N₂O EMISSIONS FROM A CULTIVATED MOLLISOL: OPTIMAL TIME OF DAY FOR SAMPLING AND THE ROLE OF SOIL TEMPERATURE\(^{(1)}\)

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SUMMARY

The correct use of closed field chambers to determine N₂O emissions requires defining the time of day that best represents the daily mean N₂O flux. A short-term field experiment was carried out on a Mollisol soil, on which annual crops were grown under no-till management in the Pampa Ondulada of Argentina. The N₂O emission rates were measured every 3 h for three consecutive days. Fluxes ranged from 62.58 to 145.99 μg N-N₂O m⁻² h⁻¹ (average of five field chambers) and were negatively related (\(R^2 = 0.34, p < 0.01\)) to topsoil temperature (14 - 20 °C). N₂O emission rates measured between 9:00 and 12:00 am presented a high relationship to daily mean N₂O flux (\(R^2 = 0.87, p < 0.01\)), showing that, in the study region, sampling in the mornings is preferable for GHG.

Index terms: soil N₂O fluxes, climate change, GHG sampling.

RESUMO: EMISSÕES DE N₂O DE UM CHERNOSSOLO CULTIVADO: O TEMPO IDEAL DO DIA PARA AMOSTRAGEM E PAPEL DA TEMPERATURA DO SOLO

O uso adequado de câmaras estáticas para determinar as emissões de N₂O no campo requer a definição da hora do dia que melhor representa a taxa de emissão média diária. Um experimento de campo de curta duração foi realizado em um Chernossolo do Pampa Ondulado.

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N2O EMISSIONS FROM A CULTIVATED MOLLISOL: OPTIMAL TIME OF DAY FOR SAMPLING... 1815

INTRODUCTION

Nitrous oxide (N2O) is the main greenhouse gas emitted by agricultural soils, with a global warming potential 310 times greater than carbon dioxide (Pérez-Ramírez, 2007). Its concentration in the atmosphere increased at a rate of 0.25 % per year between 1980 and 1998 (Houghton et al., 2001) and, according to Snyder et al. (2007), its concentration reached 319 ppb in 2005.

Closed chambers are widely used for measuring soil greenhouse gas fluxes. The most common of these procedures involves manual sampling of headspace gas from the chamber using syringes (Chen et al., 2000; Ball et al., 2002; Jantalia et al., 2008) or more advanced systems such as vacuum pumps or automated flux monitoring systems (Akiyama et al., 2000; Dobbie & Smith, 2001).

The method used to measure field N2O emissions is relatively recent, and it has no universal standard or internationally accepted guidelines (Hutchinson & Livingston, 2002). The current global estimate of agricultural N2O emissions has an uncertainty ranging from 60 to 170 % of the mean estimate (Ventera et al., 2009). Rochette & Eriksen-Hamel (2008) concluded that errors during field sampling a key concern to improve confidence levels of N2O measurements (Schindlbacher et al., 2003) requires a high number of chamber replicates to evaluate N2O fluxes at a reasonable precision, but compromises have to be made in order to limit the number of samples to manageable quantities. Soil N2O daily emission calculations are usually based on the extrapolation of a single daily measurement during a short period to represent the mean flux of a 24 h period. If the time of day that best represents the mean daily N2O flux is known, more accurate results can be expected.

In order to identify this time period, a short-term field experiment was conducted throughout three days with cultivated soils of the Argentine Pampas under no-till management. As soil temperature is a recognized key factor determining daily rates of N2O emissions (Denmead et al., 1979; Blackmer et al., 1982; Akiyama et al., 2000; di Marco et al., 2004; Jantalia et al., 2008) this variable was also considered in our experiment.

MATERIALS AND METHODS

The study was carried out on an agricultural field in the Province of Buenos Aires [34° 57' 29'' S, 60° 13' 11''], on a loamy Typic Argiudol (clay 190 g kg-1, silt 400 g kg-1) from the O'Higgins Series (INTA, 2010). Soil organic matter was determined by wet combustion (Walkley & Black, 1934) was 35.2 g kg-1 and pH in water of the A horizon was 5.7. The field had been under no-till management for the previous 15 years. The crop sequence in the three years before our measurements was maize, wheat/soybean (double cropping) and full-season soybean. Samples were taken in autumn 2010 before the harvest of the full-season soybean.

After a rainfall of 30 mm, five closed chambers (surface = 1.333 cm2, height = 12.5 cm) randomly spread across the soybean field, were inserted into the soil to a depth of 0.05 m. This size of field chambers was described as optimal by other authors (Rochette & Eriksen-Hamel, 2008) and was already used by Jantalia et al. (2008) and Alves et al. (2012). After closing the chambers, gas samples were taken after 0, 20 and 40 min, with a vacuum pump, and injected into previously evacuated 25 mL vials with a rubber stopper fixed to the vial with an aluminum flange. Once sampling was finished, the top of the chamber was removed. The process was repeated every 3 h for three consecutive days beginning at 9:00 am on the first day. The N2O collected was always measured within seven days after sampling with a GC 6890 Agilent Technologies Network gas chromatograph. The daily average soil N2O flux was calculated as the mean of the eight fluxes measured each day. Samples of the five chambers collected during three days provided 15 estimates of N2O fluxes per sampling time.

N2O flux (f) was calculated as:

\[
f = \frac{\Delta C}{\Delta t} \times \frac{V}{A} \times \frac{m}{V_m}
\]

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where $\Delta C/\Delta t$ is the change of $N_2O$ concentration in the chamber during the incubation time $\Delta t$, $V$ is the volume of the chamber (16.7 dm$^3$), $A$ is the area of soil covered by the chamber (1.333 cm$^2$), $V_m$ is the molar volume of $N_2O$ and $m$ is its molecular weight. Gas fluxes were calculated as the increase in concentration during the incubation period and expressed by the arithmetic mean and its standard deviation.

Soil temperature was measured with a thermometer inserted in the soil to a depth of 0.10 m and air temperature with a shaded thermometer at the soil surface. The concentration of $N$-$NO_3^-$ (0-0.20 m) was determined by colorimetric spectrophotometry (Markus et al., 1985). Soil bulk density (BD) was determined in 100-cm$^3$ cylinders (diameter 0.05 m) and gravimetric water content (GWC) by oven-drying at 105 $^\circ$C during 48 h. The volumetric water content (VWC) and total porosity (TP) were calculated using BD and GWC, assuming a particle density (Dp) of 2.65 Mg m$^{-3}$ (equations 2 and 3):

$$TP = 1 - \frac{BD}{Dp} \quad (2)$$  
$$VWC = GWC \times BD \quad (3)$$

The percentage of water-filled pore space (%WFPS) was calculated with the TP and VWC values (Equation 4):

$$WFPS = TP - VWC \quad (4)$$

In order to determine the time of day that best represents the mean daily $N_2O$ emission rate, regression analysis (InfoStat, 2002) was used to relate $N_2O$ data with soil and air temperature.

### RESULTS

Mean air temperature was 14.5 $^\circ$C, ranging from a minimum of 4.0 $^\circ$C to a maximum of 31.3 $^\circ$C, whereas the mean soil temperature was 16.8 $^\circ$C, with a minimum of 14.6 $^\circ$C and a maximum of 19.2 $^\circ$C. Percentage of water filled pore space (%WFPS) and $NO_3^-$ content were kept constant during the three days of measurements, averaging 0.89 m$^3$ m$^{-3}$ (89 %) and 73.4 mg kg$^{-1}$, respectively. The daily dynamics of soil and air temperature were similar in the three days of the study.

The soil temperature was slightly lower than 15 $^\circ$C on one occasion only. This temperature was the minimum value above which $N_2O$ fluxes were observed (Keeney et al., 1979). However, this low temperature did not limit $N_2O$ flux under the study conditions. The times of day when the mean daily $N_2O$ emissions were represented best were at 9:00 and 12:00 am. The coefficients of determination obtained by the regression analysis between mean daily and hourly $N_2O$ emission rates were highly significant in both cases (Table 1 and Figure 1). Other authors also found morning hours more suitable for gas sampling (Blackmer et al., 1982; Jantalia et al., 2008).

### Table 1. Relation between mean daily and mean hourly $N_2O$ emission rates

<table>
<thead>
<tr>
<th>Sampling time</th>
<th>a</th>
<th>b</th>
<th>$R^2$</th>
</tr>
</thead>
<tbody>
<tr>
<td>9:00</td>
<td>1.12***</td>
<td>-13.78</td>
<td>0.87***</td>
</tr>
<tr>
<td>12:00</td>
<td>1.13***</td>
<td>-23.73*</td>
<td>0.88***</td>
</tr>
<tr>
<td>15:00</td>
<td>0.48**</td>
<td>38.57**</td>
<td>0.35**</td>
</tr>
<tr>
<td>18:00</td>
<td>0.43***</td>
<td>36.50**</td>
<td>0.33**</td>
</tr>
<tr>
<td>21:00</td>
<td>0.82**</td>
<td>6.30</td>
<td>0.37**</td>
</tr>
<tr>
<td>24:00</td>
<td>1.31***</td>
<td>-28.24</td>
<td>0.75***</td>
</tr>
<tr>
<td>3:00</td>
<td>1.84***</td>
<td>-42.22</td>
<td>0.75***</td>
</tr>
<tr>
<td>6:00</td>
<td>0.86*</td>
<td>26.61</td>
<td>0.23*</td>
</tr>
</tbody>
</table>

$y = ax + b$. *, **, *** indicate significant adjustment of regression slopes and intercepts at $p = 0.1$, 0.05 and 0.01, respectively.

Interestingly, it was found that the dynamics of other soil N emissions (e.g. NO) were determined mainly by the season (Xunhua et al., 2003). Other daily patterns of $N_2O$ flux were reported elsewhere (Denmead et al., 1979; Blackmer et al., 1982; Akiyama et al., 2000; di Marco et al., 2004). Most of these studies reported that $N_2O$ emission rates were positively related to soil temperature (Denmead et al., 1979; Alves et al., 2012). The results obtained in this paper differ in this aspect.

$N_2O$ emission rates were poorly related to air temperature (data not shown) and were negatively related to soil temperature ($R^2 = 0.34$). $N_2O$ emission tended to increase as topsoil temperature decreased (Figure 2 and 3). We only found two cases that agreed with our results: Bailey (1976) observed an increase in $N_2O$ production when topsoil temperature
decreased from 30 to 10 °C, while Avrahami & Bohannan (2009) reported similar results in soils with low or moderate amounts of fertilizer. \(\text{N}_2\text{O}\) emissions measured on these Mollisols can be ascribed to denitrification rather than nitrification, as WFPS was always greater than 70% (Bateman & Baggs, 2005). However, the fact that \(\text{N}_2\text{O}\) emission rates decrease above 24 °C is not easy to explain. It can be speculated that during denitrification at higher temperatures, the rate of \(\text{N}_2\) production is higher than that of \(\text{N}_2\text{O}\) (Maag & Vinther, 1996). Soil anaerobiosis could be enhanced by the occurrence of horizontal pores in the topsoil of the non-tilled soils studied.

The flux of trace gases between the soil and the atmosphere is the result of three basic processes: production, consumption and transport (Conrad, 1996). Another possible factor influencing soil \(\text{N}_2\text{O}\) emissions is the continuous no-till management of these soils. Many of them have a low volume of structural pores (Taboada et al., 1998; Micucci & Taboada, 2006; Taboada et al., 2008). The formation of these pores has been attributed to the lack of mechanical disturbance as well as to the traffic of heavy farm machinery in no-till systems, resulting in shallow compaction in the form of laminar structures (Bonel et al., 2005; Sasal et al., 2006; Alvarez et al., 2009). This type of structure favors the appearance of horizontal pores, which also tend to be more tortuous and less connected (Bonel et al., 2005), thus promoting the development of anaerobiosis after heavy rain episodes.

Taking into account that diffusion capacity is related to impedance to gas diffusion, which depends on the shape and orientation of the soil pores (Fen et al., 2009), it can be also suggested that this pore arrangement, plus a WFPS percentage close to 90%, could reduce the rate at which gases move from the soil to the atmosphere (Glinski & Stepniewski, 1985). This is consistent with results of Bartelt-Hunt & Smith (2002), who found that with a decrease in soil moisture content, and therefore an increase in the percentage of air-filled pores (30 to 39%), the gas diffusion coefficient rises from 0.0133 to 0.0609 cm² s⁻¹. This decrease in gas transport rate from soil to air leads to an accumulation of \(\text{N}_2\text{O}\) molecules in the ground making them available for use by nitrous oxide reductase to produce \(\text{N}_2\), causing a decrease in \(\text{N}_2\text{O}\) rate with increasing temperature in the soil studied here.

A similar observation was described by Focht (1974), who found that as the denitrification rate increased, the quantity of \(\text{N}_2\text{O}\) evolved decreased, due to the greater increase in \(\text{N}_2\text{O}\) reduction rate than in its formation rate. Their results also support and further develop the hypothesis proposed by Bailey & Beauchamp (1973) that account for the change in relative proportion of denitrification gases produced as temperature changes. These authors reported that at 15 °C, a partial inhibition of the final reduction step of denitrification occurred (\(\text{N}_2\text{O} \rightarrow \text{N}_2\)) (Bailey, 1976).

Results indicate that to sample \(\text{N}_2\text{O}\) gases from a Mollisol, the morning hours (09:00 to 12:00 am) are more appropriate to minimize errors. Soil temperature appears to be the main factor negatively regulating the \(\text{N}_2\text{O}\) emission rate. This differs from results obtained by other authors (Denmead et al., 1979; Alves, 2011) and should be verified in the future.

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LITERATURE CITED


