NECROTIZING ENTEROCOLITIS, PATHOGENESIS AND THE PROTECTOR EFFECT OF PRENATAL CORTICOSTEROIDS

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Necrotizing enterocolitis is the most frequently occurring gastrointestinal disorder in premature neonates.

Animal models of necrotizing enterocolitis and prenatal administration of cortisone have demonstrated that cortisone may accelerate maturation of the mucosal barrier, therefore reducing the incidence of this gastrointestinal disorder.

The authors present a review of the literature of the most important risk factors associated with necrotizing enterocolitis, such as inflammatory gastrointestinal mediators, enteral feeding and bacterial colonization, and immaturity of the gastrointestinal barrier, and we emphasize the necessity for additional studies to explore the prenatal administration of cortisone as a preventive strategy for necrotizing enterocolitis.

DESCRIPTORS: Necrotizing enterocolitis. Neonate. Prematurity. Cortisone therapy. Physiopathology.

INTRODUCTION

Necrotizing enterocolitis (NEC) is a common neonatal gastrointestinal disease that affects approximately 11% of premature neonates weighing less than 1500 g¹. The average mortality is 20% to 40%, and survivors after either medical or surgical therapy can present with failure to thrive, feeding abnormalities, diarrhea, or bowel obstruction².

The etiology of NEC is multifactorial, and the most important risk factors are prematurity, hypoxia and/or intestinal ischemia, and enteral feeding and gastrointestinal bacteria colonization³.

The association of such risk factors might trigger a local inflammatory cascade with release of inflammatory mediators, resulting in NEC⁴. Alterna-

tively, an imbalance between local inflammatory mediators and an immature local defense could result in NEC defense.

Early signs of NEC are indistinguishable from sepsis neonatorum. The signs and symptoms are quite variable, ranging from feeding intolerance to evidence of sepsis, shock, peritonitis, and death. The usual presenta-

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tion includes abdominal distension, gastric residuals, bilious vomiting, and bloody stools. Lethargy, apnea, and hypoperfusion also may be a prominent feature. Physical findings found on serial examination comprise progressive abdominal tenderness, muscular guarding, and abdominal wall erythema. The presence of an abdominal mass may indicate localized perforation or progressive peritoneal irritation. However, these physical findings may be minimal and misleading, even in infants with progressive disease leading to perforation².

Since prematurity is the most important risk factor associated with NEC, possible therapeutic approaches that promote maturation of the gastrointestinal mucosal barrier, such as the prenatal administration of corticosteroids, have been explored.

1. INTESTINAL INFLAMMATORY MEDIATORS

Animal models of bowel necrosis have been established by injection of platelet-activating factor (PAF)^{5,6}, endotoxin (bacterial lipopolysaccharide or LPS)^{5,7}, and tumor necrosis factor (TNF)⁸. PAF has been implicated as the most important mediator in the pathophysiology of NEC⁵.

Platelet-activating factor is a endogenous phospholipid mediator produced by inflammatory cells, platelets, endothelial cells, and some bacteria, such as *Escherichia coli*¹⁰⁻¹⁴. PAF has a short half-life in the circulation, due to the high plasma and tissue PAF-degrading enzyme acetylhydrolase (PAF-AH) activity^{15,16}, which rapidly degrades PAF into the biologically inert lyso-PAF¹⁷.

Mesenteric or systemic administration of PAF in rats induces intestinal injury similar to NEC⁵. On the other hand, the administration of PAF receptor antagonists in animal models of NEC induced by hypoxia, endotoxin, and PAF prevented intestinal mucosal injury^{7,8,18}.

Considerable evidence indicates that the altered regulation of PAF-AH may play a role in the occurrence of NEC because:

- 1) PAF-AH activity is decreased in neonates and approaches adult enzyme activity only after 6 weeks of life¹⁹,
- 2) PAF-AH activity is deficient in sick neonates with NEC²⁰,
- PAF-AH activity can be demonstrated in breast milk (formula has none), and use of breast milk reduces the incidence of NEC^{21,20},
- The administration of PAF-AH in animal models of NEC induced by hypoxia reduced the incidence of NEC²³.

Cundell et al.²⁴ have demonstrated that the PAF receptor is an important determinant of bacterial (Streptococcus) adhesion and invasion into endothelium and epithelium.

Mackendrick et al.²⁵ have found that neonates fed enterally had higher levels of PAF and endotoxin after feeding than before feeding.

The cytotoxic effect of PAF is most likely due to reactive oxygen radical formation^{9,26-29}, and its prolonged effect seems to be due to its ability to induce TNF formation by intestine and liver³⁰ as well as to stimulate its own production³¹.

Xanthine oxidase (XO)^{29,32}, an enzyme in intestinal tissue, appears to be the major source of free radicals in reoxygenated tissue. Reperfusion of the tissue supplies molecular oxygen, which results in a burst of superoxide radical production that damages intestinal tissue by peroxidation of unsaturated lipids within the cellular and mitochondrial membranes.

PAF-induced bowel injury is not only associated with production of oxygen-derived free radicals, but also with neutrophil margination and activation and capillary leakage³³. The importance of neutrophil cells in initiating the intestinal injury has been demonstrated in leukopenic mice and rats, which were relatively protected from PAF-induced bowel injury^{34,35}.

Nitric oxide (NO) has been demonstrated to be a protective modulator for the intestinal mucosa³⁶⁻³⁹. Nitric oxide has been described as an endothelium-derived relaxing factor that promotes vasodilation and microvascular integrity, inhibits leukocyte adhesion and activation, and scavenges oxygen radicals.

Nitric oxide synthase is the enzyme responsible for NO production⁴⁰. Animal models of bowel injury induced by ischemia-reperfusion^{41,42}, endotoxin⁴³, and PAF⁴⁴ have shown that intestinal injury is markedly exacerbated by the concomitant inhibition of the NO synthase. The results of these studies suggest that an imbalance between endogenous NO and PAF production may be the factor responsible for intestinal mucosal injury.

2. ENTERAL FEEDING AND BACTERIA COLONIZATION

Enteral feeding and the pattern of intestinal colonization and bacteria adherence are risk factors for developing NEC and therefore have been studied by several researchers⁴⁵⁻⁵².

The pattern of intestinal colonization varies according to the type of enteral feeding that the neonate is receiving 48. Breast feeding causes gastrointestinal colonization predominantly by bifidobacteria (Gram-positive bacteria), which control the growth of Gram-negative bacteria 48-52. In contrast, formula-fed neonates are colonized predominantly by coliforms, enterococci, and Bacteroides spp. 50.

There are important differences between Gram-positive and Gram-negative bacteria regarding intestinal carbohydrate metabolism⁵³. The fermentation of lactose by Gram-positive bacteria yields lactic acid, which can be readily absorbed from the intestinal tract. Conversely, Gram-negative bacteria ferment lactose into hydrogen, carbon dioxide, and organic acids, which may not be cleared as readily from the intestinal lumen.

The acidification of intraluminal contents for a prolonged period of time causes a reduction of local pH, which may result in injury to the intestinal mucosal, dissociation of divalent cations resulting in increased ionized fractions, and change in the spatial configuration of intraluminal proteins. These changes in protein spatial configuration may be able to trigger a release of vasoactive substances that then alter intestinal microcirculation^{54,55}.

Bifidobacteria release less endotoxin than Gram-negative bacteria; they therefore induce the release of reduced amounts of inflammatory mediators such as interleukin-1, interleukin-6, and TNF⁵⁶.

3. IMMATURITY OF THE GASTROINTESTINAL MUCOSAL BARRIER

The immaturity of the gastrointestinal mucosal barrier can be demonstrated by:

- 1) Increased permeability of intestinal mucosal to intact proteins⁵⁷⁻⁶¹;
- 2) Reduced gastric acid secretion during the first week of life⁶²;
- Reduced concentration of proteolytic pancreatic enzymes^{63,64}, enterokinase⁶, and disaccharidase and lactase⁶⁶:
- 4) Immaturity of gastrointestinal motor activity^{66,67};
- 5) A molar ratio of carbohydrate-toprotein that is less in the newborn mucus (The total protein content in intestinal mucus from newborn rat is greater than in adult mucus.⁶⁸);
- 6) Structural modifications of intestinal microvilli; and ⁶⁹
- 7) Cellular and humoral gastro-intestinal immaturity 70,71.

4. PRENATAL CORTICOSTEROIDS AND NECROTIZING ENTEROCOLITIS

Several studies⁷²⁻⁸¹ have analyzed the maturation of intestinal mucosa following the administration of thyroid hormones⁷²⁻⁷⁴ and steroids⁷⁵⁻⁸¹.

Israel et al.⁸¹ have demonstrated in an animal model that prenatal administration of corticosteroids reduces the uptake of macromolecules from the intestinal mucosal⁸², decreases small intestinal bacterial colonization with aerobic bacteria, and reduces the incidence of bacterial translocation to the liver, resulting in a lower incidence of NEC. The association between prenatal administration of corticosteroids and decreased incidence of NEC has also been demonstrated in neonates^{79,83}.

The effect of administration of corticosteroids on different enzymes has also been demonstrated 77,78,80. Buchmiller et al. 80 have shown an en-

largement of the small intestine and a trend of increasing lactase and maltase activity after administration of corticosteroids. Engelhardt et al.⁷⁷ have demonstrated that the prenatal administration of dexamethasone stimulates maltase and sucrase activities; however, no effect was observed on catalase, superoxide dismutase, and xanthine oxidase activities. Horváth et al.⁷⁸ have shown that the prenatal administration of betamethasone stimulates Na/K – ATPase enzyme activity.

FINAL CONSIDERATIONS

The findings described above suggest that the administration of prenatal corticosteroids is associated with maturation of intestinal mucosal; however, further studies are necessary to better understand the mechanisms by which corticosteroids exert their effects.

RESUMO RHCFAP/3100

PRECIOSO AR e col. – Enterocolite necrosante: resposta imflamatória x corticoterapia pré-natal. **Rev. Hosp. Clín. Fac. Med. S. Paulo 57**(5): 243-248, 2002.

A enterocolite necrosante é a mais frequente patologia gastrointestinal adquirida no período neonatal, acometendo preferencialmente o recém-nascido prematuro. Estudos experimentais sugerem que a corticoterapia pré-natal acelera a maturação da mucosa gastrintestinal, levando a diminuição da incidência desta doença.

Os autores apresentam uma revisão da literatura em relação aos principais fatores fisiopatológicos associados a enterocolite necrosante, tais como mediadores inflamatórios gastrintestinais, nutrição enteral e colonização bacteriana e imaturidade gastrintestinal e enfatizam a necessidade de mais estudos que avaliem a influencia da corticoterapia pré-natal com fator de prevenção da enterocolite necrosante.

DESCRITORES: Enterocolite necrosante. Recém-nascido. Prematuridade. Corticoterapia. Fisiopatologia.

REFERENCES

- UAUY RD, FANAROFF AA, KORONES SB, PHILIPS EA et al. -Necrotizing enterocolitis in very low birth weight infants: biodemographic and clinical correlates. J Pediatr 1991;119:630-638.
- SCHETTINI ST, MIYOSHI MH Enterolote necrosante neonatal. Pediatria Moderna 1999;35:145-88.
- CRISSINGER KD Animal models of necrotizing enterocolitis. J Pediatr Gastroenterol Nutr 1995;20:17-22.
- CAPLAN MS, MACKENDRICK W Necrotizing enterocolitis: a review of pathogenetic mechanisms and implications for prevention. Pediatr Pathol 1993;13:357-369.
- GONZALEZ-CRUSSI F, HSUEH W Experimental model of ischemic bowel necrosis: the role of platelet-activating factor and endotoxin. Am J Pathol 1983;112:127-135.
- HSUEH W, GONZALEZ-CRUSSI F, ARROYAVE JL Plateletactivating factor-induced ischemic bowel necrosis. An investigation of secondary mediators in its pathogenesis. Am J Pathol 1986;122:231-239.
- HSUEH W, GONZALEZ-CRUSSI F, ARROYAVE JL Plateletactivating factor is an endogenous mediator for bowel necrosis in endotoxemia. FASEB J 1987;1:403-405.
- SUN XM, HSUEH W Bowel necrosis induced by tumor necrosis factor in rats is mediated by platelet-activating factor. J Clin Invest 1988;81:1328-1331.
- HSUEH W, CAPLAN MS, SUN X, TAN X et al. Platelet-activating factor, tumor necrosis factor, hypoxia and necrotizing enterocolitis. Acta Paediatr 1994;396:11-17.
- 10.BENVENISTE J Paf-aether, an ether phospho-lipid with biological activity. Prog. Clin Biol Res 1988;282:73-85.
- HANAHAN DJ Platelet-activating factor: a biologically active phosphoglyceride. Annu Ver Biochem 1986;55:483-509.
- SNYDER F Platelet-activating factor and related acetylated lipids as potent biologically active cellular mediators. Am J Physiol 1990;259:C697-708.
- DENIZOT Y, DASSA E, KIM HY, BOSSANT MJ et al Synthesis of paf-acether from exogenous precursors by the prokaryote Escherichia coli. FEBS Lett 1989;243:13-16.
- DENIZOT Y, DASSA E, BENVENISTE J, THOMAS Y Pafacether production by Escherichia coli. Biochem. Biophys Res Commun 1989;161:939-943.
- STAFFORINI DM, ELSTAD MR, MCINTYRE TM, ZIMMERMAN GA et al. - Human macrophages secrete plateletactivating factor acetylhydrolase. J Biol. Chem 1990;265:9682-9687.
- TARBET EB, STAFFORINI DM, ELSTAD MR, ZIMMERMAN GA et al. - Liver cells secrete the plasma form of plateletactivating factor acetylhydrolase. J Biol Chem 1991;266:1667-1673.

- 17. FARR RS, WARDLOW ML, COX CP, MENG KE et al. Human serum acid-labile factor is an acetylhydrolase that inactivates platelet-activating factor. Fed. **Proc** 1983;**42**:3120-3122.
- CAPLAN MS, SUN XM, HSUEH W Hypoxia causes ischemic bowel necrosis in rats: the role of platelet activating factor (PAF-acether). Gastroenterol 1990;99:979-986.
- CAPLAN M, HSUEH W, KELLY A, DONOVAN M Serum PAF acetylhydrolase increases during neonatal maturation. Prostaglandins 1990;39:705-714.
- CAPLAN MS, SUN XM, HSUEH W, HAGEMAN JR Role of platelet-activating factor and tumor necrosis factor-alpha in neonatal necrotizing enterocolitis. J Pediatr 1990;116:960-064
- MOYA FR, EGUCHI H, ZHAO B, FURUKAWA M et al. Plateletactivating factor acetylhydrolase in term and preterm human milk: a preliminary report. J Pediatr Gastroenterol Nutr 1994;19:236-239.
- LUCAS A, COLE TJ Breast milk and neonatal necrotizing enterocolitis. Lancet 1990;336:1519-1523.
- 23. CAPLAN MS, LICKERMAN M, ADLER L, DIETSCH GN et al.
 The role of recombinant platelet-activating factor acetylhydrolase in a neonatal rat model of necrotizing enterocolitis. Pediatr Res 1997;42:779-783.
- CUNDELL DR, GERARD NP, GERARD C, IDANPAAN-HEIKKILA I - Streptococcus pneumoniae anchor to activated human cells by the receptor for platelet-activating factor. Nature 1995;377:435-438.
- MACKENDRICK W, HILL N, CAPLAN M Increase in plasma platelet-activating factor levels in enterally fed preterm infants. Biol Neonate 1993;64:89-95.
- PARKS DA, BULKLEY GB, GRANGER DN et al. Ischemic injury in the cat small intestine: role of superoxide radicals. Gastroenterol 1982;82:9-15.
- VAUGHAN WG, HORTON JW, WALKER PB Allopurinol prevents intestinal permeability changes after ischemiareperfusion injury. J Pediatr Surg 1992;27:968-973.
- 28. CZYRKO C, STEIGMAN C, TURLEY DL et al. The role of reperfusion injury in occlusive intestinal ischemia of the neonate: malonaldehyde-derived fluorescent products and correlation of histology. J Surg Res 1991;51:1-4.
- PARKS DA, BULKLEY GB, GRANGER DN Role of oxygenderived free radicals in digestive tract diseases. Surgery 1983;94:415-422.
- HUANG L, TAN X, JIANG Y, REDDY J et al. PAF and endotoxin induce TNF gene expression in rat intestine and liver. FASEB J 1992;6:A1316 (abstr.).
- 31. ZHANG C, HSUEH W, CAPLAN MS, KELLY A Plateletactivating factor-induced shock and intestinal necrosis in the rat: role of endogenous platelet-activating factor and effect of saline infusion. Crit Care Med 1991;19:1067-1073.

- 32. GRANGER DN, MCCORD JM, PARKS DA et al Xanthine oxidase inhibitors attenuat ischemia-induced vascular permeability changes in the cat intestine. Gastroentrol 1986:90:80-84.
- HSUEH W, GONZALEZ-CRUSSI F Ischemic necrosis induced by platelet-activating factor: an experimental model. Meth Archiev Exp Pathol 1988;13:208-239.
- MUSEMECHE C, CAPLAN M, HSUEH W, SUN XM et al. -Experimental necrotizing enterocolitis: the role of polymorphonuclear neutrophils. J Pediatr Surg 1991;26:1047-1050
- SUN XM, HSUEH W Platelet-activating factor produces shock, in vivo complement activation, and tissue injury in mice. J Immunol 1999:147:509-514.
- MONCADA S, PALMER RMJ, HIGGS EA Nitric oxide: physiology, pathophysiology, and pharmacology. Pharmacol Ver 1991;43:109-142.
- IGNARRO LJ Biological actions and properties of endotheliumderived nitric oxide formed and released from artery and vein. Circ Res 1989;65:1-21.
- IALENTI A, IANARO A, MONCADA S, DI ROSA M Modulation of acute inflammation by endogenous nitric oxide. Eur J Pharmacol 1992;211:177-182.
- GRAF JL, VANDERWALL KJ, ADZICK NS, HARRISON MR -Nitroglycerin attenuates the bowel damage of necrotizing enterocolitis in a rabbit model. J Pediatr Surg 1997;32:283-286.
- 40. CAPLAN MS, HEDLUND E, HILL NICOLE, MACKENDRICK W - The role of endogenous nitric oxide and platelet-activating factor in hypoxia-induced intestinal injury in rats. Gastroenterol 1994;106:346-352.
- 41. KUBES P. Ischemia-reperfusion in feline small intestine: a role for nitric oxide. **Am J Physiol** 1993;**264**:G143-G149.
- AOKI N, JOHNSON G III, LEFER AM Beneficial effects of two forms of NO administration in feline splanchnic artery occlusion shock. Am J Physiol 1990;258:G275-G281.
- 43. HUTCHESON IR, WHITTLE BJR, BOUGHTON-SMITH NK -Role of nitric oxide in maintaining vascular integrity in endotoxin-induced acute intestinal damage in the rat. Br J Pharmacol 1990;101:815-820.
- 44. MACKENDRICK W, CAPLAN M, HSUEH W Endogenous nitric oxide protects aginst platelet-activating factor induced bowel injury in the rat. **Pediatr Res** 1993;**34**:222-228.
- 45. PANIGRAHI P, GUPTA S, GEWOLB IH, MORRIS JG JR Occurrence of necrotizing enterocolitis may be dependent on patterns of bacterial adherence and intestinal colonization: studies in caco-2 tissue culture and weanling rabbit models. **Pediatr Res** 1994;**36**:115-121.
- MUSEMECHE CA, KOSLOSKE AM, BARTOW SA, ALBUQUERQUE TU - Comparative effects of ischemia, bacteria, and substrate on the pathogenesis of intestinal necrosis. J Pediatr Surg 1986;21:536-538.

- 47. CAPLAN MS, MILLER-CATCHPOLE R, KAUP S, RUSSEL T et al. - Bifidobacterial supplementation reduces the incidence of necrotizing enterocolitis in a neonatal rat model.1999;117:577-583
- 48. YOSHIOKA H, ISEKI K, FUJITA K Development and differences of intestinal flora in the neonatal period in breast-fed and bottle-fed infants. **Pediatr** 1983;**72**:317-321.
- GODMAN AJ Host resistance factors in human milk. J Pediatr 1973:82:1082-1090.
- KLEESSEN B, BUNKE H, TOVAR K, NOACK J et al. Influence of two infant formulas and human milk on the development of faecal flora in newborn infants. Acta Pediatr 1995;84:1347-1356.
- OGAWA K, BEN RA, PONS S, DE PAOLO MI et al. Volatile fatty-acids, lactic acid, and pH in the stools of breast-fed and bottle-fed infants. J Pediatr Gastroenterol Nutr 1992;15:248-252
- GIBSON GR, WANG X Regulatory effects of bifidobacteria on the growth of other colonic bacteria. J Appl Bacteriol 1994;77:412-420.
- PERLMUTTER D, BOYLE JT, CAMPOS JM, WATKIN JB D-Lactic acidosis, a new metabolic complication of small bowel resection. Pediatr Res 1982;16:173 A.
- LEMANSKE RF, ATKINS FM, METCALFE DD Gastrointestinal mast cells in health and disease Part I. J Pediatr 1983;103:177-184.
- LEMANSKE RF, ATKINS FM, METCALFE DD Gastrointestinal mast cells in health and disease Part II. J Pediatr 1983;103:343-351
- 56.NICAISE P, GLEIZES A, FORESTIER F, QUERO AM et al. -Influence of intestinal bacterial flora on cytokine (IL-1, IL-6, TNF-a) production by mouse peritoneal macrophages. Eur Cytokine Netw 1993;4:133-138.
- 57. UDALL JN, PANG K, FRITZE L, KLEINMAN R et al. -Development of gastrointestinal mucosal barrier. I. The effect of age of intestinal permeability to macromolecules. Pediatr Res 1981;15:241-244.
- 58. ROBERTON DM, PAGANELIU R, DINWIDDIE R, LEVINSKY RJ Milk antigen absorption in the preterm and term neonate. **Arch Dis Child** 1982;**57**:369-372.
- BEACH RC, MENZIES IS, CLAYDEN GS, SCOPES JW -Gastrointestinal permeability changes in the preterm neonate.
 Arch Dis Child 1982;57:141-145.
- WALKER WA, ISSELBACHER KJ Uptake and transport of macromolecules but he intestine: possible role in clinical disorders. Gastroenterol 1974;67:531-550.
- WEAVER LT, LAKER MF, NELSON R Intestinal permeability in the newborn. Arch Dis Child 1984;59:236-241.
- 62.AURICCHIO S, RABINO A, MURSET G Intestinal glycosidase activities in the human embryo, fetus, and newborn. **Pediatr** 1965;**35**:944-954.

- 63. UDALL JN, BLOCH KJ, VACHINO G, WALKER WA -Development of the gastrointestinal mucosal barrier. IV. The effect of inhibition of proteolysis on the uptake of macromolecules by the intestine of the newborn rabbit. Biol Neonate 1984;45:289-295.
- 64. LEBENTHAL E, LEE PC Development of functional response in human exocrine pancreas. **Pediatrics** 1980;**66**:566.
- ANTONOWICZ I, LEBENTHAL E Developmental pattern of small intestine enterokinase and disaccharidase activities in the human fetus. Gastroenterol 1977;72:1299-1303.
- 66.PRECIOSO AR Study of gastric myoelectrical activity in neonates. Sao Paulo. 1999. – School of Medicine of University of Sao Paulo.
- BERSETH CL Gestational evolution of small intestine motility in preterm and term infants. J Pediatr 1989;115:646-651.
- 68. SHUB MT, PANG KY, SWANN DA, WALKER WA Age-related changes in chemical composition and physical properties of mucus glycoproteins from rat small intestine. Biochem J 1983;215:405-411.
- PANG KY, BRESSON JL, WALKER WA Development of the gastrointestinal mucosal barrier. III. Evidence for structural differences in microvillus membranes from newborn and adult rabbits. Biochem Biophys Acta 1983;727:201-210.
- BOUSVAROS A, WALKER WA Development and function of the intestinal mucosal barrier. In: McDonald TT, ed. Ontogeny of the immune system of the gut. CRC Press 1990.p 2-22.
- SPENCER T. MCDONALD TT The ontogeny oh human mucosal barrier immunity. In: McDonald TT, ed. Ontogeny of the immune system of the gut. CRC Press 1990.p 23-50.
- 72. JUMAWAN J, CELANO P, HOROWITZ C, LAU H et al. Effect of cortisone of L-triiodothyronine administration to pregnant rats on the activity of fetal intestinal disaccharidase and lysosomal acid b-galactosidase. Biol Neonate 1977;32:211-217.
- ISRAEL EJ, PANG KY, HARMATZ PR, WALKER WA Structural and functional maturation of rat gastrointestinal barrier with thyroxine. Am J Physiol 1987;252:762-767.

- ISRAEL EJ, PANG KY, HARMATZ PR, WALKER WA-Development of mucosal barrier function: thyroxine regulation of macromolecule uptake by the small intestine of the newborn rat. J Physiol 1973;229:681-695.
- SCHIFFRIN EJ, WALKER WA, CARTER EA, BENJAMIN J et al.
 Influence of prenatal mucosal barrier maturation on bacterial colonization in the newborn. JPGN 1993;117:271-275.
- DANIELS VG, HARDY RN, MALINOWSKA KW, NATHANIELSZ PW - The influence of exogenous steroids on macromolecule uptake by the small intestine of the newborn rat. J Physiol 1973;229:681-695.
- ENGELHARDT EL, BEGGS JC, NEU J Maturation of antioxidant enzymes in rat small intestine: lack of glucocorticoid stimulation. J Pediatr 1987;111:459-463.
- HORVÁTH K, BLOCHIN B, HILL I, VERMA R et al. The preand postnatal development of Na / K – ATPase in gastrointestinal organs of the rat: effect of betamethasone treatment. J Pediatr Gastroenterol Nutr 1993;16:412-418.
- 79. BAUER CR, MORRISON JC, POOLE W K KORONES SB et al. -A decreased incidence of necrotizing enterocolitis after prenatal glucocorticoid therapy. Pediatrics 1984;73:682-688.
- 80. BUCHMILLER TL, SHAW KS, LAM ML, STOKES R et al. -Effect of prenatal dexamethasone administration: fetal rabbit intestinal nutrient uptake and disaccharidase development. J Surg Res 1994;57:274-279.
- 81.ISRAEL EJ, SCHIFFRIN EJ, CARTER EA, FREIBERG E et al. Prevention of necrotizing enterocolitis in the rat with prenatal cortisone. **Gastroenterol** 1990;**99**:1333-1338.
- 82.SHULMAN RJ, SCHANLER RJ, LAU C, HEITKEMPER M et al. - Early feeding, antenatal glucocorticoids, and human milk decrease intestinal permeability in preterm infants. Pediatr Res 1998;44:519-523.
- 83.HALAC E, HALAC J, BEGUE EF, CASANAS JM et al. Prenatal and postnatal corticosteroid therapy to prevent neonatal necrotizing enterocolitis. A controlled trial. J Pediatr 1990;117:132-138.

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