Hypoglycemic activity of two Brazilian Bauhinia species: Bauhinia forficata L. and Bauhinia monandra Kurz.

Fábio de Sousa Menezes1,2*, Andréa Barreto Mattos Minto1, Halliny Siqueira Ruela1, Ricardo Machado Kuster1, Helen Sheridan2, Neil Frankish2

1Departamento de Produtos Naturais e Alimentos, Faculdade de Farmácia, Centro de Ciências da Saúde, Cidade Universitária, 21941-590, Rio de Janeiro, RJ, Brazil,
2School of Pharmacy and Pharmaceutical Sciences, Trinity College Dublin, University of Dublin, 23 Westland Row, Dublin 2, Ireland,
3Núcleo de Pesquisas de Produtos Naturais, Centro de Ciências da Saúde, Bloco H, Cidade Universitária, 21941-590, Rio de Janeiro, RJ, Brazil

RESUMO: “Atividade hipoglicemiante de duas espécies de Bauhinia brasileira: Bauhinia forficata L. e Bauhinia monandra Kurz.”. Extratos aquosos das folhas de Bauhinia forficata L. e Bauhinia monandra Kurz (10% p/v) foram testados em camundongos normoglicêmicos, objetivando averiguar a sua atividade hipoglicemiante. Ambos os extratos mostraram atividade hipoglicemiante na metodologia empregada. Ainda, foi possível isolar de B. forficata L. dois flavonóides, quercetina-3,7-O-α-rhamnopyranosil e kaempferol-3,7-O-α-rhamnopyranosil, sendo as estruturas estabelecidas por técnicas clássicas de RMN. Apenas o derivado da quercetina foi identificado no extrato aquoso de Bauhinia monandra por CLAE.

Unitermos: Bauhinia forficata, Bauhinia monandra, Leguminosae, atividade hipoglicemiante, flavonoides, CLAE.

ABSTRACT: The hypoglycemic activity of aqueous extracts from Bauhinia forficata L. and Bauhinia monandra Kurz leaves (10% w/v) was evaluated in normoglycemic mice. Both extracts have shown hypoglycemic activity using this methodology. It was also possible to isolate two flavonoids from B. forficata L., 3,7-di-O-α-rhamnopyranosylquercetin and 3,7-di-O-α-rhamnopyranosylkaempferol (kaempferitrin), whose structures were elucidated by usual NMR techniques. Only the quercetin derivative was identified in B. monandra aqueous extract by HPLC.

Keywords: Bauhinia forficata, Bauhinia monandra, Leguminosae, hypoglycemic activity, flavonoids, HPLC.

INTRODUCTION

Plants belonging to the genus Bauhinia are known in Brazil as “Pata-de-vaca” or “Unha-de-boi”. Their species belong to the Leguminosae family and encompass circa 300 plants cultivated in tropical areas worldwide (Miyake et al., 1986). Aqueous extracts from Bauhinia species leaves, stems and roots, mainly B. cheilantha, B. forficata, B. glabra, B. rufescens, B. splendens and B. ungalata, are widely used in Brazil and in other countries in the treatment of several diseases, specially infections, pain processes and diabetes (Achenbach et al., 1988; Ritter et al., 2002; Pereira et al., 2004; Macedo; Ferreira, 2004; Morais et al., 2005; Silva et al., 2006). Several Bauhinia species are utilized as folk medicines worldwide, including Africa, Asia, South America and Central America (Ayensu, 1978).

Phytochemical and pharmacological studies with species of this genus are ongoing and, consequently a wide range of compounds have been isolated and identified, including lactones, flavonoids, terpenoids, glicolipids, glucosyl steroids, steroids, tannins and quinines (Cechinel-Filho et al., 1996, Silva et al., 2000; Mendes et al., 2006). One early study, by Bhartiya and Gupta (1981), identified 3,4-dihydroxychalcone-4-O-β-L-arabinopyranosyl-O-α-D-galactopyranoside from B. purpurea seeds. Roots and stems of this plant are popularly used in the treatment of diarrhea in India. During the same period studies with the methanolic extract (2 g/kg) of B. racemosa flowers in gastric ulcer model induced by acetylsalicylic acid in rats have shown that this extract significantly reduced the ulcer index (Akhtar et al., 1981). A glucosyl steroid has been isolated from B. candicans aerial parts and identified as sitosterol-
3-O-α-D-riburonofuranoside (Iribarren; Pomilio, 1985). Achenbach et al. (1988), in a phytochemical study of B. manca stems have isolated 3-O-galloylepticcatequinme as its main constituent. They also identified gallic acid, cinnamic acid, β-sitosterol and β-D-glucosyl-sitosterol in significant concentrations. Alkaloids were not detected.

All the extracts from B. splendens leaves, stems, barks and roots, except the hydroalcoholic extract from its roots, have shown significant analgesic effect (10 mg/kg) in a pain model induced by acetic acid 0.6% in mice, being more efficient than salicylic acid and paracetamol. In the same study the chemical constituents from leaves, stems, barks and roots extracts were evaluated by thin layer and column chromatography. Identified compounds include ethyl gallate, rutin and quercetin (Cechinel-Filho et al., 1996).

In a more recent study, Taylor et al. (1996), on Bauhinia species used in folk medicine in Nepal, the methanolic extract from B. vahlii roots was found to be one of the most active in the treatment of virus-induced diseases with specific activity toward Herpes simplex. A later chemical study of the methanolic extract obtained from B. purpurea wood led to the isolation and identification of 6-butyl-3-hydroxy flavanone (Kuo et al., 1998). More recently the ethanolic extract from B. guianensis stem barks has shown a very good activity against Plasmodium vinckeii (50 mg/kg) (Muñoz et al., 2000) in an evaluation of one hundred and fifty six different species belonging to different families indicated by “Chacabos” (native community of Amazon Region) for the treatment of several diseases such as digestive complaints, ulcer, diarrhea, malaria, etc. Yadava and Tripathi (2000) have isolated 5,6-dihydroxy-7-methoxy-flavone-6-O-β-D-xilopyranoside from the chloroform fraction of the total ethanolic extract from B. purpurea barks. Extracts from the bark of B. guianensis stems have been shown to possess antinociceptive effects in the acetic acid writhing model in mice, as well as anti-inflammatory activity in the carrageenan, dextran and histamine paw edema model in rats (Silva; Cechinel-Filho, 2002; Falcão et al., 2005).

Based on the study of Cechinel-Filho et al. (1996) with B. splendens, Silva and Cechinel-Filho (2000) have studied B. forficata, used by the Brazilian Pharmacopoeia for the physical-chemical and botany parameters for the quality control of this species. The phytochemical study of this species showed the presence of kaempferol-3,7-dirhamnoside (kaempferitrine, lespedin) in the methanolic extracts of leaves and β-sitosterol from the hexane fraction. A comparative study carried out with all the vegetative organs of this species (leaves, roots and barks) has proved that steroids and terpenes are present in all of them, being more concentrated in the leaves. Lespedin was detected only in B. forficata leaves and because of that, this compound can be considered the chemical marker of this species for quality control purposes, present in leaves and absent in other parts.

The documented use and potential of Bauhinia species to act as hypoglycemic agents have begun to receive much attention. Bauhinia divaricata, known in Mexico as “pezunã de vaca”, has been studied regarding its hypoglycemic activity and its aqueous extract (obtained from leaves) showed a relatively significant hypoglycemic effect in normal mice (Gupta et al., 1980). It has been established that the aqueous extract from B. megalandra inhibits the glucose intestinal absorption (Gonzales-Mujica et al., 2003) in rats. However, the hypoglycemic activity of Bauhinia species has been most extensively researched in B. forficata, which is considered by the rural population in Brazil as the “true pata-de-vaca”. B. forficata Link. had its hypoglycemic activity confirmed by Juliane (1929, 1931). This species leaves are currently used in Brazil, in the form of aqueous extract (Modesto-Filho, 1988; Barbosa-Filho et al., 2005). However, a later study by Coimbra-Teixeira et al. (1992) has shown that the alcoholic leaves extract do not make the glucose blood levels lower in diabetic rats induced by streptozotocin.

Due to conflicting scientific results and the fact that Bauhinia species are widely used in Brazil as hypoglycemic plants, without any control, using popular knowledge that correlates the color of the flowers with the suggested activity power, we have decided to investigate the hypoglycemic activity of the aqueous extract of two Bauhinia species used in the State of Rio de Janeiro, Brazil. The selected species are Bauhinia forficata L. and Bauhinia monandra Kurz. A second objective is to try and isolate one or more markers for these species that could be responsible, at least in part, for their activity in normoglycemic mice, and could find potential use for quality control purposes.

**MATERIAL AND METHODS**

**Plant material**

Bauhinia forficata L. and Bauhinia monandra Kurz were collected in different parts of Rio de Janeiro State and prepared for the study. These species were identified by Professor Ângela Studart da Fonseca Vaz from the Botanical Garden of Rio de Janeiro, specialist in Leguminosae family. Two voucher samples are deposited in the Botanical Garden of Rio de Janeiro Herbarium under the numbers 3456 and 3765 respectively.

**Extracts preparation and animals**

The aqueous extracts from the two mentioned species were prepared in the concentration of 10% (w/v) using dried leaves (2 g). Further their preparation they were administered to Swiss mice (male, medium weight between 20 and 25 g). Animals were maintained in room temperature with free access to water and food.

Animal care and research protocols were in accordance with the principles and guidelines adopted by the Brazilian College of Animal Experimentation (COBEA) and approved by the Biomedical Science Institute/UFRJ-Ethical Committee for animal research.

Animal blood collection and glucose level determination

All blood samples were collected by orbital puncture, obtained from the ophthalmic plexus, in animals under anesthesia with ethyl ether. Blood was collected in heparin capillaries for micro hematocrit with capacity of 70 mL and centrifuged at 1100 rpm for 5 minutes in order to separate plasma. After its separation, an aliquot of 20 mL was collected and glucose-oxidase solution was used as blank (B). The basal glycemia (time = zero) was determined and for 12 hours before the onset and during all the experiment.

Determination of glycemia in normoglycemic animals

Mice (n = 8) were used to verify the hypoglycemic activity from the two studied species of Bauhinia which the aqueous extract was administered by oral route to male mice in a concentration of 10% (w/v).

Animals were left fasting receiving water since 12 hours before the onset and during all the experiment. The basal glycemia (time = zero) was determined and further each group received its appropriate treatment. Each control group received distilled water 0.1 mL/10g of weight, p.o and each treated group received aqueous extract (10% w/v) of one of the two tested Bauhinia species 0.1 mL/10g of weight, p.o.

The glycemia of each group was observed in 2, 4, 6 and 8 hours after the onset of the experiment with or without the administration of aqueous extract.

Separation, isolation and identification of the active compounds

The high performance liquid chromatography was made in a Shimadzu CBM 104 apparatus equipped with a LC-10AD pump and SPD-M10A DAD detector, manual injection valve Rheodyne with 20 mL of capacity, column Lichrosorb (Merk. N. 738342) RP-18 (250 x 4 mm, internal diameter 10 mm). The monitoring was in fix wavelength (254 nm). The mobile phase was formed by 1% phosphoric acid solution in water (Solvent A) and 1% phosphoric acid solution in methanol Solvent B) flowing at 1 mL/min. The gradient beginning with 30% of B growing to 40% of B in ten minutes. This composition remaining isocratic up to 20 minutes when another gradient was started going to 100% of B in 60 minutes, maximum time of the chromatographic process. Using this methodology, it could be possible to isolate from Bauhinia forficata aqueous extract, two glucosyl flavonoids: 3,7-di-O-α-rhamnopyranosylkaempferol, kaempferitrin, \( \text{RT} = 24 \text{ minutes} \) and 3,7-di-O-α-rhamnopyranosyl quercetin \( \text{RT} = 32 \text{ minutes} \).

These compounds were identified by \(^1\text{H} \) NMR (200 MHz) and \(^{13}\text{C} \) NMR (50 MHz) spectra registered in a Varian Star-200 apparatus, with deuterated solvents and TMS as internal standard. The 400 MHz \(^1\text{H} \) NMR was registered in a Bruker DRX apparatus using the same conditions. After elucidated each structure, data were compared with literature previously published one and by his way, it was possible to confirm the identity of each compound. The analysis of B. monandra aqueous extract and co-injection with the isolated molecules from B. forficata made possible to visualise or not the same compounds.

\( 3,7-\text{Di-O-α-rhamnopyranosylkaempferol} \).

\(^1\text{H} \) NMR (200 MHz, DMSO-d6, TMS): \( \delta \) (ppm) 0.8-1.1 (methyl groups of rhamnose, 2 d), 3.1-4.0 (signals concerning the H of the sugar moiety of this molecule) 5.3 (sl, anomic H from rhamnose in the position 3), 5.55 (sl, H anomic H from rhamnose in the position 7), 6.45 (sl, H-8), 6.75 (sl, H-6), 6.9 (d, \( J = 8.43 \text{ Hz} \), 2H, H-3’ and H-5’), 7.75 (d, \( J = 8.42 \text{ Hz} \), 2H, H-2’ and H-6’). \(^{13}\text{C} \) NMR (200 MHz, CD3OD, TMS): \( \delta \) (ppm) 0.9-1.3 (methyl groups of rhamnose, 2 d), 3.3-3.9 (signals concerning the H of the sugar moiety of this molecule), 5.4 (d, \( J = 1.6 \text{ Hz} \), H anomic H from rhamnose in the position 3), 5.6 (d, \( J = 1.16 \text{ Hz} \), H anomic H from rhamnose in the position 7), 6.44 (d, \( J = 2.1 \text{ Hz} \), H-8), 6.7 (d, \( J = 2.0 \text{ Hz} \), H-6), 6.94 (d, \( J = 8.80 \text{ Hz} \), 2H, H-3’ and H-5’), 7.78 (d, \( J = 8.80 \text{ Hz} \), 2H, H-2’and H-6’). \(^{13}\text{C} \) NMR (50 MHz, CD3OD, TMS): \( \delta \) (ppm) 17.7 (C-1’), 18.1 (C-6-Rha); 71.2 (C-5’-Rha); 71.6 (C-5-Rha); 71.8 (C-2-Rha); 72.1 (C-3-Rha, C-2’-Rha, C-3’-Rha); 73.1 (C-4’-Rha); 73.5 (C-4-Rha); 95.5 (C-8); 99.8 (C-6); 100.5 (C-1’-Rha); 103.5 (C-1-Rha); 107.5 (C-8a); 116.6 (C-3’); 122.3 (C-1’); 131.9 (C-2’); 136.4 (C-9'); 157.9 (C-4a); 159.7 (C-2); 161.7 (C-4’); 162.9 (C-5); 163.5 (C-7); 179.7 (C-4).

\( 3,7-\text{Di-O-α-rhamnopyranosylkaempferol} \).

\(^1\text{H} \) NMR (400 MHz, CD3OD, TMS): \( \delta \) (ppm) 0.95-1.25 (methyl groups of rhamnose, 2 d), 3.34-4.25 (signals concerning the H of the sugar moiety of this molecule), 5.37 (d, \( J = 1.37 \text{ Hz} \), H anomic H from rhamnose in the position 3), 5.56 (sl, H anomic H from rhamnose in the position 7), 6.47 (d, \( J = 2.05 \text{ Hz} \), H-8), 6.73 (d, \( J = 2.05 \text{ Hz} \), H-6), 6.92 (d, \( J = 8.21 \text{ Hz} \), H-3’), 7.33 (dd, \( J = 2.05 \text{ Hz} \) and \( J = 8.21 \text{ Hz} \), H-2’), 7.37 (d, \( J = 2.05 \text{ Hz} \), H-6’).

Statistical evaluation

Data related to the animals glycemia were
analyzed by an experimental design completely casual, applying variance analysis (ANOVA) in order to verify significant differences between the groups, with 5% of significance level (p < 0.05). To compare control and experimental groups, t-Student test was applied. Non-paired t-Student test was applied to compare the glycemia average in each measured time. The results were expressed as an arithmetic average of glycemia in each measured time, with standard deviation. Each aqueous extract was evaluated in triplicate.

RESULTS AND DISCUSSION

Evaluation of the chromatograms of the aqueous extract of *Bauhinia forficata* and *Bauhinia monandra* by UV led to the visualization of signals which we believed to be glucosyl flavonoids (or flavonoid glycosides). In the case of *Bauhinia forficata* it was possible to isolate and identify two flavonoids with the following retention times: 3,7-di-O-α-rhamnopyranosylkaempferol, kaempferitrin, (24 minutes) and 3,7-di-O-α-rhamnopyranosylquercetin (32 minutes) (Figure 1). These compounds structures were established by NMR techniques. 3,7-di-O-α-rhamnopyranosylquercetin was also identified in the aqueous extract of *Bauhinia monandra* by comparison of both UV and retention data and by co-injection using HPLC. Table 1 shows the relative amount of both flavonoids in the two extracts. The quantitative relation between kaempferitrin and 3,7-di-O-α-rhamnopyranosylquercetin observed in *B. forficata* aqueous extract used in this work is in accordance with those showed to another sample of *B. forficata* aqueous extract from Itajaí (South part of Brazil), which was also seen when they evaluated *B. forficata* fluid extract (Pinheiro et al., 2006). Flavonoids were also identified in the aqueous extract of *B. monandra*, although their structures have not been elucidated (Argolo et al., 2004).

The hypoglycemic activity was established for both species using the described methodology as it can be seen in Graphic 1. The results found in this work match some results previously published, at least for *B. forficata* (Jorge et al., 2004). In the same study, the isolated molecule (kampferitrin) was also evaluated on 14C-glucose uptake in rat soleus muscle and has shown insulinomimetic effects. The insulin-like effects of *B. forficata* aqueous extract were also evaluated in mice after scorpion envenoming. Although no insulinomimetic effect has been shown, this extract was able to enhance the venom lethality (Vasconcelos et al., 2004).

Based on these evidences, the hypoglycemic activity shown in this paper can be related to the presence of glucosyl flavonoids which have different qualitative and quantitative profiles in both extracts. Both species had a very pronounced effect in the in vitro method used to establish activity, showing them to be very promising hypoglycemic agents. It is interesting to note that no relationship was observed between the color of the flowers and the strength of the hypoglycemic activity. Also, hypoglycemic activity for other *Bauhinia* species has been shown using different methodologies (Fuentes et al., 2004; Almeida et al., 2006; Fuentes; Alarcón, 2006).

Further research is needed to permit us to

**Table 1.** Presence of 3,7-O-dirhamnosyl-kaempferol and 3,7-O-dirhamnosyl-quercetin in the studied *Bauhinia* aqueous extracts.

<table>
<thead>
<tr>
<th>Species</th>
<th>3,7-O-Dirhamnosyl-kaempferol</th>
<th>3,7-O-Dirhamnosyl-quercetin</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>B. forficata</em></td>
<td>++++</td>
<td>++</td>
</tr>
<tr>
<td><em>B. monandra</em></td>
<td>-</td>
<td>++++</td>
</tr>
</tbody>
</table>

**Figure 1.** Structure of the two isolated glucosyl flavonoids from the aqueous extract of *B. forficata.*
completely understand the way the aqueous extracts make the glucose blood level lower. It is now our intention to test the pure isolated molecules. Further studies also include the evaluation of these extracts and the extracts belonging to other Brazilian Bauhinia species, as well as their isolated compounds in beta-cell line culture (Asfari et al., 1992), to evaluate the efficacy of each extract and isolated compound in the eliciting the insulin production and release.

REFERENCES
Hypoglycemic activity of two Brazilian Bauhinia species: Bauhinia forficata L. and Bauhinia monandra Kurz.


