ISSN 1807-1929



Revista Brasileira de Engenharia Agrícola e Ambiental

v.22, n.3, p.158-163, 2018

Campina Grande, PB, UAEA/UFCG - http://www.agriambi.com.br

DOI: http://dx.doi.org/10.1590/1807-1929/agriambi.v22n3p158-163

Water salinity, nitrogen and phosphorus on photochemical efficiency and growth of west indian cherry

Francisco V. da S. Sá¹, Hans R. Gheyi², Geovani S. de Lima¹, Emanoela P. de Paiva³, Rômulo C. L. Moreira¹ & Luderlândio de A. Silva¹

- ¹ Universidade Federal de Campina Grande/Centro de Tecnologia e Recursos Naturais/Unidade Acadêmica de Engenharia Agrícola. Campina Grande, PB. E-mail: vanies_agronomia@hotmail.com ORCID: 0000-0001-6585-8161 (Corresponding author); geovanisoareslima@gmail.com ORCID: 0000-0001-9960-1858; romulocarantino@gmail.com ORCID: 0000-0002-4079-4939; luderlandioandrade@gmail.com ORCID: 0000-0001-9496-5820
- ² Universidade Federal do Recôncavo da Bahia/Núcleo de Engenharia de Água e Solo. Cruz das Almas, BA. E-mail: hans@pq.cnpq.br ORCID: 0000-0002-1066-0315
- ³ Universidade Federal Rural do Semi-Árido/Centro de Ciências Agrárias/Programa de Pós-Graduação em Fitotecnia. Mossoró, RN. E-mail: emanuelappaiva@hotmail.com ORCID: 0000-0003-4510-9205

Key words:

chlorophyll fluorescence saline stress fertilization Malphigia emarginata

ABSTRACT

The objective of this study was to evaluate the interaction between water salinity and doses of nitrogen and phosphorus on chlorophyll a fluorescence and on the growth of West Indian Cherry ('acerola') plants in the initial growth stage. The research was carried out in a protected environment, in lysimeters filled with Regolithic Neosol of loamy clay texture and low initial phosphorus content. The experiment was conducted in a randomized block design, arranged in a factorial scheme with two factors, five levels of electrical conductivity of the irrigation water - ECw (0.6, 1.4, 2.2, 3.0 and 3.8 dS m $^{-1}$) and four proportions of the recommendation of phosphorus/nitrogen - P/N (100:100, 140:100, 100:140 and 140:140% P/N of recommendation), with three replicates, and each plot consisted of one plant. Water salinity above 2.2 dS m $^{-1}$ decreases the quantum efficiency of photosystem II and the growth of west indian cherry plants. Increase in nitrogen and/or phosphorus supply improves the energy stability of west indian cherry plants by reducing the effects of salt stress on the quantum efficiency of photosystem II. The increase of 40% in the nitrogen dose increased the chlorophyll a fluorescence, but did not influence the plant growth.

Palavras-chave:

fluorescência da clorofila estresse salino adubação Malphigia emarginata

Salinidade da água, nitrogênio e fósforo na eficiência fotoquímica e crescimento da aceroleira

RESUMO

Objetivou-se avaliar a interação entre salinidade da água e doses de nitrogênio e fósforo sobre a fluorescência da clorofila a e o crescimento das plantas de aceroleira na fase inicial de crescimento. A pesquisa foi realizada em ambiente protegido, em lisímetros preenchidos com Neossolo Regolítico de textura franco-argilosa e baixo teor inicial de fósforo. O experimento foi instalado em delineamento de blocos casualizados, arranjados em esquema fatorial com dois fatores, sendo cinco níveis de condutividade elétrica da água de irrigação - CEa (0,6; 1,4; 2,2; 3,0 e 3,8 dS m⁻¹) e quatro proporções da recomendação de fósforo nitrogênio -P/N (100:100; 140:100; 100:140 e140:140% P/N da recomendação), com três repetições sendo cada parcela foi constituída de uma planta. Água de salinidade superior à 2,2 dS m⁻¹ diminui a eficiência quântica do fotossitema II e o crescimento das plantas de aceroleira. O incremento no fornecimento de nitrogênio e/ou fósforo melhora a estabilidade energética das plantas de acerola, reduzindo os efeitos do estresse salino sobre a eficiência quântica do fotossitema II. O incremento de 40% na dose de nitrogênio aumentou a florescência da clorofila a das plantas de acerola, porém não influenciou o crescimento das plantas.



Introduction

West indian cherry (*Malpighia emarginata* L.) is a tropical fruit crop whose fruits stand out for their high content of vitamin C, which varies from 1000 to 1800 mg $100g^{-1}$ (Cunha Neto et al., 2012). In Brazil, the crop is estimated to occupy an area of approximately 10,000 ha and the states with highest production are Bahia, Ceará, Paraíba and Pernambuco, responsible for 60% of the national production (Esashika et al., 2013).

Despite the expressive contribution of the west indian cherry crop to the Northeast region of the country, its yield is limited by numerous biotic and abiotic factors, particularly the availability and quality of water for irrigation, fertilization management, low rainfall and irregular rainfalls in semi-arid areas. The use of water with large amount of salts may inhibit plant growth and production, due to the reduction in the soil solution osmotic potential, possibly causing also ionic toxicity, nutritional imbalances or both, simultaneously, due to the excessive accumulation of chloride and sodium in the protoplasm (Munns & Tester, 2008; Syvertsen & Garcia-Sanchez, 2014; Sá et al., 2015). Another inconvenience is plant sensitivity to the salts, which varies according to the species, concentration and ionic nature of the salts, exposure period, phenological stage, availability of nutrients, such as N, P and K, and their interaction (Ashraf & Harris, 2004; Esteves & Suzuki, 2008).

Monitoring the effects of saline stress on plants becomes necessary to understand their physiological behavior under saline stress (Habibi & Amiri, 2013; Sá et al., 2015). In this context, there is also the importance of the interaction between water salinity and soil fertility. Given the above, this study aimed to evaluate the effect of water salinity and nitrogen/phosphorus combination on chlorophyll *a* fluorescence during the initial growth of grafted west indian cherry plants.

MATERIAL AND METHODS

The experiment was carried out in a greenhouse at the Center of Technology and Natural Resources (CTRN) of the Federal University of Campina Grande (UFCG), located in the municipality of Campina Grande-PB, Brazil (7° 15' 18" S; 35° 52' 28" W; 550 m).

The experiment was installed in a randomized block design, in 5 x 4 factorial scheme, corresponding to five levels of irrigation water electrical conductivity (0.6, 1.4, 2.2, 3.0 and 3.8 dS m^{-1}) and four proportions of the phosphorus/nitrogen (P/N) recommendation (100:100; 140:100; 100:140 and

140:140% P/N), with three replicates and one plant per plot. The recommendation corresponding to 100% of N fertilization (23.85 g of N plant year⁻¹) and P fertilization (45.0 g P_2O_5 plant year⁻¹) was based on Musser (1995).

The experiment was installed in lysimeters with capacity for 250 dm³ whose bottom part was covered with Bidim geotextile and a 5 cm layer of type-1 crushed stone. The soil used to fill the lysimeters was classified as Regolithic Neosol with clay loam texture (0-30 cm layer), from the municipality of Esperança-PB, whose physical and chemical attributes (Table 1) were determined following methodologies proposed by Donagema et al. (2011)

West indian cherry seedlings, cv. BRS 366-Jaburu, grafted on a 'Criolo' rootstock, came from the clone garden of Embrapa Tropical Agroindustry, in Pacajus, CE. The obtained seedlings were 240 days old and, during this period, were irrigated using low-salinity water (0.6 dS m⁻¹). Transplanting was performed 90 days after grafting and the water salinity treatments started 30 days after transplanting.

Solutions with lower electrical conductivity levels (ECw of 0.6 and 1.4 dS m $^{-1}$) were obtained by diluting municipal public-supply water (ECw = 1.78 dS m $^{-1}$) in rainwater, whereas the other treatments (ECw of 2.2, 3.0 and 3.8 dS m $^{-1}$) were prepared by adding NaCl, CaCl $_2$.2H $_2$ O and MgSO $_4$.7H $_2$ O salts to the public-supply water, at equivalent proportion of 7:2:1, respectively, representing the average composition of the contents of ions present in waters used for irrigation in the semi-arid region of Northeast Brazil (Medeiros et al., 2003).

Irrigations using water according to the preestablished ECw levels were performed every 3 days, replacing the evapotranspired volume to increase soil moisture to close to the maximum water retention capacity (33.42 kPa). The volume applied in each irrigation was determined according to plant water demand, based on water balance in the root zone, obtained by the difference between the applied volume minus the volume drained in the previous irrigation. To avoid the accumulation of salts in the root zone, a leaching fraction of 0.10 was applied every 50 days (Ayers & Westcot, 1999).

Fertilizations with N and P were performed according to the preestablished treatments, using urea (45% N) as source of N and single superphosphate (18% P_2O_5 , 18% Ca^{2+} , 12% S) as source of P, based on the recommendation of Musser (1995), adding 19.8 g of K_2O per plant, using potassium chloride (60% K_2O). 250 and 350 g of single superphosphate were applied as basal fertilization in the treatments of 100 and 140% of P, respectively. N fertilization was split into 24 equal applications,

Table 1. Chemical and physical-hydraulic characteristics of the soil used in the experiment

	Chemical characteristics											
pH (H ₂ O)	OM	Р	K+	Na ⁺	Ca ²⁺	Mg ²⁺	Al ³⁺	H+	ECse			
(1:2.5)	dag kg ⁻¹	(mg kg ⁻¹)		(cmol _c kg ⁻¹) (d								
5.63	1.83	18.20	0.21	0.17	3.49	2.99	0.00	5.81	0.61			
Physical characteristics												
Gra	Granulometric fraction				re (kPa)	AW	W Total navasity	Da	Dp			
Sand	Silt	Clay	Textural class	33.42	1519.5	AW	Total porosity — m³ m⁻³	(ka	dm-3)			
	g kg ⁻¹		Ciass -	dag kg ⁻¹				(kg dm ⁻³)				
572.7	100.7	326.6	CL	12.68	4.98	7.70	0.5735	1.13	2.65			

OM - Organic matter: Walkley-Black wet digestion; Ca²⁺ and Mg²⁺ extracted with 1 mol L¹ KCl at pH 7.0; Na⁺ and K⁺ extracted with 1 mol L¹ NH₄OAc at pH 7.0; CL - Clay loam; AW - Available water; Da- Apparent density; Dp- Particle density; ECse - Electrical conductivity of the soil saturation extract

at 15-day intervals along the year, using 2.21 g of urea per plant in the treatment of 100%, and 3.09 g of urea per plant in the treatment of 140%. Potassium fertilization was divided into 12 equal applications, monthly performed, using 2.75 g of KCl per plant. Fertilization treatments started as seedlings were transplanted to the lysimeters. Pests, diseases and weeds were controlled according to the need.

Initial fluorescence (Fo), maximum fluorescence (Fm), variable fluorescence (Fv = Fm-Fo) and maximum quantum efficiency of photosystem II (Fv/Fm) were evaluated at 150 days after applying the salinity levels, using a hand-held pulse-modulated fluorometer (Model OS5p - Opti Science). The Fv/Fm protocol was used, as performed by Sá et al. (2015) in mandarin hybrids. Evaluations were performed after leaf adaptation to the dark for 30 min, using the dark-adaptation clip of the device, to guarantee that all primary acceptors were oxidized, i.e., the reaction centers were open.

West indian cherry growth was analyzed based on measurements of rootstock and scion diameter to obtain the absolute growth rate (AGR) and relative growth rate (RGR), for the period between 1 (t_1) and 150 days (t_2) after applying the salinity levels (DASL), and on the number of leaves (NL). Rootstock diameter (mm) was measured at 2 cm from the soil, whereas scion diameter was determined at 2 cm above the grafting point. These data were used to calculate AGR and RGR using Eqs.1 and 2, respectively (Benincasa, 2003).

$$AGR = \frac{A2 - A1}{t2 - t1} \tag{1}$$

$$RGR = \frac{\ln A2 - \ln A1}{t2 - t1} \tag{2}$$

where:

 ${\rm A}_{\scriptscriptstyle 2}$ - variable under study, obtained at the end of the period, mm:

 $\boldsymbol{A}_{_{1}}$ $\,$ - variable under study, obtained, at the beginning of the period, mm;

 t_2 - t_1 - difference of time between the observations, days; and,

ln - natural logarithm.

Data were subjected to analysis of variance by F test; means of the P/N proportions were compared by Tukey test (p < 0.05) and means of the irrigation water salinity levels were subjected to regression analysis, using the software Sisvar, version 5.1 (Ferreira, 2011).

RESULTS AND DISCUSSION

There was significant interaction (p < 0.05) of water salinity x fertilization managements on initial fluorescence (Fo) and quantum efficiency of photosystem II (Fv/Fm) (Table 2). The absolute and relative growth rates in diameter of the rootstock and number of leaves responded to the isolated effects (p < 0.05) of irrigation water salinity, whereas maximum fluorescence and variable fluorescence were significantly influenced only by /P/N proportions (Table 2).

Initial fluorescence (Fo) of west indian cherry plants treated with the P/N proportions of 100:100% and 100:140% increased as water salinity increased, with sharp increment from the ECw levels of 1.06 and 0.96 dS m⁻¹, respectively (Figure 1A). The values relative to plants under P/N proportions of 140:100% and 140:140% did not fit to any mathematical model, showing mean values of 196.8 and 195.3 electrons quantum⁻¹, respectively. Initial fluorescence (Fo) explains the capacity of oxidation of the quinone (QA), primary receptor of electrons in the PSII (P680) reaction center; thus, increment in Fo indicates destruction of the PS (P680) reaction centers or reduction in the capacity of excitation energy transfer from the antenna to the PSII (P680), caused by the saline stress (Baker & Rosenqvst, 2004; Munns & Tester, 2008; Lucena et al., 2012; Sá et al., 2015).

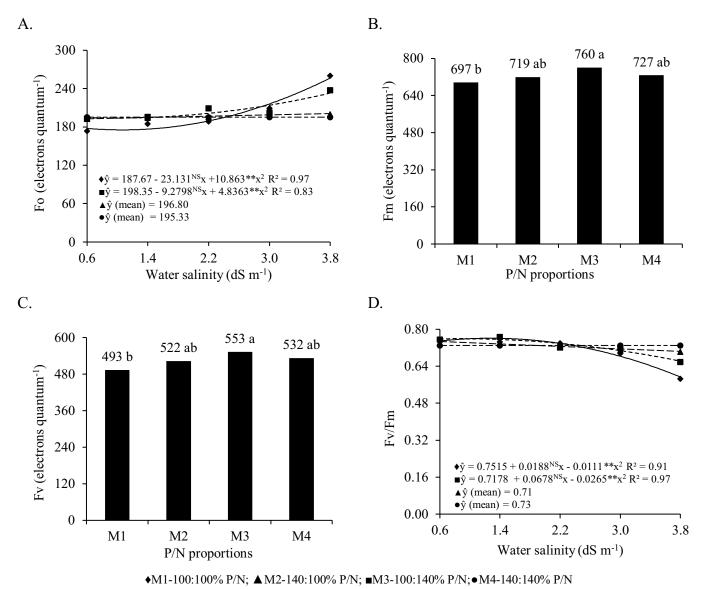
Increment of N dose in the P/N combinations stimulated maximum fluorescence (Fm) and variable fluorescence (Fv) in west indian cherry plants under 100:140% P/N, and these indices were 9.03 and 12.17% higher in the treatment 100:140 P/N compared with plants in the control treatment (100:100% P/N) (Figure 1B and C). The maximum intensity of fluorescence (Fm) refers to the state at which the PSII reaction centers reached their maximum capacity, evidencing the reduced condition of all quinone (QA) by the electrons transferred from the P680 (Baker & Rosenqvst, 2004; Mendonça et al., 2010; Silva et al., 2014). Fm directly influences the variable fluorescence, which is the potentially active energy in the PSII. Hence, the increments of Fv observed in plants of the treatment 100:140% P/N are related to the gains in variable fluorescence. Increment of fluorescence may be related to the higher chlorophyll content in the reaction centers of the treatments that received higher N doses, since N is crucial for the synthesis of the chlorophyll molecule (Taiz et al., 2017).

The increase of salinity did not significantly influence the Fv/Fm of plants fertilized with 100:140% and 140:140% P/N, which showed mean values of 0.71 and 0.73 (Figure 1D). For the treatments 100:100% P/N and 140:100% P/N,

Table 2. Summary of F test for the variables: initial fluorescence (Fo), maximum fluorescence (Fm), variable fluorescence (Fv), quantum efficiency of photosystem II (Fv/Fm), absolute growth rates in diameter of rootstock (AGR-DRS) and scion (AGR-DSC), relative growth rates in diameter of rootstock (RGR-DRS) and scion (RGR-DSC) and number of leaves (NL) of west indian cherry plants irrigated with saline water and fertilized with nitrogen and phosphorus

SV	DF -	F test significance								
		Fo	Fm	Fv	Fv/Fm	AGR-DRS	AGR-DSC	RGR-DRS	RGR-DSC	NL
Block	2	ns	*	*	ns	ns	ns	Ns	ns	*
SL	4	*	ns	ns	ns	*	ns	*	ns	*
Fertilization	3	ns	*	*	*	ns	ns	Ns	ns	ns
SL x Fertilization	12	*	ns	ns	*	ns	ns	Ns	ns	ns
CV		7.74	8.78	11.09	3.50	20.29	18.16	25.18	25.83	19.31

^{*}Significant at 0.05 probability level; ns - Not significant; SL - Water salinity level; Fertilization - Proportions of P/N; CV - Coefficient of variation



** and * - Significant at 0.01 and 0.05 probability levels, respectively; NS - Not significant. Same letters do not differ by Tukey test at 0.05 probability level

Figure 1. Initial fluorescence-Fo (A), maximum fluorescence-Fm (B), variable fluorescence-Fv (C) and quantum efficiency of PSII-Fv/Fm (D) of west indian cherry plants irrigated with saline water and fertilized with nitrogen and phosphorus doses

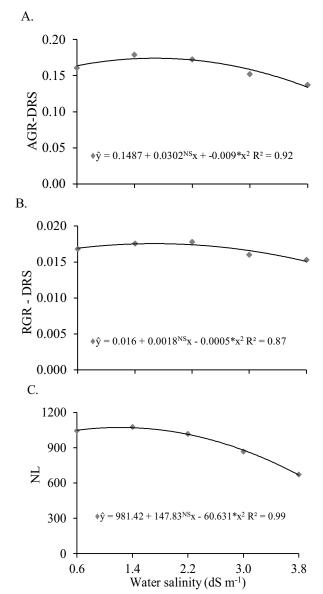
the response was quadratic, with reduction in Fv/Fm from the ECw levels of 0.85 and 1.28 dS m⁻¹, respectively, which coincided with the increment in Fo (Figure 1A and D). The increments in Fo at the highest salinity levels caused a reduction in the quantum efficiency of PSII (Fv/Fm), and this reduction was more noticeable in plants of the treatment 100:100% P/N, compared with the treatment 140:100% P/N, which may be associated with the energy compensation of the plants in the treatment 140:100%, caused by the increment in Fm (Figure 1A and B).

The reduction in quantum efficiency of photosystem II indicates alterations in the photosynthetic system caused by environmental and biotic stress factors (Baker & Rosenqvst, 2004). In general, plants subjected to the combination 100:100% P/N obtained Fv/Fm values lower than those relative to 140:100% P/N,100:140% P/N and 140:140% P/N under irrigation with water salinity above 2.2 dS m⁻¹, which indicates inhibitory action of the saline stress on the chlorophyll *a* fluorescence of these plants from this level on (Figure 1D). Hence, it becomes evident that acerola

plants that received increment in the N and/or P doses were less affected by the saline stress, because they were with a better energy stability compared with the control treatment (100:100% P/N).

Absolute and relative growth rates of the rootstock and number of leaves of west indian cherry plants were influenced by the increase in irrigation water salinity, showing a quadratic behavior, in which the highest values of absolute growth, relative growth and number of leaves, 0.17 mm day⁻¹, 0.018 mm mm⁻¹ day⁻¹ and 1,071 leaves, respectively, were obtained at ECw levels of 1.68, 1.80 and 1.22 dS m⁻¹, respectively (Figure 2A, B and C).

It is observed that the slight increment in water salinity positively influenced the growth of west indian cherry plants, probably due to the addition of calcium and magnesium, which are essential to plant development. However, from the ECw level of 2.2 dS m⁻¹ on, significant alterations occurred in comparison to the control salinity level (0.6 dS m⁻¹). Therefore, concentrations of salts in these waters higher than this level are sufficient to cause osmotic and ionic stress, which can



 * and $^{\text{NS}}$ = Significant at 0.05 probability level and not significant, respectively Figure 2. Absolute (AGR - mm day $^{-1}$) and relative (RGR -mm mm $^{-1}$ day $^{-1}$) growth rates in diameter of rootstock - DRS from 1 to 150 days (A and B) and number of leaves (C) of west indian cherry plants irrigated with saline water and fertilized with nitrogen and phosphorus at 150 days after applying the salinity levels

be confirmed by the alterations observed in chlorophyll a fluorescence (Figures 1A and D).

It can be inferred that irrigation with water salinity above 2.2 dS m⁻¹ increases the contents of salts in the root zone of west indian cherry plants, causing inhibition of growth due to the osmotic effect and toxicity by specific ions, such as sodium and chloride, since there was a reduction in the photochemical efficiency of the west indian cherry plants (Esteves & Suzuki, 2008; Munns & Tester, 2008; Syvertsen & Garcia-Sanchez, 2014; Sá et al., 2015). Growth reduction in fruit species under saline stress conditions have also been observed by Cavalcante et al. (2010) in guava plants, Souto et al. (2013) in noni plants and by Brito et al. (2014) in citrus rootstocks. These authors also attribute these reductions in plant growth to the increment in soil salinity, due to the successive irrigations with saline water, i.e., with high concentration of dissolved salts.

Conclusions

- 1. Irrigation using water with electrical conductivity up to 2.2 dS m⁻¹ is viable for west indian cherry cultivation; at higher levels, there is a reduction in quantum efficiency of photosystem II and plant growth.
- 2. Increment in nitrogen and/or phosphorus doses reduces the negative effects of the saline stress on the quantum efficiency of photosystem II of west indian cherry plants.
- 3. Increment of 40% in nitrogen dose increases chlorophyll a fluorescence of west indian cherry plants, but does not influence plant growth.

ACKNOWLEDGMENTS

To the National Council for Scientific and Technological Development (CNPq) and the Coordination for the Improvement of Higher Education Personnel (Capes) for granting the scholarships and to the National Institute of Science and Technology in Salinity (INCTSal) for the financial aid.

LITERATURE CITED

Ashraf, M.; Harris, P. J. C. Potential biochemical indicators of salinity tolerance in plants. Plant Science, v.166, p.3-16, 2004. https://doi.org/10.1016/j.plantsci.2003.10.024

Ayers, R. S.; Westcot, D. W. Qualidade da água na agricultura. 2.ed. Campina Grande: UFPB, 1999. 153p. Estudos FAO: Irrigação e Drenagem, 29

Baker, N. R.; Rosenqvist, E. Aplication of chlorophyll fluorescence can improve crop production strategies: An examination of future possibilities. Journal of Experimental Botany, v.55, p.1607-1621, 2004. https://doi.org/10.1093/jxb/erh196

Benincasa, M. M. P. Análise de crescimento de plantas: Noções básicas. Jaboticabal: FUNEP, 2003. 41p.

Brito, M. E. B.; Brito, K. S. A. de; Fernandes, P. D.; Gheyi, H. R.;
Suassuna, J. F.; Soares Filho, W. dos S.; Melo, A. S. de; Xavier,
D. A. Growth of ungrafted and grafted citrus rootstocks under saline water irrigation. African Journal of Agricultural Research,
v.9, p.3600-3609, 2014.

Cavalcante, L. F.; Vieira, M. da S.; Santos, A. F. dos; Oliveira, W.M. de; Nascimento, J. A. M. do. Água salina e esterco bovino líquido na formação de mudas de goiabeira cultivar Paluma. Revista Brasileira de Fruticultura, v.32, p.251-261, 2010. https://doi.org/10.1590/S0100-29452010005000037

Cunha Neto, J.; Rabelo, M. C.; Bertini, C. H. C. de M.; Marques, G. V.; Miranda, M. R. A. Caracterização agronômica e potencial antioxidantes de frutos de clones de aceroleira. Revista Ciência Agronômica, v.43, p.713-721, 2012. https://doi.org/10.1590/S1806-66902012000400013

Donagema, G. K.; Campos, D. V. B.; Calderano, S. B.; Texeira, W. G.; Viana, J. H. M. Manual de métodos de análise de solo. 2.ed. Rio de Janeiro, RJ: Embrapa Solos, 2011. 230p.

Esashika, T.; Oliveira, L. A. de; Moreira, F. W. Resposta da aceroleira a adubação orgânica, química e foliar num latossolo da Amazônia Central. Revista de Ciências Agrárias, v.36, p.399-410, 2013.

- Esteves, B. dos S.; Suzuki, M. S. Efeito da salinidade sobre as plantas. Oecologia Brasiliensis, v.12, p.662-679, 2008. https://doi.org/10.4257/oeco.2008.1204.06
- Ferreira, D. F. Sisvar: A computer statistical analysis system. Ciência e Agrotecnologia, v.35, p.1039-1042, 2011. https://doi.org/10.1590/S1413-70542011000600001
- Habibi, F.; Amiri, M. E. Influence of in vitro salinity on growth, mineral uptake and physiological responses of two citrus rootstocks. International Journal of Agronomy and Plant Production, v.4, p.1320-1326, 2013.
- Lucena, C. C. de; Siqueira, D. L. de; Martinez, H. E. P.; Cecon, P. R. Salt stress change chlorophyll fluorescence in mango. Revista Brasileira de Fruticultura, v.34, p.1245-1255, 2012. https://doi.org/10.1590/S0100-29452012000400034
- Medeiros, J. F. de; Lisboa, R.de A.; Oliveira, M. de; Silva Júnior, M. J. da; Alves, L. P. Caracterização das águas subterrâneas usadas para irrigação na área produtora de melão da Chapada do Apodi. Revista Brasileira de Engenharia Agrícola e Ambiental, v.7, p.469-472, 2003. https://doi.org/10.1590/S1415-43662003000300010
- Mendonça, A. V. R.; Carneiro, J. G. de A.; Freitas, T. A. S. de; Barroso, D. G. Características fisiológicas de mudas de *Eucalyptus* spp. submetidas a estresse salino. Ciência Florestal, v.20, p.255-267, 2010. https://doi.org/10.5902/198050981850
- Munns, R.; Tester, M. Mechanism of salinity tolerance. Annual Review of Plant Biology, v.59, p.651-681, 2008. https://doi.org/10.1146/annurev.arplant.59.032607.092911

- Musser, R. S. Tratos culturais na cultura da acerola. In: São José, A. R.; Alves, R. E. (ed.). Acerola no Brasil: Produção e mercado. Vitória da Conquista: DFZ/UESB, 1995. Cap.3, p.47-52.
- Sá, F. V. da S.; Brito, M. E. B.; Silva, L. de A.; Moreira, R. C. L.; Fernandes, P. D.; Figueiredo, L. C. Fisiologia da percepção do estresse salino em híbridos de tangerineira - Sunki Comum sob solução hidropônica salinizada. Comunicata Scientiae, v.6, p.463-470, 2015. https://doi.org/10.14295/cs.v6i4.1121
- Silva, L. de A.; Brito, M. E. B.; Sá, F. V. da S.; Moreira, R. C. L. M.; Soares Filho, W. dos S.; Fernandes, P. D. Mecanismos fisiológicos em híbridos de citros sob estresse salino em cultivo hidropônico. Revista Brasileira de Engenharia Agrícola e Ambiental, v.18, p.S1-S7, 2014. https://doi.org/10.1590/1807-1929/agriambi. v18nsupps1-s7
- Souto, A. G. de L.; Cavalcante, L. F.; Nascimento, J. A. M.do; Mesquita, F. de O.; Lima Neto, A. J. de. Comportamento do noni à salinidade da água de irrigação em solo com biofertilizante bovino. Irriga, v.18, p.442- 453, 2013. https://doi.org/10.15809/ irriga.2013v18n3p442
- Syvertsen, J. P.; Garcia-Sanchez, F. Multiple abiotic stresses occurring with salinity stress in citrus. Environmental and Experimental Botany, v.103, p.128-137, 2014. https://doi.org/10.1016/j.envexpbot.2013.09.015
- Taiz, L.; Zeiger, E.; Møller, I. M.; Murphy, A. Fisiologia e desenvolvimento vegetal. 6.ed. Porto Alegre: Artmed, 2017. 858p.