Possible artemisinin-based combination therapy-resistant malaria in Nigeria: a report of three cases


[1]. Department of Internal Medicine, Federal Teaching Hospital Abakaliki, State of Ebonyi, Nigeria.

ABSTRACT
Artemisinin-based combination therapy-resistant malaria is rare in Sub-Saharan Africa. The World Health Organization identifies monitoring and surveillance using day-3 parasitaemia post-treatment as the standard test for identifying suspected artemisinin resistance. We report three cases of early treatment failure due to possible artemisinin-based combination therapy-resistant Plasmodium falciparum malaria. All cases showed adequate clinical and parasitological responses to quinine. This study reveals a need to re-evaluate the quality and efficacy of artemisinin-based combination therapy agents in Nigeria and Sub-Saharan Africa.


INTRODUCTION
The World Health Organization (WHO) currently recommends artemisinin-based combination therapy (ACT) as the first-line treatment for uncomplicated Plasmodium falciparum malaria in Sub-Saharan Africa (SSA)1. The combination consists of an artemisinin derivative with a rapid effect on parasite clearance and a longer-acting drug to prevent recrudescence. The massive reductions in malaria-related morbidity and mortality in regions of high endemicity in the last decade have been in part due to the effectiveness of the ACT regimen2. However, these successes are threatened by the emergence of artemisinin-resistant strains of Plasmodium falciparum from the Thai-Cambodian border3 and Thai-Myanmar border4. Indeed, artemisinin resistance is a major threat to global health, particularly in low- and middle-income countries (LMICs), in which the disease burden is highest, substandard or counterfeit ACT compounds are widely available, and systems for the monitoring and containment of resistance are inadequate5-7.

There is little existing knowledge regarding ACT-resistant malaria in many SSA countries, including Nigeria, and the most recent reports of ACT treatment failures were in travelers who had recently visited African countries8-10. Additionally, there have been no reports of delayed parasite clearance in routine therapeutic efficacy studies conducted in Africa11. Thus, arguments for the presence of artemisinin resistance in Africa have been based solely on in vitro and/or molecular analyses of parasites collected from autochthonous patients or returning travellers4,12. However, standard in vitro tests are not reliable tools for monitoring artemisinin resistance13. In addition, none of the putative molecular markers for antimalarial drug resistance has been correlated with delayed clearance after treatment with artemisinin1. We recently encountered three cases of Plasmodium falciparum malaria that showed early treatment failure (ETF) of ACT and adequate clinical response to quinine.

CASE REPORT
Case 1
A 36-year-old female presented with a three-day history of high-grade fever with chills and a two-day history of productive cough associated with pleuritic chest pain and yellowness of the eyes. The patient is a known sickle cell disease patient. She had taken oral sulfadoxine-pyrimethamine and ciprofloxacin one day before presentation. On examination, she had a temperature of 39.5°C and was moderately pale, icteric, and mildly dehydrated. She had a respiratory rate of 44 cycles/minute, and her breath sounds were vesicular with coarse crackles in the right lower lung zone. Her pulse rate was 108 per minute and her blood pressure was 130/70mmHg. Abdominal and neurologic examinations were unremarkable. The provisional diagnosis was hemolytic crisis precipitated by sepsis from community-acquired pneumonia and poorly treated malaria in a known sickle cell disease patient. Blood samples were collected for complete blood count (CBC), film for malaria parasites, serum electrolytes, urea, and creatinine, hepatic enzymes, and blood culture assays. Chest X-ray, urinalysis, and urine culture were also performed.

The patient received intravenous ceftriaxone, oral Coartem (artemether-lumefantrine), oral erythromycin, oral paracetamol, and hematins. Her blood film showed Plasmodium falciparum parasitemia (+), and her CBC showed anemia and leukocytosis (Table 1). Urinalysis revealed nitrate (+) and proteinuria (+).
TABLE 1 - Complete blood counts of the three cases of possible ACT-resistant malaria at presentation.

<table>
<thead>
<tr>
<th>Variable</th>
<th>Case 1</th>
<th>Case 2</th>
<th>Case 3</th>
<th>Reference value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hemoglobin</td>
<td>9</td>
<td>9.3</td>
<td>5.6</td>
<td>12 – 16g/dl</td>
</tr>
<tr>
<td>WBC count</td>
<td>28</td>
<td>9.9</td>
<td>1.5</td>
<td>4 – 11 x 10^6 cells/liter</td>
</tr>
<tr>
<td>Neutrophils</td>
<td>68</td>
<td>78</td>
<td>22</td>
<td>40 – 70%</td>
</tr>
<tr>
<td>Lymphocytes</td>
<td>31</td>
<td>20</td>
<td>77</td>
<td>20 – 40%</td>
</tr>
<tr>
<td>Eosinophils</td>
<td>1</td>
<td>0.2</td>
<td>0.1</td>
<td>1 – 6%</td>
</tr>
<tr>
<td>Monocytes</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>2 – 10%</td>
</tr>
<tr>
<td>Basophils</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0 – 1%</td>
</tr>
<tr>
<td>Thrombocytes</td>
<td>180</td>
<td>162</td>
<td>178</td>
<td>150 – 400 x 10^9/liter</td>
</tr>
<tr>
<td>Malaria parasite</td>
<td>(++)</td>
<td>(+++)</td>
<td>(+)</td>
<td>positive (+) / negative (-)</td>
</tr>
</tbody>
</table>

ACT: artemisinin-based combination therapy; WBC: white blood cell.

Other requested investigations were unremarkable. After 72h of treatment, the patient was still febrile (temperature 39.4°C), though her cough and chest pain were subsiding. A repeat blood film showed the continued presence of malaria parasites. The patient was started on injections of quinine (600mg in glucose infusion) every 8h for three days, while the antibiotic treatment continued. Two days after starting the quinine, she showed a progressive reduction in temperature, and her fever resolved (temperature 37.1°C) after 72h. Oral quinine and doxycycline were then administered for the next four and seven days, respectively. Repeat blood films after days 7 and 28 showed no malaria parasitemia.

Case 2

This patient was a 33-year-old female health worker who presented with a two-day history of fever, headache, and abdominal discomfort. The fever was high-grade, intermittent, with chills, and she suffered four episodes of vomiting one day before presentation. The patient had taken sulfadoxine-pyrimethamine prior to presentation. On examination, her temperature was 39.2°C, and her respiratory rate was 38 cycles/minute. Other examination findings were unremarkable. A provisional diagnosis of uncomplicated malaria and urinary tract infection was made. Blood film for plasmodium parasitemia, CBC, urinalysis, urine culture; serum electrolytes, urea, and creatinine; and abdomino-pelvic ultrasound scan were performed. A blood smear showed severe *Plasmodium falciparum* parasitemia (+++). Her CBC showed anemia with some neutrophilia (Table 1). Other requested investigations were unremarkable. She was given analgesics, intramuscular artesunate, and oral amodiaquine for three days.

Three days later, her fever remained high (temperature 38.5°C), and she became excessively weak. A repeat blood film showed persistent *Plasmodium falciparum* parasitemia (+++), and a diagnosis of severe malaria was considered. Intravenous artesunate was started. However, her clinical state deteriorated at 48 hours after commencing artesunate, and the fever and parasitemia persisted. A diagnosis of probable ACT-resistant malaria was made. Injections of quinine (600mg infused every 8h) was begun. After two days on quinine, the patient showed a progressive reduction in temperature, which fell to 36.9°C. The injectable quinine was changed to oral quinine, and she was discharged the next day. A repeat blood smear performed on days 7 and 28 after starting quinine were both negative for malaria parasitemia.

Case 3

A 54-year-old woman presented with a six-day history of high-grade fever and a four-day history of cough. She reported having five episodes of vomiting two days before presentation. The cough was associated with pleuritic chest pain and was productive, with yellowish sputum. On examination, her temperature was 38.9°C, her respiratory rate was 48 cycles/minute, and she had coarse crepitations on the left middle lung zone. The patient was managed for community-acquired pneumonia to rule out malaria. Investigations performed included the following: chest X-ray, sputum culture, sputum for acid-fast bacilli, CBC, blood film for malaria parasites, and serum electrolytes, urea, and creatinine. She was initially given oral amoxicillin-clavulanate and clarithromycin while awaiting results. The blood smear showed *Plasmodium falciparum* parasitemia (++). Her CBC showed severe anemia with leucopenia, neutropenia, and lymphocytosis (Table 1). The sputum culture yielded an alpha-hemolytic streptococcus sensitive to ofloxacin, ceftriaxone, and amoxicillin-clavulanate. Other requested results were unremarkable. The oral antibiotics were continued, and oral Coartem (artemether-lumefantrine) was started. The patient also received a transfusion of two units of fresh whole blood.

After 72h of treatment, the patient still exhibited a fever (temperature of 38.1°C), though her cough and chest pain were resolving. A repeat blood film showed *Plasmodium falciparum* parasitemia (+). Antibiotics were continued. On the sixth day of treatment, the fever persisted, with a temperature of 37.8°C, but the cough and chest findings had resolved. However, a blood film still showed *Plasmodium falciparum* parasitemia (+); thus, a diagnosis of ACT-resistant malaria was considered. Oral quinine was given (600mg every eight hours for seven
days). The fever resolved within the next 48h, and the patient was discharged home the next day. Two blood films performed on days 14 and 28 after the initiation of quinine were negative for malaria parasites.

**DISCUSSION**

Although ACT-resistant malaria is rare in Africa, the WHO advocates increased monitoring and surveillance to identify and contain artemisinin resistance. According to the WHO classification of malaria treatment outcomes, these patients had ETF. This is particularly worrisome because the cases also fulfill the WHO definition for suspected artemisinin resistance, i.e., detectable peripheral parasitemia 72h after starting anti-malarial treatment. Because the in vivo estimation of the parasite clearance rate from peripheral blood parasite counts is becoming the standard method used for the surveillance of artemisinin resistance, it is necessary to use this method to maintain surveillance for ACT-resistant malaria.

Artemether-lumefantrine remains a very commonly used ACT for the treatment of uncomplicated *Plasmodium falciparum* malaria in SSA, and a recent pooled analysis showed a 28-day parasitological cure rate of 97% in malaria-endemic areas. Late clinical failure (LCF) of artemether-lumefantrine has been reported in three travelers who became ill after visiting Sierra Leone, Tanzania, and Congo. Because of the lipophilic nature of lumefantrine, its bioavailability increases when taken with fatty meals. Due to a lack of concurrent pharmacological evaluation, two previous reports speculated that sub-optimal lumefantrine concentrations were responsible for the observed LCF. However, pharmacological studies and genotyping in one recent report showed adequate plasma concentrations of lumefantrine and indicated that the clinical failure was due to parasite recrudescence rather than recurrent infection.

This is the first report from Nigeria showing ETF with artemether-lumefantrine. Two of the cases described received full doses of artemether-lumefantrine without good clinical response. Although the observed ETF could have been caused by the use of substandard/counterfeit artemether-lumefantrine compounds, several other patients with malaria received and responded to the same drug, and there are no current reports of in vivo resistance to artemether-lumefantrine in Nigeria. Furthermore, the second case described received and failed to respond to a combination of arteether and amodiaquine. Moreover, all the cases showed adequate clinical and parasitological response to quinine. Therefore, the ETF in the patients described may be due to ACT resistance.

This report highlights the need for closer monitoring and surveillance for AC-resistant *Plasmodium falciparum* malaria and calls for the urgent reassessment of the quality and efficacy of ACTs in Nigeria. Finally, there is a need to discover the genetic factors of the parasite that cause artemisinin resistance and to identify effective markers to monitor its spread in Nigeria and other LMICs.

**REFERENCES**