**Note**

GROWTH AND SURVIVAL OF PINK SHRIMP (*Farfantepenaeus paulensis*) POSTLARVAE IN CAGES AND PEN ENCLOSURES

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ABSTRACT: Technology for the culture of the pink shrimp *Farfantepenaeus paulensis* in low-cost cages and pen enclosures was developed together with artisanal fishermen of the Patos Lagoon estuary, southern Brazil. Although the production of postlarvae (PL) under laboratory conditions is well documented, survival and growth during the nursery phase cages and pen enclosures in the nature has not yet been analyzed. Therefore, the present study compared survival and growth of *F. paulensis* postlarvae reared either in cages or pens. Six cages (2.0 × 2.0 × 1.4 m) of PVC-coated polyester with a mesh size of 1.5 mm were stocked with 800 PL each. To assess the effect of the substrate, a 10 cm layer of sand was added to three of these cages to simulate a pen enclosure. No difference in the mean survival rate between cages (92.2%) and pens (88.7%) was found (*P* > 0.05). However, final weight of shrimp reared in pens (1.05 ± 0.05 g) was higher than those kept in cages (0.88 ± 0.12 g). The nursery phase of *F. paulensis* in cages and pens did not affect survival. Nevertheless, because of the higher growth rate of shrimp grown in pen enclosures, this kind of structure should be preferentially used in nursery rearing of this species.

Key words: Penaeidae, aquaculture, shrimp farming, nursery

INTRODUCTION

Shrimp culture has received a great deal of attention worldwide mainly because of the growing market demand and high commercial value attained by the product. In general, shrimp culture is conventionally carried out in tanks or ponds with relatively high production costs. On the other hand, in some Asian countries (Thailand, Philippines, Singapore, and India) alternative culture methods, utilizing cages and pen enclosures, have been utilized (Peña & Prospero, 1984). The application of these systems usually results in lower production costs, which allows lower economic income communities to culture shrimp (Walford & Lam, 1987).

Although cages and pens are mainly used for culturing fish, many researchers are also investigating their...

In the state of Rio Grande do Sul, many studies have been performed to determine the best conditions for the culture of the pink-shrimp *Farfantepenaeus paulensis* either under laboratory (Tsuzuki et al., 2000; Thompson et al., 2002) or natural environment conditions (Zogbi, 1997; Santos et al., 1999). All these studies indicated the high growth potential of this species. Although the technology for the production of post-larvae of the species in laboratory is well developed, there is still need for information on its culture in the nature, especially in relation to survival and growth of post-larvae during the nursery phase.

The nursery phase is an intermediate step between larviculture and grow-out, and is usually characterized by high stocking densities, high water renewal rates and feeding with top quality artificial diets (Speck et al., 1993). The main objective of the nursery phase is producing larger and sturdier juveniles, which will probably have a better chance of survival and may achieve commercial size in shorter time (Apud et al., 1983). For *F. paulensis*, the nursery phase usually begins when shrimps attain 0.06 g (PL₂), and last until they reach over 0.36 g or 36 mm of total length (Dolci & Wasiel esky, 1998). These values were stipulated to avoid shrimp escaping through the mesh of the larger pen enclosures used to grow *F. paulensis* up to commercial size (Wasiel esky, 2000).

The purpose of this work was to compare the survival and growth of *F. paulensis* post-larvae reared in cages or pens during the nursery phase in a natural environment. The percentile of the post-larvae population with weight over 0.36 g was also evaluated in order to define the best moment for transferring them to grow-out structures.

**MATERIAL AND METHODS**

The experiment was set up in Saco do Justino bay, a shallow estuarine inlet of the Patos Lagoon, Rio Grande, RS (32°03’ S, 52°05’ W), southern Brazil, during February and March 2001. The experimental period lasted 31 days. Six cages were divided in two treatments (cages and pens) with three replicates. Dimension of the cages was 2.0 × 2.0 × 1.4 m (length x width x height). Cages built with PVC-coated polyester (1.5 mm mesh size) were supported by bamboo poles. Each unit had 4 m² of bottom area and was stocked with 800 PLx, which corresponded to a density of 200 PL m². To evaluate the influence of the substrate, three cages were evenly filled with a 10 cm sediment layer order to simulate the conditions of pens. The sediment was obtained from the same estuarine inlet. This procedure was used to facilitate the evaluation of the final survival, which could be more difficult if bottomless pens were used.

The post-larvae used in the experiment were cultured during 26 days (PLx). At this age, they already acquired a higher tolerance to low salinity levels (Tsuzuki et al., 2000), which allowed them to be transferred to the experimental units after previous acclimatization. During the experimental period, post-larvae were fed twice a day, at 9h00 with a commercial diet, and at 21h00 with fresh minced crustaceans (crab and shrimp heads). Feed was provided in one feeding tray per experimental unit. Initial feeding rate was 50% of the total biomass and was periodically adjusted according to the amount of feed left in the trays.

Monitoring of growth was performed at days 1, 14, 22, 26, and 31 after stocking. One hundred shrimp from each experimental unit were randomly sampled, weighed (wet weight) and returned to the respective unit. At the end of the trial, the interval between successive growth measurements was gradually decreased to allow a closer growth monitoring as shrimp were reaching the target weight of 0.36 g.

Water temperature (0.5°C precision thermometer) and salinity (one unit precision optical refractometer) were measured daily. Water samples from each experimental unit and a control point placed 30 m away from the culture site were collected at morning (9h00) to measure dissolved oxygen (Strickland & Parsons, 1972), pH (desktop pH meter), total ammonium (UNESCO, 1983) and nitrite concentrations (Benscheneider & Robinson, 1952).

Final weight and number of shrimp of each replicate were submitted to analysis of variance (ANOVA), and if no differences were found (*P > 0.05*) results were pooled. The same procedures were used to determine differences between the abiotic data of the different treatments and the control point.

**RESULTS AND DISCUSSION**

Deterioration of water quality may cause stress to cultured organisms, negatively affecting growth and increasing their susceptibility to pathogenic agents that may cause mortality (Arulampalam et al., 1998). This is the reason why monitoring water quality is an indispensable procedure if the culture of any aquatic organisms is to be successful. In open culture systems, such as cages and pens, water minor quality problems may occur, but this will certainly depend on the water renewal rate of the place where the structures are installed.

In the present study, water temperature was 26.06 ± 1.56°C with minimum and maximum values ranging from 24.0 to 29.0°C, while salinity ranged between 4.0 and 10.0 ‰, averaging 7.28 ± 1.74 ‰. The mean con-
centration of dissolved oxygen was 7.76 (± 0.67) mg L\(^{-1}\)
and pH was 8.58 (± 0.14). Means (± SD) of nitrite and
total ammonium were 0.06 ± 0.04 mg L\(^{-1}\) and 0.08 ±
0.06 mg L\(^{-1}\), respectively. All measured water quality pa-
rameters were within the tolerance limits considered not
to affect penaeid shrimp (Vinatea, 1997).

During the first 26 days, no differences in wet
weight were found between treatments (\(P > 0.05\)). How-
ever, at the end of the trial, shrimp cultured in pens reached
a higher weight (1.05 ± 0.05 g) than those reared in cages
(0.88 ± 0.12 g) (\(P < 0.05\)). Wasielesky (2000) cultured \(F.\)
paulensis in cages and pens during 90 days in Patos La-
goan and found that the mean growth rate (g per week)
was higher in pens (0.56 g) than in cages (0.38 g), which
confirms the present results. Pen enclosures seem to allow
better growth of \(F.\) paulensis probably because higher
quantity and diversity of natural food items, especially
benthic organisms, may be present in the sediment.

During the nursery phase, survival is considered
a parameter of great importance in the success of any
shrimp production operation. Ballester et al. (2001) in-
vestigated the role of biofilm (a community of autotrophic
and heterotrophic microorganisms attached to submersed
surfaces) on the survival and growth of \(F.\) paulensis in
nursery systems in natural environment. These authors
used stocking densities of 300 shrimp m\(^{-2}\) and obtained
final survival rates between 96 and 99%. Speck et al.
(1993) reared \(F.\) paulensis post-larvae in an indoor
nursery system at the densities of 150, 300 and 600
shrimp m\(^{-2}\) and obtained survival rates of 85, 84 and 16%,
respectively. \(F.\) paulensis juveniles were reared in the
Patos Lagoon and mean survival values in cages and pens
were 90 and 95%, respectively (Wasielesky, 2000). In the
present study, the type of structure had no influence on
the survival rates of \(F.\) paulensis post-larvae (Table 1).
During the experimental period, the percentile of the
population reaching the target weight of 0.36 g was moni-
tored (Table 2), but no differences among treatments was
recorded (\(P > 0.05\)).

Waiting for all the shrimp to achieve a weight
over 0.36 g may not be an interesting strategy, because
many individuals will have reached a much larger size
by then. Therefore, it would be wiser to have them
transferred to grow-out structures as soon as possible.
The transference should not be retarded in order not
to affect final production. It seems the time that
when 95% of shrimp post-larvae reach over 0.36 g live
weight would probably be the best moment to have
them transferred to grow-out structures. This percentile
was stipulated in function of the cannibalistic nature
of this species, which could negatively influence not
only the size uniformity of the lot, but also their sur-
vival rate.

**CONCLUSION**

\(F.\) paulensis may be cultured in cages or pens dur-
during the nursery phase with no losses in terms of survival.
However, the use of pen enclosures during the nursery phase
is recommended, especially when considering that the in-
stallation costs of pens are comparatively lower than cages,
as less netting material is required per unit area. Therefore,
starting from an economic point of view, the use of pen en-
closures appears to be more attractive.

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**Table 1** - Mean (± SD) final wet weight and survival of \(F.\) paulensis post-larvae reared during 31 days in
cages or pen enclosures.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Final wet weight</th>
<th>Survival</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pen enclosures</td>
<td>1.05 ± 0.05(^a)</td>
<td>88.7 ± 2.8(^a)</td>
</tr>
<tr>
<td>Cages</td>
<td>0.88 ± 0.12(^b)</td>
<td>92.2 ± 3.6(^c)</td>
</tr>
</tbody>
</table>

Within each column, different letters means difference between treatments (\(P < 0.05\)).

**Table 2** - Mean (± SD) percentile of the population of \(F.\) paulensis post-larvae cultured in cages or pens with weight over
0.36 g during the 31 day-long experimental period.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Days</th>
<th>1</th>
<th>14</th>
<th>22</th>
<th>26</th>
<th>31</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pens</td>
<td>0</td>
<td>50.6 ± 12.05</td>
<td>84.0 ± 2.64</td>
<td>93.0 ± 2.00</td>
<td>99.6 ± 0.57</td>
<td>99.6 ± 0.57</td>
</tr>
<tr>
<td>Cages</td>
<td>0</td>
<td>53.6 ± 11.15</td>
<td>84.3 ± 4.04</td>
<td>92.6 ± 1.52</td>
<td>96.6 ± 3.05</td>
<td>96.6 ± 3.05</td>
</tr>
</tbody>
</table>

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