# Cystic fibrosis at a Brazilian center of excellence: clinical and laboratory characteristics of 104 patients and their association with genotype and disease severity

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### **Abstract**

**Objective:** To identify the clinical, laboratory and radiographic characteristics of the cystic fibrosis patients under care at Universidade Estadual de Campinas (UNICAMP) in the last decade of the twentieth century, and to investigate the association of these characteristics with genotype and severity of the disease as measured by the Shwachman score.

**Methods:** Descriptive, retrospective and cross-sectional study of the patients assisted at UNICAMP hospital's Cystic Fibrosis Clinic from July 1990 to July 2000.

**Results:** One hundred and four patients were studied; 53.8% male; 93.3% Caucasian; 89.4% presented with respiratory symptoms; 59.6% presented with digestive symptoms; 5.8% had meconium ileus; 4.8% had diabetes. The mean age at onset of symptoms was 3 months, and the mean age at diagnosis was 2 years and 4 months. At diagnosis, 69.9 and 56.6% of the patients had weight and height below  $10^{th}$  percentile, respectively; in 10.6%, sweat chloride was < 60 mEq/l. *Staphylococcus aureus* was found in 80.2%, *Pseudomonas aeruginosa* in 76.0%, and *Burkholderia cepacia* in 5.2%. ΔF508 homozygosis was observed in 18.75%, whereas 62.50% of the patients were ΔF508 heterozygous. A moderate/severe Shwachman score was found in 15.7%. Eighteen patients died in that period (17.3%). The mean age at death was 7 years and 8 months; median survival after diagnosis was 18 years and 4 months. Patients who have at least one ΔF508 mutation have more frequent alterations in fecal fat levels when compared to patients who do not have this mutation (p < 0.05). There were no differences in any parameter between ΔF508 homozygous and heterozygous patients.

**Conclusions:** The clinical and laboratory characteristics of the 104 patients studied were similar to the characteristics described for patients in other countries. Exceptions are the higher age at diagnosis and lower survival. Our results support the recommendation for early diagnosis and the need for more treatment opportunities in the population of cystic fibrosis patients.

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# Introduction

Cystic fibrosis (CF) has been widely studied by many authors from several different countries, making possible greater understanding of its pathophysiology and the introduction of new therapeutic methods and impacting significantly with reduced morbidity and increased patient survival.

Little research exists, however, evaluating cystic fibrosis populations from developing countries. As a result the treatments and public health measures offered to patients with cystic fibrosis in such countries are based on international data and don't take their specific peculiarities into account. In addition to treating patients without real knowledge of their characteristics, this situation has further implications since, when it becomes necessary to choose which drugs will be offered by the state (as it is impossible to offer all of

them) it is fundamental that this choice be based on the local population's characteristics.

The World Health Organization drew attention to this fact in 1997<sup>1</sup> making the following recommendations for actions to be taken at Cystic Fibrosis Services in developing countries: neonatal screening as a means of establishing incidence and identify those newborn babies affected; setting-up of genetic laboratories should be encouraged to identify CF genetic mutations, the development of diagnosis and treatment centers with multidisciplinary teams; the establishment of a national organization involving cystic fibrosis patients' family members, friend and other volunteers; the establishment of a national register; the dissemination of information on the condition to health professionals, public authorities and the general public and increased collaboration between the groups and other organizations (including the pharmaceutical industry).

This lack of information is observable in Brazil where little research has been done describing and analyzing the characteristics of patients with cystic fibrosis.

The objective of this study was to describe the clinical and laboratory characteristics of cystic fibrosis patients treated at the *Universidade Estadual de Campinas* (UNICAMP) hospital's Cystic Fibrosis Clinic, during the last decade of the twentieth century and check for associations with the  $\Delta$ F508 mutation and disease severity, as measured by the Shwachman.

# **Patients and Methods**

This was a retrospective, descriptive, cross-sectional cohort study of patients treated at the UNICAMP Cystic Fibrosis Clinic, between July 1990 and July 2000.

All patients who had attended at least one consultation at the Cystic Fibrosis Clinic and whose diagnosis had been confirmed by clinical history and at least two sweat tests returning chloride values greater than or equal to 60 mEq/l or by the identification of two mutations.

Electrolyte concentration in sweat was tested by means of sweat stimulation by iontophoresis with pilocarpine.<sup>2</sup>

Sputum cultures were taken to identify Staphylococcus aureus, Pseudomonas aeruginosa, Mucoid Pseudomonas aeruginosa and Burkholderia cepacia. Samples were seeded in chocolate agar, blood agar, supplemented agar, MacConkey culture medium (*Pseudomonas* specific) and thioglycollate. Mucoid strains of *Pseudomonas aeruginosa* were visually identified by their characteristic morphology (presence of liquid mucus). Selective media were not employed for Burkholderia cepacia.

The following mutations were tested for:  $\Delta$ F508, G542X, N1303K, G551D, R553X and W1282X. The  $\Delta$ F508 mutation was detected by means of polymerase chain reaction (PCR) and analyzed on 8% polyacrylamide gels, using the modifications described by Rommens et al.<sup>3</sup> Other mutations were analyzed by the PCR technique in association with restriction enzyme digestion.

Descriptive analysis employed mean, median and standard deviation values for continual variables and absolute frequency for discrete variables. Categorical variables were analyzed using the chi-square test. Fisher's exact test was applied in cases when one of the cells in the 2 X 2 tables was less than or equal to 5. Continuous and sequential measurements were compared across the two groups using the Mann-Whitney test. Survival time was analyzed using the Kaplan-Meier method. Curves were compared through the application of the Wilcoxon (Breslow) test. The significance level adopted was 5%.

This research was approved by the UNICAMP Committee for Ethics in Research.

### Results

One hundred and four patients were studied, of whom, 53.8% were male, 93.3% Caucasoid and 6.7% Negroid. Eighteen patients died during the period covered by the study. Parental consanguinity occurred in 6.2% of the population under study.

Respiratory and digestive symptoms occurred in 89.4 and 59.6% of the patients, respectively. Six had histories of meconium ileus and five developed diabetes mellitus.

The stated age of onset of symptoms varied from birth to 20 years, with a mean of 16 months and median of 3 months (Figure 1).

Age at diagnosis varied from the neonatal period to 29 years and 11 months, with a mean of 4 years and 2 months and median of 2 years and 4 months (Figure 2).

With respect of nutritional status, 69.9 and 56.6% of the patients exhibited weight and stature below the tenth percentile, respectively, at first consultation.

Room air transcutaneous hemoglobin oxygen saturation was above 95% in 59.5%, between 91 and 95% in 32.9% and below 91% in 7.6% of patients.

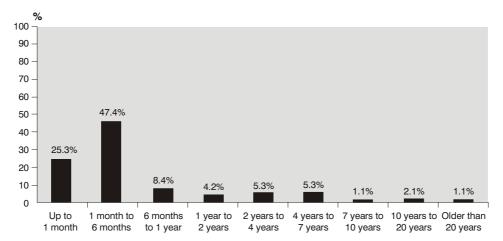
Sweat chloride concentration was below 60 mEq/l in 10.6% (11) of the patients, with 1.9% (2) exhibiting values below 40 mEq/l; 28.8% (30) of the patients presented values between 60 and 100 mEq/l and 60% (63) presented values above 100 mEq/l.

More than 80% had been colonized by Staphylococcus aureus, 76% by Pseudomonas aeruginosa, more than half by Mucoid Pseudomonas aeruginosa and 5.2% by Burkholderia cepacia (Table 1).

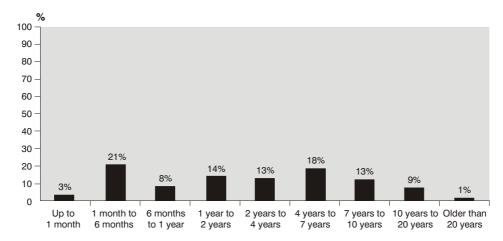
Spirometry was used for 55 patients and was normal in 27.3% (15), indicated restrictive respiratory disorder 18.2% (10), obstructive respiratory disorder 25.4% (14) and mixed respiratory disorder in 29.1% (16).

Ninety-six patients were analyzed genetically, of whom 18.75% (18) were homozygous for  $\Delta$ F508 and 62.5% (60) were  $\Delta F508$  heterozygotes. This being so, 50% of the chromosomes studied exhibited the  $\Delta$ F508 mutation.

The following percentages of other mutations were found in the 192 chromosomes studied: G542X (4.17%), N1303K (2.08%), G551D (1.04%), R553X (0.52%), W1282X (0.52%).



**Figure 1** - Distribution of patients according to the age of onset of symptoms (n = 95)



**Figure 2 -** Distribution of patients according to the age at diagnosis (n = 100)

The Shwachman score was calculated for 83 patients and returned: 57.8% (48) with an excellent or good score, 26.5% (22) with an average score and 15.7% (22) with a moderate or severe rating.

**Table 1 -** Distribution of patients according to the colonization by different bacteria (n = 96)

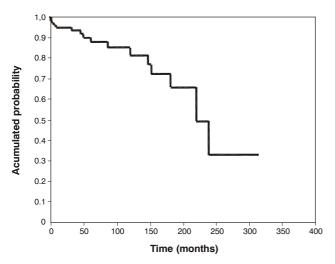
%	n
80.2	77
76.0	73
53.1	51
5.2	5
51.0	49
	80.2 76.0 53.1 5.2

During this period 18 patients (17.3%) died. Tem were male and eight female, 17 Caucasoids and one Negroid, all died from respiratory insufficiency.

Age at death varied from 6 months to 23 years and 1 month. Mean average age at death was 9 years and 5 months with a median of 7 years and 4 months.

At the point when the study was completed, median post-diagnosis survival was 18 years and 4 months and 81.39% of the patients had survived for 10 years after diagnosis. The post-diagnosis survival curve for the sample can be observed in Figure 3.

An attempt was made to test the correlation between the Shwachman score (SS) and the patients' clinical and laboratory characteristics. Eighty-three subjects were included in this analysis, of whom 54.2% (45) were male, 57.8% (48) presented an excellent or good SS, 26.5% (22) average and 15.7% (13) moderate or severe. Table 2 shows the statistical correlation of clinical and laboratory characteristics with the SS.



**Figure 3 -** The post-diagnosis survival curve for the sample analyzed using the Kaplan-Meier method

Clinical and laboratory differences between patients with cystic fibrosis with and without the  $\Delta F508$  mutation were analyzed. Ninety-six patients were studied, of whom 81% (78) had the  $\Delta F508$  mutation and 19% (18) did not present this mutation; 52% (50) were male. Table 3 presents the statistical correlation between clinical and laboratory parameters and the presence or absence of the  $\Delta F508$  mutation.

Seventy-eight patients were studied in order to evaluate the clinical and laboratory differences between patients with the  $\Delta F508$  mutation in the homozygote and heterozygote forms, of whom 23% (18) were homozygotes and 77% (60) heterozygotes for the  $\Delta F508$  mutation; 47% (37) were male. The presence of homozygosis or heterozygosis for the  $\Delta F508$  mutation did not exhibit any statistically significant correlation with any of the parameters studied, as Table 4 shows.

### **Discussion**

The clinical and laboratory characteristics of the 104 patients studied are close to values found in other, domestic and international, research.

Distribution by sex,  $race^{4-6}$  and incidence of pulmonary manifestations<sup>4,7,8</sup> was comparable with what is found in the literature.

Digestive clinical manifestations were confirmed in 59.6% of the patients and the fecal fat balance was abnormal in 67.9% of these, indicating an incidence of pancreatic insufficiency below published figures.  $^{4,7,8}$  The lower pancreatic insufficiency, compared with that found in the United States and Europe, may be the result of the lower prevalence of the  $\Delta F508$  mutation in Brazil, which is associated with this insufficiency.  $^{9-14}$ 

In the current study and in another study performed in Brazil,<sup>5</sup> the incidence of *meconium ileus* was 5.8 and 1.9%, respectively, which is much lower than values quoted in literature which vary from 15 to 20%.<sup>15,16</sup> One hypothesis

**Table 2 -** Statistical correlation between the patients' clinical and laboratory characteristics and the Shwachman score (n = 83)

Variables	р	Statistical test
Colonization by <i>Pseudomonas aeruginosa</i>	< 0.001	Fisher's exact test
Colonization by <i>P. aeruginosa</i> mucosa	< 0.001	Chi-square test
Forced vital capacity	< 0.001	Fisher's exact test
First second forced expiratory volume	< 0.001	Fisher's exact test
Oxygen saturation	< 0.001	Fisher's exact test
Number of infectious exacerbations		
during the previous years' follow-up	0.045	Chi-square test
Indications for Dornase Alpha use	0.019	Chi-square test
Indications for a regular respiratory physiotherapy program	0.017	Chi-square test
Indications for home oxygen therapy	< 0.001	Chi-square test
Sex	0.810	Chi-square test
Sweat chloride levels	0.656	Chi-square test
Colonization by Sthaphylococcus aureus	0.058	Fisher's exact test
Colonization by Burkholderia cepacia	0.431	Chi-square test
Presence of the $\Delta$ F508 mutation	0.230	Fisher's exact test
Presence of one or two $\Delta$ F508 mutations	0.669	Fisher's exact test
Current age	0.487	Chi-square test

Table 3 - Statistical correlation between the patients' clinical and laboratory characteristics and the presence or absence of the  $\Delta$ F508 (n = 96)

Variable	р	Statistical test
Fecal fat levels	0.005	Chi-square test
Sex	0.058	Chi-square test
Race	1	Fisher's exact test
Incidence of pulmonary manifestations	0.681	Fisher's exact test
Digestive clinical manifestations	0.079	Chi-square test
Incidence of <i>meconium ileus</i>	0.579	Fisher's exact test
Age at onset of symptoms	0.673	Mann-Whitney
Current weight	0.368	Fisher's exact test
Diabetes mellitus	0.570	Fisher's exact test
Colonization by Sthaphylococcus aureus	0.751	Fisher's exact test
Colonization by <i>Pseudomonas aeruginosa</i>	0.067	Fisher's exact test
Colonization by <i>P. aeruginosa mucosa</i>	0.525	Chi-square test
Colonization by <i>Burkholderia cepacia</i>	1	Fisher's exact test
Forced vital capacity	0.089	Fisher's exact test
First second forced expiratory volume	0.723	Fisher's exact test
Presence of bronchiectasis	1	Fisher's exact test
Presence of hepatomegaly	0.327	Fisher's exact test
Shwachman score	0.230	Fisher's exact test
Number of infectious exacerbations during		
the previous years' follow-up	0.298	Fisher's exact test
Indication of pancreatic enzymes	0.265	Chi-square test
Indication of respiratory physiotherapy	0.351	Fisher's exact test
Indications for Dornase Alpha use	0.540	Chi-square test
Indications for home oxygen therapy	0.255	Fisher's exact test
Death	1	Fisher's exact test
Age at death	0.606	Mann-Whitney
Survival curve	0.595	Wilcoxon's test

Table 4 - Statistical correlation between the patients' clinical and laboratory characteristics and the presence of the  $\Delta$ F508 mutation in the homozygote and heterozygote forms (n = 78)

Variable	p	Statistical test
Sex	0.803	Chi-square test
Race	0.617	Fisher's exact test
Incidence of pulmonary manifestations	0.423	Fisher's exact test
Digestive clinical manifestations	0.966	Chi-square test
Incidence of meconium ileus	0.325	Fisher's exact test
Age at onset of symptoms	0.284	Mann-Whitney
Current weight	0.570	Fisher's exact test
Diabetes mellitus	1	Fisher's exact test
Colonization by Sthaphylococcos aureus	0.493	Fisher's exact test
Colonization by Pseudomonas aeruginosa	0.746	Fisher's exact test
Colonization by P. aeruginosa mucosa	0.413	Chi-square
Colonization by Burkholderia cepacia	0.258	Fisher's exact test
Feces fat levels	1	Fisher's exact test
Forced vital capacity	0.145	Fisher's exact test
First second forced expiratory volume	0.161	Fisher's exact test
Presence of bronchiectasis	0.646	Fisher's exact test
Presence of hepatomegaly	0.734	Fisher's exact test
Shwachman score	0.669	Fisher's exact test
Number of infectious exacerbations during		
the previous years' follow-up	0.835	Fisher's exact test
Indication of pancreatic enzymes	0.529	Fisher's exact test
Indication of respiratory physiotherapy	1	Fisher's exact test
Indications for Dornase Alpha use	0.051	Chi-square test
Indications for home oxygen therapy	1	Fisher's exact test
Death	0.227	Fisher's exact test
Age at death	0.352	Mann-Whitney
Survival curve	0.175	Wilcoxon's test

for why the incidence of *meconium ileus* is lower in Brazil could be that patients with cystic fibrosis die during their first year of life, before being diagnosed.

The incidence of *Diabetes mellitus* was comparable with what has been found in other published work.<sup>4,17,18</sup>

Age at onset of symptoms, 72.7% of the patients had had some type of clinical manifestation before 6 months of age, with a median of 3 months. This demonstrates that onset is early and serves to alert us to the possibility of a CF diagnosis. Three patients' symptoms had onset after 10 years of age and one of these, after 20 years of age. This fact is evidence of the importance of diagnosis for patients whose respiratory symptoms have onset later on.<sup>19</sup>

The median of age at diagnosis was 2 years and 4 months and diagnosis was made after 10 years of age in 10% of cases. Data from the Registro Latino-americano de Fibrosis Quística, 6 including records from Brazil, show that the mean age at diagnosis was 4.2 years. In contrast, data from the Cystic Fibrosis Foundation in 2002,<sup>4</sup> reveal that the median age at diagnosis in the United States is 6 months, showing that Latin America as a whole needs to increase efforts to make early CF diagnoses, as the World Health Organization emphasizes.1

In this sample, the difference between the median age at diagnosis and the median age at onset of symptoms was 2 years and 1 month. Mitchell et al.<sup>20</sup> found an average delay between symptom onset and diagnosis of 3 years. In Brazil, studies by Maróstica<sup>21</sup> and Espinoza<sup>22</sup> show that these delays were 7 months for pediatric patients and 10.7 years in those over 15 years, respectively.

While CF does not have a cure, it does exhibit significant improvement with the treatment of symptoms.<sup>23</sup> Recent research has shown that early diagnosis reduces morbidity<sup>24</sup> and highlights the importance of starting treatment for pancreatic insufficiency and malnutrition early and also respiratory physiotherapy to maintain airways unobstructed.<sup>25</sup> One study demonstrated that infection by P. aeruginosa was the main risk factor for morbidity and mortality, pointing out that early diagnosis would aid intervention at the onset of colonization by this agent and contribute to better prognosis.<sup>26</sup> Another study also emphasizes the importance of treating P. aeruginosa aggressively when it is identified for the first time.<sup>27</sup>

The majority of states in Brazil do not currently run neonatal CF screening and its employment generates controversy. The cost of implementing screening, in a region with a disease incidence of 1:5,000, would be R\$ 25,000.00 per new case. The negative points against screening include the elevated cost, the false positive and false negative results and the fact that recent review articles have not demonstrated differences in CF progress between patients who were diagnosed by neonatal screening and those whose diagnosis was made after symptom onset. 28,29 It is suggested that greater information about CF for pediatricians, allied with easier access to diagnosis by electrolytes in sweat would be a more plausible alternative. The cost-based argument can be questioned since one recent study demonstrated that the cost of diagnosis by neonatal screening is lower than the cost of diagnosis when this is not employed.<sup>30</sup> Another study, in France, demonstrated that patients whose diagnosis was made by neonatal screening had better progress than those whose diagnosis was made after symptom onset. 31 It is known that delays identifying and treating Pseudomonas makes it eradication difficult.<sup>27</sup> Serum testing can detect colonization by Pseudomonas 6 to 12 months before cultures are positive.<sup>32</sup> In this light, diagnosis by neonatal screening would lead to the performance of routine serum testing making earlier detection and a more efficient attempt to eradicate this bacteria possible. There is also the hope that a vaccine against *Pseudomonas aeruginosa* will become available for patients with cystic fibrosis. Then neonatal screening would become less controversial since without it the chance to vaccinate some patients before colonization would be lost. Finally, we should remember that neonatal screening makes it possible to give the parents genetic counseling before another pregnancy occurs.

When neonatal screening is not employed, early diagnosis can indicate greater severity. One Brazilian study found that a combination of low Shwachman score, low birth weight and low age at diagnosis are indicative of worse prognosis. 33

During the ten-year follow-up period, 17.3% of the patients died. The median age at death was 7.8 years and median post-diagnosis survival, on the study end-date, was 18.4 years. In the United States, median survival increased from 1 year, in 1940, to 20 years in 1980.34 In 1990, CF survival in Canada and the United States was 30.9 and 27.6 years, respectively.35 Another study quotes the life expectancy of CF patients as having increased from 8 years in 1970 to 29.5 years in 1998<sup>36</sup> and, according to data from the Cystic Fibrosis Foundation, current life expectancy in the United States is 31.6 years. 4 Mortality in the United States between 1985 and 1999 for patients between 2 and 15 years old reduced by between 45 and 70%.37 Another study presents even more optimistic data and considers that, currently, for a newborn with CF, life expectancy is more than 40 years.<sup>38</sup>

In another Brazilian study, median survival, after birth, of a cohort of 111 patients diagnosed between 1970 and 1994, was 12.6 years.<sup>39</sup>

The low survival in our country, when compared with that of developed countries is very worrying and reflects levels from twenty years ago in such countries. We believe that certain factors may be contributing to this fact. Initially, the delay in making diagnoses may mean that patients start treatment in a phase where there has already been lung deterioration and the chance to treat P. aeruginosa exacerbations early may already have been missed. Furthermore, at the time of diagnosis, 70 and 56.6% respectively of our patients presented weight and stature below the tenth percentile, which values are higher than those reported in the literature, around 42 to 44%.<sup>4,40</sup> Malnutrition caused by delayed treatment may also contribute to a more rapid deterioration, since a relationship between malnutrition and worsening pulmonary function has been demonstrated.41 Another

factor which could be contributing is the small number of centers specialized in treating CF. As each center covers an area and a population that are extremely large, the difficulty patients find in accessing these centers may be contributing to compromised treatment and follow-up. According to the Cystic Fibrosis Foundation guidelines, each patient should be making four visits a year, which is not possible for many of our patients. Either increasing the number of CF treatment centers or improving the transport system for these patients could contribute to more frequent follow-up appointments and reduced morbidity and mortality.

Recent studies have demonstrated worse progress among patients with cystic fibrosis whose socio-economic status is lower. Schechter et al. found that the risk of death was 3.65 times greater among the poorest patients with cystic fibrosis, who also exhibited the most deterioration in terms of values for pulmonary function and nutritional status.<sup>42</sup> These findings were confirmed with a very large sample of patients from the Cystic Fibrosis Foundation.<sup>4</sup> A recent study demonstrated that patients with a higher socio-economic status present a 40% lower risk of mortality when compare with patients of low socio-economic status. 43 This being so, the greater morbidity and mortality of our patients may have a relationship with low socio-economic status.

Sweat chloride concentration was lower than 60 mEq/l in 11 patients (10.6%). This percentage is higher than cited in the literature where some authors relate just 2% of cystic fibrosis patients presenting sweat chloride lower than 60 mEq/I.44,45 Of these 11 patients, six (three males) had three or more sweat chloride test results lower than 60 mEq/I and diagnosis confirmed by the identification of one or two mutations. All were Caucasoids, with normal fecal fat balance and presented respiratory symptoms, but not digestive symptoms. They had all been chronically colonized by Sthaphylococcos aureus and Pseudomonas aeruginosa, five by Mucoid Pseudomonas aeruginosa and two by Burkholderia cepacia. Genetic testing found that four patients were homozygous for  $\Delta$ F508 and two for  $\Delta$ F508/N1303K.

The other five patients whose sweat chloride concentration was below 60 mEq/l in our sample presented both respiratory and digestive symptoms compatible with CF, improved with the use of pancreatic enzymes and were colonized by *Pseudomonas aeruginosa*, with one having the  $\Delta$ F508 mutation; this being so we considered CF and they received the conventional treatment.

Cystic Fibrosis diagnoses in the face of normal sweat chloride and sodium levels have also been confirmed by other authors. 46,47 In such cases attempts should be made to confirm diagnosis with the identification of two CF mutations or by measurement of nasal potential difference. 48-50 This test is not yet performed at our service. When diagnosis cannot be established based on the suggested methods, it should be based on clinical judgement, <sup>48</sup> as was the case of the five patients described.

Sixty-six percent of the patients were colonized by Pseudomonas aeruginosa, close to what is found in literature.4,51

Colonization by Mucoid Pseudomonas aeruginosa and Staphylococcus aureus, respectively, had incidences of 53 and 80%, which is similar to values found in other studies.4,7,22,52

The presence of Burkholderia cepacia in five patients (5.2%) is a worrying fact since colonization by this bacteria leads to very bad prognosis and reduces survival, 53 with one study demonstrating that this reduction is by as much as ten years in comparison with patients infected by just Pseudomonas aeruginosa. 54 Values for Burkholderia cepacia prevalence, in general, vary from 5 to 15%, 4,21,55 being higher in some regions, such as Ontario in Canada, where they reach 22%.56

Earlier Brazilian data<sup>21</sup> describe 10.7% patients presenting restrictive respiratory disorder, 25% obstructive respiratory disorder and 17.9% mixed respiratory disorder, which values are also very similar to those found in our sample.

We performed genotyping for 92.31% of our patients, which represents a good percentage when compared with the Cystic Fibrosis Foundation, who genotyped 81.4% of their patients.<sup>4</sup> In our sample, the  $\Delta$ F508 mutation was present in 50% of the 192 chromosomes studied. The other mutations were found in the following percentages: G542X (4.17%), N1303K (2.08%), G551D (1.04%), R553X (0.52%), W1282X (0.52%).

Also in Brazil, Raskin et al.<sup>57</sup> describe the most frequent mutations in the Brazilian population as being:  $\Delta F508$ (47%), G542X (5.5%), N1303K (2.6%), R553X (0.8%) and G551D (0.2%). These authors further state that 26% of CF patients are homozygous for the  $\Delta$ F508 mutation.

The most frequently encountered mutations in the United States were:  $^{58}$   $\Delta$ F508 (66.0%), G542X (2.4%), G551D (1.6%), N1303K (1.3%), W1282X (1.2%) and R533X (0.7%). In France, 59 the  $\Delta$ F508 mutation was present in 67.9% of the chromosomes of a population of 2,666 CF patients, with the other most frequent mutations being: G542X (2.5%), N1303K (2%), 1717-1G  $\rightarrow$  A(1.2%), R553X (0.8%) and G551D (0.7%). In Argentina, the  $\Delta \text{F508}$  mutation is present in 66% of the chromosomes of FC patients, 60 and, in Mexico, 7.2% of CF patients' chromosomes exhibit the G542X mutation.<sup>61</sup>

The lower incidence of the  $\Delta F508$  mutation in our country compared with data from the United States, France and Argentina, probably occurs as a result of the large amount of racial intermixing in Brazil.

Around 60% of the patients presented a Shwachman Score (SS) that was excellent or good and just 15.7% had a moderate or severe score. Henry et al.<sup>62</sup> analyzing 60 patients report their average SS as 78.87.

In this analysis, the SS exhibited a statistically significant correlation with colonization by Pseudomonas aeruginosa, colonization by Mucoid Pseudomonas aeruginosa, forced vital capacity, first second forced expiratory volume, transcutaneous hemoglobin saturation by oxygen, number of infectious exacerbations during the previous years' follow-up, indications for Dornase Alpha use, indications for a regular respiratory physiotherapy program and indications for home oxygen therapy. As these parameters are individually related with the severity of clinical status, it is considered that the SS is a good general severity evaluation method. Another Brazilian study found evidence that the SS offered positive correlation with first second forced expiratory volume. 63

In this sample, the presence of the  $\Delta$ F508 mutation was associated with abnormal fat balance and pancreatic insufficiency. These data are in agreement with international literature in which a number of different authors demonstrate that the  $\Delta F508$  mutation is associated with the presence of pancreatic insufficiency. 9-13

In this sample, homozygous  $\Delta$ F508 patients did not present any differentiating characteristic when compared with heterozygotes for the mutation. There is little extant work in the literature comparing these two subsets. Farrell & Koscik<sup>64</sup> report that sweat chloride levels are equal for the two groups.

In Conclusion, the clinical and laboratory characteristics of the 104 CF patients studied were similar to those described for the CF populations of other countries, with the exception of: pancreatic insufficiency (less frequent), previous history of *meconium ileus* (less frequent), median for age at diagnosis (higher), median of age at death and post-diagnosis survival (lower in comparison with data from the United States and Canada), malnutrition at time of diagnosis (higher than that reported in other countries), percentage of patients with normal sweat chloride concentrations (greater than that described in international literature), incidence of the  $\Delta$ F508 mutation (lower in comparison with data from the United States and Europe).

Our results also permit the conclusion that in patients who present clinical status suggestive of CF, sweat chloride concentration < 60 mEq/l does not exclude this diagnosis and genetic studies should be performed in order to attempt to define the diagnosis.

Our data allows for the conclusion that efforts towards early diagnosis and greater treatment opportunities need to be made for cystic fibrosis patients.

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# References

- 1. World Health Organization and International Cystic Fibrosis (Mucoviscidosis) Association – Implementation of cystic fibrosis services in developing countries: memorandum from a joint  $WHO/ICF\,(M)A\,meeting.\,Bulletin\,of\,the\,World\,Health\,Organization.$ 1997;75(1):10.
- 2. Gibson LE, Cooke RE. A test for concentration of electrolytes in sweat in cystic fibrosis of the pancreas utilizing pilocarpine by iontophoresis. Pediatrics. 1959;23:545-9.

- 3. Rommens JM, Kerem B, Greer W, Chang P, Tsui LC, Ray P. Rapid non radioactive detection of the major cystic fibrosis mutation. Am J Hum Genet. 1990;46:395-6.
- Cystic Fibrosis Foundation, Patient Registry. Bethesda, Maryland: 2002 Annual Data Report; 2003.
- 5. Camargos PAM, Guimarães MDC, Reis FJC. Prognostic aspects of cystic fibrosis in Brazil. Ann Trop Pediatr. 2000;20:287-91.
- Registro Latino-Americano de Fibrosis Quística (REGLAFQ). Informe del cuarto año. Buenos Aires; 1993. p. 21.
- 7. Huang N, Schidlow D, Szatrowski T, Palmer J, Laraya-Cuasay L, Yeung W, et al. Clinical features, survival rate and prognostic factors in young adults with cystic fibrosis. Am J Med. 1987;2:871-9.
- 8. Welsh MJ, Tsui LC, Boat TF, Beaudet AL. Cystic fibrosis. In: Scriver CR, Beaudet AL, Sly WS, Valle D. The Metabolic and Molecular Bases of Inherited Disease. 7th ed. New York: McGraw-Hill; 1995. p. 3799-3876.
- Kerem E, Corey M, Kerem B, Rommens JM, Markiewicz D, Levison H, et al. The relationship of the most common mutation ΔF508, N Engl J Med. 1990;323:1517-22.
- 10. Borgo G, Mastella G, Gasparini P, Zoranello A, Doro R, Pignatti PF. Pancreatic function and genetic ΔF508 in cystic fibrosis. J Med Genet. 1990;27:665-9.
- 11. Santis G, Osborne L, Knight RA, Hodson ME. Independent genetic determinants of pancreatic and pulmonary status in cystic fibrosis. Lancet. 1990;336:1081-4.
- 12. Campbell PW III, Phillips JA III, Krishnamani MR, Maness KJ, Hajinski TA. Cystic fibrosis: relationship between clinical status and  $\Delta$ F508 deletion. J Pediatr. 1991;118:239-41.
- 13. Johansen HK, Nir M, Hoib YN, Koch C, Schwartz M. Severity of cystic fibrosis in patients homozygous and heterozygous for ΔF508 mutation. Lancet. 1991;337:631-4.
- 14. McKone EF, Emerson SS, Edwards KL, Aitken ML. Effect of genotype on phenotype and mortality in cystic fibrosis: a retrospective cohort study. Lancet. 2003;361:1671-6.
- 15. Cystic Fibrosis Foundation. Clinical Practice Guidelines for Cystic Fibrosis; 1997.
- 16. Park RW, Grand RJ. Gastrointestinal manifestations of cystic fibrosis: a review. Gastroenterology. 1981;81:1143.
- 17. Finkelstein SM, Wielinski CL, Elliott GR. Diabetes mellitus associated with cystic fibrosis. J Pediatr. 1988;112:373.
- 18. Rosenecker J, Eichler I, Kuhn L, Harms HK, Von de Hardt J. Genetic determination of diabetes mellitus in Danish CF patients: prevalence and late diabetic complications. Acta Paediatr. 1994;83:72-7.
- 19. Bargon J, Rickmann J, Jacobi V, Straub R, Arnemann J, Wagner TO. Cystic fibrosis: initial diagnosis in a 39 year-old patient. Med Klin. 2000;95:697-700.
- 20. Mitcell-Heggs P, Mearns M, Batten JC. Cystic fibrosis in adolescents and adults. Quarter J Med New Series. 1976;XLV: 479-504.
- 21. Maróstica PJC. Avaliação pneumológica de pacientes portadores de fibrose cística: sua relação com grupos genéticos [tese]. Porto Alegre, Universidade Federal do Rio Grande do Sul; 1995.
- 22. Domec Espinoza MPS. Fibrose cística em jovens e adultos do Hospital das Clínicas da Unicamp [dissertação]. Campinas, Universidade Estadual de Campinas; 1998.
- 23. Tauber E, Eichler I, Gartner C, Halmerbauer G, Gotz M, Rath R, et al. Improvements of lung function in cystic fibrosis. Pediatr Pulmonol. 2002:33:263-8.
- 24. Wang SS, O'Leary LA, Fitzsimmons SC, Khoury MJ. The impact of early cystic fibrosis on pulmonary function in children. J Pediatr. 2002;141:804-10.
- 25. Wagener JS, Headley AA. Cystic fibrosis: current trends in respiratory care. Respir Care. 2003;48:234-45.
- 26. Emerson J, Rosenfeld M, McNamara S, Ramsey B, Gibson RL. Pseudomonas aeruginosa and other predictors of mortality and morbidity in young children with cystic fibrosis. Pediatr Pulmonol. 2002;34:91-100.
- 27. Bush A. Decisions facing the cystic fibrosis clinician at the first isolation of Pseudomonas aeruginosa. Paediatr Respir Rev. 2002;3:82-8.
- 28. Merelle ME, Nagelkerke AF, Lees CM, Dezateaux C. Newborn screening for cystic fibrosis. Cochrane Database Syst Rev. 2001;3:CD001402.
- 29. Farrell PM, Li Z, Kosorok MR, Laxova A, Green CG, Collins J, et al. Bronchopulmonary disease in children with cystic fibrosis after early or delayed diagnosis. Am J Respir Crit Care Med. 2003;168:1100-8.

- 30. Lee DS, Rosenberg MA, Peterson A, Makholm L, Hoffman G, Laessig RH, et al. Analysis of the costs of diagnosing cystic fibrosis with a newborn screening program. J Pediatr. 2003;142:617-23.
- 31. Siret D, Bretaudeau G, Branger B, Dabadie A, Dagorne M, David V, et al. Comparing the clinical evolution of cystic fibrosis screened neonatally to that of cystic fibrosis diagnosed from clinical symptoms: a 10-year retrospective study in a French region (Brittany). Pediatr Pulmonol. 2003;35:342-9.
- West SE, Zeng L, Lee BL, Kosorok MR, Rock MJ, Splaingard MJ, et al. Respiratory infections with *Pseudomonas aeruginosa* in children with cystic fibrosis: early detection by serology and assessment of risk factors. JAMA. 2002;287:2958-67.
- Oliveira MC, Reis FJ, Oliveira EA, Colosimo EA, Monteiro AP, Penna FJ. Prognostic factors in cystic fibrosis in a single center in Brazil: a survival analysis. Pediatr Pulmonol. 2002;34:3-10.
- 34. Maclusky I, Levison H. Cystic fibrosis. In: Chernick V, Boat TE. Kendig's Disorders of the Respiratory Tract in Children. Philadelphia: Saunders; 1990. p. 692-729.
- Kerem E, Reisman J, Corey M, Canny GJ, Levison H. Prediction of mortality in patients with cystic fibrosis. N Engl J Med. 1992;326:1187-91.
- 36. Bolyard DR. Sexuality and cystic fibrosis. Am J Matern Child Nurs. 2001;26:39-41.
- 37. Kulich M, Rosenfeld M, Goss CH, Wilmott R. Improved survival among young patients with cystic fibrosis. J Pediatr. 2003;142:631-6.
- 38. Doull IJ. Recent advances in cystic fibrosis. Arch Dis Child. 2001;85:62-6.
- Reis FJC, Camargos PAM, Rocha SF. Survival analysis for cystic fibrosis in Minas Gerais State, Brazil. J Trop Pediatr. 1998;44: 329-31
- Farrell PM, Kosorok MR, Laxova A, Shen G. Nutritional benefits of neonatal screening for cystic fibrosis. N Engl J Med. 1997;337:963-9.
- 41. Zemel BS, Jawad AF, FitzSimmons S, Stallings VA. Longitudinal relationship among growth, nutritional status and pulmonary function in children with cystic fibrosis: analysis of the Cystic Fibrosis Foundation National Patient Registry. J Pediatr. 2000;137:374-80.
- 42. Schechter MS, Shelton BJ, Margolis PA, Fitzsimmons SC. The association of socioeconomic status with outcomes in cystic fibrosis in the United States. Am J Respir Crit Care Med. 2001;163:1331-7.
- 43. O'Connor GT, Quinton HB, Kahn R, Robichaud P, Maddock J, Lever T, et al. Case-mix adjustment for evaluation of mortality in cystic fibrosis. Pediatr Pulmonol. 2002;33:99-105.
- 44. Huff DS, Huang NN, Arey JB. Atypical cystic fibrosis of the pancreas with normal levels of sweat chloride and minimal pancreatic lesions. J Pediatr. 1979;94:237.
- Stewart B, Zabner J, Shuber AP, Welsh MJ, Mccray Jr. PB. Normal sweat chloride values do not exclude the diagnosis of cystic fibrosis. Am J Respir Crit Care Med. 1995;151:899-903.
- 46. Desmarquest P, Feldman N, Tamalat A, Boule M, Fauroux B, Tournier G, et al. Genotype analysis and phenotypic manifestations of children with intermediate sweat chloride test results. Chest. 2000;118:1591-7.
- Lebecque P, Leal T, De Boeck C, Jaspers M, Cuppens H, Cassiman J. Mutations of the cystic fibrosis gene and intermediate sweat chloride levels in children. Am J Respir Crit Care Med. 2002;165:757-61.
- 48. Rosentein BJ, Cuting GR. The diagnosis of cystic fibrosis: a consensus statement. Cystic Fibrosis Foundation Consensus Panel. J Pediatr. 1998;132:589-95.

- 49. Rosentein BJ. What is a cystic fibrosis diagnosis? Clin Chest Med. 1998;19:423-41.
- 50. Ratjen F, Döring G. Cystic fibrosis. Lancet. 2003;361:681-9.
- Burns JL, Gibson RL, Mcnamara S, Yim D, Emerson J, Rosenfeld M, et al. Longitudinal assessment of *Pseudomonas aeruginosa* in young children with cystic fibrosis. J Infect Dis. 2001;183: 444-52.
- 52. Dornelas EC, Fernandes MIM, Galvão LC, Silva GA. Estudo do quadro pulmonar de pacientes com fibrose cística. J Pediatr (Rio J). 2000;76:295-99.
- 53. Soni R, Marks G, Henry DA, Robison M, Moriaty C, Parsons S, et al. Effects of *Burkholderia cepacia* infection in the clinical course of patients with cystic fibrosis: a pilot study in a Sydney clinic. Respirology. 2002;7:241-5.
- 54. Jones AM, Dodd ME, Webb AK. *Burkholderia cepacia*: current clinical issues, environmental controversies and ethical dilemmas. Eur Respir J. 2001;17:295-301.
- 55. Lewin LO, Byard PJ, Davis PB. Effect of *Pseudomonas cepacia* colonization on survival and pulmonary function of cystic fibrosis patients. J Clin Epidemiol. 1990;43:125-31.
- 56. Speert DP, Henry D, Vandamme P, Corey M, Mahenthiralingam E. Epidemiology of *Burkholderia cepacea* complex in patients with cystic fibrosis, Canada. Emerg Infec Dis. 2002;8:181-7.
- Raskin S, Philips III JA, Krishnamani MRS, Jones C, Parker RA, Rozov T. DNA analysis of cystic fibrosis in Brazil by direct PCR amplification from guthrie cards. Am J Med Gen. 1993;46:665-9.
- 58. Cystic Fibrosis Genetic Analysis Consortium Population variation of common cystic fibrosis mutations. Human Mutation. 1994;4:167-77.
- Guilloud-Batalie M, De Crozes D, Rault G, Degioanni A, Feingold J. Cystic fibrosis mutations: report from the French Registry. The Clinical Centers of the CF. Hum Hered. 2000;50:142-5.
- Saleh MC, Botelli A, Melano De Botelli M, Rezzonico CA, Argaraña CE. Cystic fibrosis: frequency of delta F508 and G542X mutations in Cordoba, Argentina. Medicina (B. Aires). 1996;56:14-6.
- 61. Villarreal MT, Chavez M, Lezana JL, Cuevas F, Carnevale A, Codova E, et al. G542X mutation in Mexican cystic fibrosis patients. Clin Genet. 1996;49:54-6.
- 62. Henry RL, Mellis CM, Petrovic L. Mucoid *Pseudomonas aeruginosa* is a marker of poor survival in cystic fibrosis. Pediatr Pulmonol. 1992;12:158-61.
- Assis I, Camargos PAM, Reis FJC, Sulmonett N, Carneiro APS. Assessing correlations between spirometry and Shwachman-Kulczycki score in children and adolescents. Pediatr Pulmonol. 2003;36:305-9.
- 64. Farrell PM, Koscik RE. Sweat chloride concentrations in infants homozygous or heterozygous for ΔF508 cystic fibrosis. Pediatrics. 1996:97:524-8.

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