First report of *Trichodinella* and new geographical records of trichodinids in Nile tilapia (*Oreochromis niloticus*) farmed in Brazil

**Primeiro relato de *Trichodinella* e novos registros geográficos de tricodinídeos em tilapia do Nilo (*Oreochromis niloticus*) cultivada no Brasil**

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**Abstract**

Massive occurrence of trichodinids is frequently accompanied by serious disease in fish farms. In this study, trichodinid species from the gills and skin of Nile tilapia (*Oreochromis niloticus*) farmed in the central-western region of Brazil (state of Goiás) were morphologically characterized. Dried slides were prepared from the parasites and were impregnated with silver nitrate (2%). Morphometric characteristics were determined and schematic drawings of the denticles were made using photomicrographs produced from the slides. Seven species of trichodinid ectoparasites (Protozoa: Ciliophora: *Trichodinidae*) were found parasitizing the gills: four of the genus *Trichodina* Ehrenberg, 1838; one of *Tripartiella* Lom, 1959; one of *Paratrichodina* Lom, 1963; and one of *Trichodinella* Štrámek–Hušek, 1953. On the body surface, three specimens of the genus *Trichodina* were identified. This study presents new geographical records of trichodinids in Brazil, thus confirming that *Trichodina centrostrigeata*, *Trichodina compacta*, *Trichodina heterodentata*, *Paratrichodina africana* and *Tripartiella orthodens* are widely distributed worldwide. Additionally, the first record of the genus *Trichodinella* in Brazil is presented.

**Keywords:** Trichodinids, trichodiniasis, parasite, taxonomy, aquaculture.

**Resumo**


**Palavras-chave:** Tricodinídeos, tricodiniásise, parasita, taxonomia, aquicultura.
Introduction

The prevalence and spread of diseases have become more and more important since aquatic food production has transitioned from being primarily based on catching wild fish to farming of increasing numbers of fish species (FAO, 2016). Captive fish and their parasites are routinely translocated around the world (BASSON & VAN AS, 1994; REINERTSEN & HAALAND, 1995; VALLADÃO et al., 2014), which may affect the health status of fish farms. Therefore, studies focusing on identifying parasites and their distribution are important.

Trichodinids are mobile peritrichous ciliated protozoa that are important within marine and inland aquaculture. More than 300 species of trichodinids have been recognized as parasites or symbionts of aquatic organisms (GONG et al., 2005; MITRA et al., 2013; MACIEL et al., 2018). They are perhaps the most frequent protozoa invading the surface of fish and have been implicated in severe disease and mortality in various parts of the world (NIKOLIĆ et al., 2003; KHAN, 2004; KHOSHNOOD & KHOSHNOOD, 2014; VALLADÃO et al., 2014).

Considering the socioeconomic importance of Nile tilapia (Oreochromis niloticus) within Brazilian aquaculture and the economic impact of parasitic diseases, very little information on these groups is available. Only three out of the eleven genera of the family Trichodinidae have so far been reported parasitizing Nile tilapias in Brazil, namely Trichodina Ehrenberg, 1838 (Trichodina centrostriigata Basson, Van As and Paperna, 1983, T. compacta Van As and Basson, 1989, T. heterodentata Duncan, 1977, T. magna Van As and Basson, 1989, T. migala Van As and Basson, 1989), Paratrichodina Lom, 1963 (Paratrichodina africana Kazubski and El-Tantawy, 1986) and Tripartiella Lom, 1959 (Tripartiella orthodens Basson and Van As, 1987) (GHIRALDELLI et al., 2006; MARTINS & GHIRALDELLI, 2008; PANTOJA et al., 2012; VALLADÃO et al., 2013, 2016; ZAGO et al., 2014; NUNES et al., 2016).

Trichodinids are usually identified through the morphology of the denticles in the adhesive disc and the development of the adoral ciliary spiral, and the denticles have very high systematic value (GONG et al., 2005). Correct identification depends mainly on the ciliary spiral, and the denticles have very high systematic value (NIKOLIĆ et al., 2003; KHAN, 2004; KHOSHNOOD & KHOSHNOOD, 2014; VALLADÃO et al., 2014).

Materials and Methods

Study area and fish

Specimens of O. niloticus (Supreme strain) were collected in June 2016, from floating net-cages in a reservoir at the fishery facilities of the Department of Animal Science of the School of Veterinary and Animal Sciences of the Federal University of Goiás, State of Goiás, central-western region of Brazil (16° 35’ 42” S; 49° 16’ 52” W).

The water quality parameters in the net-cages were measured during the fish sampling as follows (mean ± SD): the dissolved oxygen was 7.09 ± 0.39 mg L⁻¹; the temperature was 20.75 ± 0.57 °C; the pH was 7.00 ± 0.22; and the ammonia content was 0.03 ± 0.015 mg L⁻¹. The dissolved oxygen and temperature were determined using a digital oxygen meter (AT 155; Alfakit Ltd.); the pH was determined using a digital pH meter (AT 315; Alfakit Ltd.) and the ammonia concentration was determined using a digital photo colorimeter (AT 100PBS II; Alfakit Ltd.).

The experimental procedures were approved by the Ethics and Animal Welfare Committee (CEUA) of the Federal University of Goiás, under protocol number 015/2016.

Parasite diagnosis and taxonomic evaluation

Eleven fish (13.62 g ± 1.78) were collected randomly from four net-cages (0.7 m³ useful volume and density of 300 fish per cage). They were desensitized through thermal shock in water with ice (proportion 1:1) and then euthanized via medullary sectioning to conduct a parasitological survey. Samples from the gills and body mucus were collected separately through scraping. The material thus collected was deposited on glass slides and was observed while fresh under an optical microscope (40, 100 and 400 x).

Slides containing parasites were air-dried and subsequently impregnated with silver nitrate (2%) for evaluation of all their taxonomic characteristics (KLEIN, 1958). Measurements of the components of the adhesive discs and denticles were made as described by Arthur & Lom (1984) and, additionally, the numbers of denticles and pins were counted. The measurements were made on photomicrographs (1000 x) that were obtained using a Nikon® E200 optical microscope (Nikon Instruments Inc., Melville, United States) equipped with a Motic® 5.0 image capture system (Motic Instruments Inc., Hong Kong, China). These measurements were presented in micrometers as suggested by Lom (1958) and Van As & Basson (1989). All the measurements were made using the
Image Pro Plus® software media (Cybernetics Inc, Rockville, United States). The measurement data thus obtained were presented as the mean ± standard deviation (with minimum and maximum and the number of repetitions).

In order to describe the shape of the denticle, schematic drawings of the denticles were produced as proposed by Van As & Basson (1989), by means of vectorization using the CorelDraw® X8 software (Corel Corporation, Ottawa, Canada).

**Results**

*Trichodinid description: from body*

The measurements of the taxonomic characteristics of each population of trichodinids collected from the body surface of the fish are presented in Table 1.

*Trichodina compacta*

The blade was large, filling most of the space between the y-axes. The anterior margin touched the y-axes and sometimes went slightly beyond it. A prominent apophysis was observed at the anterior margin of the blade. The central part extended to half of the space between the axes and had a rounded presentation. The connection between the central part and ray had uniform thickness and was similar to the ray. The rays were generally parallel to the y-axes with greater thickness, filling almost the entire space between the y-axes. The central circle was characteristic for the species, presenting scattered darker spots (Figure 1a).

*Trichodina heterodentata*

The blade was large and filled almost the entire space between the y-axes. The anterior portion of the blade went significantly beyond the limit of the y-axes. A prominent apophysis was common at the anterior margin of the blade. The central part extended to half of the space between the axes and had a rounded presentation. The ray had a prominent apophysis in its anterior

![Figure 1. Photomicrographs of silver impregnated adhesive discs and schematic drawing of the denticles of *Trichodina compacta* (a); *T. heterodentata* (b) and *T. magna* (c) found in the body of Nile tilapia *Oreochromis niloticus* cultured in Central-West region of Brazil. Scale bars: 10 μm.](image)

*Table 1. Measurements of trichodinids from body of Nile tilapia *Oreochromis niloticus*. The data are presented as arithmetic mean ± standard deviation (minimum-maximum values; number of individuals measured).*

<table>
<thead>
<tr>
<th>Trichodinids species</th>
<th><em>Trichodina compacta</em></th>
<th><em>Trichodina heterodentata</em></th>
<th><em>Trichodina magna</em>&lt;sup&gt;*&lt;/sup&gt;</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body diameter</td>
<td>54.61 ± 3.39 (48.9-61.4; 22)</td>
<td>70.10 ± 6.15 (60.2-79.8; 18)</td>
<td>91.86</td>
</tr>
<tr>
<td>Border membrane</td>
<td>4.57 ± 0.47 (3.4-5.2; 22)</td>
<td>5.37 ± 0.70 (4.3-7.7; 18)</td>
<td>7.86</td>
</tr>
<tr>
<td>Adhesive disc</td>
<td>45.41 ± 3.30 (40.0-53.2; 22)</td>
<td>59.41 ± 6.14 (49.1-69.2; 18)</td>
<td>75.79</td>
</tr>
<tr>
<td>Denticulate ring</td>
<td>28.19 ± 2.71 (23.5-34.4; 22)</td>
<td>37.61 ± 3.92 (31.0-43.8; 18)</td>
<td>49.56</td>
</tr>
<tr>
<td>Central circle</td>
<td>13.75 ± 1.73 (10.4-18.0; 22)</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Denticule span</td>
<td>11.86 ± 0.73 (10.8-13.1; 22)</td>
<td>17.90 ± 1.41 (15.0-19.8; 18)</td>
<td>24.41</td>
</tr>
<tr>
<td>Ray length</td>
<td>4.91 ± 0.55 (4.1-5.9; 22)</td>
<td>9.60 ± 1.05 (7.5-10.9; 18)</td>
<td>13.10</td>
</tr>
<tr>
<td>Central part</td>
<td>2.51 ± 0.32 (1.9-3.0; 22)</td>
<td>2.66 ± 0.32 (2.1-3.3; 18)</td>
<td>3.63</td>
</tr>
<tr>
<td>Blade length</td>
<td>4.43 ± 0.35 (3.7-5.3; 22)</td>
<td>5.55 ± 0.62 (4.2-6.7; 18)</td>
<td>7.44</td>
</tr>
<tr>
<td>Denticule length</td>
<td>8.23 ± 0.56 (7.2-9.4; 22)</td>
<td>9.44 ± 1.06 (7.4-11.4; 18)</td>
<td>10.09</td>
</tr>
<tr>
<td>Number of denticles</td>
<td>19.29 ± 0.96 (18.21; 21)</td>
<td>23.67 ± 1.03 (22-26; 18)</td>
<td>29</td>
</tr>
<tr>
<td>Pins/denticle</td>
<td>8.32 ± 0.48 (8-9; 22)</td>
<td>10.76 ± 0.90 (10-12; 17)</td>
<td></td>
</tr>
</tbody>
</table>

<sup>*</sup>Only one parasite well impregnated of this species was found in the present study.
portion and its thickness varied from wider to thinner. The tip of the ray was sharp (Figure 1b).

**Trichodina magna**

This trichodinid had a large body size, compared with the other species. The denticle blade was tight, filling almost half of the space between the y-axes. The anterior margin of the blade was slightly flattened (straight) and did not touch the y-axes. The ray had a prominent apophysis. It was thin and curved, and pointed slightly in the anterior direction, but did not go beyond the y-axes (Figure 1c).

**Trichodinids description: from gills**

The measurements of the taxonomic characteristics of each population of trichodinids collected from the gills are presented in Table 2.

**Trichodina centrostrigeata**

The blade of the denticle was thin and the anterior and posterior margins were similar and almost parallel to each other. The blade filled a large portion of the space between the y-axes and its anterior margin went beyond the y-axes. The central part was short and rounded. The ray had uniform thickness with a sharp tip, positioned parallel or anteriorly to the y-axes, and sometimes going beyond them. This trichodinid had a central ridge that was characteristic for the species (Figure 2a).

**Trichodina compacta**

The morphological characteristics were similar to those described in specimens that were found on the body surface (Figure 2b).

**Trichodina heterodentata**

The morphological characteristics were similar to those described in specimens that were found on the body surface (Figure 2c).

**Trichodina migala**

The blade was thin and did not fill much of the space between the y-axes. The anterior portion did not touch the y-axes and, in some cases, a small apophysis was seen. The central part occupied more than half of the area between the y-axes. Ray apophyses were prominent. The ray was thin, curved, sharp and displaced in the anterior direction, sometimes going beyond the limit of the y-axes (Figure 2d).

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Table 2. Measurements of trichodinids of from gills of Nile tilapia *Oreochromis niloticus*. The data are presented as arithmetic mean ± standard deviation (minimum-maximum values; number of individuals measured).

<table>
<thead>
<tr>
<th>Characters</th>
<th><em>T. centrostrigeata</em></th>
<th><em>T. migala</em></th>
<th><em>T. heterodentata</em></th>
<th><em>T. compacta</em></th>
<th><em>Paratrichodina africana</em></th>
<th><em>T. orthodontes</em></th>
<th>Trichodinella sp.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body diameter</td>
<td>55.73 ± 5.02 (47.0-64.9; 22)</td>
<td>76.01 ± 5.66 (70.5-83.6; 4)</td>
<td>70.45 ± 6.06 (63.9-80.4; 9)</td>
<td>53.09 ± 2.20 (49.9-57.9; 9)</td>
<td>25.04 ± 2.50 (21.6-29.8; 20)</td>
<td>27.36 ± 1.35 (24.9-29.1; 9)</td>
<td>22.94 ± 2.38 (20.2-25.7; 6)</td>
</tr>
<tr>
<td>Border membrane</td>
<td>3.71 ± 0.84 (2.1-5.4; 22)</td>
<td>5.67 ± 0.50 (5.1-6.2; 4)</td>
<td>5.43 ± 0.53 (4.6-6.0; 9)</td>
<td>4.61 ± 0.53 (3.6-5.4; 9)</td>
<td>1.80 ± 0.19 (1.3-2.1; 20)</td>
<td>2.50 ± 0.31 (1.8-2.9; 9)</td>
<td>2.04 ± 0.38 (1.5-2.6; 9)</td>
</tr>
<tr>
<td>Adhesive disc</td>
<td>49.03 ± 5.31 (39.1-60.1; 23)</td>
<td>64.71 ± 6.83 (57.7-73.7; 4)</td>
<td>59.61 ± 5.58 (53.5-68.3; 9)</td>
<td>44.35 ± 2.09 (42.0-49.1; 9)</td>
<td>21.24 ± 2.42 (17.7-26.3; 20)</td>
<td>22.36 ± 1.34 (19.8-24.2; 9)</td>
<td>21.65 ± 2.84 (18.5-24.4; 6)</td>
</tr>
<tr>
<td>Denticulate ring</td>
<td>26.04 ± 2.89 (21.7-32.0; 24)</td>
<td>40.79 ± 5.50 (35.8-48.5; 4)</td>
<td>39.09 ± 6.99 (29.7-50.6; 10)</td>
<td>27.13 ± 2.13 (25.2-32.3; 9)</td>
<td>12.42 ± 1.88 (9.4-16.3; 20)</td>
<td>10.88 ± 0.88 (9.3-12.5; 9)</td>
<td>12.01 ± 1.72 (10.2-14.4; 6)</td>
</tr>
<tr>
<td>Central circle</td>
<td>-</td>
<td>-</td>
<td>13.11 ± 1.66 (10.7-16.1; 9)</td>
<td>5.82 ± 0.48 (4.7-6.7; 20)</td>
<td>7.42 ± 0.73 (6.3-8.5; 9)</td>
<td>4.37 ± 0.35 (4.0-5.0; 6)</td>
<td></td>
</tr>
<tr>
<td>N of central ridges</td>
<td>13.22 ± 1.93 (10.0-17.0; 24)</td>
<td>-</td>
<td>-</td>
<td>13.11 ± 1.66 (10.7-16.1; 9)</td>
<td>5.82 ± 0.48 (4.7-6.7; 20)</td>
<td>7.42 ± 0.73 (6.3-8.5; 9)</td>
<td>4.37 ± 0.35 (4.0-5.0; 6)</td>
</tr>
<tr>
<td>Denticle span</td>
<td>12.82 ± 1.94 (9.8-16.3; 24)</td>
<td>18.48 ± 2.02 (15.8-20.2; 4)</td>
<td>17.64 ± 1.64 (14.8-19.8; 10)</td>
<td>11.80 ± 0.87 (10.9-13.7; 9)</td>
<td>0.54 ± 0.10 (0.3-0.7; 20)</td>
<td>0.54 ± 0.06 (0.4-0.6; 9)</td>
<td>1.3 ± 0.15 (1.2-1.5; 5)</td>
</tr>
<tr>
<td>Ray length</td>
<td>4.57 ± 0.98 (2.5-6.7; 24)</td>
<td>8.30 ± 1.59 (6.0-9.7; 4)</td>
<td>9.61 ± 1.33 (7.3-11.6; 10)</td>
<td>5.38 ± 0.50 (4.7-6.2; 9)</td>
<td>3.33 ± 0.27 (2.7-3.8; 20)</td>
<td>4.87 ± 0.60 (3.7-5.7; 9)</td>
<td>2.34 ± 0.47 (1.7-2.8; 6)</td>
</tr>
<tr>
<td>Central part</td>
<td>2.42 ± 0.54 (1.4-3.3; 24)</td>
<td>3.43 ± 0.81 (2.6-4.3; 4)</td>
<td>2.46 ± 0.27 (2.1-3.0; 10)</td>
<td>2.30 ± 0.33 (1.6-2.7; 9)</td>
<td>2.42 ± 0.39 (1.7-3.4; 20)</td>
<td>2.57 ± 0.33 (2.1-3.1; 9)</td>
<td>2.21 ± 0.28 (1.9-2.6; 6)</td>
</tr>
<tr>
<td>Blade length</td>
<td>5.73 ± 0.94 (4.1-7.5; 24)</td>
<td>5.06 ± 0.87 (5.3-7.2; 4)</td>
<td>5.51 ± 0.52 (4.6-6.1; 10)</td>
<td>4.12 ± 0.51 (3.5-4.9; 9)</td>
<td>22.17 ± 1.10 (20-25; 18)</td>
<td>21.44 ± 1.01 (20-23; 9)</td>
<td>21.00 ± 1.41 (19-22; 5)</td>
</tr>
<tr>
<td>Denticle length</td>
<td>4.51 ± 0.37 (3.8-5.3; 24)</td>
<td>5.53 ± 0.48 (8.0-9.0; 4)</td>
<td>9.00 ± 1.03 (7.1-10.7; 10)</td>
<td>8.16 ± 0.51 (7.5-8.9; 9)</td>
<td>-</td>
<td>5 ± 0</td>
<td>-</td>
</tr>
<tr>
<td>Number of denticles</td>
<td>27.71 ± 1.00 (26-30; 24)</td>
<td>27.00 ± 1.41 (26-29; 4)</td>
<td>23.90 ± 2.08 (22-28; 10)</td>
<td>19.00 ± 0.76 (18-20; 8)</td>
<td>25.04 ± 2.50 (21.6-29.8; 20)</td>
<td>27.36 ± 1.35 (24.9-29.1; 9)</td>
<td>22.94 ± 2.38 (20.2-25.7; 6)</td>
</tr>
<tr>
<td>Pinn/denticle</td>
<td>9.33 ± 1.15 (8-10; 3)</td>
<td>10.00 ± 0.00 (10-10; 4)</td>
<td>10.40 ± 1.14 (9-12; 5)</td>
<td>8.89 ± 1.05 (8-10; 9)</td>
<td>1.80 ± 0.19 (1.3-2.1; 20)</td>
<td>2.50 ± 0.31 (1.8-2.9; 9)</td>
<td>2.04 ± 0.38 (1.5-2.5; 6)</td>
</tr>
</tbody>
</table>
This trichodinid had a rounded blade that was not sickle-shaped, thus differing from the genus *Trichodina*. The central part had a large and characteristic elongated part that was parallel to the central part of the adjacent denticle. The central part was thin, rounded and filled about half of the space between the y-axes. The ray was thin, with the rounded tip lying parallel to the y-axes (Figure 2e).

*Trichodinella sp.*

This trichodinid was difficult to impregnate because of its small size and minute denticle. The blade had a small projection and the anterior margin extended well into the space between the y-axes. The central part was delicate, short and did not go beyond the y-axis. The denticles were small and inserted together in the central parts and projections, which made it difficult to see them in the silver-impregnated specimens. The ray formed a characteristic short delicate curved hook (Figure 2f).

*Tripartiella orthodens*

This was a small-sized trichodinid. The anterior margin of the blade was straight and the posterior margin was rounded. In the posterior portion, this trichodinid had a prominent projection. In the anterior portion, between the blade and the central part, another prominent projection was observed, which went beyond the y-axes. The central part was small...
and occupied a small portion of the space between the y-axes. The ray was straight, with similar thickness up to its tip, and it was parallel to or was positioned slightly posteriorly to the y-axis (Figure 2g).

Discussion

In the present study four genera of trichodinids were found parasitizing Nile tilapia cultivated in Brazil: *Trichodinella* (*T. centrostripegata*, *T. compacta*, *T. heterodentata*, *T. magna* and *T. migala*), *Paratrichodinona* (*P. africana*), *Tripartiella* (*T. orthodentus*) and *Trichodinella* (*T. epizootica* sp.). All the trichodinids identified from this study have an African origin (BASSON et al., 1983; BASSON & VAN AS, 1987, 1994; VAN AS & BASSON, 1989). However, they have also been found parasitizing fish in the Americas (AGUILAR-AGUILAR & ISLAS-ORTEGA, 2015) and in Eurasia (DUNCAN, 1977; BASSON & VAN AS, 1994; MITRA & HALDAR, 2004; MITRA & BANDYOPADHYAY, 2006; MITRA et al., 2012; MOHILAL & HEMANANDA, 2012).

In Brazil, there have also been records of *T. centrostripegata* in the southeastern region (VALLADÃO et al., 2016) and in the northern region (BITTENCOURT et al., 2014); *T. compacta* in the southern region (GHIRALDELLI et al., 2006) and southeastern region (VALLADÃO et al., 2016); *T. heterodentata* in the southern region (MARTINS et al., 2010) and southeastern region (VALLADÃO et al., 2016); *T. magna* in the southern region (MARTINS & GHIRALDELLI, 2008) and southeastern region (ZAGO et al., 2014); *T. migala* in the southeastern region (VALLADÃO et al., 2016); *P. africana* in the southern region (GHIRALDELLI et al., 2006; JERÓNIMO et al., 2011), southeastern region (VALLADÃO et al., 2016), northern region (BITTENCOURT et al., 2014) and northeastern region (VALLADÃO et al., 2013); and *T. orthodentus* in the southeastern region (VALLADÃO et al., 2016). In the present study, trichodinids were identified in the central-western region of Brazil (state of Goiás) and the first record of the genus *Trichodinella* in this country was presented.

The trichodinids in this study has a very constant denticle shape that became impregnated well with silver nitrate, although in some species the denticles were somewhat less impregnated, thus making it difficult to identify them (*Paratrichodinona*, Tripartiella and *Trichodinella*). The morphometric and morphological characteristics of *T. centrostripegata*, *T. compacta*, *T. heterodentata*, *T. magna*, *T. migala*, *P. africana* and *T. orthodentus* that were identified in the present study were similar to those in previous descriptions of these species in Nile tilapias in Brazil. Nevertheless, the body diameter of *T. heterodentata* was slightly larger (64-80 versus 38-59) than those reported by Valladão et al. (2016), but was similar to those described by Duncan (1977) (64-80 versus 58-122). We consider that small differences are common, since variability in morphometric and morphological characteristics of trichodinids has been observed in other populations of these protozoa (LOM, 1958). Such differences usually relate to the developmental stages of the denticle, the host and the environmental conditions (DUNCAN, 1977; GONG et al., 2005).

The genus *Trichodinella* was described by Lom (1963) as having one incomplete turn of the two adoral ciliary rows in the adoral zone, a peculiarly shaped denticle and small dimensions, and was reported as only parasitizing the gills of fish. So far, about eight species have been identified from marine and freshwater organisms. The *Trichodinella* sp. from this study was collected from the gills and due to its small size and the minuteness of the denticles, the impregnation was not good enough to perform species identification. However, some elements of their structure, e.g. body diameter, border membrane, adhesive disc, denticulate ring, denticle span, denticle ray, central part of the denticle and blade, were fairly well preserved in the preparations. These enabled comparisons with other small trichodinids that have already been described in the literature.

The *Trichodinella* sp. of the present study seems to be identical to the species identified by Lom (1963) as *Trichodinella epizootica* Šrámek-Hušek, 1953, which occurs in the gills of different species of fish on the Eurasian continent. All the measurements of taxonomic characteristics relating to that species are very close to those of *Trichodinella* sp. in the present study. The measurements of our specimens are also similar to the *T. epizootica* populations of Basson et al. (1983) from *Cyprinus carpio*; of Albladjeo & Arthur (1989) from *Cyprinus carpio*; of Al-Rasheid et al. (2000) from *Mormyrus kannume*, of Mitra & Haldar (2004) from *Puntius gelius*; and of Basson (2010) from *Tinca tinca*.

In the literature, schematic drawings of *T. epizootica* vary significantly between each description (Figure 3). The schematic drawing of *Trichodinella* sp. identified here was very similar to the record of Mitra & Haldar (2004) (Figure 3). These differences may be due to the common variations observed in *T. epizootica* morphology within the same population or in populations from different hosts (LOM, 1963; LOM & HALDAR 1977; BASSON & VAN AS, 1987; BASSON, 2010). These variations occur mainly due to difficulties in impregnation with silver nitrate, which culminates in different interpretations by different authors. Furthermore, Figure 3 shows that the schematic drawing of *T. epizootica* described by Al-Rasheid et al. (2000) is quite similar to the schematic drawing of *Trichodinella carpi* Duncan, 1977, from Tang et al. (2005). These presented more similarities than between the records of *T. epizootica*. Measurement data and schematic drawings indicate that the parasite of our study may be of the species *T. epizootica*. However, due to the small number of well-impregnated specimens and the confusion in the literature, we have only described the genus.
Figure 3. Schematic drawing of the denticles of Trichodinella demonstrating the diversity of denticle shape among Trichodinella epizootica (redrawn from various sources: BASSON & VAN AS, 1989; MITRA & HALDAR, 2004; AL-RASHEID et al., 2000; BASSON, 2010; MITRA et al., 2013; XU et al., 1999 and TANG et al., 2005).

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