The nominotypical genus *Geniates* Kirby, 1819 is one of the 13 genera of the Neotropical tribe Geniatini. *Geniates* is recorded from northern Panama to central Chile and southern Argentina and comprises 39 species (Jameson & Hawkins 2005). The type species *Geniates barbatus* Kirby, 1819 has economical importance for cultures of *Eucalyptus robusta* Sm., *E. tereticornis* Sm. (Myrtaceae) and *Cammellia sinensis* Kuntze (Ternstroemiaceae) (tea, “chá”) in Brazil. The adult has been observed defoliating those trees and also a number of other plants: *Anacardium occidentale* L. (Anacardiaceae) (mango tree, “mangueira”), *Psidium guajava* L. (Myrtaceae) (guava tree, “goiabeira”) and *Eugenia pitanga* (O. Berg) Nied. (Myrtaceae) (Surinam cherry, “pitangueira”), *Cocos nucifera* L. (Arecaceae) (coco palm, “coqueiro”), *Mangifera indica* L. (Anacardiaceae) (mango tree, “mangueira”), *Rosa spp.* (Roscaceae), *Psidium guajava* L. (Myrtaceae) (guava tree, “goiabeira”) and *Terminalia* sp. (Combretaceae) (Anjos & Majer 2003; Costa Lima 1953; Silva et al. 1968).

Despite the economic importance of this insect group, the systematics of Geniatiini and the morphology of its immature stages are poorly known (Jameson & Hawkins 2005; Pardo-Locarno 1996). Although Ohaus (1900) recorded female pupae of *Geniates barbatus* and *G. castaneus* Burmeister, 1844, about 50 cm deep in the soil, their immature stages remain undescribed. In fact, the immature of only one species of Geniatiini is known: *Leucothyreus femoratus* Burmeister, 1844, described by Pardo-Locarno et al. (2006). Therefore the description of the larva and pupa of *Geniates* presented here represents an important addition to the knowledge of the immatures of Sacarabaeidae and to the taxonomy of Rutelinae.

**MATERIAL AND METHODS**

Ten larvae of *Geniates barbatus* were collected about 10 cm deep in the soil covered by grass (Poaceae) in two different localities of São Paulo state, Brazil. Seven larvae were collected in Granja Viana, a locality of the Cotia municipality; other three larvae were collected in Estação Biológica de Boracéia, a forest reserve in Salesópolis municipality. Larvae were reared in the laboratory of the Instituto Biológico de São Paulo (IBSP) and maintained in covered plastic pots with rich humus soil until the adult emergence.

Five larvae and two pupae were killed in boiling water and fixed in ethanol 70%. The specimens are deposited in the collection of the Museu de Zoologia da Universidade de São Paulo, state of São Paulo, Brazil (MZSP).

The specimens were examined using a stereomicroscope Carl Zeiss Stemi SV6 and microscope Carl Zeiss Axiostkop. Illustrations are produced via camera lucida attached in both equipments. Measurements were obtained by ocular micrometer.

The shape of all spiracle plates and bulla has been found to be important for a more precise identification. For this reason, the bulla ratio (dorsoventral diameter of bulla/distance of the lobes of the respiratory plate) was measured to all spiracles.


**RESULTS**

*Geniates barbatus* Kirby, 1819

**Third instar larva** (Figs. 1–23)

Description. Body (Fig. 1) length: 11.9–12.2 mm. Head (Figs. 8–11, 15) width: 4.9–5.0 mm. Epicranial and epistomal sutures present. Epicranium surface slightly rugose; two lateromedial dark spots present; each side with 5 dorsoepicranial setae (*des*), one lateroepicranial seta (*les*), 2 anteroepicranial setae (*aes*), 2 externeopical setae (*ees*), 7 ventroepicranial setae (*ves*), 3 posterofrontal setae (*pfs*), and one externofrontal seta (*efs*). Stemmata present. Clypeus and labrum rugopunctate; each side with one anteroclipeal seta (*acs*), 2 lateroclypeal setae (*lcx*), one posteralralb seta (*pls*), 4 laterobrachial setae (*lts*), one mediolabral seta (*mls*), and 2 anterolabral setae (*als*). Epipharynx (Fig. 14).

**KEYWORDS.** *Aspidolea*, Brazil; *Cyclocephala*; Insecta; Scarabaeoidea; white grubs.
Description of the third larval instar and pupa of *Geniates barbatus* Kirby

Plegmatium with 16 plegmata, each plegma with one external seta of the acanthoparia; proplegmata absent. Epizygum present and clithra absent. Corypha small, with 16–20 setae. Haptomerum prominent; zygum beak-like with apex 5-toothed; heli absent. Chaetoparia with sparse sensilla, right side with 50 internal stout setae and 54 external thin setae; left side with 9–11 anterior stout setae, 66 posteroexternal thin setae; phobae absent. Pedium smooth. Laeotorma short; pterotorma rounded and fused to laeotorma; epitorma narrow, depressed; dexiotorma narrow and internally projected. Haptolachus with 4 right setae and 13 left seate; crepis present; nesium internum (sensorial cone) prominent; nesium externum (sclerotized plate) sharply acuminated. Right mandible (Figs. 18–20) with 2 anterodorsal setae, 4 dorsal anteromolar setae, and 5–7 ventral brush-like setae; scissorial area with a simple tooth and a posterior notch; molar area with 3 transversally ridged lobes, brustia with 6 setae; ventral side with enlogate-oval stridulatory area comprised of...
approximately 43 progressively less spaced costulae; dorsal side with roughly rugose area; ventral process prominent. Left mandible (Figs. 21–23) with similar chaetotaxy to right one, but the dorsal anteromolar with a row of 11 setae; scissorial area similar to right one; molar area with 5 transversally ridged lobes, brustia with 14 setae; acia prominent, truncate and with 10 apical setae; ventral stridulatory area similar to the right one and comprised of approximately 38 costulae; rugosity dorsal area and ventral process similar to the right mandible. Maxillae (Figs. 15–17) symmetrical; cardo setose and 3-lobed; stipes setose, with the stridulatory area formed by a row of 10 teeth and an anterior truncated process; mala setose, anterior and internal setae stout, dorsal side with longitudinal depression between galea and lacinia, galea with a well developed uncus and lacinia with 3 unci; palpus 4-segmented, segment III with one external seta.
Description of the third larval instar and pupa of *Geniates barbatus* Kirby

Revista Brasileira de Entomologia 57(1): 40–46, March 2013

Figs. 15–23. *Geniates barbatus* Kirby; head, 15, venter; 16, dorsum of maxillae and labium; 17, apical detail of mala; 18–20 left mandible (venter, internal face, dorsum); 21–23 right mandible (dorsum, internal face, venter). tb, tentorial bridge.

Labbium (Figs. 15–16): submentum with a group of setae on the anterior angles and two stout setae on medial area; each mentum and prementum with 2 long medial setae and sparse short setae; palpi 2-segmented, apex of segment I with minute setae; ligula with dense external thin setae and stout internal setae flanking a small smooth central area with a sensilla. Hypopharynx (Fig. 16) with an asymmetrical sclerite with a right prominent tooth, left side with longitudinal row of 24–25 setae, and right side with a row of 6 setae. Antennae 4-segmented: I-II with sparse setae; III with one dorsal seta, 2 ventral setae, and a ventral process that have a dorsal spot; IV with one dorsal large spot, and 2 ventral small spots.

Thorax. Prothorax with a lateral sclerite and dorsally simple lobed; meso- and metathorax dorsally 3-lobed, thoracic dorsal lobes with transversal row of sparse long setae mixed with abundant small setae. Spiracle (Figs. 2–3) with C shaped respiratory...
plate, major diameter of bulla approximately three times longer than distance of the lobes of the respiratory plate. Legs (Figs. 4–6) densely setose; claws with 2 lateral setae on each side.

Abdomen with 10 segments; I-VII each forming 3 dorsal lobes, pubescence similar to the meso- and metathorax, I-VI also with small tooth-like setae; IX with sparse setae; X with sparse setae except on the posterior half of dorsum. Spiracles (Fig. 2) similar those of thorax, first segment with the bulla ratio equal 1.7, in segments II-VI ratio about 4.3, in segments VII-VIII bigger than the precedents, and with ratio about 17.5. Raster (Fig. 7) with septula poorly defined; palidia formed by 2 longitudinal irregular rows of 12–16 small tooth-like setae, and extended on ventral anal lobe. Anal opening transversal and weakly curved.

Material examined. BRAZIL, state of São Paulo, Cotia, Granja Viana, 09.viii.2009, G. Francfort leg, 4 larvae, 1 third instar exuvia (MZSP), Salesópolis, Estação Biológica de Boracéia, 27.iii.2012, D. C. Bená & J. Fuhrmann leg, 1 larva (MZSP).

Pupa (Figs. 24–28)

Description. Length: 19 mm, maximal width: 0.9 mm. Body (Figs. 24–26) oblong and light yellow with spiracles and dioneiform organs reddish brown and sclerotized. Integument macroscopically smooth and glabrous but covered by a thin and short microscopic pubescence, which gives a velvety appearance to the surface (50x of magnification); meso- and metathorax with medial area with slightly longer pubescence; apex of urotergite 9 densely pubescent.

Head (Fig. 27). Vertex visible from dorsal view. Frons smooth. Epistomal suture complete. Canthus large and weakly distinct. Eyes compressed between the anterior angles of pronotum and antennae. Antennae almost rectangular on lateral view. Clypeus transverse with curved lateral borders. Mandibles, maxillae, and palpi tubercle-like. Labium slightly convex in female and semiferiscal in males.

Thorax. Pronotum trapezoid with rounded lateral borders. Prosternal posterior process small and visible behind procoxae. Thoracic spiracle in cavity formed between the anterior and medial legs, hypomeron, and elytral thecae basis. Mesonotum shorter than pro- or metanotum; scutellar area slightly backwardly projected. Disc of metaventrite longer than wide, and without any evident anterior or posterior process. Pterotheca I supposited to pterotheca II, and anterior margin suposited by medial leg. Pterotheca II covering proximal half of posterior leg. Pro-, meso- and metacoxae contiguous medially; spurs of meso- and metatibiae internal and tubercle-like; metamfur-tibial junction slightly visible for above; male protarsus larger than female tarsus.

Abdomen. Spiracles I-IV oval and with sclerotized ring; I concealed by pterotheca II; spiracle V-VIII represented by cuticular evagination. Segment I only represented by the tergite. Dioneiform organs present in the mid-base and the mid- apex between segments I-II, II-III, III-IV, IV-V, V-VI. Tergite IX triangular and with large latero-posterior lobes; apex of lobes slightly prominent; urogonophi absent. Sternite II visible medially, female ventrite VIII with medial genital pore, male ventrite VIII with genital ampulla (Fig. 28).

Material examined. BRAZIL, state of São Paulo, Cotia, Granja Viana, 09.viii.2009, G. Francfort leg, 1 female pupa, 1 female pupal exuvia (MZSP), Salesópolis, Estação Biológica de Boracéia, 27.iii.2012, D. C. Bená & J. Fuhrmann leg, 1 male pupa (MZSP).

Biological notes. Larvae of G. barbatus collected in Cotia were associated with larvae of Cyclocephala signaticollis Burmeister, 1847 (Dynastinae, Cyclocephalini) (described by Morelli 1991), and those collected in Estação Biológica de Boracéia were associated with larvae of Aspidolea pelioptera (Burmeister, 1847) (Dynastinae, Cyclocephalini). The three species were found in superficial soil (about 10 cm deep) covered by grass, consuming the grass roots and soil with high humus content. The association is probably only a gregarious behavior around the food resources.

In the laboratory, one larva of G. barbatus was observed consuming a cadaver of a conspecific larva. It is not known if the larva killed the individual being consumed, but active sporadic cannibalism behavior has been reported for some scarab larvae (e.g. Soltani et al. 2008). On the other hand, some scarab larvae seem to attack and kill other larvae without feeding on them (e.g. Wightman 1974).

Key to the third instar larvae of Neotropical Rutelinae (modified from Pardo-Locarno et al. 2006)

1. Last antennomere with 2 or more dorsal sensory spots ................ Rutelini (see Jameson & Morón 2001) 2
   1' Last antennomere with one dorsal sensory spot .................. 2

2. Epipharynx with prominent haptorunem, without heli... 3
   2' Epipharynx with weakly prominent, rounded haptorunem, followed by transverse row of 2–4 stout heli ................. Anomalini (see Micó et al. 2003)

3. Palidia absent. Maxillary stridulatory area formed by 11 short teeth with similar shape and size ......................... Anoplognathini .... Platycycoel Dejean
   3' Palidia present. Maxillary stridulatory formed by 10 teeth with similar or different shape and size ................. Geniatini .... 4

   4' Lacinia with 3 unci. Plegmatia present. Left mandible with truncate acia. Respiratory plate V–VIII with progressively bigger diameter. Palidia long, formed by 12–16 pali ................. Geniates barbatus Kirby

DISCUSSION

Pardo-Locarno et al. (2006) characterized the Geniatini larvae based on Leucothyreus femoratus. The description of G. barbatus updates the larval diagnosis of Geniatini to: last

Revista Brasileira de Entomologia 57(1): 40–46, March 2013
Description of the third larval instar and pupa of *Geniates barbatus* Kirby

Antennomere with one dorsal sensorial spot (Rutelini have two or more; see Jameson & Morón 2001); epipharynx without heli (Anomalini have 2 to 4 well developed heli; see Micó et al. 2003; in other Rutelinae tribes heli is absent, because unciniform or teeth-like setae are not considered as heli sensu Böving 1936 and Ritcher 1948); palidia present, but reduced (raster of Anoplognathini without palidia).

This study shows that the larvae of Geniatini seem to be more similar to Anoplognathini than to Rutelini or Anomalini. Pardo-Locarno et al. (2006) proposed the separation between Geniatini (based on *L. femoratus*) and Anoplognathini (based on two species of *Platycoelia* Dejean, 1833, the single genus with described larvae in Anoplognathini; see Paucar-Cabrera & Smith 2002) using maxillary and palidal characters, as follows: Geniatini have lacinia with two unci and the maxillary stridulatory organ formed by heterogeneous teeth; meanwhile Anoplognathini have lacinia with three unci and homogeneous teeth in the maxillary stridulatory organ. Regarding these characters, the larvae of *G. barbatus* resemble more the larvae of *Platycoelia* than the larvae of *L. femoratus*.

Another common characteristic between *G. barbatus* and *Platycoelia* is the last abdominal spiracles larger than the first ones. On other hand, *Platycoelia* and *L. femoratus* have the left mandible with acuminate and glabrous acia and last antennomere with two ventral spots.

Larvae of *G. barbatus* and *L. femoratus* can be differentiated from each other by the following characters (characters of *L. femoratus* in parentheses): epipharynx with (without) plegmatia; left mandible with a truncated acia with apical setae (spine-like and glabrous); maxillary stridulatory organ with one anterior truncate process and 10 posterior acuminate teeth (with one anterior truncated process and 10 posterior teeth, the anterior five truncated); lacinia with three unci (with two unci); last antennomere with one ventral spot (two ventral spots); abdominal spiracles VII and VIII enlarged (I–VIII decreasing in length).

The pupae of *G. barbatus* and *L. femoratus* have the apex of urotergite IX prominent and upward deflected. This differentiates known Geniatini pupae from other known pupae of Rutelinae, with have the urosternite IX apex directed posteri-
orly. Moreover, the urotergite IX pubescence of *G. barbatus* and *L. femoratus* is relatively longer than in others Rutelinae. Pupae of *G. barbatus* and *L. femoratus* can be differentiated from each other by the following characters (characters of *L. femoratus* in parentheses): apex of metatibiae exposed in dorsal view (hidden by the pterothecae); dioneiform organs relatively longer (shorter); urotergite VIII transverse and shorter than urosternite IX (semispherical and longer than urotergite IX).

**ACKNOWLEDGMENTS**

I would like to thank Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) for grant support at the beginning of this project; Fundação de Amaparo à Pesquisa do Estado de São Paulo (FAPESP) (2011/20001-6) for a doctorate grant; Sônia A. Casari (MZSP), Sergio Ide (IBSP), Miguel A. Morón (Instituto de Ecologia, A.C.; Mexico), Sergio A. Vanin (Departamento de Zoologia, Universidade de São Paulo), Bruno A. S. de Medeiros (Museum of Comparative Zoology, Harvard University) and the anonymous reviewers for the valuable corrections and suggestions, and Gisele Sales Francfort and Daniela de Cassia Bená for helping with the material sampling.

**REFERENCES**


