

Anais da Academia Brasileira de Ciências (2018) 90(4): 3853-3869 (Annals of the Brazilian Academy of Sciences)

Printed version ISSN 0001-3765 / Online version ISSN 1678-2690 http://dx.doi.org/10.1590/0001-3765201820180278 www.scielo.br/aabc | www.fb.com/aabcjournal

Molecular marker assisted selection for increasing inbreeding in S, populations of cassava

PAULO HENRIQUE G.A. DE OLIVEIRA¹, ANA CLAUDIA O. BARBOSA¹, RAFAEL P. DINIZ¹, EDER J. DE OLIVEIRA² and CLAUDIA F. FERREIRA²

¹Universidade Federal do Recôncavo da Bahia, Centro de Ciências Agrárias, Ambientais e Biológicas, Rua Rui Barbosa, 710, 44380-000 Cruz das Almas, BA, Brazil ²Embrapa Mandioca e Fruticultura, Rua da Embrapa, s/n, Caixa Postal 007, 44380-000 Cruz das Almas, BA, Brazil

Manuscript received on March 15, 2018; accepted for publication on July 2, 2018

ABSTRACT

The objective of the present work was to use MAS in self-pollinated cassava populations for obtaining individuals with high inbreeding (f) aimed at rapid development of partial inbred lines. Three progenies (F0222, F1378 and F1662) were self-pollinated, generating a total of 233 S₁ individuals. The progenies and the S₁ individuals were evaluated in the seedlings and clonal evaluation trials (CETs). In the CET, plants were evaluated for the following traits: plant height (PH), root dry matter content (DMC), above ground yield (AGY), root (RY) and starch yield (SY). Twenty-seven microsatellites and five minisatellites were used to determine the level of inbreeding of the S₁ individuals. Inbreeding (f) values varied from 0.15 to 0.89 within progenies, whereas most of the S₁ individuals presented f values above 0.50. In average, 25% of the S₁ individuals were selected, which resulted in a 38% inbreeding increase in the cassava progenies. In contrast, phenotypic selection showed no differences in inbreeding increase. Furthermore, there was no correlation between the level of inbreeding and agronomic traits. MAS was efficient for the identification and selection of cassava S₁ individuals, with higher inbreeding values contributing to the decrease in the breeding cycles necessary to obtain new cassava inbred lines.

Key words: partial inbred lines, Manihot esculenta Crantz, microsatellites, self-pollination.

INTRODUCTION

Cassava is one of the main sources of carbohydrates for millions of people, mainly in Africa, Asia and Latin America, being the second most important source of starch in the world, just behind maize (Ceballos et al. 2015). The production of cassava is also strongly linked to family farming in developing countries (Halsey et al. 2008) and has great industrial potential (Oliveira et al. 2012a)

Correspondence to: Eder Jorge de Oliveira E-mail: eder.oliveira@embrapa.br due to the diverse applications in pharmaceuticals, beauty products, food, and the textile industry as well as in paper production and biofuels, among others (Saengchan et al. 2015).

Cassava cultivars for industrial purposes should contain specific characteristics, especially high root yield and dry matter content, as well as resistance to diseases (Ceballos et al. 2012, 2016, Oliveira et al. 2015). However, even though there is great market demand, few improved genotypes are available that meet the growing demands for varieties with high agronomic performance. This

possibly occurs due to the relatively short period of the cassava domestication for use as large-scale commercial crops as well as to the most diverse environmental constraints to which the crop is submitted, which means that there is a selection of varieties adapted to specific environments or traits. The task of combining different characteristics into a single variety is still overwhelming for cassava breeders. However, cassava breeding programs are relatively recent (beginning in the 1960's), and although there are reports of root yield gains, there is a constant demand for the development of new and highly productive varieties associated with different starch characteristics.

In breeding programs, an alternative for the development of cultivars with higher yield is heterosis (hybrid vigor) obtained through crossings of inbred lines (Prado et al. 2013). This strategy has been successfully used for many years in maize (Li et al. 2014) and has recently been used in autogamous species (Goff and Zhang 2013) and species that have undergone recent genetic improvement such as papaya (Vale et al. 2016). Even in completely different species, the results show a significant yield increase of hybrids obtained in these crops. In contrast, for cassavas, the exploration of heterosis has not been carried out routinely, mainly due to the fact that there is little investment in the development of inbred lines (Ceballos et al. 2012).

One of the main breeding methods used for selecting segregating populations derived from crosses between highly heterozygous elite genotypes is the phenotypic recurrent selection, which allows researchers to explore heterosis effectively (Ceballos et al. 2004, 2012), whereas the exploitation of the effects of heterosis (nonadditive effects) without the use of inbred lines belonging to different heterotic groups can be quite slow (Ceballos et al. 2015). Therefore, efforts should be made to explore the effects of dominance that are prevalent in several

productive traits in cassava, such as root yield and harvest index (Wolfe et al. 2016).

Self-pollination has already been used as a strategy to obtain segregating populations with the goal of developing cassava partial inbred lines (Freitas et al. 2016, Rojas et al. 2009). However, factors such as high crop heterozygosity, which results in progenies with a high segregation rate, difficulties in flowering and a long crop cycle, also make it difficult to obtain endogamic materials (Ceballos et al. 2017, Halsey et al. 2008). According to Oliveira et al. (2010), conventional breeding methods based on phenotypic selection in segregating populations to obtain inbred lines are costly, time consuming, space intensive and highly influenced by environmental conditions.

Another approach for the development of inbred lines would be the induction of haploids and subsequent chromosomal duplications through double haploid technology. However, the protocols for obtaining cassava haploids are limited and inefficient due to the development of a large number of abnormal individuals (Perera et al. 2014). Therefore, a feasible strategy to improve the efficiency of the selection process of homozygous individuals would be the use of integrated biotechnology tools in conventional breeding methods, such as molecular markers. Among the several molecular markers available, microsatellites (SSR) are especially important because they are codominant, multiallelic, highly polymorphic, and highly reproducible (Oliveira et al. 2006), which makes them suitable for inferences about kinship and inbreeding. SSRs have been the most used molecular markers in cassava, especially for genetic diversity analysis (Fregene et al. 2003), paternity tests (Mohan et al. 2013) and molecular marker assisted selection (MAS) (Lokko et al. 2005).

MAS has been used in plant breeding with the aim of selecting genotypes with desired characteristics directly at the DNA level (Idrees and Irshad 2014). This technique has several benefits when associated with conventional breeding methods, such as low cost, rapid generation advancement and high selection efficiency (Xu and Crouch 2008). The great advantage of MAS in comparison with conventional breeding methods is the reduction of the time to obtain the desired genotype, since it allows the selection of individuals in the early stages of development as well as the early elimination of undesirable genotypes (Morris et al. 2003). This technique has been used for several purposes, such as assisted backcrossing, quantitative trait loci (QTL) mapping, heterosis studies (Collard and Mackill 2008, Kobayashi et al. 2013) and for identifying individuals with a higher homozygosity level in segregating populations (Oliveira et al. 2010, 2012b). In cassava, the most successful application of MAS has been in the selection for resistance to cassava mosaic virus (CMD) (Okogbenin et al. 2007, Carmo et al. 2015).

The use of MAS to obtain inbred lines in segregating populations is aimed at quantifying the inbreeding level of the individuals within progenies and to select the most homozygous for new self-pollination cycles (Oliveira et al. 2010). However, there are no reports of the use of MAS to obtain inbred lines in cassava. Therefore, this is an innovative study with the goal of identifying individuals with a higher level of inbreeding in cassava segregant populations via MAS, as well as its association with agronomic traits, in order to reduce the number of self-pollination generations for obtaining partial inbred lines.

MATERIALS AND METHODS

OBTAINMENT OF SELF-POLLINATED PROGENIES

In order to obtain the S₁ segregant populations, three cassava accessions (BGM0222 = "Vermelhinha Branca", BGM1378 = "Macaxeira Branca" and BGM1662 = "IAC-14"), belonging to the Cassava Germplasm Bank (CGB) at Embrapa Mandioca e Fruticultura (Cruz das Almas, Bahia, Brazil),

were selected as parents (S₀). These accessions were selected based on their flowering rate for performing crosses and high dry matter content in the roots. For each accession, the self-pollinations were performed manually in the experimental field at Embrapa Mandioca e Fruticultura in 2014. After ripening, the fruits were collected and stored in paper bags at 8 °C. The S₁ seeds of each progeny were placed to germinate in plastic tubes containing commercial substrate, coconut fiber and vermiculite in the proportion of 2:1:1, being kept under greenhouse conditions (75% humidity, 29 $^{\circ}$ C \pm 3 $^{\circ}$ C). At 50 days after planting (\pm 30 cm in height), the seedlings were transplanted to the field without any experimental design, in a step called seedlings evaluation trials (SET).

In SET, the number of S₁ individuals for progenies F1662, F0222 and F1378 was 82, 91 and 101, respectively. The experiment was set up in 2014 at the experimental area of the Federal University of Recôncavo da Bahia (UFRB), in Cruz das Almas (Bahia, Brazil), located at 12°66′17"S latitude, 39°08′28"W longitude, and 225 m of altitude. At 12 months after planting, the individuals were selected and collected individually for the establishment of the clonal evaluation trial (CET). The only criterion used to select the individuals to compose CET was the plants' ability to produce at least five stakes of 20 cm in length.

The CET was set up at an experimental area of a private partner of Embrapa (Bahiamido) in Santo Antônio de Jesus (Bahia, Brazil), located at 13°10'56"S latitude and 39°25'30"W longitude, and 203 m of altitude. A total of 233 individuals were selected from the CET, being 76, 78 and 79 from progenies F1662, F0222 and F1378, respectively. The S₁ individuals, along with the S₀ parents, were evaluated in an augmented block design, with 233 individuals as regular treatments, distributed in four blocks and with plots of five plants. The parents (BGM0222, BGM1378, and BGM1662) and 15 clones/varieties (9783-13,

98150-06, BRS Caipira, BRS Dourada, BRS Formosa, BRS Gema de Ovo, BRS Kiriris, BRS Poti Branca, BRS Verdinha, IAC90, Cigana Preta, Correntão, Corrente, Eucalipto, and Vassoura Preta), were used as common treatments (controls). The spacing used was 0.8 m between plants and 0.9 m between rows.

DNA EXTRACTION

Young leaf tissue samples were collected from the upper third part of the plants of the three S_0 parents and S_1 individuals (274 S_1 genotypes at the SET). The leaf samples were identified and stored in an ultra-freezer at -80 °C. The genomic DNA was extracted using the CTAB method described by Doyle and Doyle (1990) with some modifications. DNA quality and quantity evaluation was performed on 0.8% agarose gels, after staining with ethidium bromide. DNA quantification was estimated by comparing the band intensity with aliquots of standard amounts of the known concentration (Lambda-Sigma DNA). The samples were diluted in Tris-EDTA (TE) and standardized to 10 ng μ L⁻¹.

AMPLIFICATION OF MICRO- AND MINISATELLITE MARKERS

PCR reactions were performed in a final volume of 15 μl containing 30 ng of genomic DNA, 0.2 mM of each primer (IDT, Integrated DNA Technologies Inc, CA, USA), 1 U of Taq DNA Polymerase (Pluthero 1993), 2 mM MgCl₂, 0.2 mM of dNTP and 1X of Tris KCl (Promega Corporation, WI, USA). Amplifications were performed on the Veritti® 96-well thermal cycler (Applied Biosystems, CA, USA) under the following conditions: denaturation at 94 °C for 5 minutes, followed by 29 cycles of denaturation at 94 °C per minute, annealing temperature (AT) specific for each primer for 60s at 72 °C per minute (Table I) and a final extension of 7 minutes at 72 °C. Electrophoresis of the amplified fragments was performed on 3% agarose gel (Invitrogen, Carlsbad, CA, USA) at 120 v for 4 hours, stained with ethidium bromide (15 mg.mL⁻¹

solution) in 0.5X Tris/Borate/EDTA buffer. The fragments were visualized under UV light and photographed with the Gel Logic 212 Pro photo documentation system (Carestream Molecular Imaging, New Haven, USA). Afterwards, the fragments were analyzed in comparison to a known base pair size marker (DNA ladder 100pb) (Fermentas, Glen Burnie, MD, USA) for the determination of the fragment size.

MOLECULAR MARKER POLYMORPHISM

Initially, 55 microsatellite primers (SSRY and EME series) and six minisatellites were tested for inbreeding analysis in S₁ cassava individuals. Microsatellites and minisatellites were previously amplified in the F0222, F1378 and F1662 parents. Primers that presented heterozygosity in the parents, with good amplification pattern and absence of nonspecific bands, were selected to be amplified in all S, individuals. After an initial evaluation, a total of 27 microsatellite loci (SSRY and EME) and five minisatellites, were used to check the polymorphism in each progeny. Sixteen microsatellites were polymorphic for progeny F1662, and 18 microsatellites were polymorphic for progenies F0222 and F1378 (Table I). Five minisatellites were used, with 4, 3 and 2 being polymorphic for progenies F022, F1378 and F1662, respectively.

AGRONOMIC EVALUATIONS

Twelve months after planting, the CET was evaluated for agronomic traits using five plants from each plot. The main traits evaluated were: plant height (PH – meter); above ground yield (AGY – t.ha⁻¹); root yield (RY – t.ha⁻¹); starch yield (SY – t.ha⁻¹); and dry matter content (DMC – %, measured using the root's specific gravity, according to Kawano et al. (1987)).

DATA ANALYSIS

Molecular data analyses were performed using the Powermarker 3.25 software (Liu and Muse

TABLE I Microsatellite (SSRY and EME series) and minisatellites markers (VNTR) were used to evaluate the inbreeding coefficient in S_1 cassava individuals in self-pollination progenies from BGM0222 (F0222), BGM1378 (F1378) and BGM1662 (F1662) cassava accessions.

	D . C	г. 1	n.	bp	AT (°C)	Markers per S ₁ progeny		
Markers	Reference	Forward	Reverse			F0222	F1378	F1662
SSRY101	Mba et al. (2001)	ggagaataccaccgacagga	acagcagcaatcaccattte	213	55		X	
SSRY103	Mba et al. (2001)	tgagaaggaaactgcttgcac	cagcaagaccatcaccagttt	272	55		X	X
SSRY105	Mba et al. (2001)	caaacatctgcacttttggc	tegagtggettetggtette	225	55	X		
SSRY106	Mba et al. (2001)	ggaaactgettgeacaaaga	cagcaagaccatcaccagttt	270	55		X	X
SSRY13	Mba et al. (2001)	gcaagaattccaccaggaag	caatgatggtaagatggtgcag	234	55	X		
SSRY143	Mba et al. (2001)	geteatgaactgageettea	agcagatccaaatcactgaaa	153	55	X		
SSRY165	Mba et al. (2001)	aaatgagttgcaaaggccaa	ggtaaacaaatgatgtggtgttc	243	55			X
SSRY168	Mba et al. (2001)	acagecacacttgtteteea	ctgcaatctccaacagcaac	277	45	X	X	
SSRY170	Mba et al. (2001)	tctcgatttggtttggttca	tcatccttgttgcagcgtta	299	55	X		X
SSRY175	Mba et al. (2001)	tgactagcagacacggtttca	gctaacagtccaataacgataagg	136	55		X	
SSRY179	Mba et al. (2001)	caggctcaggtgaagtaaagg	gcgaaagtaagtctacaacttttctaa	226	55	X	X	X
SSRY182	Mba et al. (2001)	ggaattctttgcttatgatgcc	ttcctttacaattctggacgc	253	55		X	
SSRY28	Mba et al. (2001)	ttgacatgagtgatattttcttgag	gctgcgtgcaaaactaaaat	180	55		X	
SSRY30	Mba et al. (2001)	ccatccactagaaactttaaaagca	caactcagcggagctttttc	220	55		X	
SSRY49	Mba et al. (2001)	tgaaaatctcactggcattattt	tgcaaccatagtgccaagc	300	55	X	X	X
SSRY68	Mba et al. (2001)	gctgcagaatttgaaagatgg	cagctggaggaccaaaaatg	287	55		X	X
SSRY8	Mba et al. (2001)	agtggtttgagaagactggtga	tttccaaaatggaacttcaaa	288	45	X		X
SSRY81	Mba et al. (2001)	ggcgatttcatgtcatgctt	tgattttctgcgtgatgagc	204	55	X		
SSRY82	Mba et al. (2001)	tgtgacaattttcagatagcttca	caccatcggcattaaactttg	211	55			X
SSRY83	Mba et al. (2001)	tggctagatggtgattattgctt	tgettactetttgatteeaeg	239	55	X		
SSRY93	Mba et al. (2001)	tttgttgctcacatgaaaacg	cagatttcttgtggtgcgtg	289	55			X
SSRY94	Mba et al. (2001)	aggatggacttggagatgga	ggtggaagtaaggctgttagtg	268	55		X	X

Markers	Reference	Forward	Davious	hn	AT (°C)	Markers per S ₁ progeny		
Markers	Reference	Forward	Reverse	bp		F0222	F1378	F1662
EME189	Kunkeaw et al. (2011)	cagagcacatccagaaattgtt	gaaatagatcaagtgccccatc	180	60	X	X	
EME205	Kunkeaw et al. (2011)	ccagagcgtataactggaac	tgcaggagtgtggatatggtt	250	55	X		X
EME260	Kunkeaw et al. (2011)	gttggagttgtagttgctgc	catgggctgtgaaaatgaact	160	58	X	X	X
EME395	Kunkeaw et al. (2011)	tcaaaggtatcggggaggtag	gtttaccccactaacatcgcat	200	58	X	X	
EME425	Kunkeaw et al. (2011)	cctccacaacettatcaatca	eggtagecatagecataaca	140	55			X
VNTR4	C.D. Carmo, Unpublished	aatcatatcaggggctggtg	cgagggaaatgctgacctt	220	58	X	X	
VNTR5	C.D. Carmo, Unpublished	ttgettecaatetteteaca	gatcaaaacgggcttgaaat	233	58		X	X
VNTR71	C.D. Carmo, Unpublished	tgcagataaactccaaaagtaagaa	getteatggttgaggetett	589	60	X	X	X
VNTR72	C.D. Carmo, Unpublished	gcttaggcgggaagaaaatg	tgttcaactgcctctctttgc	828	58	X		
VNTR90	C.D. Carmo, Unpublished	gaaccgtgaacagtaaccgata	cccggctgtttcaataaaat	835	60	X		

bp: size in base pairs; AT: annealing temperature; X: Primer selected for inbreeding analysis in each progeny.

2005) to determine the expected heterozygosity (He), observed heterozygosity (Ho) and inbreeding coefficient (f) in all S₁ individuals and the three S₀ parents. The inbreeding coefficient was determined following the method proposed by Wright et al. (1965) in which .

Population structure within selfings was investigated using the Euclidean distance matrix, which shows the extent of diversity between the individuals. Then, the principal coordinate (PCO) analysis was performed using the R function dudi.pco (R Core Team 2017). The PCO analysis indicated no visible population structure (Figure 1), and therefore it was not necessary to calculate the kinship matrix as a variance-covariance matrix to correct for random genotypic effects.

The analysis of the phenotypic data was carried out using the linear mixed model, y = Xf + Zg + Wb + e with the following model, whereas: y – is the phenotypic data; f – is the vector of means of the individuals (fixed effects); g – is the vector

of the genotypic effects of the individuals (random effects); b – is the vector of the environmental block effects (random effects); and e – is the vector of residue (random effect). The incidence matrices of the random effects are represented by uppercase letters.

Pearson's correlation was estimated using the *corrgram* package of the R software (R Core Team 2017), based on the best linear unbiased prediction (BLUP) and inbreeding coefficient values of the S_1 individual. The selection of the S_1 individuals based on BLUP analysis of the phenotypic traits was performed using the following selection index: SI = (PH*5) + (AGY*5) + (RY*10) + (SY*10) + (DMC*10).

RESULTS

GENETIC PARAMETERS AND INBREEDING ANALYSIS OF S, INDIVIDUALS

The distribution of inbreeding coefficients (f) was very similar among the three progenies. The highest proportion of S_1 individuals presented f between

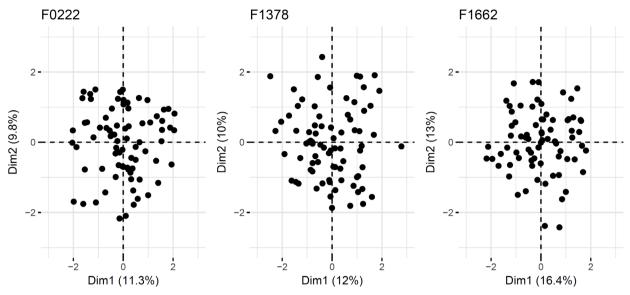


Figure 1 - Principle coordinate analysis for 78, 79 and 76 S₁ progenies derived from parent BGM0222 (F0222), BGM1378 (F1378), and BGM1662 (F1662), respectively, based on mini and microsatellites markers.

0.40 and 0.60, representing 50%, 56% and 41% of the total individuals from progenies F0222, F1378 and F1662, respectively (Figure 2). The distribution of S_1 individuals with f ranging from 0.60 to 0.79 (high inbreeding coefficient) was 24% (F0222), 20% (F1378) and 27% (F1662). In contrast, only 1.25% of the individuals from progenies F0222 and F1378, presented f estimates above 0.80.

The observed heterozygosity (Ho) of the $\rm S_1$ individuals presented similar distribution to the inbreeding coefficient for the three progenies, possibly because it is an important component on the f formula. Approximately, 50%, 45% and 41% of the individuals in progenies F0222, F1378 and F1662, respectively, presented individuals with Ho between 0.40 and 0.60 (Figure 3). The distribution of individuals with high Ho (> 0.60) ranged from 24% to 32% in progenies F0222 and F1378, respectively. In contrast, the lowest Ho values were 0.17, 0.11 and 0.25 in $\rm S_1$ individuals of progenies F0222, F1378 and F1662, respectively (Table II).

Regarding the He and Ho analyses, there was very small variation in He values in all progenies (ranging from 0.46 to 0.50 and mean of 0.49), while Ho variation was larger (ranging from 0.35)

to 0.65) (Table III). The lowest Ho values were observed for markers SSRY83 (F0222), VNTR71 (F1378) and SSRY170 (F1662) (0.35, 0.37 and 0.38, respectively), while the loci with the highest Ho were SSR08 (0.61) in progeny F0222; as well as SSRY103, SSRY106, SSR168, SSRY175, SSRY28, SSRY49 and VNTR05 (0.60 to 0.65) in progeny F1378 and SSRY49 (0.58) in progeny F1662. Although the F1378 progeny had the highest number of loci with Ho above 0.60, the Ho mean was similar among the progenies, i.e., 0.51 (F0222), 0.51 (F1378) and 0.52 (F1662).

Considering the chromosomal location of the mini- and microsatellite markers, in general, the distribution of the inbreeding coefficient and Ho were more similar in F1662 (f and Ho ranging from 0.40 to 0.62 and 0.38 to 0.60, respectively), in comparison with the other progenies (Figure 4). In contrast, the parameters' variation in F0222 (f = 0.39 to 0.65 and Ho = 0.35 to 0.61, respectively) and F1378 (f = 0.36 to 0.63 and Ho = 0.37 to 0.64, respectively) progenies was not high enough to raise the hypothesis of unbalanced homozygosity in different chromosomes. Although a small number of molecular markers was used, results indicate that

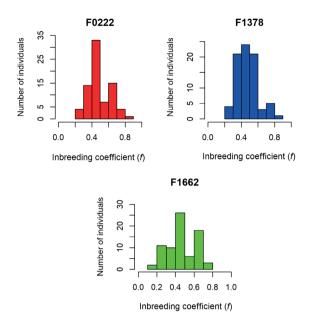


Figure 2 - Distribution of the inbreeding coefficient (f) of the S₁ cassava individuals from the self-pollination of the accession BGM0222 (F0222), BGM1378 (F1378) and BGM1662 (F1662).

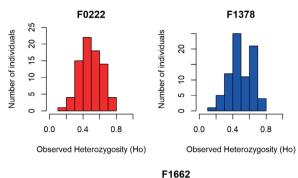
TABLE II Minimum, maximum and average values of observed heterozygosity (Ho) and inbreeding coefficient (f) in S_1 individuals from different self-pollinated progenies.

Progeny	Parameter	Minimum	Maximum	Average		
F0222	Но	0.17	0.78	0.51		
	f	0.22	0.83	0.49		
F1378	Но	0.11	0.72	0.51		
	f	0.25	0.89	0.49		
F1662	Но	0.25	0.81	0.52		
	f	0.15	0.74	0.49		

the genotyping with the mini- and microsatellites was well distributed in the different cassava chromosomes (11 of the 18 chromosomes of *M. esculenta*), which contributes to a better coverage of the level of homozygosity in this species.

ANALYSIS OF INBREEDING AND AGRONOMIC PERFORMANCE BASED ON DIFFERENT SELECTION CRITERIA

Considering the genotypic information of the molecular markers, S_1 individuals with the highest level of inbreeding coefficient ($f \ge 0.60$) within



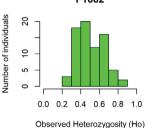


Figure 3 - Distribution of the observed heterozygosity (Ho) of the S_1 cassava individuals from the self-pollination of the accessions BGM0222 (F0222), BGM1378 (F1378) and BGM1662 (F1662).

progeny were identified and selected for the next self-pollination cycles. The selection intensity applied was 25%, 21% and 27% in progenies F0222, F1378 and F1662, thus making it possible to select 20, 17 and 21 S₁ individuals, respectively, with higher endogamy.

The comparison between the means of the agronomic traits (plant height, above ground yield, roots and starch yield and root dry matter content) and the S, individuals selected based on the inbreeding coefficient ($f \ge 0.60$), showed that there was no significant difference in the means for most of these traits, except for above ground yield (AGY) in progeny F1378, in which the S, individuals selected for presenting high f also showed high means (Figure 5). Therefore, it is possible to select S₁ individuals in cassava progenies with high inbreeding level without significant losses in AGY. In contrast, as for all selective processes, the selection of the S₁ individuals with the highest inbreeding coefficient resulted in a reduction in phenotypic variation for plant height (F1378 and

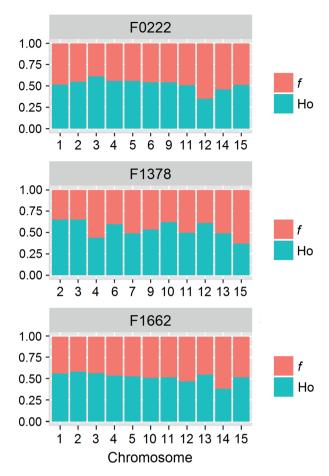


Figure 4 - Distribution of observed heterozygosity (Ho) and inbreeding coefficient (*f*) of the S₁ cassava individuals from the self-pollination of accession BGM0222 (F0222), BGM1378 (F1378) and BGM1662 (F1662), considering the chromosomal location of the molecular markers used for genotyping.

F1662), above ground yield (F0222 and F1662) and root and starch yield (F1662) (Figure 5).

If the selection of S₁ individuals was performed based on the selection index (agronomic performance) rather than the inbreeding coefficient, as is traditionally done in breeding programs, an increase in the trait's means would be expected, especially for dry matter content (F1378 and F1662), above ground yield and root yield (F1378) and starch yield (F0222 and F1378) (Figure 6). In this case, the performance for most of the agronomic traits in the three progenies would be preserved, but there would be a reduction in the inbreeding coefficient in the F0222 and F1662 progenies,

which would certainly contribute to an increase in the number of self-pollinations necessary to obtain inbred lines.

RELATIONSHIP BETWEEN INBREEDING AND AGRONOMIC TRAITS

Considering the distribution of the inbreeding coefficient in three different groups, Group 1 (0.10 < f < 0.39), Group 2 (0.40 < f < 0.59) and Group 3 (> 0.60), there was great phenotypic difference between S, progenies within this inbreeding classification (Figure 7). There was no linear relationship between inbreeding and agronomic performance, since, in some cases, the average of Group 2 was lower (dry matter content in the roots, above ground yield and root yield - F1378) or higher (root dry matter content, root and above ground yield in the F1662 progeny and root and above ground yield in the progeny F0222) in comparison to the other groups. In addition, the variations in the agronomic traits were very similar between the different inbreeding groups, indicating a lack of association between the agronomic performance of the S₁ progenies and their inbreeding level, based on molecular analysis.

Except for the correlation between PH and DMC and between AGY and DMC that did not differ from zero, the other correlation estimates were positive and different from zero (Figure 8). There was a strong positive correlation between RY and SY (0.99). In contrast, there was a positive correlation of moderate magnitude between AGY and RY (0.53); AGY and SY (0.52); PH and AGY (0.48); PH and RY (0.30); PH and SY (0.30); DMC and RY (0.21); and DMC and SY (0.28).

DISCUSSION

MOLECULAR MARKERS USED TO SELECT PARTIAL INBRED LINES

The determination of the inbreeding coefficient by the Ho and He analysis of different molecular

TABLE III

Expected (He) and observed heterozygosity (Ho) of microsatellite loci in S₁ cassava individuals originated from self-pollination of accessions BGM0222 (F0222), BGM1378 (F1378) and BGM1662 (F1662).

	F022	2		F1378				F1662			
Primer	Но	Не	Cr	Primer	Но	Не	Cr	Primer	Но	Не	Cr
EME189	0.54	0.49	9	EME189	0.56	0.50	9	EME205	0.56	0.50	1
EME205	0.49	0.50	1	EME260	0.50	0.48	11	EME260	0.51	0.49	11
EME260	0.51	0.50	11	EME395	0.56	0.49	6	EME425	0.57	0.49	5
EME395	0.54	0.50	6	SSRY101	0.49	0.49	7	SSRY103	0.47	0.49	12
SSRY08	0.61	0.50	3	SSRY103	0.60	0.50	12	SSRY106	0.47	0.49	12
SSRY105	0.44	0.50	14	SSRY106	0.62	0.49	12	SSRY165	0.55	0.50	13
SSRY13	0.53	0.50	2	SSRY168	0.63	0.50	6	SSRY170	0.38	0.49	14
SSRY143	0.51	0.50	11	SSRY175	0.65	0.49	3	SSRY179	0.51	0.49	4
SSRY170	0.48	0.50	14	SSRY179	0.44	0.50	4	SSRY49	0.58	0.50	2
SSRY179	0.56	0.50	4	SSRY182	0.51	0.48	9	SSRY68	0.57	0.50	4
SSRY49	0.57	0.50	2	SSRY28	0.61	0.50	12	SSRY8	0.56	0.50	3
SSRY68	0.56	0.49	4	SSRY30	0.49	0.48	13	SSRY82	0.49	0.49	5
SSRY81	0.56	0.46	5	SSRY49	0.65	0.50	2	SSRY93	0.57	0.49	3
SSRY83	0.35	0.50	12	SSRY68	0.44	0.50	4	SSRY94	0.53	0.50	11
VNRT04	0.52	0.49	11	SSRY94	0.56	0.49	11	VNTR05	0.51	0.49	10
VNRT71	0.51	0.50	15	VNTR04	0.43	0.48	11	VNTR71	0.52	0.50	15
VNTR72	0.54	0.50	1	VNTR05	0.62	0.50	10				
VNTR90	0.49	0.50	11	VNTR71	0.37	0.47	15				
Average	0.51	0.49		Average	0.51	0.49		Average	0.52	0.49	

Cr: chromosomal region of the molecular markers.

markers is an efficient way of determining inbreeding, especially in self-pollination, full-and half-sibling populations (Goudet and Keller 2002). The inbreeding coefficient in the S₁ cassava progenies varied from 0.00 to 0.89 in the different self-pollinated populations. However, most S₁ individuals presented inbreeding values ranging from 0.40 to 0.60. On average, inbreeding was very close to 0.50 (0.48 for F2222, 0.49 for F1378 and F1662). These results are in agreement with the expected average reduction of 50% of heterosis and, therefore, the heterozygosity of the loci (Wu et al. 2016).

Considering only one self-pollination cycle, it was possible to identify S_1 individuals with inbreeding coefficients above the expected mean (f > 0.70). This result is of utmost importance to accelerate the development of cassava partial

inbred lines, since, after using only a few molecular markers, the results demonstrated the efficiency of MAS for identifying the most homozygous individuals within S₁ segregant populations.

The use of MAS to obtain inbred lines is widely disseminated and has been successfully used in several crops, such as papaya (Oliveira et al. 2010, 2012b), soybean (Song et al. 2014), wheat (Vinod et al. 2015), rice (Hasan et al. 2015), and castor bean (Machado et al. 2016). Most of these studies indicated the possibility of applying MAS to select transgressive individuals in different species and breeding populations. The early selection of the most homozygous individuals certainly will have a positive impact on reducing the time required for the development of these partial inbred lines in comparison to the phenotypic selection used in conventional breeding.

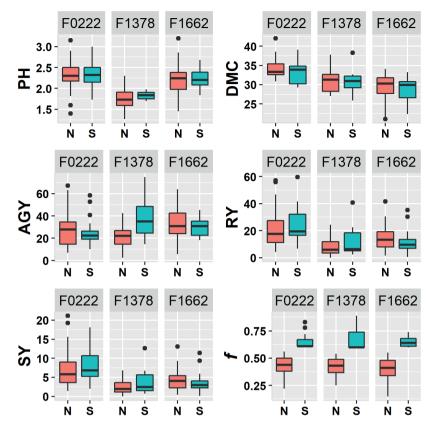


Figure 5 - Boxplot of the selection of S_1 cassava individuals from the self-pollination of the accessions BGM0222 (F0222), BGM1378 (F1378) and BGM1662 (F1662), based on the inbreeding coefficient (f). N: group of unselected individuals with f < 0.60. S: group of individuals selected with $f \ge 0.60$. PH: plant height (m); DMC: root dry matter content (%); AGY: above ground yield (t.ha⁻¹); RY: root yield (t.ha⁻¹); SY: starch yield (t.ha⁻¹).

Despite the efforts and investments made in the development of molecular markers for establishment of efficient MAS in cassava breeding, few practical results have been reported for selection of the desirable individuals (Oliveira et al. 2012a). Nonetheless, some examples of successful use of MAS in cassava breeding refer to the introduction of the CMD resistance gene in African germplasm using resistance sources from Latin America (Okogbenin et al. 2007). In other crops such as rice, MAS implementation has contributed for increasing the genetic gain in comparison to phenotypic methods, since it has reduced, on average, 40% of the costs for the development of superior rust-resistant genotypes

(Kuchel et al. 2005). Additionally, Morris et al. (2003) concluded that MAS did not present an effective cost reduction compared to conventional selection methods in maize, but it was more efficient for reducing the time required to obtain inbred lines.

In our work, selection based on the phenotypic data did not result in significant differences in the mean of the selected and unselected individuals for most of the traits, except for RY and SY in the F0222 progeny, and AGY, RY and SY in the F1378 progeny (Figure 6). By contrast, selection based on individuals with $f \ge 0.60$ resulted in an average increase of approximately 38% in the inbreeding mean of the progenies, without

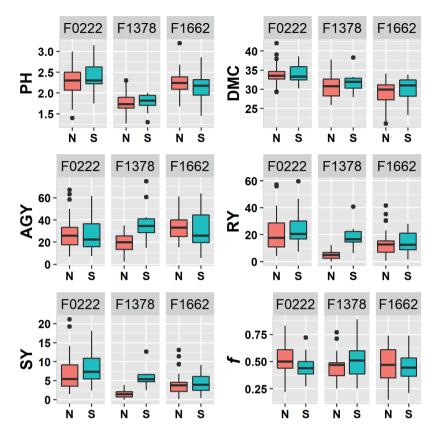


Figure 6 - Boxplot of the selection of S_1 cassava individuals from the self-pollination of accessions BGM0222 (F0222), BGM1378 (F1378) and BGM1662 (F1662), based on the selection index. S and N: S_1 selected and unselect individuals by the selection index, respectively. PH: plant height (m); DMC: root dry matter content (%); AGY: above ground yield (t.ha⁻¹); RY: root yield (t.ha⁻¹); SY: starch yield (t.ha⁻¹).

dramatically affecting the agronomic performance of the selected individuals (Figure 5). In contrast, by default, cassava and other cross-pollinated crops have a strong correlation between root yield and inbreeding (Ceballos et al. 2015, Freitas et al. 2016, Kaweesi et al. 2016). Therefore, probably the absence of differences in the phenotypic performance of the S₁ individuals with drastic differences in the inbreeding coefficients is due to the number of molecular markers used. However, the results obtained through MAS can greatly contribute in the selection of the most homozygous individuals to compose the S₂ generation and also result in genetic gains when associated with the phenotypic evaluations.

Although the use of inbred lines is an important objective in cassava breeding, its attainment is difficult, since conventional methods of self-pollination are expensive, time-consuming and can still be influenced by environmental effects (Ceballos et al. 2004). As a consequence, the development of cassava inbred lines could take around 14 years. Conversely, the use of MAS can be very efficient in identifying the most homozygous individuals, which can be selected early and then quickly submitted to a new self-pollination cycle to generate Sn populations with fixed loci.

Morris et al. (2003) evaluated the cost-benefit of MAS in comparison to the phenotypic selection for the development of maize inbred lines with

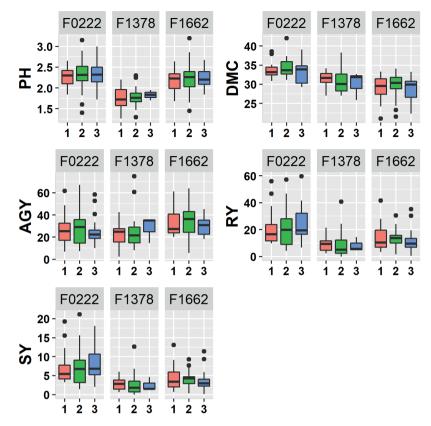


Figure 7 - Boxplot of the agronomic performance of S_1 cassava individuals from the self-pollination of accessions BGM0222 (F0222), BGM1378 (F1378) and BGM1662 (F1662), based on the inbreeding coefficient grouping (f). Group 1 (0.10 < f < 0.39), Group 2 (0.40 < f < 0.59), and Group 3 (f > 0.60). PH: plant height (m); DMC: root dry matter content (%); AGY: above ground yield (t.ha⁻¹); RY: root yield (t.ha⁻¹), SY: starch yield (t.ha⁻¹).

introgression of a dominant allele by backcross. The authors stated that 96% of the genome of the recurrent parent was recovered with three cycles of backcross assisted by microsatellites. However, for the phenotypic selection, six backcrosses were required to recover this same proportion of the recurrent parent genome. Therefore, MAS made it possible to reduce three cycles of backcross, which led to saving time and human resources. In another study in rice, MAS allowed the researchers to obtain highly homozygous genotypes with resistance to flooding in three cycles of backcrosses (BC₃F₁), whereas, by phenotypic selection, this would only be possible in the fifth backcross cycle (BC₅F₁) (Iftekharuddaula et al. 2012). Therefore,

the inherent advantages of MAS are very clear, and orphan crops such as cassava can greatly benefit from this tool when combined with conventional breeding.

EFFECT OF INCREASING INBREEDING ON AGRONOMIC TRAITS

There was no correlation between the inbreeding coefficient and the agronomic traits in the S₁ cassava individuals. One hypothesis to explain this result may be the lack of linkage between the molecular markers and the expression of these agronomic characteristics. According to Balloux et al. (2004), the number of markers used to accurately determine the correlations between agronomic performance

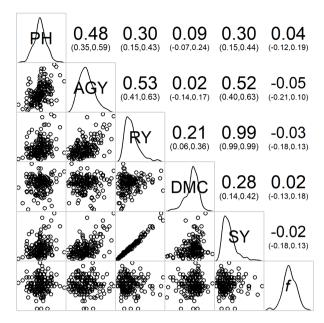


Figure 8 - Pearson correlation estimates for the inbreeding coefficient and agronomic traits in S₁ cassava individuals. PH: plant height (m); DMC: root dry matter content (%); AGY: above ground yield (t.ha⁻¹); RY: root yield (t.ha⁻¹); SY: starch yield (t.ha⁻¹); f: inbreeding coefficient.

and inbreeding may be a limiting factor, since a small number of markers may be inadequate to provide sufficient information on heterozygosity in the entire genome. Particularly in the case of cassava, the incorporation of SNPs via genotyping-by-sequencing (GBS) techniques to identify inbreeding in segregating progenies could eliminate the problem of low genomic coverage, since SNPs are widely distributed in the cassava genome (1 SNP for each 121bp – Pootakham et al. 2014).

The absence of a strong association between inbreeding and agronomic attributes identified by molecular markers in cassava indicates that it is possible to obtain genotypes with a higher level of inbreeding without great losses in agronomic performance, although this information should be studied in the S₂ populations. In contrast, some studies regarding inbreeding depression in S₁ cassava individuals have reported that, depending on the progeny evaluated, there are significant losses in some agronomic traits, such as root yield,

above ground yield, harvest index and dry matter content due to inbreeding depression (Rojas et al. 2009, Kawuki et al. 2011, Freitas et al. 2016). However, according to these same authors, the self-pollination of cassava accessions also allows the procurement of transgressive individuals with higher phenotypic values in comparison to the parents.

As for the agronomic traits, the results obtained in this study show associations of medium to high magnitude, especially between RY × SY, RY × AGY, AGY × SY and PH × AGY. These results are in agreement with other studies carried out for cassava, which presented correlations between agronomic, morphological and biochemical characteristics with magnitudes and direction similar to those observed in the present study (Ojulong et al. 2008, Ntawuruhunga and Dixon 2010, Gu et al. 2013, Oliveira et al. 2016).

PERSPECTIVES FOR MAS IMPLEMENTATION AND INBREEDING EXPLOITATION IN CASSAVA

Most of the cassava breeding programs use the recurrent phenotypic selection method to drive segregating populations generated from intraspecific crosses between heterozygous individuals (Ceballos et al. 2004), although the use of partial inbred lines has also been an alternative to generate transgressive clones (Freitas et al. 2016). However, recently, the efficiency of cassava breeding programs using conventional breeding strategies has been questioned, since, according to Ceballos et al. (2016), less than 1% of the clones generated reach the final stages of evaluation for the farmer's recommendation. Therefore, the low predictive ability of the parents' breeding values used for crossing is a factor that must be overcome (Ceballos et al. 2016). The use of inbred lines has been pointed out as an alternative to improve the prediction of the individuals' breeding values, besides allowing the use of genomic tools to improve the genetic gain with the selection.

The search for cassava inbred lines is relatively new in breeding programs; however, the potential gains of its use have been debated in the literature (Ceballos et al. 2015, Freitas et al. 2016, Kaweesi et al. 2016). Some theoretical benefits of the use of inbreeding in cassava are: reduction of the genetic load with the elimination of deleterious alleles from the populations; the elucidation of the genetic control of the main agronomic traits; the possibility of using intra- and interpopulation methods in a practical and fast way; the definition of heterotic groups to better explore heterosis; and the use of backcrosses to easily exchange alleles of interest in elite genotypes, especially monogenic or recessive genes (Ceballos et al. 2004).

The development of cassava inbred lines could lead to changes in the way the crop is propagated, since the use of seeds from pure inbred lines would allow the complete reproduction of the genotype of the cultivars/accession (as clonal propagation). In this case, the use of seeds for planting could favor the reduction of pathogens, which are frequently present in the cutting stakes, and allowing the initial evaluation of the clones with replicates in the first stages of selection; whereas this could increase selection efficiency in early cassava breeding stages, since, in general, there is a low correlation of cassava genotype performance in the clonal evaluation and preliminary yield evaluation trials, which directly affects the selection of superior clones and the elimination of individuals with lower genetic potential.

In addition to the aforementioned factors, the attainment of partially inbred cassava lines will depend on the flowering rate of the selected genotypes. It is known that breeding programs' selections tend to advance individuals who are upright when focusing on mechanized planting systems, since both cultural practices and harvest are facilitated, in addition to the fact that the multiplication rate of these genotypes is high. This leads to the selection of plants that do not branch, and since flowering is

directly related to the level of branching of the clones (Ceballos et al. 2017), the plants undergo negative selection for flowering. Therefore, advancement of self-pollination generations to obtain cassava inbred lines should be made through the association between molecular (MAS) and phenotypic selection considering the main agronomic traits and, whenever possible, taking into account genotypes with a high flowering rate.

Therefore, the present study demonstrated that the use of MAS was efficient in determining the inbreeding coefficient in S₁ cassava individuals, whose results can be applied to loci fixation in segregating cassava progenies. This enables reduction of the time required to obtain partially inbred lines and to better exploit the inherent advantages of this kind of genetic material.

CONCLUSIONS

Molecular-assisted selection is efficient in identifying individuals with high inbreeding level, providing a selection of approximately 25% of the individuals in cassava self-pollinated progenies. This early identification of the more homozygous individuals hastens the advancement of generations, allowing cost and time reductions compared to conventional selection for the development of inbred lines. There was no correlation between the inbreeding level and agronomic traits in cassava self-pollinated progenies; this is very important, since it is possible to obtain S₁ individuals with high inbreeding and desirable agronomic attributes.

ACKNOWLEDGMENTS

The authors thank the Fundação de Amparo à Pesquisa do Estado da Bahia (FAPESB), Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) and Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) for the financial assistance and scholarship support.

REFERENCES

- BALLOUX F, AMOS W AND COULSON T. 2004. Does heterozygosity estimate inbreeding in real populations? Mol Ecol 13: 3021-3031.
- CARMO CD, SILVA MS, OLIVEIRA GAF AND OLIVEIRA EJ. 2015. Molecular-assisted selection for resistance to cassava mosaic disease in *Manihot esculenta* Crantz. Sci Agric 72: 520-527.
- CEBALLOS H, IGLESIAS CA, PÉREZ JC AND DIXON AGO. 2004. Cassava breeding: opportunities and challenges. Plant Mol Biol 56: 503-516.
- CEBALLOS H, JARAMILLO JJ, SALAZAR S, PINEDA LM, CALLE F AND SETTER T. 2017. Induction of flowering in cassava through grafting. J Plant Breed Crop Sci 9: 19-29.
- CEBALLOS H, KAWUKI RS, GRACEN VE, YENCHO GC AND HERSHEY C. 2015. Conventional breeding, marker-assisted selection, genomic selection and inbreeding in clonally propagated crops: a case study for cassava. Theor Appl Genet 128: 1647-1667.
- CEBALLOS H, KULAKOW P AND HERSHEY C. 2012. Cassava breeding: current status, bottlenecks and the potential of biotechnology tools. Trop Plant Biol 5: 73-87.
- CEBALLOS H, PÉREZ JC, BARANDICA OJ, LENIS JI, MORANTE N, CALLE F, PINO L AND HERSHEY CH. 2016. Cassava Breeding I: the value of breeding value. Front Plant Sci 7: 1-12.
- COLLARD BCY AND MACKILL DJ. 2008. Marker-assisted selection: an approach for precision Plant Breed in the twenty-first century. Philos Trans R Soc Lond B Biol Sci 363: 557-572.
- DOYLE JJ AND DOYLE JL. 1990. Isolation of plant DNA from fresh tissue. Focus 12: 13-15.
- FREGENE MA ET AL. 2003. Simple sequence repeat marker diversity in cassava landraces: genetic diversity and differentiation in an asexually propagated crop. Theor Appl Genet 107: 1083-1093.
- FREITAS JPX, SILVAS VS AND OLIVEIRA EJ. 2016. Inbreeding depression in cassava for productive traits. Euphytica 209: 137-154.
- GOFF SA AND ZHANG Q. 2013. Heterosis in elite hybrid rice: speculation on the genetic and biochemical mechanisms. Curr Opin Plant Biol 16: 221-227.
- GOUDET J AND KELLER L. 2002. The correlation between inbreeding and fitness: does allele size matter? Trends Ecol Evol 17: 201-202.
- GU B, YAO Q, LI K AND CHEN S. 2013. Change in physicochemical traits of cassava roots and starches associated with genotypes and environmental factors. Starch-Stärke 65: 253-263.
- HALSEY M, OLSEN KM, TAYLOR NJ AND AGUIRRE PC. 2008. Reproductive biology of cassava. (*Manihot esculenta* Crantz) and isolation of experimental field trials. Crop Sci 48: 49-58.

- HASAN MM, RAFII MY, ISMAIL MR, MAHMOOD M, RAHIM HA, ALAM MA, ASHKANI S, MALEK MA AND LATIF MA. 2015. Marker-assisted backcrossing: a useful method for rice improvement. Biotechnol Biotechnol Equip 29: 237-254.
- IDREES M AND IRSHAD M. 2014. Molecular markers in plants for analysis of genetic diversity: A review. Eur Acad Res 2: 1513-1540.
- IFTEKHARUDDAULA KM, SALAM MA, NEWAZ MA, AHMED HU, COLLARD BCY, SEPTININGSIH EM, SANCHES DL, PAMPLONA AM AND MACKILL DJ. 2012. Comparison of phenotypic versus marker-assisted background selection for the SUB1 QTL during backcrossing in rice. Breed Sci 62: 216-222.
- KAWANO K, FUKUDA WMG AND CENPUKDEE U. 1987. Genetic and environmental effects on dry matter content of cassava root. Crop Sci 27: 69-74.
- KAWEESI T, KYALIGONZA V, BAGUMA Y, KAWUKI R AND FERGUSON ME. 2016. Inbreeding enhances field resistance to cassava brown streak viruses. J Plant Breed Crop Sci 8: 138-149.
- KAWUKI RS, NUWAMANYA E, LABUSCHAGNE MT, HERSELMAN L AND FERGUSON ME. 2011. Segregation of selected agronomic traits in six S₁ cassava families. J Plant Breed Crop Sci 3: 154-160.
- KOBAYASHI A, SONODA J, SUGIMOTO K, KONDO M, IWASAWA N, HAYASHI T, TOMITA K, YANO M AND SHIMIZU T. 2013. Detection and verification of QTLs associated with heat-induced quality decline of rice (*Oryza sativa* L.) using recombinant inbred lines and nearisogenic lines. Breed Sci 63: 339-346.
- KUCHEL H, YE G, FOX R AND JEFFERIES S. 2005. Genetic and economic analysis of a targeted marker-assisted wheat breeding strategy. Mol Breed 16: 67-78.
- KUNKEAW S, YOOCHA T, SRAPHET S, BOONCHAN AWIWAT A, BOONSENG O, LIGHTFOOT DA, TRIWITAYAKORN K AND TANGPHATSORNRUANG S. 2011. Construction of a genetic linkage map using simple sequence repeat markers from expressed sequence tags for cassava (*Manihot esculenta* Crantz). Mol Breed 27: 67-75.
- LI Y ET AL. 2014. Contributions of parental inbreeds and heterosis to morphology and yield of single-cross maize hybrids in China. Crop Sci 54: 76-88.
- LIU K AND MUSE SV. 2005. PowerMarker: an integrated analysis environment for genetic marker analysis. Bioinform 21: 2128-2129.
- LOKKO Y, DANQUAH EY, OFFEI SK, DIXON AG AND GEDIL MA. 2005. Molecular markers associated with a new source of resistance to the cassava mosaic disease. Afr J Biotechnol 4: 873-881.
- MACHADO EL, SILVA SA, FERNANDES SL AND BRASILEIRO HS. 2016. Genetic variability and homozygosity in a F₄ castor bean population by microsatellite markers. Bragantia 75: 307-313.

- MBA REC, STEPHENSON P, EDWARDS K, MELZER S, NKUMBIRA J, GULLBERG U, APEL K, GALE M, TOHME J AND FREGENE M. 2001. Simple sequence repeat (SSR) markers survey of the cassava (*Manihot esculenta* Crantz) genome: towards an SSR-based molecular genetic map of cassava. Theor Appl Genet 102: 21-31.
- MOHAN C, SHANMUGASUNDARAM P AND SENTHIL N. 2013. Identification of true hybrid progenies in cassava using simple sequence repeat (SSRY) markers. Bangladesh J Bot 42: 155-160.
- MORRIS M, DREHER K, RIBAUT JM AND KHAIRALLAH M. 2003. Money matters II: costs of maize inbred line conversion schemes at CIMMYT using conventional and marker-assisted selection. Mol Breed 11: 235-247.
- NTAWURUHUNGA P AND DIXON AG. 2010. Quantitative variation and interrelationship between factors influencing cassava yield. J Appl Biosci 26: 1594-1602.
- OJULONG H, LABUSCHANGNE MT, FREGENE M AND HERSELMAN LA. 2008. Cassava clonal evaluation trial based on a new cassava breeding scheme. Euphytica 160: 119-129.
- OKOGBENIN E ET AL. 2007. Marker-assisted introgression of resistance to cassava mosaic disease into Latin American germplasm for the genetic improvement of cassava in Africa. Crop Sci 47: 1895-1904.
- OLIVEIRA EJ, OLIVEIRA GAF, COSTA JL, OLIVEIRA VJS, DANTAS ACVL, DANTAS JLL AND PADUA JG. 2012b. Genetic diversity and marker-assisted inbreeding in papaya. Sci Hortic 147: 20-28.
- OLIVEIRA EJ, PÁDUA JG, ZUCCHI MI, VENCOVSKY R AND VIEIRA MCL. 2006. Origin, evolution and genome distribution of microsatellites. Genet Mol Biol 29: 294-307.
- OLIVEIRA EJ, RESENDE MDV, SANTOS VS, FERREIRA CF, OLIVEIRA GAF, SILVA MS, OLIVEIRA LA AND VILDOSO-AGUILAR CI. 2012a. Genome-wide selection in cassava. Euphytica 187: 263-276.
- OLIVEIRA EJ, SANTANA FA, OLIVEIRA LA AND SANTOS VS. 2015. Genotypic variation of traits related to quality of cassava roots using affinity propagation algorithm. Sci Agric 72: 53-61.
- OLIVEIRA EJ, SANTOS PEF, PIRES AJV, TOLENTINO DC AND SILVA SV. 2016. Selection of cassava varieties for biomass and protein production in semiarid areas from Bahia. Biosci J 32: 661-669.
- OLIVEIRA EJ, SILVA AS, CARVALHO FM, SANTOS LF, COSTA JL, AMORIM VBO AND DANTAS JLL. 2010. Polymorphic microsatellite marker set for *Carica papaya* L. and its use in molecular-assisted selection. Euphytica 173: 279-287.
- PERERA PIP, ORDOÑEZ CA, LOPEZ-LAVALLE LB AND DEDICOVA B. 2014. A milestone in the doubled haploid pathway of cassava. Protoplasma 251: 233-246.
- PLUTHERO FG. 1993. Rapid purification of high-activity Taq DNA polymerase. Nucleic Acids Res 11: 4850-4851.

- POOTAKHAM W, SHEARMAN JR, RUANG-AREERATE P, SONTHIROD C, SANGSRAKRU D, JOMCHAI N, YOOCHA T, TRIWITAYAKORN K, TRAGOONRUNG S AND TANGPHATSORNRUANG S. 2014. Large-scale SNP discovery through RNA sequencing and SNP genotyping by targeted enrichment sequencing in cassava (*Manihot esculenta* Crantz). PLoS One 9: 1-19.
- PRADO SA, GANBIM BL, NOVOA AD, FOSTER D, SENIOR ML, ZINSELMEIER C, OTEGUI MAE AND BORRAS L. 2013. Correlations between parental inbred lines and derived hybrid performance for grain filling traits in maize. Crop Sci 53: 1636-1645.
- R CORE TEAM. 2017. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0. Available at: http://www.R-project.org/.
- ROJAS MC, PÉREZ JC, CEBALLOS H, BAENA D, MORANTE N AND CALLE F. 2009. Analysis of inbreeding depression in eight S₁ cassava families. Crop Sci 49: 543-548.
- SAENGCHAN K, NOPHARATANA M, LERDLATTAPORN R AND SONGKASIRI W. 2015. Enhancement of starch-pulp separation in centrifugal-filtration process: Effects of particle size and variety of cassava root on free starch granule separation. Food Bioprod Process 95: 208-217.
- SONG B, SHEN L, WEI X, GUO B, TUO Y, TIAN F, HAN Z, WANG X, LI W AND LIU S. 2014. Marker-assisted backcrossing of a null allele of the α-subunit of Soybean (Glycine max). Breed 133: 638-648.
- VALE EM, REIS RS, SANTA-CATARINA R, PEREIRA MG, SANTA-CATARINA C AND SILVEIRA V. 2016. Comparative proteomic analysis of the heterosis phenomenon in papaya roots. Sci Hortic 209: 178-186.
- VINOD NM, SHARMA JB, TOMAR RS, SIVASAMY M AND PRABHU KVP. 2015. Marker-assisted backcross breeding to combine multiple rust resistance in wheat. Plant Breed 134: 172-177.
- WOLFE MD, KULAKO WP, RABBI IY AND JANNINK JL. 2016. Marker-based estimates reveal significant non-additive effects in clonally propagated cassava (*Manihot esculenta*): implications for the prediction of total genetic value and the selection of varieties. G3-Genes Genom Genet 6: 1-47.
- WRIGHT S. 1965. The interpretation of population structure by F-statistics with special regard to systems of mating. Evolution 19: 395-420.
- WU HX, HALLINGBÄCK HR AND SÁNCHEZ L. 2016. Performance of seven tree breeding strategies under conditions of inbreeding depression. G3-Genes Genom Genet 6: 529-540.
- XU Y AND CROUCH JH. 2008. Marker-assisted selection in plant breeding: from publications to practice. Crop Sci 48: 391-407.