Luteal dynamics in goats: morphological and endocrine features

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**ABSTRACT** - The aim of this study was to establish the morphologic and endocrine characteristics of luteal dynamics in goats. It was used Toggenburg female goats that showed natural estrus in a 48-hour interval. After estrus, ultrasonographic evaluations of the ovaries were daily performed during 21 days using a portable device (5MHz probe). Blood sample was collected for plasma progesterone (P\(_4\)) determination. Corpora lutea were detected for the first time on day 5 and progressively increased in size until D9 (1.26 ± 0.08 cm\(^2\)), with no variation on subsequent days. In females with one ovulation, the first visualization of the corpora lutea was earlier than in those with multiple ovulation (4.54 ± 0.18 vs 5.74 ± 0.25 days). At the moment of the first visualization, luteal area was smaller in animals with single ovulation. Plasma P\(_4\) concentration progressively increased until day 9 and it did not show significant increase until luteolysis, characterized by a sharp decrease in P\(_4\) concentration, reaching values below 1 ng/mL in 24 hours. The luteal area slowly and gradually decreased in size. It was observed a significant positive correlation between P\(_4\) concentration and area during luteogenesis and luteolysis (r = 0.63 and r = 0.50, respectively). When corpus luteum reached its maximum size (D9), female with more than one corpora lutea, with a greater luteal tissue area, did not show P\(_4\) concentration higher than those with one ovulation (5.92 ± 0.59 vs 7.04 ± 0.79 ng/mL). These results show that luteal dynamics in Toggenburg goats follow a similar pattern to those observed in other goat breeds and luteal tissue growth was positively correlated with corpora lutea functionality.

Key Words: corpus luteum, goats, progesterone, ultrasonography

**Introduction**

The growth of goat industry in Brazil is stimulating the use of assisted reproductive technologies for this specie. However, the development of these techniques requires basic knowledge on ovarian physiology, which is still limited in small ruminants, especially when compared to other domestic species such as cows.
Corpus luteum is a transitory gland responsible for syntheysis and release of progesterone (P₄), a steroidal hormone essential for early embryo development and pregnancy support (Smith et al., 1994). Correct and accurate corpus luteum identification is crucial for the success of assisted reproduction programs. Previous studies in other species reported that the presence of a functional corpus luteum positively influenced the outcome of superovulation protocols (González-Bulnes et al., 2002) and estrus synchronization (Brito et al., 2003; Uribe-Velásquez et al., 2010).

A positive correlation between luteal tissue area and concentration of plasma P₄ was previously described for cows (Siqueira et al., 2009), sheep (Davies et al., 2006) and goats (Orita et al., 2000). In bovines, the higher number of corpus luteum is not related to an increase in concentration of plasma P₄ (Mann et al., 2007). In sheep and goats, maximal plasma P₄ concentration was related to the breed (Fonseca & Torres, 2005).

In small ruminants, ovarian function was evaluated and studied mainly by surgical techniques (Camp et al., 1983). These surgical techniques, also performed in embryo transfer procedures, cannot be repeatedly used because they almost always lead to some extent of adherence formation (Gonçalves et al., 2001). The use of transrectal ultrasonography in small ruminants was a hallmark in the 1990s, and important studies on ovarian physiology could be done. The ultrasonography technique allowed the identification of ovarian structures in goats with a sensitivity and specificity similar to those observed in surgical methods (Simões et al., 2005). The first sonographic studies on ovarian function, however, focused on follicular dynamics (Medan et al., 2003), and only recently, luteal activity was systematically studied in this specie (Simões et al., 2007).

The aim of this study was to characterize luteal dynamics in Toggenburg goats by ultrasonography during estrous cycle and early pregnancy, and the correlations among morphological changes in the corpus luteum and P₄ plasma concentrations.

### Material and Methods

The experiment was conducted during the breeding season (March and April). According to Köppen, the region climate is classified as Cwa, characterized by dry winters and rainy summers, annual temperature average from 18°C to 23°C and annual precipitation between 2000 and 2600 mm³.

In this study, twenty-three nulliparous Toggenburg goats (*Capra hircus*) with average age of eight months, body weight 33.5 ± 1.2 kg and body condition score 3.5 ± 0.07 (1 to 5 scale) were used. These animals did not have reproductive pathologies detectable by ultrasonography or vaginoscopy and showed estrus behavior, detected by teasers, within a 48-hour interval. After estrus detection, all females were bred twice, every day, by a sexually mature buck until the end of estrus. Fourteen goats became pregnant and two females were ruled out because of the presence of follicular cyst.

The animals were confined in collective pens, feed with Napier grass (*Pennisetum purpureum v. Taiwan*) and concentrated. Water and mineral salt were provided ad libitum.

A portable ultrasound device (Aloka SSD 500®, Aloka Co. Japan) equipped with an adapted 5MHz linear rectal transducer was used in this study. This adaptation consisted on the attachment of two hard sticks to the transducer cable. The ultrasonographic evaluations were performed always by the same technician and in the same period of the day.

Ovarian sonographic evaluations started on the day of estrus detection (Day 0) and were performed daily during 21 consecutive days. On the day of estrus detection, the single or codominant follicles were identified and measured, and evaluated until ovulation. The ovulation moment was considered as the day in which the previously identified dominant follicle was no longer present in the ovary. After ovulation, the subsequent evaluations aimed to characterize the periods of luteogenesis, luteal function and luteolysis. During these evaluations the corpus luteum area was measured on its larger diameter.

Luteal tissue area (cm²) was considered the difference between total corpus luteum area and the luteal cavity area, when it was present. For animals with double (n=7) or triple (n=1) ovulation, luteal tissue area in each day of the cycle was considered as the sum of luteal tissue area from all corpus luteum present in both ovaries.

Just before each ovarian exam a blood sample was collected for further plasma P₄ determination. Samples were collected by jugular vein puncture using vacuolized 5 mL tubes containing EDTA. Blood samples were kept in an isothermal container (5°C) and they were immediately sent to the laboratory to be processed. The samples were then centrifuged at 4°C at 894 g for 20 minutes. Plasma was separated and frozen (−20°C) until P₄ evaluation.

The plasma P₄ concentration was determined by radioimmunoassay technique (RIA) using commercial kits (Coat a Count® – DPC-Med Lab, Rio de Janeiro, Brazil) and performed at the Endocrinology laboratory of the Universidade Estadual Paulista Julio de Mesquita Filho (Unesp), Botucatu, São Paulo State, Brazil.
Luteal tissue area and plasma P4 concentrations were checked for normality with the Lilliefors test and for homogeneity by the Cochran and Bartlett test. The effect of post-ovulation day on luteal tissue area and plasma P4 progesterone was evaluated by analysis of variance (ANOVA), and comparisons among means determined by Tukey’s test (5% probability). The luteogenesis and luteolysis periods were characterized according to the presence of significant differences on luteal tissue area and plasma P4 concentrations. Associations among variables were determined by Pearson correlation method. Statistical analysis was performed in SAEG – Sistema de Análises Estatísticas (SAEG, 2007).

Data from all animals (n=21) were used to evaluate luteogenesis, and from the nonpregnant animals (n=7) to evaluate luteolysis after day 16. To analyze luteal tissue area and P4 concentration during luteolysis, data were normalized to the moment in which plasma P4 dropped to values below 1 ng/mL.

**Results and Discussion**

Aiming to establish the moment of the first corpus luteum detection and to track corpus luteum morphological changes along afterwards, ultrasonographic evaluations started after estrus detection. At ovulation, the mean diameter of the dominant follicle was 7.4 ± 0.1 mm, similar to the size previously reported on Toggenburg goats (7.7 ± 1.3 mm; Maffili et al., 2006), as well as in others breeds such as Serrana (7.1 ± 0.1 mm; Simões et al., 2006), Shiba (7.8 ± 0.2 mm; Medan et al., 2005) and Saanen (6.9 ± 1.8 mm; Maffili et al., 2005). Ultrasoundography is considered an efficient tool to identify ovarian structures. The sensibility and efficiency of this technique were previously demonstrated by Viñoles et al. (2004) in ewes and by Simões et al. (2005) in goats. In the present study, the mean moment of first corpus luteum visualization was Day 5 of estrous cycle (ranging from Day 4 to 7). In animals with single ovulation the first identification was performed earlier (P<0.05) than in animals with multiple ovulation (Table 1).

Dickie et al. (1999) and Viñoles et al. (2004) reported that corpus luteum identification was more difficult in sheep with multiple ovulations, which suggest that sensitivity of ultrasonographic evaluation is reduced in these cases. In the present study, the first visualization of the corpus luteum was done earlier in animals with single ovulation, although they had a smaller total luteal tissue area at this moment. On the day of the first visualization, however, luteal area was not different between animals with single or multiple ovulations when each corpus luteum was individually considered (0.56 ± 0.06 and 0.53 ± 0.05 cm², respectively; P>0.05). Among goats with more than one ovulation (n = 8), only one animal had ovulations occurring in distinct ovaries. The presence of more than one corpus luteum in the same ovary may have made corpus luteum individualization more difficult, delaying identification of each corpus luteum in animals with multiple ovulations.

Previous studies demonstrated that the first visualization of the corpus luteum in goats can be performed between Day 2 and Day 3 of estrous cycle (Orita et al., 2000; Riesemberg et al., 2001; Medan et al., 2003; Simões et al., 2007) when higher frequency (7.5 MHz) transducers are used. The present study showed that corpus luteum identification can be performed with lower frequency (5 MHz) transducers, which have a broader range of applications and is largely used in bovines, although first visualization was only possible two days later (on Day 5).

In small ruminants embryo transfer procedures are performed by surgical techniques at around Day-7 of estrous cycle. The use of ultrasonography prior to these surgical techniques would avoid unnecessary surgical procedures in those cases when embryo donors or recipients did not respond to the hormonal treatment, i.e., do not present a functional corpus luteum, optimizing the use of animals available and avoiding their precocious discard due to surgical consequences. Detection of corpora lutea as early as Day-5, as shown in the present study, demonstrates that ultrasonography can be performed before embryo transfer procedures to evaluate presence, number and position of corpus luteum.

After corpus luteum first detection, morphological features could be tracked in a daily basis and correlated to progesterone production, which is the main indicator of corpus luteum function. In the present study, size of luteal

<p>| Table 1 - Day and luteal tissue area (cm²) when corpus luteum was first visualized in goats with single and multiple ovulation |
|-----------------|-----------------|-----------------|</p>
<table>
<thead>
<tr>
<th>N° of ovulation</th>
<th>Day of first visualization (days)</th>
<th>Luteal area at day of first visualization (cm²)†</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 (n = 13)</td>
<td>4.54 ± 0.18b</td>
<td>0.56 ± 0.06b</td>
</tr>
<tr>
<td>≥2 (n = 08)</td>
<td>5.74 ± 0.25a</td>
<td>0.83 ± 0.11a</td>
</tr>
<tr>
<td>Mean (n = 21)</td>
<td>5.00 ± 0.19</td>
<td>0.63 ± 0.07</td>
</tr>
</tbody>
</table>

a,b - Different letter in the same column differ (P<0.05, ANOVA).
† In those animals with multiples ovulations, luteal tissue area is the sum of all corpus luteum presented in the ovaries.
tissue area progressively increased from detection to Day 9 (growth rate of 0.16 cm²/day, P<0.05), achieving a mean size of 1.26 ± 0.08 cm², and entered in a plateau phase, with no significant increase on subsequent days (Fig. 1). Previous studies in goats reported that corpus luteum reached its maximum size between Day 8 and Day 11 (Camp et al., 1983; Medan et al., 2003; Simões et al., 2007), and this range may be related to differences in breed or in the technique used (surgery and ultrasonography). The morphological changes during this period were closely related to the endocrine profile. An increase on plasma P₄ concentration until Day 9 was also detected (from 0.07 ± 0.05 to 6.31 ± 0.46 ng/mL, P<0.05; Figure 1), and after that, no significant increase was observed until the onset of natural luteolysis, when these concentrations began to decrease reaching values lower than 1 ng/mL in 24 hours (Figure 2).

These combined results demonstrate that luteal dynamic on Toggenburg goats was characterized by a growth phase (luteogenesis) which was associated to an increase in corpus luteum functional maturity, as demonstrated by plasma P₄ concentrations. These changes are the result of an intensive cellular proliferation and biochemical changes which occur in the corpus luteum during luteogenesis period, aiming the synthesis and release of progesterone (Smith et al., 1994; Sangha et al., 2002). The association between growth of luteal tissue and steroidogenesis observed in the current study was similar to what was observed in other goat breeds (Orita et al., 2000; Medan et al., 2003; Simões et al., 2007), and domestic ruminants such as cows (Siqueira et al., 2009) and sheep (Davies et al., 2006). The plateau phase observed both in corpus luteum size and P₄ production on day 9, however, differs from luteal dynamics in cows, in which functional maturity is achieved after maximum corpus luteum size (Viana et al., 1999). The meaning of this difference needs further investigation, but we can hypothesize that functional maturity is attained earlier in the goat corpus luteum, based on the same earlier prostaglandin response capacity in this specie (Rubianes & Menchaca, 2003) when compared to bovine (Weems et al., 2006).

The plateau phase was followed by a regression phase (luteolysis), characterized by a decrease both in luteal tissue area and plasma P₄ concentration. The luteolysis process consists of a loss in steroidal capacity (functional luteolysis) and regression (structural luteolysis) of the corpus luteum (McCracken et al., 1999). At luteolysis, the decrease on plasma P₄ concentrations occurred faster and more abruptly when compared to the decrease observed in luteal tissue area (92 and 10% of reduction, respectively), demonstrating the temporal difference between functional and structural luteolysis, which was also described in other goat breeds (Simões et al., 2007) and also in cows (Viana et al., 1999). This temporal difference can be explained by the fact that in the luteolysis mechanism, the very first changes induced by PGF₂α are a disruption of steroidogenic pathway activity and a reduction in availability of P₄ precursors by reduction on the corpus luteum blood flow, which reduces plasma P₄ concentrations to values below 1 ng/mL within 24 hours. The structural luteolysis is a more gradual process because it is mediated by an apoptotic cascade, with luteal cells death and fagocitosis, and replacement by fibroblasts (Niswender, 2000).

The difference in morphological and functional luteolysis is also reflected in the greater correlation between luteal tissue area and the progesterone found in the present study during luteogenesis (r = 0.63; P<0.05) when compared to luteolysis (r = 0.50; P<0.05). The significant overall correlation of corpus luteum morphology and function during estrous cicle was also reported in goats (r = 0.80;
Orita et al., 2000), cows (r = 0.69; Siqueira et al., 2009) and sheep (r = 0.59; Davies et al., 2006), and it demonstrates that ultrasonography is a useful tool to estimate luteal function in those species.

Despite the positive correlation observed between luteal tissue area and plasma P4 concentration during corpus luteum formation and regression periods, the results presented on Table 2 show that physiological capacity to synthesize and release P4 was not greater in those animals bearing more than one corpus luteum.

In cows (Mann et al., 2007) and sheep (Bartlewski et al., 1999), an increase in plasma P4 concentrations associated with the number of ovulations was not observed either. In contrast, Jarrell & Dziuk (1991) only detected a greater plasma P4 concentration in pregnant goats with more than one corpus luteum between Day 3 and Day 25 of estrous cycle, but after Day 30 this difference was no longer observed. This inconsistency can be related to other sources of variation in P4 production, such as breed, nutrition and body size, or even to the fluctuating nature of P4 production, which is reflected in greater variation coefficients when compared to luteal size (9.9% vs. 3.6%; respectively). In fact, on Day 9, when plasma P4 concentrations reached their maximum value, a significant correlation between luteal tissue area and plasma P4 concentration was not observed.

Luteal cavities were frequently found in the present study, being present as a central anechoic cavity in 83% of corpus luteum (25/30). On the first visualization day, the mean area of these cavities was 0.29 ± 0.20 cm², representing 45.3% of total area of the corpus luteum. These cavities progressively regressed in size (P<0.01) until Day 11, when reached an area smaller than 0.1 cm²; representing less than 7% of total area of the corpus luteum. The luteinization of the follicle is a process that occurs from the outside to inside. When this process is not complete, a luteal cavity is formed. This cavity is a central structure filled by a clear serous transudate (Singh et al., 1997).

Previous studies also reported a high frequency of corpus luteum with a central anechoic cavity in goats (Simões et al., 2005 – 65%; Simões et al., 2007 – 78%), sheep (Dickie et al., 1999 – 12% a 68%; González-Bulnes et al., 2000 – 33.3%), and cows (Kastelic et al., 1990 – 79%; Singh et al., 1997 – 72%; Siqueira et al., 2009 – 66%). Despite its elevated frequency, a correlation between the presence of these cavities and duration of estrous cycle or plasma P4 concentrations was not observed; therefore corpus luteum cavities have no functional relevance (Kastelic et al., 1990, González-Bulnes et al., 2000). However, from a practical point of view, Viñoles et al. (2004) reported that, in early diestrus, the corpus luteum with a large central cavity can be misidentified as an ovarian follicle, delaying or even making the correct identification of the corpus luteum more difficult. Yet, as in goats, it is not possible to directly assess the reproductive tract in a non-invasive basis, the presence of an anechoic central cavity in the corpus luteum help to localize the ovaries by ultrasonography.

**Conclusions**

Despite the limitations related to ultrasonographic exam in small ruminants, the identification, measurement and daily monitoring of corpus luteum can be performed by rectal ultrasonography using a 5MHz linear transducer; and the luteal tissue area can be used to estimate corpus luteum function, although progesterone production is not affected by the number of corpus luteum. Luteal dynamics in Toggenburg goats is characterized by a luteogenesis, a plateau and a luteolysis phase, as previously observed on other domestic ruminant species. Ultrasonography can be considered as a potential tool to evaluate corpora lutea in embryo donors and recipients.

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