Chicken Serologic Response to *Salmonella enterica* Serotype Typhimurium assessed by Elisa

**ABSTRACT**

This study evaluated two enzyme-linked immunosorbent assays (ELISA) in the detection of chicken serologic response against *Salmonella enterica* serotype Typhimurium. The assays have used as detecting antigen the soluble bacterial proteins of a non-flagellated strain of *Salmonella* Typhimurium (AgTM), and antibody conjugated to peroxidase or alkaline phosphatase. According to the results, optimal dilutions of antigen (concentration 5.49 mg/mL) and serum samples in both assays were 1:20,000 and 1:1,000, respectively. In such conditions, the ELISA/AgTM was able to detect serological response to *Salmonella* Typhimurium. Cross-reactions to *Salmonella* serotypes Gallinarum and Pullorum were seen, but not with other serotypes such as Enteritidis.

**INTRODUCTION**

Breeding flocks must be free of *Salmonella* and early detection is extremely important to prevent the disease and its dissemination (Barrow, 1994). Bacteriological methods are required and have been the traditional means of obtaining such prevention. However, serological methods may be used to detect infection caused by a number of serotypes (Barrow, 1992). After the first enzyme linked immunosorbent assay (ELISA) to detect *Salmonella enterica* subspecies arizonae infection has been described in turkeys (Nagaraja et al., 1984, Nagaraja et al., 1986), other reports have indicated the usefulness of different indirect ELISAs to detect circulating antibodies (Barrow, 1991). Since the development of ELISA in the 1980s, they have been increasingly used in poultry flocks in the serological detection of invasive *Salmonella* serotypes associated with human food poisoning, such as *S*. Typhimurium and *S*. Enteritidis (Barrow et al., 1989; Hassan et al., 1990).

Oral infection with invasive serotypes often leads to the production of circulating antibodies, mainly IgG antibodies (Barrow, 1992). The advantages of serology are that *Salmonella* induce the production of high circulating IgG and the titers persist for up to 45 weeks after infection (Hassan et al., 1990; Barrow, 1995). Any degree of cross-reaction between LPS from groups B and D has been demonstrated with sera from chickens infected with *S*. Typhimurium (Barrow, 1992; Barrow et al., 1992). Antigens of variable specificity have been largely used for preliminary screening, including soluble protein antigen produced by sonication of whole cells (Hassan et al., 1990). Few studies have been reported on the use of this antigen in regard to *S*. Typhimurium infections. Barrow et al. (1989) examined sera from seven flocks, four of which had bacteriological evidence of *S*. Typhimurium infection. High antibody titers were detected in three flocks by the soluble protein antigen from sonicated whole cells, but in none of the three flocks considered to be non-infected.

**Keywords**

Chicken, ELISA, enzyme, *Salmonella* sp, serology.

**Acknowledgements**

Mr. Antônio José dos Santos, Mrs. Aparecida Rodrigues Baptista and Mrs. Maria de Lourdes Feres Tamanini for laboratorial support and FAPESP and CNPq for financial support.

Financial support from: FAPESP and CNPq.
Oliveira GH, Berchieri Jr A, Montassier HJ

Chicken Serologic Response to *Salmonella enterica* Serotype Typhimurium assessed by Elisa

The Brazilian poultry industry is based on imported grandparent flocks and the National Health Program establishes that the flocks must be free of *S. Gallinarum* and *S. Pullorum*, whereas *S. Enteritidis* and *S. Typhimurium* must be controlled.

The present research was undertaken to assess an ELISA as a tool to search chicken serological response to *S. Typhimurium*. The ELISA was established using soluble proteins as antigen and antibodies conjugated to either alkaline phosphatase or peroxidase.

**MATERIAL AND METHODS**

**Reference serum samples**

Adult laying hens (18 weeks of age) with no signs of clinical salmonellosis and showing negative results in bacteriologically inspection and slide agglutination test were given 0.5mL of inactivated cultures of *Salmonella* (10⁶CFU/mL) intramuscularly. Serological response was individual for each of the following *Salmonella* serotypes: Enteritidis, Typhimurium, Gallinarum, Pullorum, Infectis, Montevideo, Binza, Livingstone, Anatum, Stanley, Eimsbuettel, Ealing and Virchow. Cultures were prepared in 10mL of Bacto-Brain Heart Infusion Agar and were inactivated with 0.2mL of 40% formaldehyde. Birds were inoculated five times with each *Salmonella* serotype at week intervals. Blood was collected three weeks after the last inoculation and serum samples were stored at -20°C. Serum samples from SPF chickens negative to *Salmonella* were kindly provided by Dr. Paul A. Barrow (Institute for Animal Health, UK).

**ELISA Procedure**

**1- Peroxidase**

The procedure was carried out as described by Hassan *et al.* (1990) and Barrow *et al.* (1992) with some modifications. The soluble protein detecting antigen was prepared by sonication (Branson Sonifer 250 – USA), using 8 cycles of - 85watts with 30 s-intervals. Washed whole cells of a non-flagellated *Salmonella* from SPF chickens negative to *Salmonella* were used. The ELISA was established using soluble proteins as antigen and antibodies conjugated to peroxidase. The ELISA can be done using different antigens. Both blocking buffer was prepared with 10% skimmed milk (Molico, Nestlé, São Paulo, Brazil) in carbonate-bicarbonate buffer. Blocking was performed for 45minutes at 37°C. Serum samples were diluted in PBST with 10% skimmed milk and added to the plates. It was then added rabbit anti-chicken IgG peroxidase conjugate (Sigma A-9046) diluted to 1:2,000 in PBST. Finally, ortho-phenylenediamine (10mg/mL) (OPD - Sigma P-8287) and 100µL of hydrogen peroxide were diluted in 25mL of citrate phosphate buffer pH 4.9-5.2 (0.1M C₆H₈O₇, 0.2M NaHPO). The color reaction was stopped using 2N HCl after 15 min at room temperature. Absorbance was read at 490nm using an ELISA plate reader (Microplate Reader 550, Bio Rad, USA). Each serum sample was tested in duplicate and the mean was calculated for further analysis.

**2- Alkaline Phosphatase**

The steps in alkaline phosphatase ELISA were similar to those described above for peroxidase ELISA, except for the conjugate and substrate. In this case, it was used a rabbit anti-chicken IgG alkaline phosphatase conjugate (Sigma A-9171) diluted to 1:1,000 in PBST and p-nitrophenyl phosphate (5mg/mL) (pNPP - Sigma N-9389) was used as substrate, diluted in 5mL of diethanolamine buffer pH 9.8 (100mM diethanolamine, 500nM MgCl₂). Besides, the color reaction was developed for 30 min at room temperature and then it was stopped with 3M NaOH. Absorbance was read at 405nm.

**RESULTS AND DISCUSSION**

*Salmonella* Typhimurium was the most common serotype involved in animal and human foodborne salmonellosis before the outbreaks of *Salmonella* Enteritidis (Barrow, 1992) and it is still frequently isolated (Almeida *et al.*, 2000; Zancan *et al.*, 2000). Therefore, the first studies to prevent avian salmonellosis based on monitoring programs were focused on searching for serological response to *Salmonella* Typhimurium. Although fecal excretion of enteric *Salmonella* is intermittent, serologic response lasts for several months (Hassan *et al.*, 1990) and a serologic test might be useful to demonstrate whether a poultry flock has been or is contaminated by *Salmonella* Typhimurium. The enzyme-linked immunosorbent assay (ELISA) has been used routinely in the poultry industry for monitoring programs of various diseases. Thus, it is not difficult to introduce a new ELISA test for *Salmonella* monitoring. The ELISA can be done using different antigens. Both
LPS and flagellum antigens produce strong reactions. ELISA using LPS as antigen detects IgG produced against LPS in birds infected by *Salmonella* that belong to serogroups B and D (Chart et al., 1990), due to the common antigen O12. Serologic response to flagellum antigen is less persistent and might be detected up to four months (Hassan et al., 1990; Baay & Huis in't Veld, 1993). The ELISA using soluble proteins of *Salmonella Typhimurium* as antigen was similar to the assay prepared with LPS as antigen. The results were very comparable; however, soluble protein antigen preparation is less laborious (Hassan et al., 1990; Barrow et al., 1992).

According to the present results (Table 1), peroxidase and alkaline phosphatase assays should be performed using the antigen diluted to 1:20,000 (5.49 mg/mL) for both conjugates and the serum sample diluted to 1:1,000. Positivity was considered when readings were higher than, or equal to, 1.0 (OD ≥ 1.00). In this condition the test is suitable to detect serologic response to *Salmonella Typhimurium*. Cross-reaction was observed between *Salmonella* and the serotypes Gallinarum and Pullorum, but not with *S. Enteritidis* whichever conjugate was used. Barrow (1991) and Barrow (1992) reported no cross-reaction with other enterobacteria (*Escherichia coli*, *Klebsiella*, *Proteus* and *Citrobacter*), corroborating our results. Cross-reactions with *Salmonella* serotypes from groups B and D should be of interest, since *Salmonella* of these groups have been isolated from poultry and are of concern to public health (Zancan et al., 2000, Gama et al., 2003; Ribeiro et al., 2003). Therefore, ELISA/AgTM is suitable to indicate evidences of avian salmonellosis and may be used as a screening test for further bacteriological examination.

###REFERENCES


###Table 1 - Enzyme-linked immunosorbent assays (ELISA/AgTM)1. Determination of single dilution of chicken serum to differentiate serological response to *Salmonella Typhimurium* from other *Salmonella* serotypes.

<table>
<thead>
<tr>
<th>Positive sera to <em>Salmonella</em> serotypes</th>
<th>Peroxidase conjugate</th>
<th>Reciprocal dilution of sample of sera</th>
<th>Alkaline phosphatase conjugate</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gallinarum (D)</td>
<td>1.136</td>
<td>1.148</td>
<td>Gallinarum (D)</td>
</tr>
<tr>
<td>Ealing (O)</td>
<td>0.092</td>
<td>0.119</td>
<td>Ealing (O)</td>
</tr>
<tr>
<td>Livingstone (C)</td>
<td>0.141</td>
<td>0.144</td>
<td>Livingstone (C)</td>
</tr>
<tr>
<td>Anatum (E)</td>
<td>0.354</td>
<td>0.229</td>
<td>Anatum (E)</td>
</tr>
<tr>
<td>Stanley (B)</td>
<td>0.350</td>
<td>0.328</td>
<td>Stanley (B)</td>
</tr>
<tr>
<td>Infantis (C)</td>
<td>0.263</td>
<td>0.246</td>
<td>Infantis (C)</td>
</tr>
<tr>
<td>Embothrium (C)</td>
<td>0.165</td>
<td>0.191</td>
<td>Embothrium (C)</td>
</tr>
<tr>
<td>Virchow (C)</td>
<td>0.189</td>
<td>0.129</td>
<td>Virchow (C)</td>
</tr>
<tr>
<td>Binza (E)</td>
<td>0.401</td>
<td>0.257</td>
<td>Binza (E)</td>
</tr>
<tr>
<td>Typhimurium (B)</td>
<td>1.231</td>
<td>1.298</td>
<td>Typhimurium (B)</td>
</tr>
<tr>
<td>Enteritidis (D)</td>
<td>0.627</td>
<td>0.285</td>
<td>Enteritidis (D)</td>
</tr>
<tr>
<td>Pullorum (D)</td>
<td>1.197</td>
<td>1.927</td>
<td>Pullorum (D)</td>
</tr>
<tr>
<td>Montevideo (C)</td>
<td>0.110</td>
<td>0.154</td>
<td>Montevideo (C)</td>
</tr>
<tr>
<td>Negative serum</td>
<td>0.038</td>
<td>0.056</td>
<td>Negative serum</td>
</tr>
</tbody>
</table>

1- AgTM diluted to 1:20,000; *Positive samples: OD ≥ 1.0.
Hassan JO, Barrow PA, Mockett APA, McLeod S. Antibody response to experimental *Salmonella* typhimurium infection in chickens measured by ELISA. The Veterinary Record 1990; 126:519-522.


