Seroepidemiology of *Sarcocystis neurona*, *Toxoplasma gondii* and *Neospora* spp. among horses in the south of the state of Minas Gerais, Brazil

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Abstract

The present study used the indirect fluorescent antibody test (IFAT) to determine the seroprevalence of *Sarcocystis neurona*, *Toxoplasma gondii* and *Neospora* spp., and evaluated the variables associated with these infections among 506 apparently healthy horses, reared in the south of the state of Minas Gerais, Brazil. This study was conducted between April 2012 and October 2013. Among the horses, the true prevalence of *S. neurona* was 26% (95% CI: 22.0-30.4%), *T. gondii* 19.9% (95% CI: 15.5-24.8%) and *Neospora* spp. 23.9% (95% CI: 19.9-28.1%); and among the farms, 88.3% (95% CI: 70.7-96.1%), 71.6% (95% CI: 41-92.8%) and 85% (95% CI: 70.7-96.1%), respectively. Regarding mixed infection, 17 horses (3.4%) were seropositive for both *S. neurona* and *T. gondii*, 16 (3.2%) for *T. gondii* and *Neospora* spp. and 14 (2.8%) for *S. neurona* and *Neospora* spp. The associations between seropositivity and variables relating to the structure of the farm, management and health were analyzed using the logistic regression analysis, through the generalized estimating equations (GEE). The results suggest that the south of Minas Gerais is an enzootic area for *S. neurona*, *T. gondii* and *Neospora* spp. among horses, with prevalence of asymptomatic subclinical or chronic infections.

Keywords: Apicomplexa, Sarcocystidae, indirect immunofluorescence, risk factors.
Introduction

Brazil possesses one of the largest horse herds in the world, with approximately 5.3 million head. Horse-rearing is an important source of income and employment. In the southern region of the state of Minas Gerais, horse rearing occupies an important space in the local economic, touristic, historic and cultural scene, since this was the birthplace of the Mangalarga Marchador horse breed, which is internationally admired (IBGE, 2013).

Equine protozoal myeloencephalitis, caused by *Sarcocystis neurona*, is a disease characterized by a neurological condition in which horses participate as an accidental host. In Brazil, the seroprevalence of *S. neurona* among horses has been estimated in 37.7% (Western blot) (LINS et al., 2012) and 33.7% (ELISA) (PIVOTO et al., 2014) in Rio Grande do Sul, and 8.75% (IFAT) in Rio de Janeiro (STELMANN, 2014). Worldwide seroprevalence varies between 0% and 89.2% (DUBEY et al., 2015).

Toxoplasmosis, the disease caused by *Toxoplasma gondii*, is a zoonosis of global interest due to its impact on public health. It can be transmitted through consumption of raw or undercooked meat, including horse meat (DUBEY & SU, 2009). The levels of antibodies for *T. gondii* among horses in Brazil has been estimated in 12.8% (IFAT) in Minas Gerais (NAVES et al., 2005), 2.5% (IFAT) in Mato Grosso do Sul (LASKOSKI et al., 2015) and 28.4% (IFAT) in states of Alagoas, Paraíba, Pernambuco, Rio Grande do Norte and Minas Gerais (GENNARI et al., 2015). Worldwide, seroprevalence varies between 0% and 72.2% (ALSHAHERY & MANSOUR, 2012; MATSUO et al., 2014).

Neosporosis is a parasitic disease caused by the protozoon *Neospora caninum*, which can infect several mammals, including horses, and by *N. hughesi*, which infects horses and is an uncommon etiological agent for equine protozoal myeloencephalitis (GOODSWEN et al., 2013). In Brazil, the prevalence of anti-*Neospora* spp. antibodies among horses has been estimated in 57.6% (IFAT) in São Paulo (STELMANN et al., 2011), 25.7% (IFAT) in Santa Catarina (ABREU et al., 2014), and 1.87% (IFAT) in Rio de Janeiro (STELMANN, 2014) and, abroad, from 0% to 35% (DUBEY et al., 1999; PATITUCCI et al., 2004).

These three protozoa of the phylum Apicomplexa, family Sarcocystidae, are transmitted through consumption of water and food contaminated by sporocysts, in the case of *S. neurona*, and sporulated oocysts, in the cases of *T. gondii* and *Neospora* spp., which are released by the definitive hosts: respectively, poussums, felids and canids (LANGONI et al., 2007; DUBEY & SU, 2009; DUBEY et al., 2015). The infection is usually asymptomatic among horses, but may sporadically present nervous symptoms in the cases of *S. neurona*, *T. gondii* and *N. hughesi*, and reproductive disorders, when involving *T. gondii* and *N. caninum* (LOCATELLI-DITTRICH et al., 2006; GENNARI et al., 2015).

Because of the importance of horse-rearing in the south of the state of Minas Gerais, determining the prevalences of *S. neurona*, *T. gondii* and *Neospora* spp., along with the factors associated with these infections, is fundamental for implementing effective control measures, so as to reduce the exposure of the animals and prevent the economic losses caused by these parasites. Moreover, this type of investigation increases the knowledge on the distribution of these agents among the horse population nationally.

Thus, due to the scarcity of studies in Brazil and lack of information regarding equine infection in the southern part of the state of Minas Gerais, the present study had the objectives of determining the prevalence of *S. neurona*, *T. gondii* and *Neospora* spp., and identifying possible factors associated with seropositivity among Mangalarga Marchador horses.

Materials and Methods

Study area and animals

Blood samples were gathered from 506 Mangalarga Marchador horses that had been reared on 53 farms that were distributed among 27 municipalities in the microregions of Andrelândia, Lavras, São João Del Rey, São Lourenço and Varginha, in the southern portion of the state of Minas Gerais (Figure 1). The study was conducted between April 2012 and October 2013 after approval had been obtained from the Ethics Commission for Animal Use (COEA).
of the Federal University of Lavras (UFLA), and was registered at this institution under number 023/11.

The farms were chosen randomly based on guidance from the Brazilian Association of Mangalarga Marchador Horse Breeders (Associação Brasileira de Criadores de Cavalos Mangalarga Marchador; ABCCM). In this sampling, the proportions between the mesoregions and the sizes of farms and their structures were maintained in order to attain homogeneous representativeness. At least ten horses from each farm were required for the prevalence calculation on S. neurona, T. gondii and Neospora spp. In addition, pregnant mares and foals of less than six months of age were not included in the sampling. The reasons were that pregnant mares are more sensitive to manipulation, while foals younger than six months may present colostral antibodies (passive immunity), which would interfere with the serological diagnosis. The blood samples were obtained by means of jugular venipuncture and were collected in Vacutainer tubes. All the animals appeared to be healthy. After centrifugation at 500 g for 10 minutes, the serum samples from 506 horses were stored in individually identified cryotubes and frozen at −20 °C until the serological tests were performed.

The definition of the number of horses (n) needed to estimate the prevalence was based on the formula of the Pan-American Zoonosis Center (CEPANZO, 1979), which is appropriate for calculating samples for research on chronic diseases: \[ n = \left( \frac{p (100-p) \times Z^2}{d^2 \times p/100} \right) \]. Considering an estimated prevalence (p) of 50%, a 95% confidence interval (z) of 1.96 and an acceptable error margin (d) of 10%, the minimum number (n) of horses was 384.

**Information-gathering on the farms**

A questionnaire-based interview was carried out with either the owners or the person responsible for each farm. The questions related to herd health, the breeder, management and the structure of the farm. Moreover, questions asking about any history of reproductive problems and the neurological condition of the herds were also devised. This information served as independent variables for subsequent analyses that evaluated the associations between seropositivity to the protozoa and factors relating to sociocultural information, herd health and management practices observed on the farms. In total, 305 variables were analyzed, which were obtained by means of the questionnaire.

**Serological tests**

The indirect fluorescent antibody test (IFAT) was the method chosen for investigating anti-S. neurona, T. gondii and Neospora spp. IgG antibodies. This is considered to be the gold-standard test for serologically diagnosing the latter two protozoa (LANGONI et al., 2007; DUBEY & SCHARES, 2011).

In investigating anti-S. neurona antibodies, the IFAT was performed as described by Johnson et al. (2013). The antigens were S. neurona merozoites, produced in cell cultures. Ten microliters of S. neurona merozoite solution was placed in each one of the 12 slide wells for IFAT (about 10 merozoites / field at 40x magnification in an optical microscope). After slides had been dried at room temperature, they were stored in a freezer at −20 °C until the time of use. The IFAT for T. gondii was performed using the technique described by Langoni et al. (2007) and the antigens were the tachyzoites of the parasite. After successive inoculations, the tachyzoites were recovered through rinsing the peritoneal cavity of mice with a saline solution. Ten microliters of a solution containing tachyzoites was placed in each of the 12 slide wells for IFAT (about 10 tachyzoites / field at 40x magnification in an optical microscope). After the slides had been dried at room temperature, they were stored in a freezer at −20 °C until the time of the serological tests. In the present study, N. caninum tachyzoites were used as the antigen in the IFAT, and their antibodies presented cross-reactions with N. hughesi (GONDIM et al., 2009). The IFAT for Neospora spp. was performed in accordance with the technique described by Paré et al. (1995), using glass slides containing tachyzoites of N. caninum as the antigen (Laboratório Imunodot, Jaboticabal, Brazil).

An equine anti-IgG conjugate (SIGMA, Saint Louis, MO, USA) was used for performing the IFAT, at a dilution of 1:64, along with positive (serum samples of mares seropositive to S. neurona, T. gondii and Neospora spp.) and negative (serum samples of newborn foal before colostrum ingestion and serum samples of periparturient mare, both seronegative to respective protozoa) controls for the respective protozoa. The serum samples were considered positive if they presented complete fluorescence of the S. neurona merozoites at a dilution of 1:80 (DUBEY et al., 2015), and of the T. gondii and Neospora spp. tachyzoites, respectively, at dilutions of 1:64 (ABREU et al., 2014) and 1:50 (LOCATELLI-DITTRICH et al., 2006). Both the IFAT and the preparation of slides containing S. neurona and T. gondii antigens were performed at the Parasite Disease Laboratory of the Federal University of Lavras (UFLA).

**Statistical analyses**

The EpiData software was used to tabulate the data relating to the variables studied. These data were later ranked and analyzed using the statistical software SPSS 20.0 statistical software (IBM, 2011). In calculating true prevalence (TP) for the anti-S. neurona antibodies, values of 94% and 88.9% were used for sensitivity (SE) and specificity (SP), respectively, as prescribed by Duarte et al. (2003). In the case of TP for T. gondii, the SE and SP values were, respectively, 90.9% and 100% (LU et al., 2013). In turn, TP for Neospora spp. presented values of 91.0% and 99.0%, respectively, for SE and SP, in accordance with Björkman & Uggl (1999).

Subsequently, the SE and SP between the herds were used to calculate the TP at herd level (NOORDHUIZEN et al., 2001), by means of the epidemiological calculators of the Epitools software (SERGEANT, 2011). The variables relating to the structural, sanitary and management characteristics of the farms, along with the inherent characteristics of the animals, were firstly subjected to the chi-square test or Exact Test of Fisher (less than five observations in any category) using the SPSS 20.0 software (IBM, 2011), so as to demonstrate their association with the seropositivity of the studied agents. This association, when statistically significant, was quantified by the odds ratio and its confidence interval at 95% (CI 95%), which estimates the risk relationship between
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Results

Farm profile

Among the 506 animals analyzed, 343 were female (67.8%) and 163 were male (32.2%). There were 61 foals (12.1%) and 445 adults (87.9%). Among the 53 farms, horse-rearing was the main activity on only 21 of them (39.6%). Regarding size, 27 farms (51.0%) covered more than 100 ha, with a mean area of 199.1 ha. Forty-eight farms (90.6%) had at least one seropositive horse, with TP of 85% (95% CI: 70.7-96.1%), with at least one positive horse on 45 farms (84.9%). Interaction between the different species of animals occurred in 39 (73.6%) herds, to a greater or lesser degree. Regarding the farming system, 36 farmers (67.9%) maintained their animals exclusively in picket-fenced enclosures. Considering food management, 47 farms (88.7%) used grass and also offered feed to the animals, among which 21 (44.7%) manufactured the concentrate on-site. Regarding sanitary management, 50 farmers (94.3%) vaccinated their animals, and 31 (58.5%) had veterinary assistance. Most of the farms used artificial insemination techniques (62.3%). Horse-rearing was the main source of income on 13 farms (24.5%), and 33 (62.3%) had been carrying this activity for over 20 years.

Seroprevalence and variables associated with infection

Sarcocystis neurona. Out of the 506 horses, 117 (23.0%) presented anti-S. neurona antibodies, with true prevalence (TP) of 26.0% (95% CI: 22.0-30.4%). Out of the 53 properties analyzed, 44 had at least one seropositive horse, with TP of 88.3% (95% CI: 74.4-91.6%). As shown in Table 1, the properties that included forest reserves or that bordered a reserve presented a high risk (p < 0.05) of having horses seropositive for S. neurona. Farms that manufactured their own feed also had greater risk (p < 0.05) of having infected horses.

Toxoplasma gondii. In the case of T. gondii, 107 (21.1%) horses were seropositive, representing a TP of 19.9% (95% CI: 15.5-24.8%). On 45 farms, at least one horse was seropositive, indicating a TP of 71.6% (95% CI: 41.0-92.8%). Farms that had more than 100 horses presented lower risk (p < 0.05) of having infected horses. Farms that vaccinated their horses against tetanus and influenza, and those that had an area for treating, cleaning and caring for the animals, presented lower risk (p < 0.05) of infection. Horses over the age of five years were significantly (p < 0.05) more liable to infection by T. gondii (Table 2). There was a significant association (p < 0.05) between the use of embryo transferece and seropositivity, both through the Exact Test of Fisher or chi-square test and through the multivariate analysis using the generalized estimating equations (GEE). In this latter test, horses on farms that did not use embryo transferece had twice as much risk of infection (95% CI: 1.086-3.003) as farms that did use this technique (p = 0.023).

Neospora spp. The TP for Neospora spp. was 23.9% (95% CI: 19.9-28.1%), with 105 seropositive horses. On the farms, the TP was 85% (95% CI: 70.7-96.1%), with at least one positive animal on 41 farms. Farms in the southern part of the state of Minas Gerais that offered hay to their horses presented lower risk

Table 1. Epidemiological variables associated with antibodies for S. neurona (IFAT:1:80), in the univariate analysis (P < 0.05), among horses reared in the southern part of the state of Minas Gerais, Brazil.

<table>
<thead>
<tr>
<th>Factors</th>
<th>Sarcocystis neurona</th>
<th>Odds ratio (OR)</th>
<th>95% CI</th>
<th>P*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Positive</td>
<td>Negative</td>
<td>n</td>
<td>%</td>
</tr>
<tr>
<td>Feed manufactured on the farm (n = 47**)</td>
<td>13</td>
<td>92.59</td>
<td>25</td>
<td>7</td>
</tr>
<tr>
<td>No</td>
<td>13</td>
<td>65.00</td>
<td>7</td>
<td>35.00</td>
</tr>
<tr>
<td>Yes</td>
<td>25</td>
<td>92.59</td>
<td>2</td>
<td>7.41</td>
</tr>
<tr>
<td>Farm includes or borders a forest reserve (n = 53)</td>
<td>2</td>
<td>33.33</td>
<td>3</td>
<td>66.67</td>
</tr>
<tr>
<td>No</td>
<td>2</td>
<td>33.33</td>
<td>3</td>
<td>66.67</td>
</tr>
<tr>
<td>Yes</td>
<td>42</td>
<td>87.50</td>
<td>6</td>
<td>12.50</td>
</tr>
</tbody>
</table>

IFAT: indirect fluorescent antibody test; CI: confidence interval. *Exact Test of Fisher (P < 0.05). **Six farms did not use feed. There was no significant association (P > 0.05) between seropositivity and the following variables: sex; age; keeping horses in stables; and purpose of horse-rearing.
(p < 0.05) of having animals seropositive for *Neospora* spp. Farms on which the horses were in contact with other animals, or in which horses over the age of five years were present, presented greater risk (p < 0.05) of infection (Table 3). Horses on farms on which cattle and cats were also present showed greater risk (p < 0.05) of being infected by *Neospora* spp., from the multiple logistic regression through the generalized estimating equations (GEE) (Table 4).

### Mixed infection

Mixed infection by *S. neurona* and *T. gondii* occurred in 3.4% (17/506) of the horses, while 3.2% (16/506) were positive for both *T. gondii* and *Neospora* spp. Horses infected by *S. neurona* and *Neospora* spp. represented 2.8% (14/506) of the sample. Infection involving the three agents occurred among only 1.4% (7/506) of animals. In 11.3% (6/53) of the farms there was positivity for all

### Table 2. Epidemiological variables associated with antibodies for *T. gondii* (IFAT 1:64), in the univariate analysis (P < 0.05), among horses reared in the southern part of the state of Minas Gerais, Brazil.

<table>
<thead>
<tr>
<th>Factors</th>
<th>Positive</th>
<th>Negative</th>
<th>Odds ratio (OR)</th>
<th>95% CI</th>
<th>P*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total number of horses on the farm (n = 53)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>0.033</td>
</tr>
<tr>
<td>&gt;100 animals</td>
<td>6</td>
<td>4</td>
<td>40.00</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>&lt;100 animals</td>
<td>39</td>
<td>4</td>
<td>9.31</td>
<td>6.493</td>
<td>1.272-33.333</td>
</tr>
<tr>
<td>Vaccine against tetanus (n = 53)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>0.038</td>
</tr>
<tr>
<td>Yes</td>
<td>3</td>
<td>3</td>
<td>50.00</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>No</td>
<td>39</td>
<td>4</td>
<td>89.36</td>
<td>10.64</td>
<td>8.403</td>
</tr>
<tr>
<td>Vaccine against influenza (n = 53)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>0.014</td>
</tr>
<tr>
<td>Yes</td>
<td>12</td>
<td>6</td>
<td>66.66</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>No</td>
<td>33</td>
<td>5</td>
<td>94.28</td>
<td>5.72</td>
<td>8.264</td>
</tr>
<tr>
<td>Use of embryo transfer (n = 53)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>0.017</td>
</tr>
<tr>
<td>Yes</td>
<td>17</td>
<td>7</td>
<td>70.83</td>
<td>29.17</td>
<td>1</td>
</tr>
<tr>
<td>No</td>
<td>28</td>
<td>1</td>
<td>96.15</td>
<td>3.85</td>
<td>8.771</td>
</tr>
<tr>
<td>Use of a treatment, cleaning and care area for the animals (n = 53)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>0.050</td>
</tr>
<tr>
<td>Yes</td>
<td>20</td>
<td>7</td>
<td>74.07</td>
<td>25.93</td>
<td>1</td>
</tr>
<tr>
<td>No</td>
<td>25</td>
<td>1</td>
<td>96.15</td>
<td>3.85</td>
<td>8.771</td>
</tr>
<tr>
<td>Age of the animals (n= 485**)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>0.011</td>
</tr>
<tr>
<td>&lt; 5 years</td>
<td>36</td>
<td>185</td>
<td>16.29</td>
<td>83.71</td>
<td>1</td>
</tr>
<tr>
<td>&gt; 5 years</td>
<td>68</td>
<td>196</td>
<td>25.76</td>
<td>74.24</td>
<td>1.783</td>
</tr>
</tbody>
</table>

IFAT: indirect fluorescent antibody test; CI: confidence interval. *Exact Test of Fisher and chi-square test (P < 0.05). **The ages of 21 horses were not informed.

There was no significant association (P > 0.05) between seropositivity and the following variables: presence of cats; rearing purpose and system; sex; and rearing of horses and cattle together.

### Table 3. Epidemiological variables associated with antibodies for *Neospora* spp. (IFAT 1:50), in the univariate analysis (P < 0.05), among horses reared in the southern part of the state of Minas Gerais, Brazil.

<table>
<thead>
<tr>
<th>Factors</th>
<th>Positive</th>
<th>Negative</th>
<th>Odds ratio (OR)</th>
<th>95% CI</th>
<th>P*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Supply of hay (n= 53)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>0.045</td>
</tr>
<tr>
<td>Yes</td>
<td>13</td>
<td>8</td>
<td>61.90</td>
<td>38.10</td>
<td>1</td>
</tr>
<tr>
<td>No</td>
<td>28</td>
<td>12</td>
<td>87.50</td>
<td>12.50</td>
<td>4.310</td>
</tr>
<tr>
<td>Contact between horses and other animals (n= 53)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>0.008</td>
</tr>
<tr>
<td>No</td>
<td>7</td>
<td>7</td>
<td>50.00</td>
<td>50.00</td>
<td>1</td>
</tr>
<tr>
<td>Yes</td>
<td>34</td>
<td>5</td>
<td>87.18</td>
<td>12.82</td>
<td>6.800</td>
</tr>
<tr>
<td>Age of the animals (n= 485**)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>0.034</td>
</tr>
<tr>
<td>&lt; 5 years</td>
<td>37</td>
<td>184</td>
<td>16.74</td>
<td>83.26</td>
<td>1</td>
</tr>
<tr>
<td>&gt; 5 years</td>
<td>65</td>
<td>199</td>
<td>24.62</td>
<td>75.38</td>
<td>1.624</td>
</tr>
</tbody>
</table>

IFAT: indirect fluorescent antibody test; CI: confidence interval. *Exact Test of Fisher and chi-square test (P < 0.05). **The ages of 21 horses were not informed. There was no significant association (P > 0.05) between seropositivity and the following variables: presence of dogs on the property; sex of the animals; and occurrence of either reproductive or neurological disorders in the equine herds.
three agents, and all farms presented horses infected by at least one protozoan. However, 47.2% (239/506) of the horses were seronegative for all three parasites.

**Discussion**

**Farm profile**

Most of the farms presented interaction between different animals, mostly between horses and cattle and/or dogs, which were the two predominant species on these farms. Moreover, the herds were mostly located in pastures and near woodlands, thus allowing contact with wild fauna and, consequently, favoring exposure of the horses to the parasites studied in this investigation (ABREU et al., 2014; DUBEY et al., 2015). Most of the horses were reared for commercial purposes, thus allowing a large flow of animals, which favors dispersion of the agents (MOURA et al., 2013). The majority of the farms showed good sanitary practices, such as vaccinating the animals, sanitizing the installations, storing food adequately and having veterinarian assistance. This demonstrated the owners’ concern for caring for the herd’s health, which could have influenced the seroprevalence rates of the agents analyzed and the absence of clinical symptomatology among the animals.

**Seroprevalence and variables associated with infection**

*Sarcozystis neurona*. The prevalence of *S. neurona* among horses (26.0%) was within the range observed in previous studies conducted in Brazil, 37.7% (Western blot) in Rio Grande do Sul (LINS et al., 2012) and 8.3% (IFAT) in Rio de Janeiro (STELMANN, 2014), and worldwide, same 26.1% (Western blot) in Argentina (MORE et al., 2014) and 27.6% (IFAT) in EUA (PUSTERLA et al., 2014). The high prevalence among herds (88.3%) can be explained by the fact that most horses were reared in picket-fenced enclosures, thus facilitating their contact with wild fauna, which predisposed the animals towards infection by *S. neurona* (MORLEY et al., 2008). In addition, the presence of woodlands near the pastures in the absolute majority of the farms (90.57%), probably increased the risk of contact between horses and wild animals, such as possums (DUBEY et al., 2015).

The possum (*Didelphis albiventris*) is a broadly distributed animal in South America for which primary and secondary woodlands are its natural habitat (ORR, 1986). Given its high synanthropy, this species is a potential disseminator of pathogens between domestic animals and man (MULLER et al., 2005). This would explain the association between the presence of forest reserves either on the farms or bordering the farms and seropositivity for *S. neurona*, which has also been reported in other studies (MORLEY et al., 2008; DUBEY et al., 2015). The greater risk of infected animals found on farms that manufactured their own feed suggests that the storage of the raw material used in the formulation was inadequate, thus allowing access by wild animals (possums) to the food (MORLEY et al., 2008).

*Toxoplasma gondii*. The prevalence of anti-*T. gondii* antibodies among the horses (19.9%) was within the range observed in serological studies carried in Brazil, 31.5% (IFAT) in states of Mato Grosso, Mato Grosso do Sul, Paraná and São Paulo (VIDOTTI et al., 1997), and 2.1% (IFAT) in Rio de Janeiro (STELMANN, 2014), and same 17.7% (IFAT) in Paraná (FINGER et al., 2013), and abroad, 2.6% (IFAT) in South Korea (GUPTA et al., 2002) and 40.5% (IFAT) in Egypt (GHAZY et al., 2007). The high prevalence among farms (71.6%) indicated that there was wide dispersion of the parasite in the region. Thus, we can infer that positive farms (those with at least one seropositive equine) had potential risk factors for infection with *T. gondii*. Considering the intense movement of horses between farms, as most farms had animals that participated in commercial events, which may have predisposed the animals towards infection by *T. gondii*, as suggested by Kouam et al. (2010).

In the present study, the lower risk of seropositivity for *T. gondii* that was found on farms with more than 100 horses can be partly explained by the fact that these larger farms were technologically driven and their owners were better informed. On the other hand, the precarious manner of storing the food offered to the horses (DUBEY & SU, 2009), as well as environmental conditions favorable towards development and survival of parasites (KOUM et al., 2010), helps to explain the greater risk of contamination of horses on small farms.

Although there was no direct relationship with the form of transmission of *T. gondii*, the lower risk of seropositivity observed in farms that had their animals vaccinated against tetanus and

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**Table 4. Epidemiological variables associated with antibodies for Neospora spp. (IFAT 1:50), in the multiple logistic regression analysis using the generalized estimate equations, among horses reared in the southern part of the state of Minas Gerais, Brazil.**

<table>
<thead>
<tr>
<th>Variables</th>
<th>Neospora spp.</th>
<th>Adjusted Odds Ratio (OR)</th>
<th>CI 95% (OR)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Positive</td>
<td>Negative</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Presence of cattle (n= 53)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>No</td>
<td>2</td>
<td>3</td>
<td>40.00</td>
<td>60.00</td>
</tr>
<tr>
<td>Yes</td>
<td>39</td>
<td>9</td>
<td>.8125</td>
<td>18.75</td>
</tr>
<tr>
<td>Presence of cats (n= 53)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>No</td>
<td>21</td>
<td>10</td>
<td>67.74</td>
<td>32.26</td>
</tr>
<tr>
<td>Yes</td>
<td>15</td>
<td>1</td>
<td>93.75</td>
<td>6.25</td>
</tr>
</tbody>
</table>

IFAT: indirect fluorescent antibody test; CI: confidence interval. QIC (Quasilikelihood under the Independence model Criterion): 448.2; QICC: 446.4 (The QIC and QICC was used as GEE logistic regression goodness of fit measure, considering that when QICC approximates QIC the GEE model is correctly specified and that the model with the smaller QIC and QICC statistics was preferred).
influenza, or that used a specific area to care for the animals, allows the inference that the owners are concerned about the health of their animals. Thus, use of hygienic care and sanitary measures, including those that are protection factors against other agents and not directly against *T. gondii*, indirectly contribute towards reducing the risk of exposure or preventing immunosuppressant coinfections, which would be able to increase the risk of infection though this parasite (TASSI, 2007).

Greater seropositivity among horses over the age of five years can be explained by the fact that older animals have had more chances of becoming exposed to *T. gondii* over the course of their lives and, consequently, of becoming infected and producing antibodies against this protozoon.

The farms in the south of the state of Minas Gerais that used embryo transfer techniques presented lower risk of horses becoming infected by *T. gondii*. Embryo transfer is a modern form of animal reproduction technology and is not directly related to transmission of *T. gondii*. However, we can infer that these properties were more technologically-oriented, with use of good hygiene-sanitary practices capable of controlling various illnesses on the farm, and that either directly or indirectly, these practices contributed towards reducing the risk of horse infections by *T. gondii*. Thus, these farms were better prepared to implement preventive measures against *T. gondii*, such as safe storage of food and hygiene of installations (DUBEY & SU, 2009). Thus, these farms avoided environmental conditions that would favor development and survival of parasite oocysts, such as warm and humid locations (LANGONI et al., 2007; KOUAM et al., 2010).

Although the presence of cats on the farm is a risk factor (DUBEY & SU, 2009), this situation was not observed in the present study, as also reported by Stelmann (2014). The presence of seronegative cats on the farms and/or intense traffic of animals in the region could explain the absence of this correlation (LANGONI et al., 2007). There was no significant association between sex, rearing purpose or rearing system (stable or pasture) and seropositivity, and thus our results were in agreement with those of Stelmann (2014), although in disagreement with those of Kouam et al. (2010).

*Neospora* spp. The seroprevalence of *Neospora* spp. (23.9%) among the horses was within the range observed in other studies (IFAT) carried out in Brazil, 57.6% (IFAT) in São Paulo (STEMMANN et al., 2011) and 4.1% (IFAT) in Santa Catarina (MOURA et al., 2013), and same 25.7% (IFAT) in Santa Catarina (ABREU et al., 2014), and abroad, 8.9% (IFAT) in Israel (KLGHER et al., 2007) and 28.0% (IFAT) in Italy (CIARAMELLA et al., 2004), and same 24.0% (ELISA) in Czech Republic (BÁRTOVÁ et al., 2010). The wide distribution of the infection among the farms (85.0%) was probably due to the environmental conditions of the southern part of the state of Minas Gerais, typical of Brazilian tropical regions, which favor sporulation of *Neospora* spp. oocysts (MOURA et al., 2013).

The association between seropositivity for *Neospora* spp. and “no supply of hay” suggests that farms that are able to provide hay for their horses are more technologically-oriented and their owners are better informed. Thus, these owners take greater care regarding storage and supply of food, thereby minimizing the risk of animal infection. Moreover, hay is a source of roughage, which indicates that on the farms where hay is supplied as food, there is greater confinement of the animals in stalls and lower contact with pastures. Thus, there is a lower risk of contact between the horses in stables and the definitive hosts (ABREU et al., 2014).

Greater positivity of *Neospora* spp. among horses above the age of five years suggests that horizontal transmission predominates, considering that only 16.7% (37/221) of the animals below the age of five years were infected. This lower efficiency of congenital transmission among horses, in comparison with cattle, can be partly explained by placentation differences (PITEL et al., 2003).

In the south of the state of Minas Gerais, horse farms with the presence of cattle presented a significant association with infection by *Neospora* spp. These results corroborate the findings of Moura et al. (2013) and Abreu et al. (2014), thus suggesting that cattle are able to maintain the agent on the farms, since dogs commonly have access to bovine meat and their placental remains.

The significant association between seropositivity for the agent and the presence of cats in the farms suggests that there is greater presence of rodents on these farms. These animals can act as intermediate hosts for *Neospora* spp. and are able to maintain the parasite's cycle on the farm and infect dogs, which due to their carnivorous habit, hunt these rodents. A similar result was observed by Stelmann (2014), who correlated equine neosporosis with access of cats to food. Although there are studies that have correlated infection by *Neospora* spp. with the presence of dogs and contact between them and horses (MOURA et al., 2013), this was not shown on these farms in the southern part of the state of Minas Gerais. This result may suggest that the dogs were probably not infected, or were not in the habit of feeding off placental remains and tissues of the animals.

In the present study there was no significant association between sex and seropositivity to *Neospora* spp., which was also observed previously by Moura et al. (2013). Similarly, animals with a history of either reproductive or neurological disorders did not present any significant correlation with the infection. In the case of reproductive disorders, there was also no association with positivity in the study by Moura et al. (2013). However, correlations between reproductive alterations and infection by the parasite were reported by Locatelli-Dittrich et al. (2006) and Abreu et al. (2014).

**Conclusions**

The results from the present study allow it to be stated that the southern part of the state of Minas Gerais is an enzootic area for *S. neurona*, *T. gondii* and *Neospora* spp., with a wide distribution of these parasites among Mangalarga Marchador horse farms. However, asymptomatic subclinical or chronic infection predominates in the region. Horizontal transmission seems to be the main form of infection for these parasites, and different factors are associated with seropositivity among horses. More research is needed in order to establish precisely which factors increase the risk of transmitting these agents, as well as to determine which species of *Neospora* (*N. caninum* or *N. hughesi*) infects horses.
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