MOLECULAR CONFIRMATION OF MAIZE RAYADO FINO VIRUS AS THE BRAZILIAN CORN STREAK VIRUS

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ABSTRACT: Maize rayado fino virus (MRFV), present in various countries in Latin America, has shown similarities to corn streak virus that occurs in Brazil, regarding pathogenic, serological and histological characteristics. In the current report both virus were molecularly compared to confirm the similarities between them. MRFV was identified by nucleic acid hybridization in samples of maize tissues exhibiting symptoms of “corn stunt” disease, collected from two Brazilian States – São Paulo and Minas Gerais. The coat protein gene and 3’ non-translated region of MRFV were amplified from infected tissues by reverse transcription-polymerase chain reaction (RT-PCR) using MRFV-specific primers, and were characterized by nucleotide sequence and phylogenetic analysis of the cloned PCR products. Phylogenetic analysis of the relationships between the Brazilian isolates and isolates obtained from Latin America and the United States reveals a close relationship to isolates from Brazil, Peru and Bolivia. Results support the proposal that the Brazilian corn streak virus be regarded as an isolate of MRFV and provide evidence for the presence of MRFV in “corn stunt” disease in Brazil.

Key words: MRF virus, corn viruses, phytoplasma, corn stunt

INTRODUCTION

Corn stunt, a disease identified in maize in the Southern US, and Central and South America, is considered to be a syndrome induced by a complex constituted by a phytoplasma, a spiroplasma, and a virus (Maize rayado fino virus-MRFV), which can all be transmitted simultaneously by their leafhopper vector, mainly Dalbulus maidis De Long & Wolcott (Gamez, 1973; Nault & Bradfute, 1979; Nault et al., 1980). MRFV is the type member of the genus Marafivirus, family Tymoviridae, which includes Oat blue dwarf virus (Gamez & Leon, 1988). The sequence of MRFV has already been reported (Hammond & Ramirez, 2001).

The disease has been detected in all Central American countries, including Costa Rica, El Salvador, Guatemala, Honduras, Nicaragua, Panama, as well as Mexico, Colombia, Peru, Venezuela, Uruguay, the United States, and Brazil (Gamez, 1980; Gamez et al., 1979; Kogel et al., 1996). In Brazil, rayado fino is known as “corn streak” (Costa et al., 1971) and the virus is vectored by D. maidis (Kitajima et al., 1976). In Central
America, local maize lines were usually tolerant to MRFV, but the virus caused significant losses in newly introduced varieties. In Brazil, the virus is important in the “safrinha” system, and together with corn stunt spiroplasma and maize bushy stunt phytoplasma, MRFV formed a complex referred to as “corn stunt” causing significant losses (Massola et al., 1999).

The molecular epidemiology of MRFV was previously examined by performing a phylogenetic analysis of the coat protein gene of fourteen isolates collected from several countries in Latin America and the United States (Hammond et al., 1997). The nucleotide sequence was highly conserved among the isolates, yet three phylogenetic clusters were revealed. The current report describes the characterization of the coat protein gene of three virus isolates obtained of samples from the São Paulo and Minas Gerais regions of Brazil. The phylogenetic relationships of the Brazilian isolates to those previously characterized were examined. A brief report appeared earlier (Hammond & Bedendo, 2001).

MATERIAL AND METHODS

Sampling - A total of 37 maize leaf samples exhibiting symptoms of corn stunt were collected and preserved from four Brazilian States during 1995 and 1996 (Bedendo et al., 1997).

Nucleic acid extractions, hybridizations, and RT-PCR - Total nucleic acid extracts were prepared from dried leaf tissue using a previously described method (Hammond et al., 1997). MRFV was detected in samples spotted onto nylon membranes (Kogel et al., 1996). MRFV-specific cDNA was synthesized by reverse transcription in a reaction containing 2-3 µL total nucleic acid, primer MRFV-10 (5’ GCCCAACAGGCTTTAGGCCGGACCTGACTACC 3’, complementary to a region in the 3’ NTR of MRFV) in reaction conditions previously described (Hammond & Bedendo, 2001). PCR reactions contained an aliquot of first-strand cDNA and primers MRFV-09 (5’ CCTCTTCTGATCCCTCCTGTGTGCTGGAGACG 3’) and MRFV-10 (Hammond et al., 1997). PCR products were analyzed by gel electrophoresis to confirm their size and were cloned directly into the pCR2.1 plasmid vector (Invitrogen).

Nucleotide sequencing and sequence comparisons - The nucleotide sequences of the 633 bp MRFV fragments inserted in the pCR2.1 plasmids were obtained using an automated DNA sequencer (Applied Biosystems International Model 373A). Sequence data were analyzed using Lasergene software by DNASTAR for the Apple Macintosh. Multiple sequence alignments were performed using the Clustal Method Version 5 (Higgins et al., 1992). Cladistic analyses were performed with the computer program phylogenetic analysis using parsimony (PAUP) version 3.1 written by Swofford (Swofford, 1993) on a Power Macintosh Model 8100. Uninformative characters were excluded from the analysis and the phylogenetic trees were constructed by a branch and bound search, using random stepwise addition. Bootstrap analyses (100 replicates) were performed to estimate the support for the inferred phylogenies (Felsenstein, 1987). Gen bank accession numbers of previously sequenced isolates are provided in Hammond et al. (1997). Oat blue dwarf virus (OBDV- Gen bank accession no. U87832) was used as the outgroup (Edwards et al., 1997).

RESULTS AND DISCUSSION

Sixteen out of 37 samples tested for the presence of MRFV by nucleic acid hybridization using an MRFV-specific probe were positive for MRFV (data not shown). Reverse transcription-polymerase chain reaction (RT-PCR) products were obtained from three of the samples (Brazil 17, 26, and 48) using primers MRFV 9 and MRFV 10 and consisted of the expected 633 bp fragment (data not shown). No amplification occurred from healthy plants. The fragment was subsequently cloned into the plasmid vector pCR2.1 for further analysis. Single clones representing each isolate were selected for nucleotide sequencing. The 633 bp fragment encodes nt. 3-533 of the coat protein gene and 99 bp of the 3’NTR (Hammond et al., 1997).

The isolates from Brazil share 98% similarity among themselves and from 90 to 97% similarity to other MRFV isolates. Phylogenetic relationships between the isolates were inferred from the dendrogram (Figure 1). The Brazilian isolates cluster with the isolates from Peru and Bolivia, which are geographically adjoining Brazil.

The results obtained from sequence analysis of RT-PCR products of Brazilian corn streak virus confirm the results obtained previously with serology (Gamez et al., 1979; Kitajima et al., 1975), ie., isolates of the virus obtained from São Paulo and Brasilia were found to be related to, but distinguishable from, isolates from Costa Rica, Mexico, Guatemala, El Salvador, Honduras, Nicaragua, Panama, Peru, Uruguay, and Colombia. This research supports the proposal that Brazilian corn streak virus be regarded as an isolate of MRFV.

The samples analyzed in this study originate from the Ribeirão Preto region of São Paulo and the National Center for Corn Research, Sete Lagoas region of Minas Gerais. Samples 17 and 26 were collected from São Paulo in areas approximately 100 km apart; sample 48 was collected in Minas Gerais at approximately 400 km from samples 17 and 26.

The contribution of MRFV to the corn stunt symptoms exhibited by maize plants is unknown. The usual symptoms of MRFV infection on maize consist of numerous short and long chlorotic stripes, wilting of...
young plants, general chlorosis, and stunting. Symptoms are most noticeable at the base of leaves, with an increase in severity in the younger leaves (Gamez, 1980; Nault et al., 1980). Symptom severity is not always proportional to viral replication within the host and may be influenced by environmental conditions or germ plasm (Bustamente et al., 1998). In addition, the symptoms of MRFV infection could be masked by the symptoms produced by co-infection with corn stunt agents. Further examination of the occurrence of MRFV in Brazil and its association with corn stunt disease may answer these questions. The use of existing resistance or tolerance germ plasm (Bustamente et al., 1998) or the development of resistant germ plasm through biotechnology will benefit control measures and reduce losses to the diseases caused by MRFV.

Figure 1 - Phylogenetic tree constructed using the branch-and-bound method of parsimony analysis of the nucleotide sequences. The tree was constructed using OBDV as outgroup. Branch lengths are proportional to the number of inferred character state transformations. Vertical branches are arbitrary. Numbers above the line indicate nucleotide differences between branch nodes. Bootstrap (confidence) values are shown below the line on the branches (100 replications, 50% majority rule consensus tree). GenBank accession numbers are: Guatemala (U97724), Costa Rica (U97721), Venezuela (U97730), Mexico (U97725), United States (U97729), Bolivia (U97717), Peru (U97726), Colombia (U97719), OBDV (U87832), Brazil 17 (AF186177), Brazil 48 (AF186178), Brazil 26 (AF186179).

REFERENCES


