HEMOLYMPHATIC COMPONENTS IN VECTORS OF Trypanosoma cruzi: STUDY IN SEVERAL SPECIES OF THE SUBFAMILY TRIATOMINAE (HEMIPTERA: REDUVIDAE).

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SUMMARY

The members of the subfamily Triatominae (Hemiptera: Reduviidae) comprise a great number of species of medical importance in the transmission of the **T. cruzi** (American trypanosomiasis).

The aim of this study was to contribute to the knowledge about the chemical composition in proteins, lipids, lipoproteins, and carbohydrates of vectors of Chagas' disease corresponding to twelve members of the subfamily Triatominae. This study was carried out in ninphs of the fifth instar and adult males of the species: T. delpontei, T. dimidiata, T. guasayana, T. infestans, T. mazzotti, T. pallidipennis, T. patagonica, T. platensis, T. rubrovaria, T. sordida of the Triatoma genus, and D. maximus and P. megistus of the Dipatalogaster and Panstrongylus genera respectively.

The results show on one hand, qualitative differences in the protein composition, and on the other hand, similarity in the lipoprotein profiles. Lipids, proteins, and carbohydrates did not show significant differences between species or/and stages.

KEY WORDS: Chagas' disease; Vectors; Proteins; Lipids; Lipoproteins; Carbohydrates.

INTRODUCTION

The study of the hemolymphatic compounds provides valuable information for the understanding of relationships of one insect with another of the same or different species ^{17,34}.

LOUGTHON & WEST (1965) studied the characteristics of development and distribution of the hemolymphatic proteins in Lepidoptera ²². The electrophoretic profile has been established in various insects of medical importance, particularly in different members of the subfamily Triatominae (Hemiptera: Reduviidae) ^{1,23,25,31}.

The major sugar found in the hemolymph of insects is a-trehalose, a non reducing glucose-glucose disacharide, responsible for providing energy of rapid consumption for essential activities ³³.

The lipids, another indispensable metabolic compound, are involved in many processes such as locomotion, reproduction, embryogenesis and

metamorphosis. They also function as hormones in metabolic regulation and as structural components⁸. Lipophorin (HDLp), the major lipoprotein in insects, has been reported as one structure unparalleled in nature by their capacity for lipid binding. The understanding of these biochemical compounds is of great importance in insects which greatly influence upon society through destruction of crops and transmission of diseases ²⁶.

At present, most investigations are concerned principally with two species, that is, Manduca sexta (holometabolous) and Locusta migratoria (hemimetabolous)²⁶, but the information about triatomine bugs is scarce, showing a high variation, probably as consequence of the different physiological situations, nutrition conditions, sex or infection by Blastocrithidia triatomae⁷. This parasite in Triatoma infestans can produce changes in the hemolymphatic profile protein ⁴, delay and high mortality in their development²⁷. Also, modifica-

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tions in the protein pattern have been reported by parasitic actions upon Heliothis zea² and Spodoptera frugiperda¹¹ as well as metabolic modifications characterized by increase of lipids and diminution of proteins in locust¹⁵. HAYAKAWA (1987) described one inhibitor factor of the lipid transport in parasitized insects¹⁴.

Nowadays, the knowledge of the lipid and lipoprotein composition of Chagas' disease vectors, in standard conditions is related to T. infestans³⁰ and Rhodnius prolixus¹³.

The study of the metabolic compounds carried out in our laboratory in members of the subfamily Triatominae demanded one previous analysis in standard conditions of the proteic, lipidic, lipoprotein, and carbohydrate compounds. The present study comprises twelve species corresponding to three genera with epidemiologic importance and of different geographic distribution: ten species corresponding to Triatoma genus and two species belonging to Dipetalogaster and Panstrogylus genera. This study was carried out in fifth instar and adult insects.

MATERIALS AND METHODS

Insects:

Genus Triatoma (Laporte, 1832)

T. delpontei (Romaña & Abalos, 1947).

T. dimidiata (Latrelle, 1811).

T. guasayana (Wygodzinsky & Abalos, 1949).

T. infestans (Klug, 1834).

T. mazzotti (Usinger, 1941).

T. pallidipennis (Stal, 1872).

T. patagonica (Del Ponte, 1929).

T. platensis (Neiva, 1913).

T. rubrovaria (Blanchard, 1843).

T. sordida (Stal, 1859).

Genus Panstrongylus (Berg, 1879)

P. megistus (Burmester, 1835).

Genus Dipetalogaster (Usinger, 1939).

D. maximus (Uhler, 1894).

The experiments were carried out using groups with ten insects, fifth instar (NV) and adult males respectively, reared and maintained in the insectary at 28± 1°C, 60-70 % humidity, 16/8 h day/night cycle, free of infection by T. cruzy or B. triatomae and fed fortnightly on hens. Each group was synchronically moulted, fed and then allowed to fast for 5 days prior to hemolymph collection.

Hemolymph collection: to collect hemo-

lymph, the insects were anesthetized by cooling on ice. The legs were cut off and the insects were placed in a precooled centrifuge tube, and centrifuged at 90 g at 4°C for 1 minute. The hemolymph was then pooled into a precooled tube containing 10 mM ethylenediaminetetracetic acid (disodium salt) (Na₂EDTA), 10 mM dithiothreitol and phenylmethylsulfonyl fluoride (PMSF), N-a-ptosyl-1-lysine chloromethyl ketone (TLCK) and aprotinin 1 mM in order to avoid proteolysis¹⁰. Then it was centrifuged at 10.000 g at 4°C for 10 minutes to remove hemocytes. For the carbohydrate determination, the hemolymph was collected only with Na EDTA and immediately used. The volume of hemolymph collected in each group ranges between 120 µl and 250 µl for NV and from 150 µl to 420 µl for adult males.

Protein, lipid and carbohydrate determinations: protein concentration was quantified by the Bradford method ³, the total lipid content was estimated according to FRINGS at al. ¹² and that of carbohydrates, according to SCOTT et al. ²⁸.

Electrophoresis: in all the cases, 120 μg of proteins were submitted to polyacrylamide gel electrophoresis on 7% acrylamide gels, according to Davis procedure. Scans were performed on a Helena-Scan densitometer. The pre-stained with Sudan Black lipoproteins were fractionated by electrophoresis on polyacrylamide as described previously 6.

Statistical analysis: Student's "t" tests for observations were utilized in all instances 29 . Results are reported as mean \pm SEM.

RESULTS

Overall protein values for adult males and NV of the ten species of Triatoma genus were not significantly different (2.8 \pm 0.5 g/dl and 2.5 \pm 0.4 g/dl respectively). This also applies to **D.maximus** and **P. megistus** (Table 1).

The estimation of total lipids and carbohydrates in the standard conditions did not reveal significant differences. The highest value for the lipids was found in the Triatoma genus for both instars (Table 1).

On the other hand, the fractionation of the hemolymphatic lipoprotein pre-stained with Sudan

Table 1
Protein, lipid and carbohydrate composition in hemolymph of fifth instar (NV) and adult (male) from species of three genera of the subfamily Triatominae (Hemiptera: Reduviidae).

Genus	Protein (g/dl)		Lipid (mg/dl)		Carbohydrate (mg/dl)	
	NV	male	NV	male	NV	male
Triatoma ¹	2.5±0.4(*)	2.8 ± 0.5	108±24	103±31	50.4±8.2	51.2±6.8
Dipetalogaster ²	1.9±0.4	2.2±0.3	95±23	83±18	55.5±7.0	57.2±6.3
Panstrongylus ³	2.6 ± 0.5	2.8 ± 0.4	79±25	87±31	54.5±8.2	53.0±3.8

(*): mean ± SEM, from four determinations carried out in duplicate in groups of ten insects.

(1): T.delpontei; T.dimidiata; T.guasayana; T.infestans; T.mazzotti; T.pallidipennis; T.patagonica;

T.platensis; T.rubrovaria; T.sordida. (2): D.maximus (3): P.megistus.

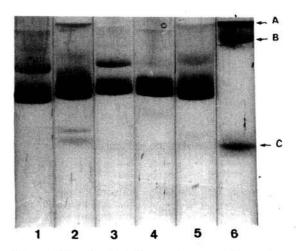


Figure 1: Hemolymphatic lipoproteins separated by polyacrylamide gel electrophoresis.

1-D.maximus (σ); 2-D.maximus (NV); 3-P.megistus (σ); 4-T.infestans (σ); 5-T.platensis (NV); 6-Human serum (A:VLDL, B:LDL, C:HDL).

Black B showed four fractions for NV and two for adults, in the three genera. These fractions compared with a human normal serum, revealed one migration between HDL (High Density Lipoprotein) and LDL (Low Density Lipoprotein) (Fig.1)

The electrophoretic profiles of the proteic components that characterized the different species showed qualitative differences (Fig.2 A,B and Fig.3). For the Triatoma genus, the number of the components ranges between 13 and 18 for NV and from 14 to 20 for adults; T. mazzotti and T. guasayana being the ones with the lowest number of bands presented in this instar, while T. pallidipennis, T. dimidiata, and T. patagonica have the greater number of bands, that is, they have 18. In adults, the species showing the lowest

number of bands was **T. guasayana** with 14 and **T. infestans** showed the greatest number with 20. **D. maximus** and **P. megistus** in the fifth instar show 17 and 18 proteic bands respectively whereas adults of **D. maximus** showed 18 and **P. megistus** showed 21.

DISCUSSION

Biochemistry and molecular biology of insects have nowadays become important fields of research where proteic, hormonal, and lipoproteic systems show characteristics similar to mammals so that these systems are considered as biochemical models ¹⁹.

Most information found in the literature deals with very few species of insects, particularly **M. sexta, L. migratoria,** and **Bombyx mory** ⁸. Comparison of the data obtained is very difficult due to the differences in the physiological state, feed conditions and/or fast, sex or methodology employed ^{18,32}. Another important factor is the probable infection by parasites which has been proved to produce modifications in the hemolymphatic components ^{2,4,11} as well as to influence their development and life cycle ²⁷.

We have studied insects from two stages in three genera of vectors of Chagas' disease where some variables were established in order to be able to make comparisons amongst the different stages and genera. This type of study contributes to a better knowledge of the aspects related with the generation of energy, specially the necessary energy for flying. Although the members of the subfamily Reduviidae are not considered "good flyers", recent reports by **LEHANE & SCHOFIELD** (1982)

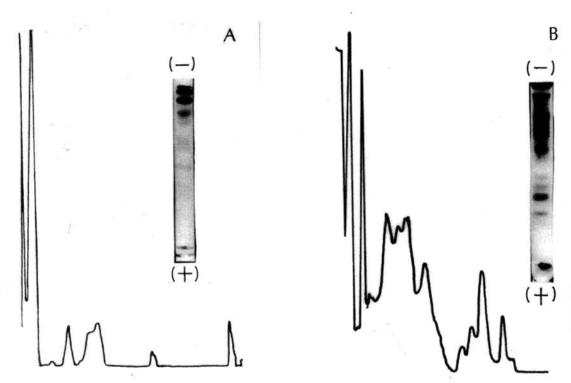


Figure 2: Protein patterns obtained by gel electrophoresis of hemolymph from T.cruzi vectors and scan densitometer. A: - Panstrongylus megistus (NV); B: Dipetalogaster maximus.(σ)

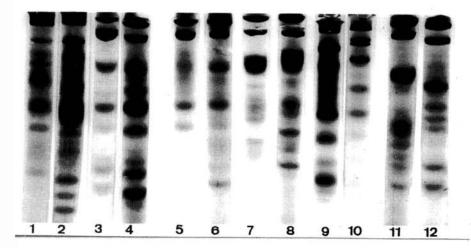


Figure 3: Proteins patterns obtained by gel electrophoresis of hemolymph from Triatoma genus. T. infestans: 1-(NV), 2-(\(\sigma\); T. rubrovaria: 3-(NV),4-(\sigma\); T. guasayana:5-(NV),6-(\(\sigma\); T. mazzotti:7-(NV),8(\(\sigma\); T. patagonica:9-(NV),10-(\(\sigma\), T. pallidipennis:11-(NV),12-(\(\sigma\)).

point to the possibility that they can fly several kilometers. (Results obtained from experiments performed in the field ²¹). In agreement with these reports the presence of structural components of the nervous system - photoreceptors-responsible for the stabilization of flight in **T.infestans** has also been reported ¹⁶.

The analysis of the total proteins did not give significant differences among the genera (Table 1)

and the results corresponding to the **T. infestans** species agree with those informed for NV²⁴, whereas the differences found with that reported by **PERASSI** (1972) could be attributed to the longer time elapsed from feeding till the hemolymph was obtained ²⁵.

In the present report, the densitometric evaluation of the obtained proteic profiles allowed to establish qualitative differences among the species and the stages from Dipetalogaster and Panstrongylus genera (Fig. 2 A,B) and for the members of the Triatoma genus (Fig.3).

The values of total lipids and carbohydrates were not significantly different for the three genera in their two stages (Table 1). The highest value of total lipids corresponded to **T. infestans** in agreement with that reported for the adults stage of this species ³⁰, whereas the values of carbohydrates were in general lower than those reported for phitophague insects ³³.

The hemolymphatic lipoproteins have been informed in a variable number between four and seven depending on the insect studied ³².

For the members of the subfamily Reduviidae here reported were shown two to four bands in polyacrylamide; the NV being the stage which showed a higher number. One of these bands was broad which may be indicating a high molecular heterogeneity. All fractions showed mobility between HDL and LDL human lipoproteins (Fig.1). This broad band, present in both stages and in three genera, corresponds to the HDLp fraction obtained by ultracentrifugation whose fractionation on Sepharose 6B showed a bimodal distribution 5. The additional components shown in the NV stages may correspond to the VHDLp (Very High Density Lipophorin) specific of each stage 19.

The general absence of marked differences in the data from the different analyses studied is not surprising, since the insects studied belong to the same subfamily where several parameters were standardized.

It should be noted that suitable knowledge about the biochemical potentialities of the generation of the energy for flying and demonstration of this ability in several species of triatomines ²⁰, could be relevant in order to design new control programmes and to analyse the aspects related to the transmission of the Chagas' disease.

RESUMO

Componentes hemolinfáticos em vetores do Trypanosoma cruzi: Estudo em doze espécies da subfamília Triatominae (Hemiptera : Reduviidae).

A subfamília Triatominae (Hemíptera:

Reduviidae) abrange um grande número de espécies de importância médica na transmissão de parasitoses provocadas pelo **T. cruzi** (Tripanosomiase americana).

Com a finalidade de contribuir ao conhe-cimento da composição química em proteínas, lípides, lipoproteínas e carboidratos da hemolinfa de vetores da doença de Chagas, se apresentam os achados correspondentes a 12 espécies de 3 gêneros, membros da subfamília Triatominae. O estudo foi efetuado com ninfas do 5° estádio e adultos machos das espécies: T. delpontei, T. dimidiata, T. guasayana, T. infestans, T. mazzotti, T. pallidipennis, T. patagonica, T. platensis, T. rubrovaria e T. sordida, o gênero Triatoma, e D. maximus e P. megistus, dos gêneros Dipetalogaster e Panstrongylus, respectivamente.

Os achados mostraram diferenças qualitativas na composição proteica, sendo semelhantes os perfis lipoproteicos. Os lípides, proteínas e carboidratos totais não apresentam diferenças significativas entre espécies e/ou gêneros, e/ou estádios.

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REFERENCES

- ADAMS, R. & RYCKMAN, R.A. A comparative electrophoresis of the Triatoma rubida complex (Hemiptera: Reduviidae: Triatominae). J.med.Ent., 6: 1-7, 1969.
- BARRAS, D.J.; KISNER, R.T.; LEWIS, W.J. & JONES, R.T. -Effects of the parasitoid, Microplitis croceipes on the hemolymph proteins of the corn earworm, Heliothis zea. Comp. Biochem. Physiol., 43B: 941-947, 1972.
- BRADFORD, M.M. A rapid sensitive method for the quantification of microgram quantities of protein utilizing the principles of protein dye binding. Ann. Biochem., 72: 248-253, 1976.
- CANAVOSO, L.E.; RUBIOLO, E.R. & PIAZZA, L.A. Modificaciones del perfil proteico en la hemolinfa de
 Triatoma infestans, infectados por cepas de
 Trypanosoma cruzi y Blastocrithida triatomae
 (Trypanosomatidae). In: JORNADAS CIENTIFICAS
 DE LA SOC. BIOLOGIA DE CORDOBA, 6., Córdoba,
 1990. p.36.
- CANAVOSO, L.E. & RUBIOLO, E.R. Lipophorin (HDLp) in vectors of Trypanosoma cruzi. Mem. Inst. Oswaldo Cruz, 86 (suppl.1): 244, 1991.

- CARNERO, R.A. & RUBIOLO, E.R. Presencia de lipoproteínas en orina de diabéticos con nefropatías. Acta Bioquim. clin. lat. amer.., 23: 11-216, 1989.
- CERISOLA, J.A.; DEL PRADO, C.E.; ROHWEDDER, R. & BOZZINI, J.P. - Blastocrithidia triatomae nsp found in Triatoma infestans from Argentina. J. Protozool., 18: 503-506, 1971.
- CHAPMAN, M.J. Animal lipoproteins: chemistry, structure and comparative aspects. J.Lipid Res., 21: 789-853, 1980.
- DAVIS, B.J. Disk, electrophoresis. II. Method and aplication to human serum proteins. Ann. N.Y. Acad. Sci., 121: 404, 1964.
- EDELSTEIN, C. & SCANU, A.M. Precautionary measures for collecting blood destined for lipoprotein isolation. Meth. Enzimol., 128: 151-155, 1986.
- FERKOVICH, S.M.; GREANY, P.D. & DILLARD, C.-Changes in haemolymph proteins of the fall Armyworm, Spodoptera frugiperda (J.E. Smith), associated with parasitism by the braconid parasitoid Cotesia marginiventris (Cresson). J.Insect Physiol., 29: 933-942, 1983.
- FRINGS, C.S. & DUNN, R.T. A colorimetric method for determination of total serum lipids based on the sulpho-phospho-vanillin reaction. Amer.J.Path., 3: 89-91, 1970.
- GONDIM, K.C.; OLIVEIRA, P.L.; COELHO, H.S.L. & MASUDA, H. - Lipophorin from Rhodnius prolixus: purification and partial characterization. Insect. Biochem., 19: 153-161, 1989.
- HAYAKAWA, Y. Inhibition of lipid transpor in insects by a parasitic factor. Comp. Biochem. Physiol., 87B: 279-283, 1987.
- HORWOOD, M.A. & HALES, D.F. Fat body changes in locust, Chortoicetes terminifera (Walker) (Orthoptera: Acrididae); parasitized by Nementrinid fly. Arch.Insect. Physiol., 17: 53-63, 1991.
- INSAUSTI, T.C. Ocellar pathways in Triatoma infestans. Mem. Inst. Oswaldo Cruz, 86 (suppl 1): 244, 1991.
- KIMURA, M. & OTHA, T. Proteins polimorfism as a phase of molecular evolution. Nature, 229: 4167-4169, 1971.
- KRAMER, S. J. & DE KORT, C.A.D. Juvenile hormone carrier lipoproteins in the hemolymph of the colorado potato beetle, Leptinotarsa decembineata. Insect. Biochem., 8: 87-92, 1978.
- LAW, J.H. & WELLS, M.A. Insects as biochemical model. J. biol. Chem., 264: 16335-16338, 1989.
- 20. LEHANE, M.J. & SCHOFIELD, C.J. Preliminary report

- on flight by some triatomine bugs. Trans.roy.Soc.trop.Med.Hyg., 70: 526, 1976.
- LEHANE, M.J. & SCHOFIELD, C.J. Flight initiation in Triatoma infestans (Klug) (Hemiptera:Reduviidae). Bull. ent. Res., 72: 497-510, 1982.
- LOUGHTON,B. & WEST, A. The development and distribution of hemolymph protein in Lepidoptera. J.Insect Physiol., 11: 911-932, 1965.
- MISSELINNJ, G.; KARCHER, D.; DE KEYSER, F. & VAN SANDE, M. Studies on hemolymph proteins of insects of medical importance by means of microelectrophoresis in agar gel. In: Protides of the biological fluids. Proceeding of the 7th Colloquium Bruges, 1959. Peeters Ed. Amsterdam, Elsevier Publishing, 1960. p.127-135.
- PAZ,D.; GIMENEZ, H. & TOMATIS, M.E. Hemolinfa de Triatoma infestans: cambios en volumen y proteinas totales despues de una ingesta. Chagas, 5: 23-28, 1988.
- PERASSI, R. Protein components of the haemolymph of triatomid bugs (Hemiptera:Reduviidae). Bol. chil. Parasit., 27: 74-80, 1972.
- RYAN, R.O. -Dynamics of insect lipophorin metabolism. J. Lipid Res., 31: 1725-1739, 1990.
- 27. SCHAUB, J.A. Development time and mortality in larvae of the reduviid bugs Triatoma infestans and Rhodnius prolixus after coprophagic infection with Blastocrithidia triatomae (Trypanosomatidae). J.invertebr.Path.,51: 23-31, 1988.
- SCOTT,T.A. & MELVIN, E.H. Determination of dextran with anthrone. Analyt.Chem.,25: 1656-1661, 1953.
- SNEDECOR, G.W. & COCHRAN, W.G. Statistical methods. Iowa, The Iowa State University Press, 1971.
- TIERNO, M.A. & BRENNER, R.R. Bioquímica del ciclo evolutivo del Triatoma infestans (Vinchuca).I. Composición de los lípidos y ácidos grasos. Acta physiol. lat. amer., 28: 41-52, 1978.
- VAN SANDE, M. & KARCHER, D. Species differentiation of insects by hemolymph electrophoresis. Science,131: 1103-1104, 1959.
- WITMORE, E. & GILBERT, L.I. Hemolymph proteins and lipoproteins in Lepidoptera: a comparative eletrophoretic study. Comp.Biochem.Physiol., 47B: 63-78, 1974.
- WIGGELESWORTH, V.B. The principles of insect physiology. London, Chapman and Hall, 1970. Chap.10, p. 426.
- WYATT,G.R. & PAN, M.L. Insect plasma proteins. Ann.Rev.Biochem., 47: 779-817, 1978.

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