Fatty acid profile of meat, diurnal changes in volatile fatty acids, rumen fluid parameters, and growth performance in Korean native (Hanwoo) steers fed high- and low-forage diets supplemented with chromium-methionine

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ABSTRACT - The objective of this study was to determine the effects of forage level in diets supplemented with chromium-methionine (Cr-Met) on rumen fluid parameters, meat fatty acid composition, and performance of Korean beef (Hanwoo) steers. Twenty-three Hanwoo steers were used in this experiment. A completely randomized design and repeated measurements were used to analyze the data set. Beef steers were fed diets containing high (10 head; average body weight (BW) = 525.1±27.5; forage:concentrate (F:C) = 60:40) (60F) and low (13 head; average BW = 531.8±32.4; F:C = 40:60 ratio) (40F) forage diets supplemented with Cr-Met for 60 d. Dry matter intake, BW, and feed efficiency were not different between the two treatment groups. Fatty acid composition of meat including myristate, stearate, and gamma linoleate was not different between the two groups; however, palmitate, palmitoleate, and linoleate were higher in 60F group than 40F group. Ammonia-N showed a higher trend in 40F group, whereas pH demonstrated higher values in 60F group. Ruminal acetate was higher in 60F group than 40F group and maintained a high trend throughout the sampling time, whereas no differences were found in ruminal propionate, butyrate, and valerate between two groups. A high-forage diet (60%) improves meat quality and has no adverse effects on performance of Hanwoo steers.

Key Words: forage level, Hanwoo steers, meat quality

Introduction

Previous studies in our laboratory have reported improvements in carcass characteristics in Korean native (Hanwoo) steers (Sung et al., 2015), meat fatty acid profile, and performance of Holstein steers (Song, et al., 2013; Ghassemi Nejad et al., 2016) receiving chromium-methionine (Cr-Met) supplementation. In those studies, 400 ppb/d of Cr-Met supplementation was reported as the optimum level. On the other hand, Sung et al. (2015) reported higher marbling score, rib-eye area, and meat yield index in Hanwoo steers fed a high-forage diet supplemented with 400 ppb/d of Cr-Met compared with the low-forage diet group. However, to the best knowledge of the authors, the effects of Cr-Met supplementation on daily pattern of rumen fluid parameters including volatile fatty acids (VFA), pH, ammonia-N, and fatty acid composition of meat from Hanwoo steers fed high-and low-forage diets is yet to be investigated.

Monounsaturated fatty acids (MUFA) and saturated fatty acids (SFA) are the main components of the great majority of fatty acids in beef fat. Furthermore, beef fats are among the richest natural sources of conjugated linoleic acid (CLA) and trans-vaccenic acid (Chin et al., 1992), which have been shown to have health benefits for humans (Belury, 2002). Any increase in ruminal starch digestion should be followed by an increase in organic acids that are later converted to glucose, which is a precursor for marbling (Kitchalong et al., 1995; Lindemann et al., 2008). Any discussion of marbling levels should also include the genetic influence. Hence, there might be differences between Holstein and Hanwoo steers. Previous research has shown that forage-finished cattle produce beef with more CLA and n-3 fatty acids compared with grain-finished cattle (French et al., 2000). Melton et al. (1982) found that grass-fed cattle beef had a decreased concentration of MUFA and a greater concentration of SFA compared with grain-fed cattle; however, one study found that grass-fed cattle beef had less SFA and more MUFA than grain-fed cattle beef (French et al., 2000). Moreover, it has been reported that Cr supplementation can be employed to manipulate the quality of meat due to its biological function on body fat and muscle metabolism (Kitchalong et al., 1995; Sung et al., 2015); however, factors such as the level of Cr supplementation and its source, nutrients, chromium levels in the basal diet,
breed and species may intervene with these functions (Sung et al., 2015; Ghassemi Nejad et al., 2016). Decreasing diet energy density through increasing forage level may also increase ruminal acetate:propionate ratio and methane energy loss (Merchen et al., 1986; Kinser et al., 1988; Zinn et al., 1994). Therefore, the effects of high- and low-forage diet when supplemented with Cr-Met in diet of Hanwoo steers on rumen fluid characteristics and fatty acid profile of beef were investigated in this study.

**Material and Methods**

Animal procedures were approved by the Ethical Committee for Animal Research of Kangwon National University. The experiment site was the Gangwon province (37°55'03.64"N, 127°46'22.71"E) in Republic of Korea. Twenty-three Korean native (Hanwoo) steers were used in this experiment and assigned to two treatment groups. The feeding amount of Cr-Met (Innobio Co., Ltd., Shiheung, Korea) to animals was limited to 400 ppb/cow/d. The duration for the study was two months. Feed was total mixed ration (TMR) including commercial concentrate and forage (alfalfa hay, bermudagrass hay, and rice straw) comprising 16±0.05% crude protein (CP) and 78±12.3 total digestible nutrients (TDN). Treatments included what follows: steers fed diets that contained 60% concentrate and 40% forage supplemented with 400 ppb/d of Cr-Met (average body weight (BW) = 531.8±32.4; low-forage diet) (40F); and steers fed diets that contained 40% concentrate and 60% forage supplemented with 400 ppb/d of Cr-Met (10 head; average BW = 525.1±27.5; high-forage diet) (60F).

Forages and concentrate were mixed and offered twice daily to methods of AOAC (1990; Table 1). Neutral detergent fiber; ADF - acid detergent fiber. DM - dry matter; OM - organic matter; CP - crude protein; NDF - neutral detergent fiber; ADF - acid detergent fiber.

Table 1 - Chemical composition of feeds

<table>
<thead>
<tr>
<th>Feed</th>
<th>DM (%)</th>
<th>OM</th>
<th>CP</th>
<th>NDF</th>
<th>ADF</th>
</tr>
</thead>
<tbody>
<tr>
<td>Concentrate</td>
<td>89.2</td>
<td>91.2</td>
<td>16.5</td>
<td>35.1</td>
<td>12.6</td>
</tr>
<tr>
<td>Forage</td>
<td>89.1</td>
<td>90.7</td>
<td>10.2</td>
<td>64.5</td>
<td>48.2</td>
</tr>
</tbody>
</table>

Concentrate used was commercial pellet concentrate on the farm. Forage included alfalfa hay (55%) + bermudagrass (35%) + rice straw (10%).

Volatile fatty acids (VFA; acetate, propionate, butyrate and valerate) of rumen were measured using a gas chromatograph (GC, shimadzu GC-17A, 30 m × 0.25 mm × 0.25 µm column). After filtration of the rumen fluid, metaphosphoric acid was added to precipitate proteins, and formic acid (5%) was used to eliminate the disturbing “ghosting” effect. The clear supernatant obtained after centrifugation (3000 rpm for 10 min) was injected directly into the column. The analysis of one rumen sample requires about 8 min, in which the temperatures of injector, column, and detector were -20 °C, 100 °C, and 230 °C, respectively. Depending on the implementation of the column temperature programing, helium (He) as a carrier gas with a flow rate of 7 mL/min and hydrogen (H₂) gas and air oxygen with a flow rate of 15 mL were used. The split ratio was 1:30, and 1 µL of the sample was used. For the quantitative determination, a nonautomatic integrator was used. The individual VFA were calculated by comparing with a standard.

Upon completion of the field experiment, five steers per group were slaughtered to measure the fatty acid composition of beef from the loin side. Samples from each steer were frozen at -20 °C for 12 h and thawed prior to analysis. According to the method of lipid extraction (Folch et al., 1957), 6 g of sample and chloroform/methanol (2:1) solution were homogenized in a 25 mL homogenizer (Dix 6000, Heidolph, Germany) at 1100 × g for 30 s. Next 6 mL of 0.88% KCl solution were added to the homogenate, followed by centrifugation at 2500 × g (GS-6R Centrifuge, Beckman, Ramsey, MN, USA) for 10 min. The fluid was filtered through filter paper and lipid was concentrated

using a nitrogen gas concentrator (MGS-2200, Eyelaa Tokyo Rikakikai Co., Ltd, Tokyo, Japan) following the method of AOAC (1990). Each of the fatty acid methyl ester standards (Sigma-Aldrich Co., Saint Louis, MO, USA) was qualitatively compared with retention time, and the analytic conditions were used for gas chromatography (Agilent 6890N, Agilent Technologies, Santa Clara, CA, USA). For this, a sample of beef and tissue was kept for the split ratio of 1:10. The oven injector was heated with 220 °C. A carrier of gas 1 mL/min was heated at 150 °C for one min. A column HP-Innowax (30 m length × 0.32 id × 0.25 µm thickness) was kept for a detector temperature of 275 °C. The oven maintained the temperature of 200 °C to 250 °C at 3 °C/min and 250 °C for 5 min.

Statistical analysis was carried out using the GLM procedure of SAS (Statistical Analysis System, version 9.1) for a completely randomized design. Duncan’s multiple range test was used for ranking treatment means within a significant F test and means were considered significantly different at P<0.05. Means with probabilities between 0.05<P<0.10 reflected a tendency to difference. All the data are reported as the sample mean±the standard deviation. Pairwise comparisons between means of different groups were performed using a t-test. The difference between two subsets of data is considered statistically significant if the t-test gives a significance level P (P-value) less than 0.05.

Rumen fluid parameters including pH, ammonia-N, acetate, propionate, butyrate and valerate were analyzed using PROC MIXED of SAS (Statistical Analysis System, version 9.1) for repeated measurement analysis (Littell et al., 1998) and the means were compared for significance by Tukey’s test. The statistical model used for analyses was as follows:

\[ y_{ijk} = \mu + \alpha_i + d_{ij} + \tau_k + (\alpha \tau)_{ik} + e_{ijk}, \]

in which \( y_{ijk} \) is the response at time \( k \) on animal \( j \) in treatment group \( i \); \( \mu \) is the overall mean; \( \alpha_i \) is the fixed effect of treatment; \( d_{ij} \) is the random effect of animal \( j \) in treatment group \( i \); \( \tau_k \) is a fixed effect of time \( k \); \( (\alpha \tau)_ik \) is a fixed interaction effect of treatment \( i \) with time \( k \); and \( e_{ijk} \) is random error at time \( k \) on animal \( j \) in treatment \( i \). Variance and covariance assumption structures (AR(1), UN, CS, etc.) were tested, then AR(1) was selected as the best covariance structure for final analysis.

**Results**

Dry matter intake (DMI), body weight, ADG, and FE were not different (P>0.05) between the two groups (Table 2).

Rumen ammonia-N increased for the first 3 h of the post-feeding period and then decreased; however, no difference (P>0.05) was observed between the two treatment groups (Figure 1). Ruminal propionate, butyrate, and valerate showed no differences (P>0.05) between the two treatment groups (Figure 2).

In the current study, no significant differences (P>0.05) in composition of fatty acids (myristate, stearate, oleate, and gamma linoleate) were observed. However, linoleate...
C18:2n6, P<0.05), palmitate (C16:0, P = 0.04), and palmitoleate (C16:1n7, P = 0.03) were higher in the 60F group than the 40F group. Oleate was higher (P = 0.08) in 60F group than 40F group (Table 3).

Cr-Met had no effect (P>0.05) on saturated, unsaturated, or unsaturated/saturated fatty acids ratio values (Table 3).

**Discussion**

The similar DM intake in both groups may explain the lack of differences in body weight of steers and consequently ADG and FE between 40F and 60F groups (Table 2). It appears that the 20% difference in forage ratio between the two groups could not affect DM intake. Ohh and Lee (2005) concluded that Cr-Met supplementation may not improve body weight of animals. However, by contrast, it has been reported that a higher forage ratio may lead to lower DM intake and lower weight gain in steers in a longer period of growth (Song et al., 2013; Sung et al., 2015).

A timely decrease in pH post-feeding was predictable in both groups; however, in 60F group, these changes were slighter than in 40F group (Figure 1). Because of the higher concentrate ratio in 40F, a higher amount of soluble carbohydrate may result in lower pH. Ghassemi Nejad et al. (2012, 2016) reported lower pH in Brown Swiss heifer and Holstein steers fed a higher concentrate ratio. Normally, pH begins to decline post-feeding and continues to fall down after 3 h. Other factors can influence pH decreasing trend, including but not limited to synchronization between

**Table 3 - Fatty acid profile of meat from Hanwoo steers fed high- and low-forage diets supplemented with chromium-methionine**

<table>
<thead>
<tr>
<th>Item</th>
<th>Treatment 40F</th>
<th>Treatment 60F</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>C14:0 (myristate)</td>
<td>3.6±0.5</td>
<td>3.9±0.3</td>
<td>0.53</td>
</tr>
<tr>
<td>C16:0 (palmitate)</td>
<td>28.3±1.3b</td>
<td>29.4±1.4a</td>
<td>0.04</td>
</tr>
<tr>
<td>C16:1n7 (palmitoleate)</td>
<td>4.2±0.7b</td>
<td>5.5±0.9a</td>
<td>0.03</td>
</tr>
<tr>
<td>C18:0 (stearate)</td>
<td>10.3±1.4</td>
<td>10.2±1.2</td>
<td>0.73</td>
</tr>
<tr>
<td>C18:1n9 (oleate)</td>
<td>46.9±2.3</td>
<td>48.2±2.0</td>
<td>0.08</td>
</tr>
<tr>
<td>C18:2n6 (linoleate)</td>
<td>2.6±0.6b</td>
<td>3.6±0.7a</td>
<td>0.03</td>
</tr>
<tr>
<td>C18:3n6 (gamma-linoleate)</td>
<td>0.3±0.08</td>
<td>0.3±0.04</td>
<td>0.35</td>
</tr>
<tr>
<td>SFA</td>
<td>46.3±1.7</td>
<td>46.4±2.3</td>
<td>0.45</td>
</tr>
<tr>
<td>UFA</td>
<td>54.7±3.1</td>
<td>54.6±2.7</td>
<td>0.57</td>
</tr>
<tr>
<td>UFA/SFA</td>
<td>1.2±0.1</td>
<td>1.2±0.1</td>
<td>0.91</td>
</tr>
</tbody>
</table>

SFA - saturated fatty acids; USA - unsaturated fatty acids.

Means ± standard deviation.
ab - values within a row with different letters differ significantly at P<0.05.

![Figure 2](image-url) - Diurnal changes in ruminal acetate, propionate, butyrate, and valerate acids in Korean beef steers fed low- and high-forage diets supplemented with chromium-methionine (hourly basis).
carbohydrate and protein, rate of passage of rumen flora, digesta flow rate, and microbial protein production (Sung et al., 2015; Ghassemi Nejad et al., 2013).

A slight increase in ammonia-N content of rumen 3 h post-feeding and then a decline were expected due to the availability of nitrogen for both feeding groups. Microbial protein synthesis depends on the availability of nitrogen (or amino acids) and on the degradable carbohydrate content, or on the content of organic matter, and the synchronization in rumen degradation between protein and carbohydrates. If there are high amounts of degradable nitrogen or if the content and rate of carbohydrate degradation in the rumen is reduced or not synchronized with the degradation of protein, losses of nitrogen and/or energy in the rumen could occur (Ghassemi Nejad et al., 2014, 2015).

Ruminal acetate was higher (P<0.05) in 60F compared with 40F group, and remained higher post-feeding (Figure 2). The higher acetate in rumen of 60F group can be explained by the higher forage ratio resulting in higher pH values. Higher pH values in 60F group favor rumen fiber digestibility and maintain stability of pH in due time (Sung et al., 2015). Moreover, a higher forage ratio in the diet causes higher secretion of saliva, which fosters higher and stable rumen pH. The content of volatile fatty acids and the acetate/proprionate ratio may change due to different forage: concentrate ratios. Higher acetic acid may be observed in the rumen fluid of steers fed diets with higher forage ratios, as described by several researchers (Merchen et al., 1986; Kinser et al., 1988; Zinn et al., 1994). Therefore, a large amount of forage in 60F, which may have resulted in more salivary flow to rumen, provides favorable conditions for the better microbial growth. This may explain the higher acetate production in rumen fluid in 60F compared with 40F group. In 40F group, a higher concentrate ratio is a major factor for lower ruminal pH and consequently lower acetate production. Zinn and Plascencia (1996) found no significant differences in pH, acetate, propionate, and butyrate between feedlot cattle fed various levels of alfalfa (10% compared with 30%).

The meat fatty acid composition has a promising role in the quality of beef for consumers (Lindemann et al., 2008; Sung et al., 2015; Ghassemi Nejad et al., 2016). Chromium can influence fat mobilization from body stores to meet nutrient requirements. Hence, alterations in some fatty acid compositions can be expected. Chromium-methionine has a high potential to be absorbed through increased digestion in the gastro-intestinal tract (Ohh et al., 2004; Sung et al., 2015; Ghassemi Nejad et al., 2016). The finishing diet strongly influences the fatty acid composition of beef (Smith et al., 2009; Ghassemi Nejad et al., 2016). Zea et al. (2007) reported higher saturated fatty acids (SFA) in animals fed concentrates. Furthermore, Smith et al. (2009) reported that grain feeding stimulates the activity of adipose tissue stearoyl-CoA desaturase in marbling adipose tissue and lowers ruminal isomerization/hydrogenation of dietary polyunsaturated fatty acids (PUFA), resulting in a noticeable increase in monounsaturated fatty acids (MUFA) in beef over time. However, the current study found no differences in the SFA, unsaturated fatty acids (UFA), or UFA/SFA values, which were previously reported in Holstein steers by Ghassemi Nejad et al. (2016).

Conclusions

Chromium-methionine supplementation in high-forage (60%) diets can improve meat quality regarding fatty acid composition by increasing the palmitic, palmitoleic, and linoleic acid contents.

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