Chemical composition and cardiovascular effects induced by the essential oil of Cymbopogon citratus DC. Stapf, Poaceae, in rats

Flávia V. Moreira,¹ Joana F. A. Bastos,¹ Arie F. Blank,² Péricles B. Alves,³ Márcio R. V. Santos*¹

¹Departamento de Fisiologia, Centro de Ciências Biológicas e da Saúde, Universidade Federal de Sergipe, Av. Marechal Rondon s/n, Rosa Elze, 49.100-000 São Cristóvão-SE, Brasil,
²Departamento de Agronomia, Universidade Federal de Sergipe, Av. Marechal Rondon s/n, Rosa Elze, 49.100-000 São Cristóvão-SE, Brasil,
³Departamento de Química, Universidade Federal de Sergipe, Av. Marechal Rondon, s/n, Rosa Elze, 49.100-000 São Cristóvão-SE, Brasil.

INTRODUCTION

The essential oils are a mixture of volatile substances composed mainly of terpenes in addition to some other non-terpene components. These volatile substances are commonly found in aromatic plants and their therapeutic potentials have been lately evaluated (Kris-Etherton et al., 2002; Edris, 2007; Paduch et al., 2007).

Studies in animals have demonstrated beneficial properties of essential oils in the cardiovascular system as antithrombotic, antiplatelet, endothelial protective, vasorelaxant and hypotensive activities (Lahlou et al., 2005; Edris, 2007). Recent reports have showed that cardiovascular effects of essential oils also occur in humans as improvement in coronary flow (Shiina et al., 2008) and hypotensive and bradycardic effects (Dayawansa et al., 2003).

Cymbopogon citratus (DC.) Stapf, Poaceae, is an aromatic medicinal plant popularly known as “capim-
Leaves of Cymbopogon citratus (DC.) Stapf, Poaceae, were collected from the garden of medicinal plants from Federal University of Sergipe (Brazil). A voucher specimen was deposited at Herbarium-ASE of Federal University of Sergipe (code 9391). EOCC was obtained from the fresh leaves by hydrodistillation in a Clevenger apparatus for 8 h and stored at 4 °C. When required, EOCC was dissolved in a saline/cremophor (0.1% v/v) solution, for \( \textit{in vivo} \) experiments, or distilled water/cremophor (0.1% v/v) solution, for \( \textit{in vitro} \) experiments, at desired concentrations.

The drugs used were: acetylcholine chloride (ACH), \( \text{L} \)-phenylephrine (Phe), NG-nitro-\( \text{L} \)-arginine methyl ester hydrochloride (L-NAME), atropine sulphate, indomethacin (INDO), tetraethylammonium (TEA) and cremophor (all from Sigma), sodium thiopental (Cristália), heparin sodium salt (Roche) and nifedipine (NIF) (RBI). The INDO was dissolved with sodium bicarbonate (NaHCO\(_3\)) to 2.5% in saline (m/v), while others drugs were freely dissolved in Tyrode solution or saline. All stock solutions were maintained at 0 °C and diluted to the desired concentration with Tyrode solution (\( \textit{in vitro} \) experiments), when necessary. NaHCO\(_3\) and EtOH in used concentrations showed no effect on control experiments (data not shown).

EOCC was analyzed by gas chromatography coupled with mass spectrometry (GC/MS) according to these experimental conditions: capillary column DB-5MS (30m x 0.25mm x 0.25 mm i.d), electron impact 70 eV; helium (99.999%) was used as carrier gas at a constant flow of 1.2 mL min\(^{-1}\) and an injection volume of 0.5 µL (dilution in ethyl acetate); injector temperature 250 °C; detector temperature 280 °C. The oven temperature was programmed from 50 °C (isothermal for 2 min), with an increase of 4 °C/min., to 200 °C, then 10 °C/min to 300 °C, ending with a 10 min isothermal at 300 °C. Mass spectra were taken at 70 eV; a scan interval of 0.5 s and fragments from 40 to 550 Da. Quantitative analysis of the chemical constituents was performed by flame ionization gas chromatography (FID), under same conditions GC-MS. Identification of individual components of the essential oil was performed by computerized matching of the acquired mass spectra with those stored in NIST21 and NIST107 mass spectral library of the GC-MS data system. Retention indices (RI) for all compounds were determined according to literature (Van Den Dool & Kratz, 1963) for each constituent as previously described (Adams, 2007). Male Wistar rats (200-300 g) were used in all experiments. They were housed in conditions of controlled temperature (21±1 °C) and exposed to a 12 h light-dark cycle with free access to food (Purina-Brazil) and tap water. All procedures described in the present work are approved by the Animal Research Ethics Committee from Universidade Federal de Sergipe (CEPA n° 31/2007).

The measurements of the mean arterial pressure (MAP) and heart rate (HR) were performed as described by Guedes et al. (2004). The MAP and HR were recorded before and after administration of EOCC (1, 5, 10 and 20 mg/kg, i.v., randomly), and after treatment with atropine (2 mg/kg; i.v.; 15 min.), L-NAME (20 mg/kg; i.v.; 30 min), INDO (5 mg/kg, i.v. 30 min) or sodium thiopental (45 mg/ kg; i.p., 60 min), separately. The change of MAP and HR for each dose was expressed as percentage of baseline values.

For \( \textit{in vitro} \) experiments, the rings of rat superior mesenteric artery (1-2 mm) were isolated and maintained according to the technique described by Santos et al. (2006).

The presence of functional endothelium was assessed by the ability of acetylcholine to evoke more than 70% of relaxation against phenylephrine-induced contraction.

Values were expressed as the mean±SEM. The results were analyzed or with repeated measures one or two-way ANOVA followed by Bonferroni post-test. The pD\(_2\) values were obtained by non-linear regression. All procedures were performed by using Graph Pad Prism 3.02™.

RESULTS AND DISCUSSION

In Brazil, many hypertensive patients with associated cardiovascular diseases drink daily tea of Cymbopogon citratus, and this study demonstrated possible benefits of the essential oil extracted of this plant on the cardiovascular system. Our results demonstrated that OECC appears to have a calcium-blocking property as many drugs used in the treatment of hypertension such as amilodipine, nifedipine and verapamil (Sociedade Brasileira de Cardiologia, 2007). The analysis of the EOCC by GC/MS demonstrated the presence of eight constituents that are showed in Table 1. The major constituents were geranial (43.08%), neral (32.19%) and mircene (17.58%) (Table 1). The percentages obtained in the present work were similar to those demonstrated in other studies for the same species.
(Blanco et al., 2009; Guimarães et al., 2008). Furthermore, it did not find in the literature any study demonstrating the activity of these constituents on the cardiovascular system.

In non-anesthetized rats, the injections of EOCC (1, 5, 10 and 20 mg/kg; i.v.) induced an intense and transitory hypotension associated with bradycardia (Figure 1A).

It is established that the stimulation of cardiac muscarinic receptors from sinoatrial node by vagal action induces intense bradycardia, which can be followed by hypotension due to a decrease in cardiac output (Peterson et al., 1984). It is also well known that activation of endothelial muscarinic receptors induces intense vasodilatation due to release of endothelium-derived relaxing factors, mainly NO and PGI₂ (Furchgott & Zawadzki, 1980; Moncada & Higgs, 1993). This activation can cause decrease in peripheral vascular resistance and, consequently, hypotension. The involvement of the muscarinic receptors in these responses was evaluated through experiments in the animals pre-treated with atropine (2 mg/kg), a non-selective antagonist of these receptors. In these animals, the hypotension was significantly attenuated only in dose of 20 mg/kg, while bradycardia was completely abolished in the doses of 5, 10 and 20 mg/kg (Figure 1A). This suggests that the bradycardia seems to be caused by cardiac muscarinic activation.

In order to check a possible indirect effect of EOCC through of central nervous system and vagus nerve, we performed experiments by using rats anesthetized with sodium thiopental (45 mg/kg), a general anesthetic

<table>
<thead>
<tr>
<th>Constituents</th>
<th>R1</th>
<th>%</th>
</tr>
</thead>
<tbody>
<tr>
<td>6-Methyl-5-hepten-2-one</td>
<td>982</td>
<td>0.57</td>
</tr>
<tr>
<td>Mircene</td>
<td>988</td>
<td>17.58</td>
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<tr>
<td>Linalool</td>
<td>1100</td>
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<tr>
<td>Epoxide of rosefurano</td>
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<td>0.51</td>
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<tr>
<td>Neral</td>
<td>1239</td>
<td>32.19</td>
</tr>
<tr>
<td>Geraniol</td>
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</tr>
<tr>
<td>Geranial</td>
<td>1269</td>
<td>43.08</td>
</tr>
<tr>
<td>Acetate of geraniol</td>
<td>1379</td>
<td>0.62</td>
</tr>
<tr>
<td>Total identified</td>
<td>100.00</td>
<td>100.00</td>
</tr>
</tbody>
</table>

Table 1. Chemical composition of the essential oil of *Cymbopogon citratus* (DC.) Stapf, Poaceae, leaves

*Percentages obtained by FID peak-area normalization (DB-5 column); *Retention Indices (DB-5 column)

![Figure 1](image-url)
with vagalitic and central vasomotor depressant actions (Morgan et al., 2006). In this condition, only bradycardia was significantly attenuated in the doses of 5, 10 and 20 mg/kg (Figure 2), suggesting that, at least in part, the bradycardia induced by EOCC appears to involve components of central nervous system.

Interestingly, despite the bradycardia has been completely abolished by atropine and attenuated by sodium thiopental, the hypotension induced by the doses of 5 and 10 mg/kg was not affected. Thus, the hypotension does not appear to be caused by decrease of cardiac output due to intense bradycardia and a possible decrease of the peripheral vascular resistance may also is contributing to the expression of this effect.

A possible role of the NO and PGI2 in the EOCC-induced hypotension was investigated. In animals treated with l-NAME (20 mg/kg), a nitric oxide (NO) synthase inhibitor (Moncada & Higgs, 1993) or INDO (5 mg/kg), an inhibitor of cyclooxygenase (COX) (Furchgott & Zawadzki, 1980), the hypotension and bradycardia were not significantly altered (Figure 1B), suggesting that NO and PGI2 appear not be participating of these effects.

To reinforce the hypothesis of that hypotensive response could be due to a decrease in peripheral vascular resistance caused by a possible vasorelaxation, we performed experiments in isolated rings of the rat superior mesenteric artery. In rings with intact endothelium, EOCC (1-3000 µg/mL) produced relaxations in concentration-dependent manner of tension induced by 10 µM of Phe (pD2 = 2.52±0.10; Emax = 103±10%; n = 6). These vasorelaxations were not different of those obtained in the denuded-endothelium rings (pD2 = 2.34±0.15; Emax =107±4%; n = 6) (Figure 3A), suggesting that this effect appears not to be mediated by endothelium.

These initial in vitro findings are in agreement with those obtained in the in vivo experiments, which demonstrated that hypotension induced by the EOCC appears not to be mediated by endothelial factors, such as NO or PGI2, but appears be caused by a vasodilatation. Furthermore, our results are analogous to others studies that had demonstrated that several essential oils present also a potent hypotensive effect through a direct vasorelaxant effect (Lahlou et al., 2005; Guedes et al., 2004).

It well known that the maintenance of smooth muscle contraction depends on Ca2+ influx from extracellular space through voltage and/or receptor operated calcium channels (VOCC and/or ROCC, respectively) (Karaki & Weiss, 1988). It is well also reported that the increase of external K+ concentration (KCl 80 mM) induces smooth muscle contraction through VOCC activation and subsequent calcium release from sarcoplasmic reticulum (Karaki & Weiss, 1988). The high K+-induced contraction is inhibited by Ca2+ channel blockers or by removal of external Ca2+. This contraction is, therefore, entirely dependent Ca2+ influx (Karaki & Weiss, 1988). Thus, we evaluated the EOCC effect on endothelium-denuded rings pre-contracted with Ca2+-depolarizing solution (KCl 80 mM). This set of experiments revealed that EOCC also induce vasorelaxations similarly to those obtained in rings pre-contracted with phenylephrine (pD2 = 2.04±0.12; Emax =101±7%; n = 6) (Figure 3B), which suggest that EOCC appears to be aging through the inhibition of the Ca2+ influx through VOCC.

Furthermore, in preparations without endothelium pre-contracted with Phe, EOCC (1000 mg/mL) did not induce an additional effect on the maximal vasorelaxation of NIF (10 mM). Interestingly, the same was observed when NIF (10 mM) was added on the maximal vasorelaxation of the EOCC (1000 mg/mL) (Figure 4), demonstrating that EOCC could be acting through a similar pathway of NIF, i.e., blocking dihydropyridine sensitive L-type VOCC (Hagiwara et al., 1993). Although the results show a majority participation of the Ca2+ channels, the possible involvement of K+ channels can not be discarded.

According to the literature, potassium channels are the dominant ion conductive pathways in vascular

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**Figure 2.** Hypotensive and bradycardic responses induced by in bolus administration of EOCC (1, 5, 10 and 20 mg/kg, i.v.) in non-anesthetized rats before and after sodium thiopental anesthesia (45 mg/kg, i.p., 30 min). Values are mean±SEM of six experiments. The data were analyzed with repeated measures two-way ANOVA followed by Bonferroni post-test. *p<0.05, **p<0.01 and ***p<0.001 vs Control.
A non-selective blocker of these channels (Cook, 1989). In this condition, EOCC was able to induce relaxations ($pD_2 = 2.18\pm0.08$; $E_{\text{max}} = 105\pm4\%$; $n = 6$) that was not significantly different of the control ($pD_2 = 2.34\pm0.15$; $E_{\text{max}} = 107\pm4\%$; $n = 6$) (Figure 3B). This result suggests that $K^+$ channels do not seem to be involved in the vasorelaxant effect induced by the EOCC.

Take together, these results demonstrate that the EOCC induces hypotension associated with bradycardia in muscle cells. The electrochemical gradient for $K^+$ ions is such that opening of $K^+$ channels results in diffusion of this cation out of the cells and membrane hyperpolarization. This effect closes Ca$^{2+}$ channels and leads to vasodilatation (Jackson, 2000).

Thus, the participation of the $K^+$ channels in the EOCC-induced vasorelaxation was investigated by using rings without functional endothelium, pre-contracted with Phe (10 $\mu$M), in the absence or presence of 100 $\mu$M of TEA, a non-selective blocker of these channels (Cook, 1989). In this condition, EOCC was able to induce relaxations ($pD_2 = 2.18\pm0.08$; $E_{\text{max}} = 105\pm4\%$; $n = 6$) that was not significantly different of the control ($pD_2 = 2.34\pm0.15$; $E_{\text{max}} = 107\pm4\%$; $n = 6$) (Figure 3B). This result suggests that $K^+$ channels do not seem to be involved in the vasorelaxant effect induced by the EOCC.

Figure 3. Vasorelaxant effect of EOCC (1-3000 $\mu$g/mL, cumulatively) or vehicle in rings of rat superior mesenteric artery: A. pre-contracted with Phe (10 $\mu$M) in the control condition (with endothelium) and after removal of endothelium; B. without endothelium pre-contracted with Phe (10 $\mu$M), pre-contracted with KCl 80 mM or pre-contracted with Phe (10 $\mu$M) after incubation with TEA (0.1 mM, 30 min). Values are mean±SEM of six experiments. The data were analyzed with one-way ANOVA followed by Bonferroni post-test.
normotensive non-anaesthetized rats. The hypotension appears to be caused by a decrease in peripheral vascular resistance, while bradycardia seems to be due to an activation of cardiac muscarinic receptors, involving, in part, compounds of central nervous system. Furthermore, the EOCC induces vasorelaxation in rat mesenteric artery possibly due to an inhibition of the Ca\(^{2+}\) influx through voltage-operated Ca\(^{2+}\) channels. This plant seems presents a potential clinical use for hypertension treatment, however, further studies are necessary to evaluate it safety and therapeutic margin before the human use.

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