



## Evaluating the hydrophilic antioxidant capacity in different citrus genotypes

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### Abstract

Antioxidants are the body's defense system against the damage caused by reactive oxygen species, formed naturally during many physiological activities. In vegetables and fruits, various antioxidant compounds such as vitamin C, polyphenols, flavonoids, and carotenoids have been identified. Because fruits and vegetables are the primary antioxidant sources in our daily diet, it is necessary to determine their antioxidant capacity. Citrus fruit consumption per capita has steadily increased over the world over the last 30 years. Citrus fruits are high in vitamin C as well as other active ingredients like phenols and flavonoids that are beneficial to human health. Using carotenoid complement and pigmentation genetic diversity, the objective of this research was to see how vitamin C and carotenoids contributed to the capacity of hydrophilic antioxidants of the citrus fruits' pulp. Six citrus cultivars were chosen for this purpose: two sweet orange genotypes, Valencia Ruby and Valencia Late; two grapefruit genotypes, Star Ruby and Marsh; and two mandarin genotypes, Nadorcott and Clementules. In proportion to their color singularity, total carotenoid composition and content in fruit pulp differed dramatically. A good and clear connection was found between hydrophilic antioxidant capacity and vitamin C concentration in the pulp of various fruit species, as measured by DPPH and ABTS tests. The proportion of vitamin C to the total HAC was calculated to be between 15% and 30%.

**Keywords:** vitamin C; citrus fruit; carotenoids; antioxidant capacity.

**Practical Application:** In the current research it was aimed to see how vitamin C and carotenoids contributed to the capacity of hydrophilic antioxidants of the citrus fruits' pulp.

### 1 Introduction

Citrus fruit is the world's most traded horticulture product and one of the most widely cultivated horticultural commodities (Herrera-Barros et al., 2021; Rangel et al., 2011). Around two-thirds of the overall citrus, output comes from the United States, China, the Mediterranean nations, and Brazil (Lacirignola & D'Onghia, 2009). Citrus fruits are among the most valuable cultivars in world commerce, and they're in great demand for food additives in meals and drinks, freeze concentrates, juice processing, and fresh consumption all over the world (Czech et al., 2020; Xu et al., 2008). Colour is a crucial determinant in consumer acceptance and internal and external quality across the many citrus cultivars and species (Hoppu et al., 2021). The genus citrus comes in a wide range of colors both on the outside and inside (Ballistreri et al., 2019; Monselise, 2019): from the new sweet orange cultivars or pink or red lycopene-accumulating grapefruits, and

orange of mandarins to the citrons, many grapefruits' cultivars, pummelos, and yellow color of lemons (Alquezar et al., 2013; Rodrigo et al., 2013). Pectin, carotenoids, anthocyanins (blood oranges), limonoids, coumarins, phenolic acids, flavonoids, mineral elements, Vitamins E, C, and A are just a few of the phytochemicals found in citrus fruits (Zou et al., 2016). Citrus fruit intake has been linked to significant health advantages as well as a lower risk of chronic illnesses, according to findings from multiple in vivo and in vitro investigations (Ahmed & Azmat, 2019; Aune, 2019; Durazzo et al., 2019; Ma et al., 2020; Wallace et al., 2020).

The antioxidant activity of such substances has been linked to their positive impacts on health (Rodrigues et al., 2011; Shahidi & Ambigaipalan, 2015; Yashin et al., 2017). Antioxidants

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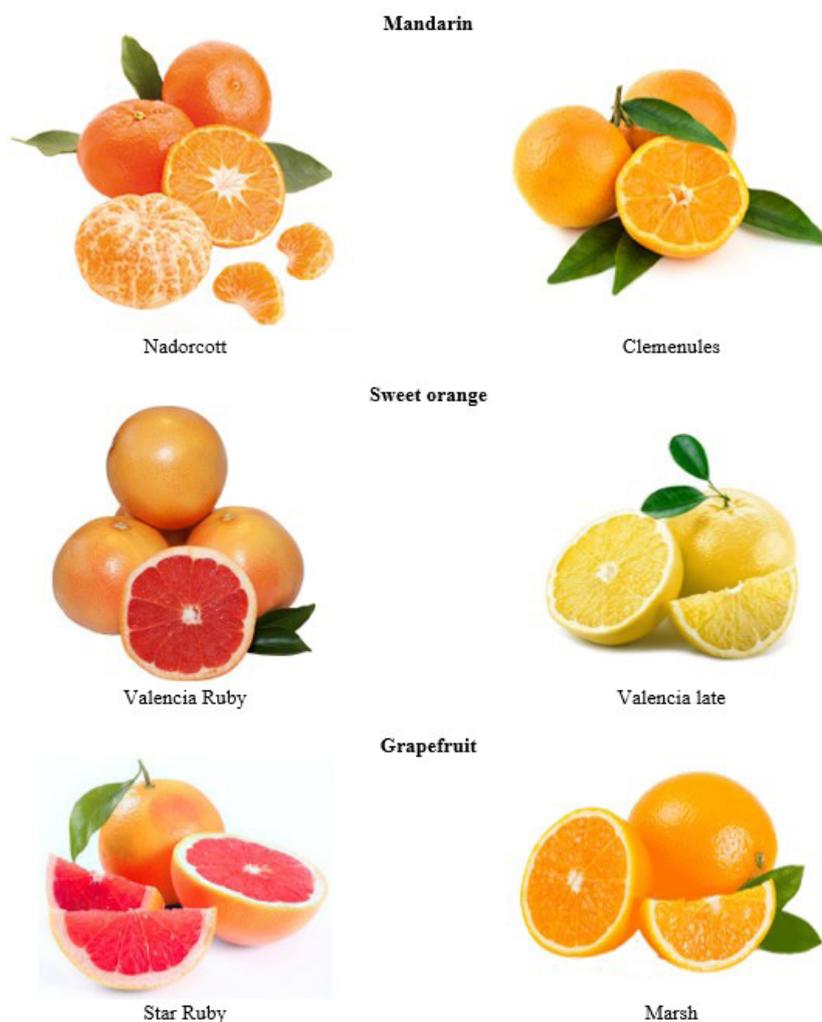
are organic or artificial substances that quench or scavenge nitrogen species or reactive oxygen, such as free radicals, to postpone or avoid physiological oxidants' oxidative cell damage (Lobo et al., 2010; Sen et al., 2010). Citrus fruit contains carotenoids, which are one of the significant phytochemicals. With a wide range of carotenoids in regards to amounts and types across various cultivars and species, citrus fruits are among the highly sophisticated suppliers of carotenoids (Kato, 2012). A unique mix of hydrophilic molecules is responsible for the antioxidant activity of citrus fruit (Buljeta et al., 2021). Carotenoids, which are effective antioxidants that scavenge peroxy radicals and singlet molecular oxygen, serve a vital function in plants in shielding biological systems from oxidative damage and photooxidative processes (Kancheva & Kasaikina, 2013; Krause, 2019). Over the years, a considerable number of researchers have investigated the citrus fruit extracts' antioxidant properties concentrating on antioxidant evaluation of particular solvents by the bioactive components of a single extraction, regardless of each component family's preferential solubility in the mixture of extraction (Durazzo, 2017; Gómez-Mejía et al., 2019; Jayaprakasha et al., 2008). The results of multiple studies reveal that overall phenolic compounds, vitamin C content,

and antioxidant activity in the juice and pulp of various citrus cultivars and species are all positively correlated (Brito et al., 2014; Dong et al., 2019; Ramful et al., 2011; Elkhatim et al., 2018). Regardless of the fact that certain carotenoids have long been known to have antioxidant properties, their proportional role to total antioxidant capacity (TAC) in foods remains debated (Koszewska & Kuzak, 2021).

The main objective of this work has been to assess the hydrophilic antioxidant capacity (HAC) fractions using the ABTS and DPPH radical scavenging tests, employing the genetic variation in citrus fruits' carotenoid and coloring makeup. For such purpose, we investigated Vit C content, as well as carotenoids composition and content, in fruits from two genotypes of oranges with differing pulp coloring, two mandarins, and two grapefruits.

## 2 Material and methods

Two mandarin genotypes, Nadorcott and Clemenules, two grapefruit genotypes, Star Ruby and Marsh, and two sweet orange genotypes, Valencia Ruby and Valencia Late, were chosen for their distinctive pulp colors (Figure 1).



**Figure 1.** Image of citrus cultivars' mature fruit utilized in this research.

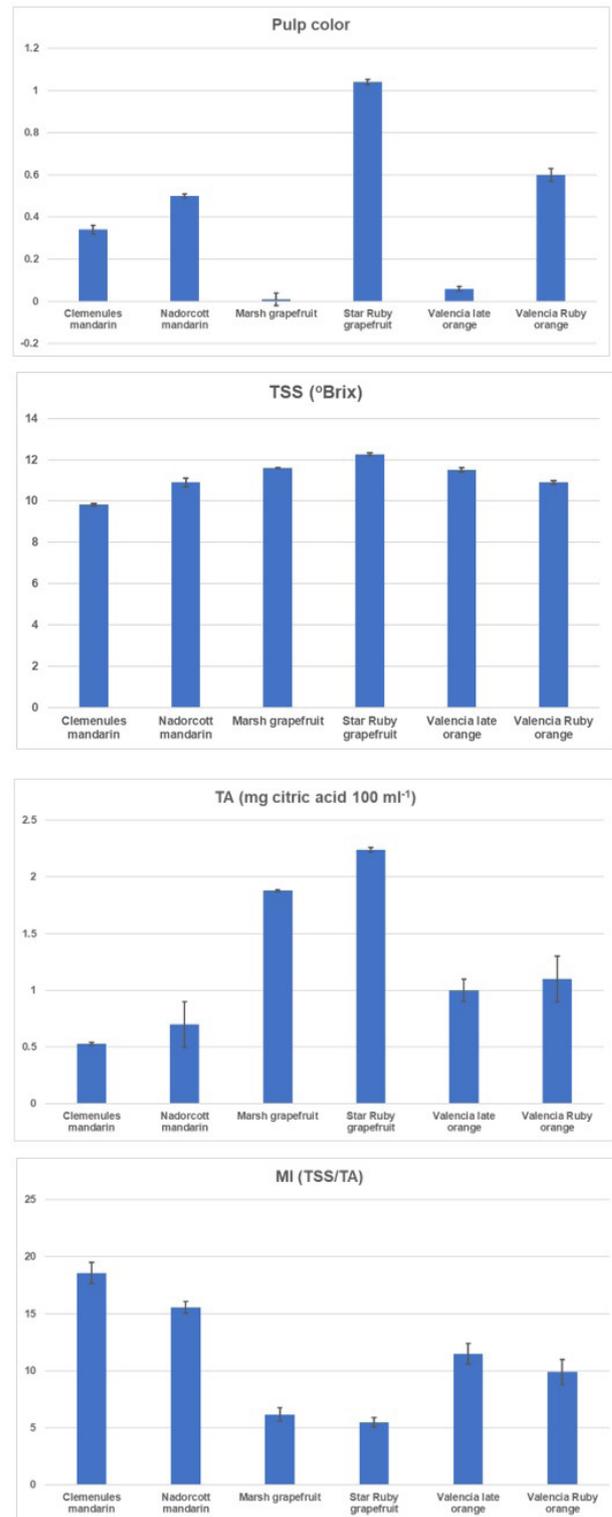
Fruits from mature trees grown within the typical growing and agronomic conditions were gathered for each genotype. Fruits have been chosen based on size consistency and the absence of any exterior defect or injury. After the pulp color was determined, pulp tissue was prepared by excising tiny cube portions of around 1 cm<sup>2</sup> comprising free of segment membranes juice vesicles. Utilizing a Braun MPZ22 Citrus Juicer, the remaining pulp was squeezed for juice and filtered through a 0.8 mm metal sieve. Liquid nitrogen was used to quickly freeze the samples, which were then kept at -80 °C, awaiting examination. A Minolta Chroma Meter CR-400 was used to measure the color of the pulp from three distinct locations.

The color of citrus fruit was determined using a well-known relation, the Hunter *a/b* ratio was used to express color, and the Hunter input variables *a* and *b* were calculated (Carmona et al., 2010; Venkatram et al., 2017). Color index data are also the averages of 10 fruits minimum  $\pm$  SD for each cultivar. A digital refractometer was used to measure the total titratable acidity (TA) and total soluble solids (TSS) of the juices (Teerachaichayut & Ho, 2017; Włodarska et al., 2017). The TSS/TA relation was used to determine the maturity index (MI). Each sample's carotenoid composition was determined by employing high-performance liquid chromatography (HPLC) (Ahamad et al., 2007; Hodisan et al., 1997). According to Alós et al. (2021), total Vit C was evaluated following ascorbic acid extraction. The antioxidant capacity was determined using the ABTS test. TAC was measured using the method published by Curi et al. (2021), with a few tweaks, allowing for the determination of antioxidant capacity due to hydrophilic chemicals. With slight modifications, Girennavar et al. (2007) DPPH free radical test was also used to assess the hydrophilic antioxidant activity. As Trolox equivalent antioxidant capacity (TEAC), a variety of bioactive substances' relative antioxidant capacity (RAC) was determined by Müller et al. (2011). We calculated the correlation of Vit C and carotenoids contained in the samples' hydrophilic extracts to the capacity of antioxidant using these results as a benchmark. Following the assessment of the HAC, as well as the amounts of various chemicals, their contribution to the capacity of antioxidant was estimated. XLSTAT software's one-way analysis of variance was used to calculate the statistical significance, and any substantial variations across cultivars at  $p < 0.05$  were determined using Tukey's test.

### 3 Results and discussion

Figure 2 shows the fruit quality metrics (MI, TA, TSS, and pulp color or *a/b* Hunter) of the six citrus genotypes used in this investigation.

The key factors for choosing these genotypes were the color of the pulp (Hunter ratio) and variations across crops of the same species. As a result, Nadorcott's orange coloring was brighter than Clemenules' mild orange pigmentation when it came to mandarin fruit. Clemenules mandarin, on the other hand, has less strong internal and exterior color compared to other mandarin cultivars, although having great fruit quality and economic value. Compared to Clementine mandarin, the TSS of Nadorcott was somewhat greater, implying a fairly later-ripening, while in both genotypes, the MI at reaping season



**Figure 2.** MI (Maturity Index), TA (Total Acidity), TSS (Total Soluble Solids), and Hunter ratio (color index) of the pulp of citrus genotypes studied (mean  $\pm$  standard deviation,  $n = 3$ ).

was identical. Contrary to Marsh grapefruit pulp's customary pale yellowish coloring, Star Ruby grapefruit has a distinctive red coloration. Internal MI, on the other hand, was comparable in both grapefruits (Figure 2), showing that, apart from fruit

coloring, the aging process under Mediterranean circumstances tends to follow an identical pattern in both cultivars. The pale orange tint of Valencia's late orange fruit contrasts sharply with the reddish coloring of the pulp taken at the same stage of maturity and under the same climatic environments, demonstrating the phenotypic difference. Having moderate to low color and carotenoid concentration, Valencia late is a well-known orange cultivar that is grown all over the world. Internal maturation metrics in Valencia oranges were statistically equal, showing that the mutant's additional ripening mechanisms unaffected (Figure 2). The carotenoid composition and content of the six citrus genotypes fruits studied were compared to the variations in pulp coloring. Amongst the six genotypes, pulp's total carotenoid concentration differed greatly (Table 1).

Nadorcott's pulp has 3.2 times more carotenoid concentration than Clemenules'. In comparison with their corresponding counterpart, carotenoids were found to be much greater in orange and the red-fleshed grapefruit. Relative to the red Star Ruby, the carotenoids in the white Marsh grapefruit were significantly low. In the Valencia Ruby's pulp, the total carotenoids have been over

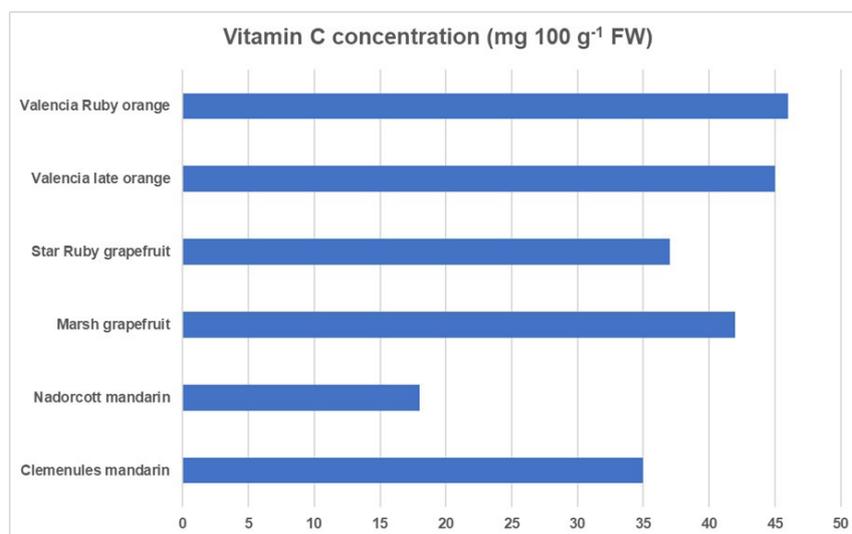
20 times greater than in Valencia's pulp (Table 1). Despite the fact that other carotenoids were practically similar in Clemenules, the Nadorcott mandarin pulp had 4 and 6 times more violaxanthin and  $\beta$ -cryptoxanthin, respectively. The deeper orange coloration detected in the Nadorcott pulp relative to Clemenules may be explained by these fundamental variations in these xanthophylls (Table 1). The major carotenoid in mandarin juice and pulp is  $\beta$ -cryptoxanthin, which gives the fruit its bright orange color. Vitamin C is probably best known for its antioxidant properties in plant cells, and it's a big part of the citrus fruit's health advantages. The content of vitamin C in the six cultivars pulp used for this investigation is shown in Figure 3.

The antioxidant properties of the citrus varieties were studied to determine the role of carotenoid concentration and Vit C to the citrus fruit's antioxidant capacity. ABTS and DPPH scavenging assays were used to estimate the HAC fraction (Figure 4).

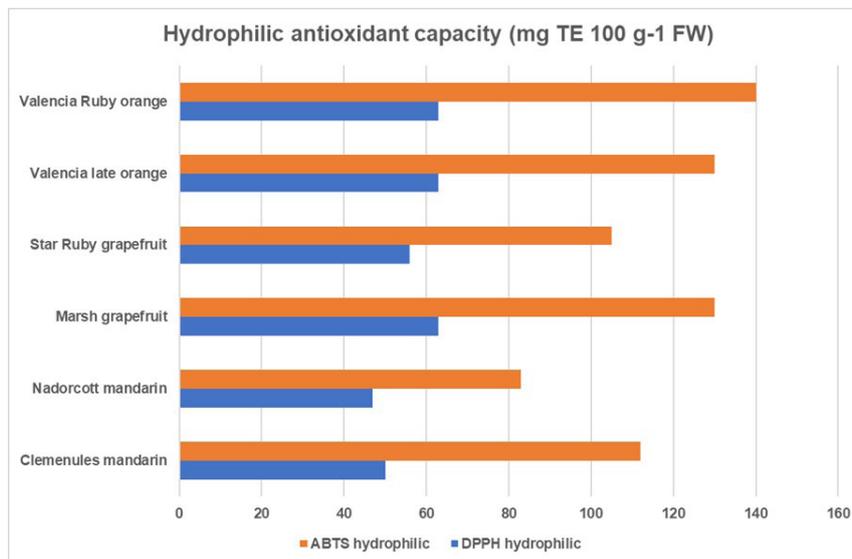
For the six citrus cultivars investigated, the value of  $R^2$  of 0.91 and 0.85 for the ABTS and DPPH tests, correspondingly, show a good correlation between Vit C and HAC. According to these findings,

**Table 1.** Composition and content of carotenoids in the pulp of the samples under study (mean  $\pm$  SD).

Carotenoid (mg 100 g <sup>-1</sup> FW)	Oranges		Grapefruits		Mandarins	
	Valencia late	Valencia Ruby	Marsh	Star Ruby	Clemenules	Nadorcott
Antheraxanthin	0.14 $\pm$ 0.01	0.11 $\pm$ 0.03	0.01 $\pm$ 0.01	0.02 $\pm$ 0.02	0.10 $\pm$ 0.01	0.32 $\pm$ 0.02
$\beta$ -Carotene	nondetected	0.04 $\pm$ 0.01	nondetected	0.57 $\pm$ 0.02	traces	0.24 $\pm$ 0.12
$\beta$ -Cryptoxanthin	0.06 $\pm$ 0.02	0.03 $\pm$ 0.01	nondetected	0.01 $\pm$ 0.01	0.31 $\pm$ 0.03	1.83 $\pm$ 0.50
$\xi$ -Carotene	0.03 $\pm$ 0.01	0.06 $\pm$ 0.01	nondetected	0.03 $\pm$ 0.01	0.07 $\pm$ 0.01	0.26 $\pm$ 0.13
Luteoxanthin	0.06 $\pm$ 0.02	0.02 $\pm$ 0.01	nondetected	nondetected	0.02 $\pm$ 0.01	0.11 $\pm$ 0.03
Lycopene	nondetected	0.86 $\pm$ 0.02	nondetected	0.74 $\pm$ 0.08	nondetected	nondetected
Phytoene	0.1 $\pm$ 0.01	13.31 $\pm$ 0.11	0.03 $\pm$ 0.01	0.81 $\pm$ 0.02	0.26 $\pm$ 0.03	0.27 $\pm$ 0.14
Phytofluene	0.02 $\pm$ 0.01	1.84 $\pm$ 0.36	0.01 $\pm$ 0.01	0.32 $\pm$ 0.01	0.31 $\pm$ 0.27	0.28 $\pm$ 0.13
Violaxanthin	0.38 $\pm$ 0.02	0.23 $\pm$ 0.01	0.01 $\pm$ 0.01	nondetected	0.25 $\pm$ 0.01	1.00 $\pm$ 0.04
Zeaxanthin	0.07 $\pm$ 0.02	0.06 $\pm$ 0.01	nondetected	0.03 $\pm$ 0.03	0.04 $\pm$ 0.01	0.09 $\pm$ 0.02
Total carotenoids	0.86 $\pm$ 0.03	16.51 $\pm$ 0.28	0.06 $\pm$ 0.01	2.60 $\pm$ 0.15	1.44 $\pm$ 0.26	4.69 $\pm$ 1.06



**Figure 3.** Vitamin C levels of the citrus cultivars' pulp under study.



**Figure 4.** In the citrus cultivars' pulp, the HAC fraction was measured using ABTS and DPPH.

vitamin C content in the pulp of sweet oranges, grapefruits, and mandarins is linked to their HAC. In comparison, both ABTS and DPPH tests revealed a near-complete lack of association between carotenoid concentration and hydrophilic activity.

#### 4 Conclusion

In vitro, antioxidant tests could be valuable markers for estimating the healthful qualities of the consumable section of fruits. Regrettably, due to the absence of standardization, the techniques' differences, and the potential of chemicals found to quench or scavenge various radicals, inconsistencies can arise, leading to an underestimation or overestimation of the particular contribution of the various hydrophilic constituents of a food sample. To assess HAC in the citrus cultivars' pulp throughout this research, we employed two antioxidant assays with different carotenoid composition, content, and coloring. The findings revealed a link between vitamin C concentration and HAC. The proportion of vitamin C to the total HAC was calculated to be between 15% and 30%.

#### References

- Ahamad, M. N., Saleemullah, M., Shah, H. U., Khalil, I. A., & Saljoqi, A. U. R. (2007). Determination of beta carotene content in fresh vegetables using high performance liquid chromatography. *Sarhad Journal of Agriculture*, 23(3), 767-770.
- Ahmed, W., & Azmat, R. (2019). Citrus: an ancient fruits of promise for health benefits. In M. Sajid & A. Amanullah (Eds.), *Citrus: health benefits and production technology* (pp. 19-30). London: IntechOpen. <http://dx.doi.org/10.5772/intechopen.79686>.
- Alós, E., Rey, F., Gil, J. V., Rodrigo, M. J., & Zacarias, L. (2021). Ascorbic acid content and transcriptional profiling of genes involved in its metabolism during development of petals, leaves, and fruits of orange (*Citrus sinensis* cv. Valencia Late). *Plants*, 10(12), 2590. <http://dx.doi.org/10.3390/plants10122590>. PMID:34961061.
- Alquezar, B., Rodrigo, M. J., Lado, J., & Zacarias, L. (2013). A comparative physiological and transcriptional study of carotenoid biosynthesis in white and red grapefruit (*Citrus paradisi* Macf.). *Tree Genetics & Genomes*, 9(5), 1257-1269. <http://dx.doi.org/10.1007/s11295-013-0635-7>.
- Aune, D. (2019). Plant foods, antioxidant biomarkers, and the risk of cardiovascular disease, cancer, and mortality: a review of the evidence. *Advances in Nutrition*, 10(Suppl. 4), S404-S421. <http://dx.doi.org/10.1093/advances/nmz042>. PMID:31728499.
- Ballistreri, G., Fabroni, S., Romeo, F. V., Timpanaro, N., Amenta, M., & Rapisarda, P. (2019). Anthocyanins and other polyphenols in citrus genus: biosynthesis, chemical profile, and biological activity. In R. R. Watson (Ed.), *Polyphenols in plants: isolation, purification and extract preparation* (2nd ed., pp. 191-215). London: Elsevier. <http://dx.doi.org/10.1016/B978-0-12-813768-0.00014-1>.
- Brito, A., Ramirez, J. E., Areche, C., Sepúlveda, B., & Simirgiotis, M. J. (2014). HPLC-UV-MS profiles of phenolic compounds and antioxidant activity of fruits from three citrus species consumed in northern Chile. *Molecules*, 19(11), 17400-17421. <http://dx.doi.org/10.3390/molecules191117400>. PMID:25356563.
- Buljeta, I., Pichler, A., Šimunović, J., & Kopjar, M. (2021). Polyphenols and antioxidant activity of citrus fiber/blackberry juice complexes. *Molecules*, 26(15), 4400. <http://dx.doi.org/10.3390/molecules26154400>. PMID:34361554.
- Carmona, L., Rodrigo, M. J., & Zacarias, L. (2010). Exploring the involvement of ethylene in the regulation of color changes in citrus fruit. *Acta Horticulturae*, 934, 879-885.
- Curi, P. N., Schiassi, M. C. E. V., Pio, R., Peche, P. M., Albergaria, F. C., & Souza, V. R. (2021). Bioactive compounds and antioxidant activity of fruit of temperate climate produced in subtropical regions. *Food Science and Technology*, 41(3), 607-614. <http://dx.doi.org/10.1590/fst.23420>.
- Czech, A., Zarycka, E., Yanovych, D., Zasadna, Z., Grzegorzczak, I., & Klys, S. (2020). Mineral content of the pulp and peel of various citrus fruit cultivars. *Biological Trace Element Research*, 193(2), 555-563. <http://dx.doi.org/10.1007/s12011-019-01727-1>. PMID:31030384.
- Dong, X., Hu, Y., Li, Y., & Zhou, Z. (2019). The maturity degree, phenolic compounds and antioxidant activity of Eureka lemon [*Citrus limon* (L.) Burm. f.]: a negative correlation between total phenolic content, antioxidant capacity and soluble solid content. *Scientia Horticulturae*, 243, 281-289. <http://dx.doi.org/10.1016/j.scienta.2018.08.036>.

- Durazzo, A. (2017). Study approach of antioxidant properties in foods: update and considerations. *Foods*, 6(3), 17. <http://dx.doi.org/10.3390/foods6030017>. PMID:28264480.
- Durazzo, A., Lucarini, M., Souto, E. B., Cicala, C., Caiazzo, E., Izzo, A. A., Novellino, E., & Santini, A. (2019). Polyphenols: a concise overview on the chemistry, occurrence, and human health. *Phytotherapy Research*, 33(9), 2221-2243. <http://dx.doi.org/10.1002/ptr.6419>. PMID:31359516.
- Elkhatim, K. A. S., Elagib, R. A., & Hassan, A. B. (2018). Content of phenolic compounds and vitamin C and antioxidant activity in wasted parts of Sudanese citrus fruits. *Food Science & Nutrition*, 6(5), 1214-1219. <http://dx.doi.org/10.1002/fsn3.660>. PMID:30065822.
- Girenavar, B., Jayaprakasha, G. K., Jadegoud, Y., Gowda, G. A. N., & Patil, B. S. (2007). Radical scavenging and cytochrome P450 3A4 inhibitory activity of bergaptol and geranylcoumarin from grapefruit. *Bioorganic & Medicinal Chemistry*, 15(11), 3684-3691. <http://dx.doi.org/10.1016/j.bmc.2007.03.047>. PMID:17400460.
- Gómez-Mejía, E., Rosales-Conrado, N., León-González, M. E., & Madrid, Y. (2019). Citrus peels waste as a source of value-added compounds: extraction and quantification of bioactive polyphenols. *Food Chemistry*, 295, 289-299. <http://dx.doi.org/10.1016/j.foodchem.2019.05.136>. PMID:31174761.
- Herrera-Barros, A., Tejada-Tovar, C., & Gonzalez-Delgado, A. D. (2021). Comparative assessment of Al<sub>2</sub>O<sub>3</sub>-modified biomasses from agricultural residues for nickel and cadmium removal. *Journal of Water and Land Development*, 49(4-6), 29-34.
- Hodisan, T., Socaciu, C., Ropan, I., & Neamtu, G. (1997). Carotenoid composition of *Rosa canina* fruits determined by thin-layer chromatography and high-performance liquid chromatography. *Journal of Pharmaceutical and Biomedical Analysis*, 16(3), 521-528. [http://dx.doi.org/10.1016/S0731-7085\(97\)00099-X](http://dx.doi.org/10.1016/S0731-7085(97)00099-X). PMID:9589412.
- Hoppu, U., Puputti, S., & Sandell, M. (2021). Factors related to sensory properties and consumer acceptance of vegetables. *Critical Reviews in Food Science and Nutrition*, 61(10), 1751-1761. <http://dx.doi.org/10.1080/10408398.2020.1767034>. PMID:32441536.
- Jayaprakasha, G. K., Girenavar, B., & Patil, B. S. (2008). Radical scavenging activities of Rio Red grapefruits and sour orange fruit extracts in different in vitro model systems. *Bioresource Technology*, 99(10), 4484-4494. <http://dx.doi.org/10.1016/j.biortech.2007.07.067>. PMID:17935981.
- Kancheva, V. D., & Kasaikina, O. T. (2013). Bio-antioxidants—a chemical base of their antioxidant activity and beneficial effect on human health. *Current Medicinal Chemistry*, 20(37), 4784-4805. <http://dx.doi.org/10.2174/09298673113209990161>. PMID:24274817.
- Kato, M. (2012). Mechanism of carotenoid accumulation in citrus fruit. *Journal of the Japanese Society for Horticultural Science*, 81(3), 219-233. <http://dx.doi.org/10.2503/jjshs1.81.219>.
- Koszewska, J., & Kuzak, Ł. (2021). Exogenous NaHS treatment alleviated Cd-induced stress in *Ocimum basilicum* plants through modulation of antioxidant defense system. *Journal of Water and Land Development*, 50(6-9), 1-9.
- Krause, G. H. (2019). The role of oxygen in photoinhibition of photosynthesis. In C. H. Foyer & P. M. Mullineaux (Eds.), *Causes of photooxidative stress and amelioration of defense systems in plants* (pp. 43-76). Boca Raton: CRC Press. <http://dx.doi.org/10.1201/9781351070454-2>.
- Lacirignola, C., & D'Onghia, A. M. (2009). The Mediterranean citriculture: productions and perspectives. In A. M. D'Onghia, K. Djelouah & C. N. Roistacher. *Citrus tristeza virus and Toxoptera citricidus: a serious threat to the Mediterranean citrus industry* (pp. 13-17, Options Méditerranéennes: série B. Etudes et Recherches, Vol. 65). Bari: CIHEAM.
- Lobo, V., Patil, A., Phatak, A., & Chandra, N. (2010). Free radicals, antioxidants and functional foods: impact on human health. *Pharmacognosy Reviews*, 4(8), 118-126. <http://dx.doi.org/10.4103/0973-7847.70902>. PMID:22228951.
- Ma, G., Zhang, L., Sugiura, M., & Kato, M. (2020). Citrus and health. In M. Talon, M. Caruso & F. G. Gmitter Jr. (Eds.), *The genus citrus* (pp. 495-511). Duxford: Elsevier. <http://dx.doi.org/10.1016/B978-0-12-812163-4.00024-3>.
- Monselise, S. P. (2019). Citrus and related genera. In A. H. Halevy (Ed.), *CRC handbook of flowering* (pp. 275-294). Boca Raton: CRC Press. <http://dx.doi.org/10.1201/9781351072540-38>.
- Müller, L., Fröhlich, K., & Böhm, V. (2011). Comparative antioxidant activities of carotenoids measured by ferric reducing antioxidant power (FRAP), ABTS bleaching assay (αTEAC), DPPH assay and peroxy radical scavenging assay. *Food Chemistry*, 129(1), 139-148. <http://dx.doi.org/10.1016/j.foodchem.2011.04.045>.
- Ramful, D., Tarnus, E., Aruoma, O. I., Bourdon, E., & Bahorun, T. (2011). Polyphenol composition, vitamin C content and antioxidant capacity of Mauritian citrus fruit pulps. *Food Research International*, 44(7), 2088-2099. <http://dx.doi.org/10.1016/j.foodres.2011.03.056>.
- Rangel, C. N., Carvalho, L. M. J., Fonseca, R. B. F., Soares, A. G., & Jesus, E. O. (2011). Nutritional value of organic acid lime juice (*Citrus latifolia* T.), cv. Tahiti. *Food Science and Technology*, 31(4), 918-922. <http://dx.doi.org/10.1590/S0101-20612011000400014>.
- Rodrigo, M. J., Alquézar, B., Alós, E., Lado, J., & Zacarias, L. (2013). Biochemical bases and molecular regulation of pigmentation in the peel of citrus fruit. *Scientia Horticulturae*, 163, 46-62. <http://dx.doi.org/10.1016/j.scienta.2013.08.014>.
- Rodrigues, E., Poerner, N., Rockenbach, I. I., Gonzaga, L. V., Mendes, C. R., & Fett, R. (2011). Phenolic compounds and antioxidant activity of blueberry cultivars grown in Brazil. *Food Science and Technology*, 31(4), 911-917. <http://dx.doi.org/10.1590/S0101-20612011000400013>.
- Sen, S., Chakraborty, R., Sridhar, C., Reddy, Y. S. R., & De, B. (2010). Free radicals, antioxidants, diseases and phytomedicines: current status and future prospect. *International Journal of Pharmaceutical Sciences Review and Research*, 3(1), 91-100.
- Shahidi, F., & Ambigaipalan, P. (2015). Phenolics and polyphenolics in foods, beverages and spices: antioxidant activity and health effects—a review. *Journal of Functional Foods*, 18, 820-897. <http://dx.doi.org/10.1016/j.jff.2015.06.018>.
- Teerachaichayut, S., & Ho, H. T. (2017). Non-destructive prediction of total soluble solids, titratable acidity and maturity index of limes by near infrared hyperspectral imaging. *Postharvest Biology and Technology*, 133, 20-25. <http://dx.doi.org/10.1016/j.postharvbio.2017.07.005>.
- Venkatram, A., Padmavathamma, A. S., Rao, B. S., Sankar, A. S., Manorama, K., & Vijaya, D. (2017). Effect of antioxidants on drying, yield, hunter color L\*, a\*, b\* and moisture content of dried-on-vine raisins prepared from seedless varieties of grape (*Vitis vinifera* L.). *International Journal of Pure & Applied Bioscience*, 5(4), 1127-1134. <http://dx.doi.org/10.18782/2320-7051.5701>.
- Wallace, T. C., Bailey, R. L., Blumberg, J. B., Burton-Freeman, B., Chen, C. O., Crowe-White, K. M., Drewnowski, A., Hooshmand, S., Johnson, E., Lewis, R., Murray, R., Shapses, S. A., & Wang, D. D. (2020). Fruits, vegetables, and health: a comprehensive narrative, umbrella review of the science and recommendations for enhanced public policy to improve intake. *Critical Reviews in Food Science and Nutrition*, 60(13), 2174-2211. <http://dx.doi.org/10.1080/10408398.2019.1632258>. PMID:31267783.
- Włodarska, K., Pawlak-Lemańska, K., Górecki, T., & Sikorska, E. (2017). Classification of commercial apple juices based on multivariate analysis of their chemical profiles. *International Journal of Food*

- Properties*, 20(8), 1773-1785. <http://dx.doi.org/10.1080/10942912.2016.1219367>.
- Xu, G., Liu, D., Chen, J., Ye, X., Ma, Y., & Shi, J. (2008). Juice components and antioxidant capacity of citrus varieties cultivated in China. *Food Chemistry*, 106(2), 545-551. <http://dx.doi.org/10.1016/j.foodchem.2007.06.046>.
- Yashin, A., Yashin, Y., Xia, X., & Nemzer, B. (2017). Antioxidant activity of spices and their impact on human health: a review. *Antioxidants*, 6(3), 70. <http://dx.doi.org/10.3390/antiox6030070>. PMID:28914764.
- Zou, Z., Xi, W., Hu, Y., Nie, C., & Zhou, Z. (2016). Antioxidant activity of citrus fruits. *Food Chemistry*, 196, 885-896. <http://dx.doi.org/10.1016/j.foodchem.2015.09.072>. PMID:26593569.