STUDIES ON ANTI-COMPONENT 5 ANTIBODIES IN ANIMALS INFECTED WITH TRYPANOSOMA CRUZI

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A competitive antibody enzyme immunoassay, using a monoclonal antibody against the species-specific Trypanosoma cruzi antigen 5, was used to investigate the presence of anti-component 5 antibodies in sera of opossums, dogs, rabbits and rats infected with this parasite. The sera from 51 Venezuelan patients with Chagas' disease were also tested. About 90% of the infected subjects showed significant levels of anti-component 5 antibodies. Nevertheless, these antibodies were not detected in the sera of dogs, rats and opossums infected with T. cruzi. Some sera from infected rabbits presented significant results but close to the limit of positivity of the test. These findings suggest that the immune response in animals naturally or experimentally infected with T. cruzi is different from that observed in human Chagas' disease.

Key words: Trypanosoma cruzi, Antibodies, Immunoassay.

The association of Trypanosoma cruzi with T. rangeli¹¹ ²³ or leishmanial infections¹² ¹³, has been reported many areas of Central and South America. As there are important antigenic cross-reactivities between these different parasites³ ⁸ the use of classical serological tests for the diagnosis of Chagas’ disease becomes ineffective in these coendemic areas. In an attempt to gain better specificity for diagnosis, several technical approaches have been used⁹ ¹⁰ ¹⁴ ¹⁵ ¹⁶ ¹⁷ ¹⁸ ¹⁹ ²⁰.

In previous studies, Afchain et al³ have identified a species-specific antigen called component 5, in the T. cruzi parasite. Recently, by using murine monoclonal antibodies directed against this component¹⁸ ¹⁹, Lemesre et al¹⁶ proposed a competitive enzyme immunoassay (CEIA) for the specific diagnosis of Chagas’ disease. The purpose of the present work was to use the CEIA to investigate anti-component 5 antibodies in sera of dogs, rabbits, opossums and rats infected with T. cruzi and to compare it with that find in patients with American trypanosomiasis.

MATERIALS AND METHODS

Animal sera

Twenty-eight mongrel dogs of both sexes, 2-5 months old, were inoculated intraperitoneally with 4-6 x 10⁵ T. cruzi trypomastigotes of the 12SF strain per kg of body weight as described by Andrade et al⁴. They all developed positive parasitaemias at 5-7 days after inoculation and sera were collected at different times of both the acute and the chronic phase of the infection. Although parasitaemia became negative at the chronic stage, xenodiagnosis and/or the complement fixation test remained positive for all animals. A total of 40 serum samples of infected dogs was used in this study. The sera from 14 dogs before inoculation were used as control.

Thirteen New Zealand white rabbits, weighing 1200-1500 g, were inoculated intraperitoneally with 10⁷ blood stream forms of T. cruzi (12 SF strain) per kg of body weight. Parasitaemias in the acute phase, and positive xenodiagnosis and indirect immunofluorescence in the chronic phase confirmed the infection. A total of 24 serum samples collected at several times after inoculation was used in this study. Seven pre-inoculation sera from the same rabbits were used as control.

Fifteen male Fischer rats, weighing 180-200 g were inoculated intraperitoneally with 1 ml heparinized rat blood containing 10⁴ T. cruzi trypomastigotes of the Tehuantepec strain as described by Rodriguez et al²⁰. The infection was monitored by the presence of parasites in the blood and by a positive passive hemagglutination test¹⁹. A total of 15 serum samples was collected at different times after infection. The sera from 8 normal Fischer rats were used as control.

The sera from 25 opossums (Didelphis albitratus), captured in an endemic area for Chagas’ disease (Castro Alves, Bahia state, Brazil) have also been studied. Twelve were naturally infected with T. cruzi as confirmed by a positive xenodiagnosis. The sera from 14 opossums, captured in a non-endemic area for Chagas’ disease (Jacobina, Bahia state, Brazil) and with a negative xenodiagnosis, were used as control. Two of them showed a positive culture for Leishmania spp.
Human sera

The sera from 51 Venezuelan patients with chronic Chagas' disease and a positive serology for *T. cruzi* (immunofluorescence, complement fixation test, haemagglutination and ELISA) were generously provided by Dr. Rosa M. Mubsch, Universidad de Carabobo, Facultad de Ciencias de la Salud, Maracay, Venezuela. Control sera were obtained from 20 healthy subjects with a negative serology for *T. cruzi*.

Monoclonal antibody (mAb)

Hybridomas secreting mAb against component 5 has been described by Orozco et al. Ascitic fluid were produced in Balb/c mice and the mAb (II-190/30) was purified by a 50% ammonium sulfate precipitation followed by a ion exchange chromatography on DEAE-Trisacryl (IBF, Villeneuve la Gaillarde, France). Labelling of purified mAb with alkaline phosphatase (grade I from calf intestine, Boehringer, Mannheim, West Germany) was performed by the one-step glutaraldehyde method of Avrameas. Briefly: to 5mg of enzyme were added 1ml of the mAb solution (2mg/ml) and 10μl of 20% glutaraldehyde solution. The mixture was allowed to react for 2 h at room temperature and then dialysed against PBS.

Competitive antibody enzyme immunoassay (CEIA)

It was performed as described by Lemesre et al. Briefly, polypropylene beads (Seroa, Monaco) were coated overnight at room temperature with a component 5-enriched *T. cruzi* antigen. After 3 washes in PBS containing 0.1% Tween 20, the beads were incubated for 2 hr with PBS-0.1% bovine serum albumin, washed 3 times with PBS-Tween and once with PBS. CEIA was carried out in disposable polystyrene tubes. Coated beads were incubated for 3 hr at 37°C in 350 μl of PBS containing the labeled mAb (final dilution 1/800) and the test sera (final dilution 1/20). After 3 washes in PBS-Tween, the beads were transferred to another tube and the labelled antibody fixed to them was revealed by adding 300 μl of the enzyme-substrate (1 mg/ml 4-para-nitrophenylphosphate in 0.5 M Na₂CO₃, 0.001 M MgCl₂ buffer pH 10.4). After contact for one hour at 37°C, the colour development was stopped by adding 300 μl NaOH 2N and absorbance was measured at 405 nm. As the CEIA involves the competitive inhibition of binding of alkaline phosphatase-conjugated mAb by anti-component 5 antibodies present in test sera low extinction values indicate a positive result.

Immuno-electrophoresis

The presence of anti-component 5 antibodies in sera of some animals infected with *T. cruzi* was also investigated by immuno-electrophoresis as described by Aachine et al.

RESULTS

The presence of anti-component 5 antibodies in sera of dogs, rats, opossums, rabbits and patients infected with *T. cruzi* was investigated by the competitive antibody enzyme immunoassay (CEIA) with enzyme-conjugated anti-component 5 mAb (Fig. 1). The limit of positivity for each animal species studied was calculated in each control group as the mean subtracted of twice the standard deviation. The results were then slightly corrected in order to have the same cut-off value as that observed with normal human sera. Therefore, the data obtained for each group of animals infected with *T. cruzi* could be compared in the same Figure 1 with the results from the patients with Chagas' disease (positive control). In these conditions, anti-component 5 antibodies were not detected in sera of dogs, rats and opossums infected with *T. cruzi*. In fact, only 4 out of 40 sera of dogs, 1 out of 13 opossums with negative xenodiagnosis, and 3 out of 15 sera of rats showed significant results which were near the limit of positivity. In the group of rabbits infected with *T. cruzi*, about the half of
the tested sera presented a significant positive value, but most of them was closed to the limit of positivity. There was no association between these results and the phase of the infection.

To analyse further the presence of anti-component 5 antibodies in experimental Chagas’ disease, the sera of the infected animals showing a significant result in the CEIA were tested by immunoelectrophoresis against a crude antigenic extract of *T. cruzi*. Although most of the sera from infected animals recognised different components in the *T. cruzi* extract, none of them reacted with the component 5 (data not shown).

**DISCUSSION**

A previous study demonstrated that more than 90% of patients with Chagas’ disease presented significant high levels of anticomponent 5 antibodies. In the present investigation, by using the same methodology, the CEIA with enzyme-conjugated mAb we have confirmed these findings in another group of patients but failed to detect significant levels of anti-component 5 antibodies in dogs, rats, opossums and rabbits infected with *T. cruzi*.

The interest in component 5 begun when Afchain et al. showed that this antigen represents one of the major immunoelectrophoretic lines recognized by the sera of most of the patients with Chagas’ disease. Furthermore, it has been demonstrated that component 5 is a species-specific antigen of *T. cruzi*. The generation of mAb against this antigen allowed the identification of a 72,000 Mr glycoprotein and its maturation products of 51,000, 43,000 and 24,000 Mr as the molecules containing the antigenic determinants of component 5. In order to avoid the cross-reactivities frequently observed between *T. cruzi* and others trypanosomatidae, Breniere et al. and Lemesre et al. have successfully used different techniques to detect anti-component 5 antibodies in sera of patients infected with *T. cruzi*. These methods have proved to be useful for the differential immunodiagnosis of human Chagas’ disease.

The importance of different mammalian species as reservoir hosts in American trypanosomiasis has been showed by several investigators. As there area many areas in Central and South America in which Chagas’ disease is coendemic with other trypanosomatidae infections, the detection of anticomponent 5 antibodies in wild reservoirs would be important for epidemiological purposes. Nevertheless, in the present study by using a highly sensitive technique, the CEIA, we were unable to find this antibody in opossums naturally infected with *T. cruzi*. In addition, dogs and rats experimentally infected with this parasite did not present significant levels of anti-component 5 antibodies. Finally, the detection of lower levels of this antibody in some sera of rabbits infected with *T. cruzi* could not be confirmed by immunoelectrophoresis, a less sensitive technique.

In previous studies, Kirchhoff et al. have shown that the expression of a surface antigen identified by a mAb differs among strains and clones of *T. cruzi*. This suggests that strain-specific immune responses can occur during the infection. Nevertheless, the anti-component 5 mAb generated against the Tehuantepec strain of *T. cruzi* also reacted with the 12 SF strain (Andrade et al.: in preparation) used here to infect dogs and rabbits. Moreover, in the present work, the rats were infected with the same strain used to produce the mAb. Therefore, the possibility of a strain-specific immune response against component 5 as an argument to explain the absence of anti-component 5 antibodies in dogs, rats and rabbits infected with *T. cruzi* is excluded.

The presence of anti-component 5 antibodies in sera of patients with Chagas’ disease and their absence in animals naturally and experimentally infected with *T. cruzi* argue in favour of a different immune response during the animal infection as compared to that in human beings. Probably, component 5 is not immunogenic in various animals infected with *T. cruzi*. It is noteworthy that sera from rabbits and mice immunized with antigenic extracts of *T. cruzi* recognized strongly this antigen. Our results can be related to a recent observation by Lemesre et al. showing that the humoral immune response in mice infected with *Leishmania chagasi* is different from that noticed in patients with kala-azar. All these findings argue against the use of animals experimentally infected with trypanosomatidae as immunological or immunopathological models of the disease.

**RESUMO**

Foi usado um teste imunoenzimático competitivo para investigar a presença de anticorpos anticomponente 5 nos soros de sargentões, cães, coelhos e ratos infectados com o Trypanosoma cruzi. Neste teste, foi utilizado um anticorpo monoclonal contra o antígeno 5 que é específico do *T. cruzi*. Também foram testados os soros de 51 pacientes venezuelanos com doença de Chagas.

Apar desses anticorpos não serem detectados nos soros de cães, ratos e sargentões infectados com o *T. cruzi*, alguns soros de coelhos infectados apresentaram resultados positivos porém em níveis próximos aos do limite de positividade do teste. Esses achados sugerem que a resposta imune em animais naturalmente ou experimentalmente infectados com o *T. cruzi* é diferente daquela que é observada na doença de Chagas humana.

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REFERENCES


