited to plants belonging to Rutales, especially Meliaceae and Rutaceae.^{6,7} Therefore, these compounds have been important for systematic classification of the genus in the Rutaceae family.^{3,8,9} Recently, it was evaluated the antinociceptive activity of hydrolysable tannins isolated from the methanol extract from leaves, anti-inflammatory activity and cytotoxic of pyranochromones from dichloromethane extract from root barks of *D. vandellianum*.^{2,10}

Natural products continue to be a major source of new drugs, and they may be used as prototypes for design of lead molecules.^{11,12} From 1940 to 2014, 175 small molecules were approved for cancer treatment, nearly half were natural products or directly derived from them.¹³ In this context, literature reports have been described different classes of secondary metabolites with antitumor activity, such as limonoids^{14,15} and alkaloids.^{16,17} Although, several limonoids have been found to exhibited a range of biological activities like insecticidal, insect antifeedant and growth regulating activity on insects.^{6,18}

Considering the chemical and pharmacological potential of the *D. vandellianum*, an investigation of chloroform extract from inflorescences was performed leading to the isolation of a new A-*seco*limonoid, dictyolomol (1). The structure of this compound and the corresponding acetyl derivative **1a** were elucidated by 1D and 2D NMR, HRESIMS and IR. In addition, a known quinolone alkaloid previously described was identified as casimiroine (**2**), isolated from the same species.⁹ The compounds obtained were screened for their cytotoxic activities.

RESULTS AND DISCUSSION

The inflorescences of *D. vandellianum* were used to obtain a chloroform extract, which was fractionated by silica gel column chromatography and further purified to afford a new limonoid (1) and casimiroine (2) (Figure 1), previously found on this species.⁹



Figure 1. Dictyolomol (1), O-acetyl derivative (1a) and casimiroine (2) isolated from the D. vandellianum

Compound 1 was isolated as a white powder with molecular formula of C₂₆H₂₈O₉, as deduced from the HRESIMS analysis $(m/z 485.1807 [M + H]^+$, calcd. for $C_{26}H_{29}O_9$, m/z 485.1806), implying 13 indices of hydrogen deficiency. The ¹H NMR spectrum of **1** showed resonances for four tertiary methyl groups at $\delta_{\rm H}$ 0.85 (s, H-30), 1.10 (s, H-28), 1.36 (s, H-18), and 1.43 (s, H-29). Characteristic signals of a β -substituted furan moiety were observed at $\delta_{\rm H}$ 6.52 (br d, 1H, J = 1.4 Hz, H-22), 7.68 (t, 1H, J = 1.4 Hz, H-23) and 7.73 (m, 1H, H-21). The presence of two singlets at $\delta_{\rm H}$ 4.36 (s, 1H, H-15) and 5.59 (s, 1H, H-17) are in accordance with the presence of the D-ring with a 14,15-epoxy group, typical for limonoids from D. vandellianum.^{2,3} Furthermore, signals at $\delta_{\rm H}$ 4.17 (dd, 1H, J = 11.9, 2.0 Hz, H-19 β), 4.59 $(d, 1H, J = 11.9 \text{ Hz}, H-19\alpha), 5.99 (d, 1H, J = 10.0 \text{ Hz}, H-2), and 6.93$ (dd, 1H, J = 10.0, 2.0 Hz, H-1) were attributed to A-ring seco limonoid with an α , β -unsaturated lactone. The chemical shifts of the protons at $\delta_{\rm H}$ 4.68 (d, 1H, J = 5.0 Hz, H-7) and 5.63 (d, 1H, J = 5.0 Hz, OH-7) were attributed by comparison with literature data.^{19,20} The existence

Table 1. NMR spectroscopic data of 1 and 1a (J in Hz)

position	1 (DMSO- d_6) ^a		1 (Pyr- d_5) ^b		1a (Pyr- d_5) ^b	
	$\delta_{\scriptscriptstyle m H}$	$\delta_{ m c}$	$\delta_{\scriptscriptstyle \mathrm{H}}$	$\delta_{ m c}$	$\delta_{ ext{ H}}$	$\delta_{ m c}$
1	6.93 dd (10.0, 2.0)	149.1	6.96 d (9.6)	149.0	6.99 br d (9.6)	148.6
2	5.99 d (10.0)	121.2	6.36 d (9.6)	123.1	6.36 d (9.6)	123.5
3		161.7		162.5		162.5
4		61.8		62.7		64.1
5		73.3		74.7		74.8
6		205.4		206.2		199.7
7	4.68 d (5.0)	81.0	5.08 s	83.6	5.99 s	83.2
8		45.5		47.3		45.5
9	3.05 t (9.4)	41.0	3.36 t (8.8)	43.2	3.43 t (8.8)	43.7
10		44.9		46.4		46.3
11	1.80 m	17.1	1.95 m	18.7	2.01 m	19.0
12	1.67 m	25.8	1.83 m	27.3	1.90 m	28.0
12	1.32 m		1.53 m		1.53 m	
13		38.5		39.8		39.9
14		72.1		72.7		71.6
15	4.36 s	55.9	5.24 s	56.8	4.30 s	56.5
16		167.4		168.2		168.2
17	5.59 s	77.5	5.88 s	78.6	5.86 s	78.6
18	1.36 s	18.5	1.51 s	18.7	1.48 s	19.0
19	4.59 d (11.9)	69.4	4.81 d (12.0)	70.6	4.82 br d (12.0)	70.5
19	4.17 dd (11.9, 2.0)		4.72 d (12.0)		4.75 d (12.0)	
20		120.3		121.5		121.3
21	7.73 m	142.0	7.74 s	142.4	7.74 s	142.5
22	6.52 br d (1.4)	110.3	6.54 s	110.9	6.53 s	110.9
23	7.68 t (1.4)	143.2	7.67 s	144.1	7.67 s	144.2
28	1.10 s	22.0	1.10 s	22.0	1.31 s	22.9
29	1.43 s	20.9	1.53 s	21.7	1.54 s	22.1
30	0.85 s	14.4	1.23 s	15.1	1.25 s	15.6
OH-7	5.63 d (5.0)					
Ac-7					2.21 s	20.8
						170.3

^a500 and 125 MHz. ^b400 and 100 MHz.

of hydroxyl proton signal indicates that no chemical exchange with water molecules, which appears as a DMSO- d_6 impurity, has occurred. This can be explained by the occurrence of intramolecular hydrogen bonding involving the hydroxyl and carbonyl group.

In the ¹³C NMR spectrum of compound **1** were observed 26 signals. The chemical shifts at $\delta_{\rm C}$ 110.3, 120.3, 142.0, and 143.2 were assigned to carbons C-22, C-20, C-21, and C-23, respectively, of the β -substituted furan moiety. The presence of the 14,15-epoxy *D*-lactone was confirmed by the signals observed at $\delta_{\rm C}$ 55.9 (C-15), 72.1 (C-14), 77.5 (C-17), and 167.4 (C-16). The signals at $\delta_{\rm C}$ 81.0 and 205.4 were in agreement with a hydroxyl at C-7 and carbonyl group at C-6. The chemical shifts at $\delta_{\rm C}$ 69.4, 121.2, 149.1, and 161.7, were attributed to carbons C-19, C-2, C-1, and C-3, respectively, from the A-ring *seco* limonoid with an α , β -unsaturated lactone. The presence of two non-hydrogenated carbons at $\delta_{\rm C}$ 61.8 and 73.3 revealed the presence of an epoxy group and were assigned to C-4 and C-5. Methyl

groups were observed at $\delta_{\rm C}$ 14.4, 18.5, 20.9, and 22.0, corresponding to C-30, C-18, C-29, and C-28, respectively.

In its HMBC spectrum, H-17 ($\delta_{\rm H}$ 5.59) was correlated to C-18 ($\delta_{\rm C}$ 18.5) and C-14 ($\delta_{\rm C}$ 72.1); H-12 ($\delta_{\rm H}$ 1.72) was correlated to C-14 ($\delta_{\rm C}$ 72.1) and C-17 ($\delta_{\rm C}$ 77.5) thus confirming 1.72/25.8 for H-12/C-12. The insertion of a hydroxyl group at C-7 was established by the correlations from H-7 ($\delta_{\rm H}$ 4.68) to the carbons Me-30 ($\delta_{\rm C}$ 14.4) and C-5 ($\delta_{\rm C}$ 73.3), and from OH-7 ($\delta_{\rm H}$ 5.63) to C-8 ($\delta_{\rm C}$ 45.5) and C-6 ($\delta_{\rm C}$ 205.4). The presence of a 4,5-epoxy group was confirmed by the correlation between H-28 ($\delta_{\rm H}$ 1.10) and the carbons C-4 ($\delta_{\rm C}$ 61.8) and C-5 ($\delta_{\rm C}$ 73.3). H-28 was also correlated to C-29 ($\delta_{\rm C}$ 20.9), and H-29 ($\delta_{\rm H}$ 1.43) to C-28 ($\delta_{\rm C}$ 22.0), corroborating the presence of two methyl groups linked to C-4.

The presence of the lactone ring was verified by the correlations observed between H-19 ($\delta_{\rm H}$ 4.17) and carbons C-3 ($\delta_{\rm c}$ 161.7) and C-5 ($\delta_{\rm c}$ 73.3). Another important correlation was found between H-2

 $(\delta_{\rm H}$ 5.99) and C-10 ($\delta_{\rm C}$ 44.9), consistent with a A-ring *seco* limonoid (Figure 2).



Figure 2. Key HMBC correlations of compound 1

The relative configurations of **1** was established by the analysis of the NOESY spectrum (Figure 3). The correlations of Me-18/H-15 suggested the β -orientation of the 14,15-epoxy group. The correlation of H-15/H-7, along with the correlation of OH-7/Me-30 revealed the 7 β -equatorial position of the hydroxyl group. The correlations of H-7/H-9, H-9/H-1 and H-1/Me-29 led to the assignment of the α -orientation for these protons. In order to determine the C-15 and C-7 configuration, the acetylation of **1** was carried out (Ac₂O/pyr), which gave the *O*-acetyl derivative **1a**. Comparison of the ¹H NMR spectra of **1** and **1a** obtained in pyridine- d_5 (Table 1) showed a shielding of H-15 of **1a**, which suggests that H-15 has a *cis* relationship with H-7.⁹

Thus, after analysis of the spectra, it was possible to establish that compound **1** is a new natural compound with an A-*seco*-limonoid structure, named dictyolomol.



Figure 3. Key NOESY correlations of compound 1

Compounds 1, 1a and 2 were evaluated for their cytotoxicities toward two human cancer cell lines (MCF-7 and HeLa). However, the compounds did not demonstrate cytotoxicity against either of these cell lines.

CONCLUSIONS

The phytochemical study of *D. vandellianum* led to the isolation of a new A-*seco*-limonoid, dictyolomol (1), and the known alkaloid, casimiroine (2), which was already reported from this species. The cytotoxicity evaluation against MCF-7 and HeLa of these compounds and the corresponding acetyl derivative **1a** was investigated. The results were negative for all compounds.

EXPERIMENTAL

General experimental procedures

IR were recorded on a BOMEM-MB 100 spectrophotometer. 1Dand 2D-NMR experiments were performed using a Bruker Avance III HD (400 and 100 MHz for ¹H and ¹³C, respectively) and Varian NMR (500 and 125 MHz for ¹H and ¹³C, respectively) spectrometer. The residual peaks of the deuterated solvents were taken as reference points. High-resolution mass spectrometry (HRMS) analyses were performed on a microTOFII ESI-TOF mass spectrometer. Isolation, purification, and analysis of the chemical constituents were performed using chromatographic methods, including column chromatography (CC), thin-layer chromatography (TLC) and high-performance liquid chromatography (HPLC). A preparative RP C₁₈ column (ACE; 250 mm × 21.2 mm × 5 µm) was used for the separation. Silica gel (Silicycle[®], particle size 0.040-0.063 mm) was used in CC. Commercial silica gel plates (Whatman) were used in TLC, with a 0.25-mm-thick layer on an aluminum support (20 × 20 cm). The substances were analyzed by using ultraviolet radiation at wavelengths of 254 and 366 nm (Boitton brand apparatus) and by impregnating the plates in glass chambers saturated with iodine vapor.

Plant material

The specimens were collected in Morro do Chapéu, Bahia, Brazil (11°13'43"S, 41°09'28"W) in March 2016. Access registration in the National Management System of Genetic Patrimony and Associated Traditional Knowledge (SISGEN) was obtained under No. A1F8819. The vouchers specimens were identified by Prof. Maria Lenise Silva Guedes and deposited at the Herbarium Alexandre Leal Costa (ALCB), Institute of Biology, Federal University of Bahia with the registration number 123484.

Extraction and isolation

Inflorescences of *D. vandellianum* (706.1 g) were macerated three times in chloroform for seven days. After evaporation of the solvent under reduced pressure the crude extract (3.5 g) was fractionated by silica gel (70 g) column chromatography (petroleum ether, 100%, 5 fractions; chloroform, 100%, 17 fractions; CHCl₃:MeOH, 98:2, 4 fractions; CHCl₃:MeOH, 95:5, 4 fractions; MeOH, 100%, 3 fractions). 33 fractions of 200 mL were collected and pooled in 17 groups according to their chromatographic profile on TLC. The fraction group 13-14 (400 mg), obtained after regrouping and drying, was suspended in chloroform and the precipitate separated to obtain the compound **1** (140 mg). The fraction 10 was purified by preparative liquid chromatography with MeCN–H₂O (30:70, v/v) to obtain **2** (3.28 mg).

Dictyolomol (1): White powder; $[\alpha]_{^{25}D}^{25} + 116$ (*c*. 0.1, MeCN); IR (KBr) ν_{max} 3430, 1738, 1700, and 1058 cm⁻¹; ¹H and ¹³C NMR data (500 MHz and 125 MHz, respectively, in DMSO-*d*₆), see Table 1; HRESIMS 485.1807 [M + H]⁺ (calcd. for C₂₆H₂₉O₉, *m/z* 485.1806).

Acetylation of dictyolomol (1)

Acetylation was carried out by dissolving the compound 1 (8 mg) in pyridine and Ac₂O (1:2) at room temperature. After 24 h, the reaction mixture was carefully added into cold water and extracted successively with CHCl₃. The organic layer was washed with aqueous HCl solution (5%, v/v), to eliminate pyridine, and with H₂O. Then, the CHCl₃ phase was evaporated to dryness using a rotatory evaporator to obtain **1a** (4.49 mg).

Cytotoxicity assays

MCF-7 (human breast carcinoma cell line) was cultured in RPMI 1640 medium, and HeLa (human epithelioid cervix carcinoma cell line) was cultured in DMEM medium, both culture medium were supplemented with 10% fetal bovine serum (SBF), 100 U mL⁻¹ penicillin, and 100 μ g mL⁻¹ streptomycin at 37 °C in a humidified atmosphere with 5% CO₂. Cells were seeded into 96-well plates at a density of 3 × 105 cell well⁻¹. Following a 24 h period, cells were

incubated with the compounds (25 μ M) dissolved in DMSO (0.4%). After culturing for 72 h, the supernatant was discarded, and the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) solution (5 mg mL⁻¹) was added and incubated for another 3 h. The deposited formazan was dissolved with sodium dodecyl sulfate (SDS) (100 μ L).²¹ Positive control was DMSO (20%). The optical densities were measured using a microplate reader (Synergy HT, BioTek[®]). Data were analyzed with GraphPad Prism 5.0.

SUPPLEMENTARY MATERIAL

NMR spectra for compounds **1** and **1a** can be found at http://quimicanova.sbq.org.br in PDF format, with free access.

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