ENRICHING NUTRITIVE VALUE OF CASSAVA ROOT
BY YEAST FERMENTATION

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ABSTRACT: Cassava (Manihot esculenta) is extensively cultivated throughout the tropics and subtropics regions due to its ability to grow in diverse soil conditions and minimal management. Experiments were made to study the cassava root fermentation by yeasts in order to enhance the nutritive value of their products (fresh pulp and chips). Both cassava chip (CC) and fresh cassava root pulp (FCR) samples were fermented by Saccharomyces cerevisiae in solid-liquid media fermentation conditions during 132 hours and dried at 30°C. Products were analyzed for proximate composition, mineral composition, essential aminoacids and antinutrient content. There were increases (p < 0.01) in protein (30.4% in CC and 13.5% in fermented cassava root -FCR) and fat contents (5.8% in CC and 3.0% in FCR). S. cerevisiae fermented cassava products had very low hydrocyanic acid (HCN) contents (CC, 0.5 mg kg⁻¹ and FCR, 47.3 mg kg⁻¹). There was a remarkable increase in lysine content in the fermented cassava chip (FCC). The best acceptability and organoleptic attributes (color, texture and aroma) of enriched cassava chip were achieved after 132h of bioprocessing. The results of this study suggest that FCC can be nutritionally improved with S. cerevisiae for animal feeding.

Key words: Saccharomyces cerevisiae, fermentation, cassava chip, fresh cassava root pulp, protein, animal feed

INTRODUCTION

Cassava (Manihot esculenta, Crantz) is extensively cultivated throughout the tropics and subtropics regions due to its ability to grow in diverse soil conditions and minimal management (Wanapat, 2003; Wanapat et al. 2006; Wanapat & Khampa, 2007). The root is composed almost entirely of carbohydrate which can be used as important food source. However, it contains cyanogens (1- 3% CP) depending on cultivars (Stupak et al., 2006) and large amount of cyanogenic glucosides in the cassava flour (Cumbana et al., 2007) which could limit cassava root utilization for consumption and for livestock feeding. The laminarin and lotaustralin cassava’s cyanogenic compounds are changed to hydrocyanic acid (HCN) by the action of the laminarase enzyme when roots are crushed or sliced (Wanapat et al., 1999; Cardoso et al., 2005). The common process...
of cassava root sun drying does not sufficiently re-
move the cyanide content. Bradbury (2006) and
Cumbana et al. (2007) reported that the total cyanide
content in cassava flour was reduced to three or six
folds when soaking in water in an open vessel for 5
hours at (30°C). The process of protein enrichment
of animal feed using the microorganisms in a semi-
solid culture to improve the nutritional value of the for-
age palm for ruminants feeding has been evaluated
(Araujo et al, 2008; Vendruscolo et al, 2009). Ferment-
tation of cassava peels by pure culture of S. cerevisiae
could increase its protein content from (2.4%) in
nonfermented cassava to (14.1%) in fermented prod-
ucts (Antai & Mbongo, 1994). The fermented cassava
flour with S. cerevisiae enhanced the protein level
(from 4.4% to 10.9%) and decreased the amount of
cyanide content (Oboh & Kindahunsi, 2005). The ob-
jective of this study is to investigate enrichment of cas-
sava root pulp and chip by fermentation using S. cer-
erevisiae for animal feeding.

MATERIAL AND METHODS

Material
Cassava roots used in these experiments were
freshly harvested from the field. The commercial baker
yeast (Saccharomyces cerevisiae), manufactured by
Berly Speciality Industries, Company Limited, Bangkok,
Thailand was used in the fermentation processes.
Commercial grade urea and sugar cane molasses were
purchased from the local shop.

Yeast inoculants preparation
Baker yeast was cultured in the cylinder vessels
which contained solution of 20% molasses (w/v) and
4% urea (w/v). The products were incubated at room
temperature and oxygen was supplied by an air pump
for 60 h.

Sample preparation
A completely randomized design (CRD) with a 2
× 2 factorial arrangement of treatments was used.
The model included cassava source, method of fer-
mentation and their interactions as fixed effects, and
experimental run as random effects. Treatments
were: (i) unfermented cassava chip (UFCC), (ii) fer-
mented cassava chip (FCC), (iii) unfermented fresh
cassava root pulp (UFCR), and (iv) fermented fresh
cassava root pulp (FFCR). There were three repli-
cates for each treatment. In the second and fourth
treatments, CC and FCR were washed, grated, after
which 100 g of processed pulp was spread on a tray
(about 50 cm diameter) to an average layer thickness
of 2 cm. Yeast inoculum was mixed and inoculated
into 0.5 kg of the mash (CC and FCR) as the starter
and 250 mL nutrient solution [urea (48 g) and mo-
lasses (24 g)] were added. Fermentation was con-
ducted during 132 hours at 25°C under an air rela-
tive humidity between 40% and 50%. The first and
third treatments were unfermented cassava mash,
which served as the control. Both the fermented and
unfermented cassava mash were sun-dried for three
days at a 30°C average temperature and milled into
yeast fermented cassava products as presented in
Figures 1 and 2.

Chemical analysis
Dry matter (DM) of yeast fermented and non fer-
mented cassava products were analyzed for DM by
drying at 105°C for 12 h in a forced air oven, ash and
nitrogen contents were determined by micro–Kjeldahl
method (AOAC, 1997). Attempts were also made to
calculate the amount of solubilized protein-N in each
extraction by using the Lowry procedure (Lowry et
al., 1951). The samples were also analyzed for neu-
tral-detergent fiber (NDF) and acid-detergent fiber
(ADF) (Soest et al., 1994). One of the 5 mL samples
were used for cyanide concentration measurement by
spectrophotometrical (SpectroSC, LaboMed, inc.
USA) with the 2, 4-quinolinediol-pyridine reagent (Lam-
bert et al., 1975). The mineral (Ca and P) contents
were determined on aliquots of the solutions of the ash
by established flame atomic absorption spectrophotom-
etry procedures using a Perkin-Elmer atomic absorp-
tion spectrophotometer. Amino acids were determined
spectrophotometrically according to procedure of
Chinhard (1952).
Nutritive value of cassava

Analysis of data

All data were statistically analyzed using the analysis of variance of a completely randomized design with a 2 × 2 factorial arrangement of treatments (cassava source and method of fermentation) using the GLM procedure (SAS, 1988). Treatment means were statistically compared using Duncan’s New Multiple Range Test.

RESULTS AND DISCUSSION

The cassava fermentation products had brownish color and good aroma after sun-drying (Figures 1 and 2). DM of FCC and FCR were 85.3% and 65.7%, respectively and the mean protein value of *S. cerevisiae* fermented cassava products, FCR and CC were 18.9% and 30.4%, respectively (Table 1). They were remarkably higher when in comparison to unfermented cassava products. The increase in growth and proliferation of the fungi or bacterial complex in the form of single cell proteins may possibly account for the apparent increase in the protein content as also found by Antai & Mbongo (1994); Oboh (2002). Moreover, Correia et al. (2007) reported that *S. cerevisiae* was assessed to increase protein levels of pineapple waste (PW) by solid state bioprocessing (SSB) with and without nitrogen supplementation while PW (10 g) was inoculated with *S. cerevisiae*. Optimum protein content (22% dry basis), which is 3.5-fold of the original protein content, reached at 48 h of incubation when 0.25% (NH₄)₂SO₄ was added to the medium. This high protein content could be attributed to the ability of the *S. cerevisiae* to secrete some extracellular enzymes such as amylases, linamarase and cellulase into the cassava mash during their metabolic activities, which would lead to yeast growth (Oboh & Akindahunsi, 2003). The protein content of the product (Table 1) was similar to the products fermented with *S. cerevisiae* and *Lactobacillus* spp. in solid media using cassava peels (Oboh, 2006). This high protein cassava product could very well serve as a protein source in animal diets provided it is economically viable.

Fat content of FCR and CC increased (p < 0.01) after fermentation. The reason for the unusually high fat content increase could not be yet well explained. However, it could partly be attributed from cell increase of the fermentation. In addition, there could be possible transformation of carbohydrate to fat as occurred in *Leuconostoc mesenteriodes* (Padmaja et al., 1993) to be highly lipolytic. In fact, this highly lipolytic microorganism was able to hydrolyse 5% olive oil added to yeast agar medium. Similar result was found in cassava fermentation by Fagbemi & Ijah (2006). Akindumila & Glatz (1998) reported that certain fungi can produce microbial oil during fermentation. The decrease in carbohydrate could be attributed to the possible transformation of some of the carbohydrate, which could be used as carbon sources for synthesis of protein or fat (Lehninger, 1987). However, there was a substantial increase in lipid constituent of the fermenting cassava mash that had been previously inoculated. This increase was a result of the growth of yeast cells added as inocula to the cassava mash. However, there was no changes in NDF, ADF and OM contents of the fermented cassava products.

In increases in protein content have been obtained in the fermented products (Tables 1 and 2). In addition, there was a decrease in the HCN content when compared with the unfermented cassava products. Levels of the residual cyanide present in both FCR (47.3 mg kg⁻¹) and CC (0.5 mg kg⁻¹) were remarkably low when compared with the normal cyanide content of the unfermented cassava. These levels were considered safe for animal feeding. Oboh et al. (2002) and Oboh & Akindahunsi (2003) found that cyanide concentration in the cassava peels fermented with waste-water from fermented cassava pulp was low, when compared with the normal cyanide content of cassava products in Nigeria [19.0 mg kg⁻¹ (gari), 25 mg kg⁻¹ (fufu)], and with that of the cyanide content of some micro-fungi fermented cassava products (9.1–17.2 mg kg⁻¹). This suggest that baker’s yeast is capable of utilizing cyanogenic glycosides and the breakdown products, thus explaining why it is one of the natural flora involved.
in cassava fermentation during processing (Tweyongyere & Katongole, 2002).

Cassava fermentation with \textit{S. cerevisiae} resulted in a higher lysine content than those in unfermented cassava (Table 2). This indicate that the increase in lysine content of the cassava mash was a result of solid and liquid fermentation in which further increased the number of the yeast cells in cassava mash. Working with a mixed culture of the cellulolytic fungus \textit{Trichoderma viride} and the yeast \textit{Candida utilis}, Adebawo et al. (2000) and Toshihiro et al. (2007) used acid and alkali treatment, and gamma irradiation in which amino acids were found to be enhanced. In contrast, Ca and P contents were not changed, are in agreement with the ones reported by Akindahunsi et al. (1999).

**CONCLUSION**

The baker’s yeast, a cheap and non-pathogenic microorganism used in this study, could efficiently increase the protein and lysine contents of \textit{CC} and reduced the level of cyanide by fermentation. Therefore, yeast fermentation, especially in cassava root pulp and cassava chip, could potentially be used to enhance their nutritive value as animal diets, especially their protein and mineral contents of these products.

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**REFERENCES**


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Table 1 - Chemical composition and nutritive value of fermented cassava products (%DM).

<table>
<thead>
<tr>
<th>Item</th>
<th>CC\textsuperscript{1}</th>
<th>FCC\textsuperscript{4}</th>
<th>FCR\textsuperscript{2}</th>
<th>FFCR\textsuperscript{6}</th>
<th>SEM\textsuperscript{7}</th>
</tr>
</thead>
<tbody>
<tr>
<td>DM</td>
<td>89.4\textsuperscript{a} 85.3\textsuperscript{a}</td>
<td>68.9\textsuperscript{b} 65.7\textsuperscript{b}</td>
<td>1.95</td>
<td></td>
<td></td>
</tr>
<tr>
<td>OM</td>
<td>95.8</td>
<td>97.6</td>
<td>95.9</td>
<td>96.2</td>
<td>0.88</td>
</tr>
<tr>
<td>CP</td>
<td>3.4\textsuperscript{a} 32.5\textsuperscript{b}</td>
<td>3.2\textsuperscript{a} 21.1\textsuperscript{a}</td>
<td>1.2</td>
<td></td>
<td></td>
</tr>
<tr>
<td>NPN</td>
<td>0.2\textsuperscript{a} 2.1\textsuperscript{b}</td>
<td>0.3\textsuperscript{a} 2.2\textsuperscript{b}</td>
<td>0.15</td>
<td></td>
<td></td>
</tr>
<tr>
<td>TP\textsuperscript{8}</td>
<td>3.2\textsuperscript{a} 30.4\textsuperscript{b}</td>
<td>2.8\textsuperscript{a} 18.9\textsuperscript{b}</td>
<td>0.11</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fat</td>
<td>2.7\textsuperscript{a} 5.8\textsuperscript{b}</td>
<td>2.3</td>
<td>3.0\textsuperscript{a}</td>
<td>0.02</td>
<td></td>
</tr>
<tr>
<td>NDF</td>
<td>7.7</td>
<td>7.3</td>
<td>7.8</td>
<td>7.5</td>
<td>0.04</td>
</tr>
<tr>
<td>ADF</td>
<td>6.1</td>
<td>5.8</td>
<td>6.2</td>
<td>6.0</td>
<td>0.05</td>
</tr>
</tbody>
</table>

\textsuperscript{a,b}Means with different superscripts differ \((p < 0.01)\), \textsuperscript{1}CC = Cassava chip, \textsuperscript{2}FCR = Fresh cassava root, \textsuperscript{3}UCC = Unfermented cassava chip, \textsuperscript{4}FCC = Fermented cassava chip, \textsuperscript{5}UFCR = Unfermented fresh cassava root, \textsuperscript{6}FFCR = Fermented fresh cassava root pulp, \textsuperscript{7}Standard error of the mean, \textsuperscript{8}True protein = Crude protein (CP) - NPN

Table 2 - Cyanide, lysine, Ca and P contents of fermented cassava products.

<table>
<thead>
<tr>
<th>Item</th>
<th>CC\textsuperscript{1}</th>
<th>FCC\textsuperscript{4}</th>
<th>FCR\textsuperscript{2}</th>
<th>FFCR\textsuperscript{6}</th>
<th>SEM\textsuperscript{7}</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cyanide (mg kg\textsuperscript{-1})</td>
<td>3.4\textsuperscript{a} 0.5\textsuperscript{a}</td>
<td>68.6\textsuperscript{b} 47.3\textsuperscript{b}</td>
<td>0.88</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lysine g 100 g\textsuperscript{-1} protein</td>
<td>3.8\textsuperscript{a} 8.5\textsuperscript{b}</td>
<td>3.9\textsuperscript{a} 5.5\textsuperscript{a}</td>
<td>0.53</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ca (g kg\textsuperscript{-1})</td>
<td>1.35 0.9</td>
<td>1.31 1.26</td>
<td>1.5</td>
<td>1.5</td>
<td></td>
</tr>
<tr>
<td>P (g kg\textsuperscript{-1})</td>
<td>0.7 0.64</td>
<td>0.73 0.69</td>
<td>0.6</td>
<td>0.6</td>
<td></td>
</tr>
</tbody>
</table>

Means with different superscripts differ \((p < 0.01)\), \textsuperscript{1}CC = Cassava chip, \textsuperscript{2}FCR = Fresh cassava root pulp, \textsuperscript{3}UCC = Unfermented cassava chip, \textsuperscript{4}FCC = Fermented cassava chip, \textsuperscript{5}UFCR = Unfermented fresh cassava root, \textsuperscript{6}FFCR = Fermented fresh cassava root pulp, \textsuperscript{7}Standard error of the mean.


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