Hypolipidemic activity of *Moringa oleifera* Lam., Moringaceae, on high fat diet induced hyperlipidemia in albino rats

**Pankaj G. Jain,* Savita D. Patil, Nitin G. Haswani, Manoj V. Girase, Sanjay J. Surana**

**INTRODUCTION**

Cardiovascular disease is leading cause of death in India as well as in western countries. Hyperlipidemia is a collective term used to describe human conditions when a plasma level of one or more classes of lipids, namely cholesterol, triacylglycerides, phospholipids and fatty acids increases above normal levels. Hyperlipidemia is one of the major causes of the development of cardiovascular disorders (Raida et al., 2008).

Various parts of the *Moringa oleifera* Lam., Moringaceae, tree have been studied for several pharmacological actions. The aqueous extract of leaves of *M. oleifera* was reported for the wound healing (Rathi et al., 2006) and antiurolithiasis activity (Karadi et al., 2006). The methanolic crude extract of *M. oleifera* shows antibacterial activity (Nantachit, 2006). The bark extract has been shown to possess antifungal and antitubercular activity (Bhatnagar et al., 1961). The ethanolic and aqueous extract of pods was found to be hypotensive (Faizi et al., 1998).

In many cultures of the world, herbal remedies are increasingly being employed in an attempt to achieve the same purpose. In India, for instance, the leaves of *Moringa oleifera* Lam. is claimed to possess cholesterol-reducing effect and is used to treat patients with heart disease and obesity. For this reason it was decided to resolve this claim by investigating the effects of the methanolic extract of leaves of *Moringa oleifera* Lam. (English: Horseradish plant or drumstick tree) on the lipid profile of the Wistar rat using experimental hyperlipidaemic animal model.
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MATERIAL AND METHODS

**Plant material**

The plant material was collected from local area of Shirpur in Dhule district, Maharastra, India. At the time of collection whole plant including leaves, pods and flower were collected and authentication was made by Dr. D.A. Patil, Professor, HOD, Department of Botany S.S.V.P.S’s College of Science; Dhule (Maharashtra). The voucher specimen (number 190647) was deposited for future reference.

**Extraction**

Leaves of the plant were removed and dried under shade in a room. After ten days of drying leaves were pulverized and sieved with a 40 # sieve. *Moringa oleifera* powder was extracted in methanol by using Soxhlet apparatus. The extract was dried in vacuum evaporator below 40 °C and sterilized through Whatmann filter no. 42 and reconstituted in saline.

**Phytochemical investigation of extract**

Different chemical constituents present in methanolic extract were subjected to the tests by Kokate (1994) and Trease & Evans (1997).

**HPTLC study**

β-sitosterol important chemical constituent of extract was detected using HPTLC fingerprint obtained using a Camag Linomat applicator and detector. The plates were observed under UV light after spraying with vanillin-con HCl reagent.

**Animals**

Healthy male albino Wistar rats of 180-200 g body weight were used in this study. The rats were housed in polypropylene cages under standard conditions (12 h light and dark cycles, at 25±30 °C and 35-60% humidity). Standard palleled feed (Amrut Pellets for rat, Pune, India) and tap water were provided *ad libitum*. Study was approved by Institutional Animal Ethical Committee CPCSEA, India (Registration No.651/02/C/CPCSEA).

**Chemicals**

Cholesterol, cholic acid (Loba-chemie Ltd), triacylglyceride, HDL kits (RFCL Diagnostic Pvt. Ltd), simvastatin (Aitemis biotech Ltd.), methanol were used in this study.

Preparation of the hyperlipidemic diet

The high fat diet was prepared by mixing calculated amounts of 2% cholesterol, 1% cholic acid and 1 mL coconut oil. The Parachute coconut oil was chosen because of its high saturated fat content which aggravates the atherogenic profile in the rats (Hassarajani et al., 2007).

**Cholesterol fed diet hyperlipidemia**

Animals were divided into six groups (n=6). Normal group (Group I) received normal diet *ad libitum*. Control group (Group II) received high fat diet (HFD). After fifteen days groups IV, V, VI received high fat diet along with the plant methanolic extract in the dose of 150, 300 and 600 mg/kg, *p.o.*, for thirty days. Group III received simvastatin the standard drug in the dose of 4 mg/kg *p.o.* after fifteen days along with high fat diet. After thirty days blood was withdrawn by cardiac puncture, allowed to clot for 45 min and serum was separated by centrifugation and lipid profile was done by using commercially available kits, with the help of ELISA reader (Biotech USA) (Hassarajani et al., 2007).

**Fecal cholesterol excretion**

Fecal matter was collected during the last three days of treatment period. The dried and powered fecal matter was extracted with CHCl₃;MeOH (2:1). The resultant extract was then analyzed for cholesterol contents in a manner similar to that the serum. The cholesterol excreted in the fecal matter (mg/g) was calculated.

**HMG-CoA reductase activity**

For the estimation of HMG-CoA reductase activity five group of rats were used and each group containing six animals (Venugopala & Ramakrishnan, 1975).

Group I: Vehicle control (0.5 mL carboxy methyl cellulose solution) for seven days; Group II: Test drug treated (150 mg/kg) for seven days; Group III: Test drug treated (300 mg/kg) for seven days; Group IV: Test drug treated (600 mg/kg) for seven days; Group V: Simvastatin treated (4 mg/kg) for seven days.

The liver tissue was removed as quickly as possible and a 10% w/v homogenate was prepared in saline arsenate solution. The homogenate was deproteinized using an equal volume of dilute perchloric acid and allow to stand for 5 min, followed by centrifugation. To 1 mL of the filtrate, 0.5 mL of freshly prepared (alkaline hydroxylamine reagent in the case of HMG-CoA) was added. It was mixed and 1.5 mL of ferric chloride reagent was added after 5 min. The absorbance was read after 10 min at 540 nm vs a similarly treated saline arsenate blank.
The ratio of HMG-CoA/mevalonate was calculated.

**Biochemical assays for lipids**

Cholesterol, HDL and triacylglyceride levels were estimated from serum by CHOD-PAP method and GPO-PAP method (Devi & Sharma, 2004), respectively. LDL and VLDL-cholesterol were calculated following the method by Johnson et al. (1997). While the atherogenic index was calculated by using the method described by Muruganandan et al. (2005).

**Statistical analysis**

Results are expressed as mean±SEM (standard error mean) and subjected to one-way analysis of variance (ANOVA) followed by Dunnett's test and values with p<0.05 were considered to be statistically different.

**RESULTS AND DISCUSSION**

The percentage yield of the methanolic extract was found to be 47.2% w/w. Phytochemical investigation was performed and the following compounds were identified: glycoside, steroids flavonoids and alkaloids. A 36.57% of β-sitosterol was present in crude extract of *Moringa oleifera* quantified through HPTLC results (Figure 1).

There was significant increase in the levels of serum cholesterol, triacylglyceride and LDL-c and a decrease in level of HDL-c in the high fat diet fed animals when compared to normal fed rats. Elevated level of blood cholesterol especially LDL-c is a known major risk factor for CHD whereas HDL-c is cardio protective. Treatment with methanolic extract, at three different doses significantly decreased the levels of total cholesterol and LDL-c as compare to the controls (Tenpe et al., 2007) (Table 1).

Atherogenic index indicates the deposition of foam cells or plaque or fatty infiltration or lipids in heart, coronaries, aorta, liver and kidneys. The higher the atherogenic index, the higher is the risk of the above organs for oxidative damage (Mehta et al., 2003). Atherogenic index was significantly reduced in the *M. oleifera* treated groups.

HFD causes the oxidative stress which finally increases production of reactive oxygen species. An increasing scientific literature provides ample direct or indirect evidence that overproduction of ROS can induce cellular damage via oxidation of critical cellular components such as membrane lipids, proteins, and DNA.

Since the result of the study indicated that the methanolic extract of *M. oleifera* has beneficial effect on lipid profile we have investigated its mechanism of action. Cholesterol homeostasis is maintained by the two processes viz. cholesterol biosynthesis in which HMG-Co-A reductase catalyzes rate limiting process and cholesterol absorption of both dietary cholesterol and cholesterol cleared from the liver through biliary secretion (Hassarajani et al., 2007).

The HMG-Co-A/mevalonate ratio has an inverse relationship to the activity HMG-Co-A reductase. The result of the study indicated that the activity of the enzyme is significantly depressed by the methanolic extract of *M. oleifera* as is evidenced by the increase in the ratio. There was significant increase in the ratio of HMG-CoA/mevalonate in the test drug at the dose of 600 mg/kg (p<0.05) and in the standard drug at the dose of 4 mg/kg (p<0.01) as compare to the control group (Table 2). Methanolic extract at doses of 300 mg/kg (p<0.05), 600 mg/kg (p<0.01) and standard drug simvastatin (p<0.01) shows significant increase in cholesterol excretion when compared with HFD control group (Figure 2).

**Figure 1.** HPTLC peak of standard β-sitosterol (A) and of *Moringa oleifera* Lam., Moringaceae (B) at 580 nm.
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Table 1. Effect of methanolic extract of *Moringa oleifera* Lam. on lipid profile in cholesterol fed hyperlipidemic rats

<table>
<thead>
<tr>
<th>Groups</th>
<th>Cholesterol (mg/dl)</th>
<th>Triacylglyceride (mg/dl)</th>
<th>VLDL (mg/dl)</th>
<th>LDL (mg/dl)</th>
<th>HDL (mg/dl)</th>
<th>Atherogenic index</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal#</td>
<td>84.40±2.45</td>
<td>122.2±6.78</td>
<td>24.44±1.35</td>
<td>15.16±1.99</td>
<td>45.20±1.50</td>
<td>0.89±0.09</td>
</tr>
<tr>
<td>Control (HFD)</td>
<td>196.8±7.01</td>
<td>200.2±5.31</td>
<td>40.04±1.06</td>
<td>116.2±7.45</td>
<td>40.60±1.29</td>
<td>3.87±0.25</td>
</tr>
<tr>
<td>Simvastatin 4 mg/kg</td>
<td>99.20±2.74&quot;</td>
<td>86.60±3.54&quot;</td>
<td>17.32±0.71&quot;</td>
<td>22.68±8.44&quot;</td>
<td>59.20±2.45&quot;</td>
<td>0.69±0.10&quot;</td>
</tr>
<tr>
<td>MeOH Extract 150 mg/kg</td>
<td>183±5.13</td>
<td>182±2.89&quot;</td>
<td>36.40±0.58&quot;</td>
<td>105.6±6.21</td>
<td>41±1.82</td>
<td>3.51±0.30</td>
</tr>
<tr>
<td>MeOH Extract 300 mg/kg</td>
<td>142.6±2.92&quot;</td>
<td>169.2±3.96&quot;</td>
<td>33.84±0.80&quot;</td>
<td>61.36±3.40&quot;</td>
<td>47.40±1.21&quot;</td>
<td>2.01±0.08&quot;</td>
</tr>
<tr>
<td>MeOH Extract 600 mg/kg</td>
<td>121.2±3.88&quot;</td>
<td>157.4±4.55&quot;</td>
<td>31.48±0.91&quot;</td>
<td>38.72±4.06&quot;</td>
<td>51±1.52&quot;</td>
<td>1.38±0.09&quot;</td>
</tr>
<tr>
<td>Simvastatin 4 mg/kg</td>
<td>121.2±3.88&quot;</td>
<td>157.4±4.55&quot;</td>
<td>31.48±0.91&quot;</td>
<td>38.72±4.06&quot;</td>
<td>51±1.52&quot;</td>
<td>1.38±0.09&quot;</td>
</tr>
</tbody>
</table>

#Regular diet water *ad libitum*; values are expressed as mean±SEM, n=6; Data was analyzed by one way ANOVA followed by Dunnet test; **p<0.01, very significant; *p<0.05, significant.

Table 2. Effect of methanolic extract of *Moringa oleifera* on HMG-CoA reductase activity.

<table>
<thead>
<tr>
<th>Group</th>
<th>Absorbance ratio (mean±SEM)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>1.489±0.1512</td>
</tr>
<tr>
<td>Methanolic extract 150 mg/kg</td>
<td>1.513±0.139</td>
</tr>
<tr>
<td>Methanolic extract 300 mg/kg</td>
<td>1.718±0.217</td>
</tr>
<tr>
<td>Methanolic extract 600 mg/kg</td>
<td>2.126±0.233</td>
</tr>
<tr>
<td>Simvastatin 4 mg/kg</td>
<td>3.133±0.223</td>
</tr>
</tbody>
</table>

Values are expressed as mean±SEM, n=6; data was analyzed by one way ANOVA followed by Dunnet test; **p<0.01, very significant; *p<0.05, significant.

Figure 2. Effect of methanolic extract (150, 300, 600 mg/kg) and simvastatin drug 4 mg/kg on the excretion of cholesterol in rats each value represents the mean±of six rats, *p<0.05, **p<0.01 vs. control.

Plant sterols inhibit the absorption of dietary cholesterol, but the resulting decrease in serum cholesterol has been slight (Saluja et al., 1978). Although *M. oleifera* has been shown to contain β-sitosterol in HPTLC, β-sitosterol is a plant sterol with a structure similar to that of cholesterol, except for the substitution of an ethyl group at C24 of its side chain. The cholesterol lowering effect may be due to this inhibition in reabsorption of cholesterol from endogenous sources in association with a simultaneous increase in its excretion into feces in the form of neutral steroids. The increased fecal excretion of cholesterol observed with *M. oleifera* treated group as compared to the HFD control group.

Conclusively, the observed cholesterol reducing action of the methanolic extract of *Moringa oleifera* Lam. indicates the hypolipidemic activity.

REFERENCES


