

Inflammatory Phenotype by OCT Coronary Imaging: Specific Features Among De Novo Lesions, In-Stent Neointima, and In-Stent Neo-Atherosclerosis

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Abstract

Background: Coronary stenosis can be caused de novo atherosclerosis, in-stent restenosis, and in-stent neoatherosclerosis, three entities that develop from a diverse pathophysiological milieu.

Objective: This study aims to investigate, using optical coherence tomography (OCT), whether or not coronary lesions related to these processes differ in their local inflammatory profile.

Methods: Retrospective analysis of patients with diagnosed or suspected coronary lesions who had undergone OCT imaging for clinical reasons. Macrophage and intra-plaque neovascularization were assessed by OCT and used as surrogates of local inflammation. A significance level of < 0.05 was adopted as statistically significant.

Results: From the 121 lesions, 74 were de novo, 29 were restenosis, and 18 were neoatherosclerosis. Neovascularization was found in 65.8% of de novo, 10.3% in restenosis, and 94.4% in neoatherosclerosis ($p < 0.01$ for all). The volume of neovascularization was different among lesion types (950 vs. 0 vs. 6220, respectively [median values in $1000 \times \mu\text{m}^3/\text{mm}$]; $p < 0.01$ for all), which were significantly higher in neoatherosclerosis and lower in restenosis. The presence of macrophages differed among the lesions (95.9% in de novo vs. 6.9% in restenosis vs. 100% in neoatherosclerosis [$p < 0.01$ for all]). Moreover, the intensity of macrophagic infiltration was different among lesion types (2.5 vs. 0.0 vs. 4.5, respectively [median values of macrophage score]; $p < 0.01$ for all), significantly higher in neoatherosclerosis and lower in restenosis.

Conclusion: When compared using coronary OCT, de novo atherosclerosis, in-stent restenosis, and neoatherosclerosis presented markedly different inflammatory phenotypes.

Keywords: Coronary Restenosis; Atherosclerosis; Stents.

Introduction

Coronary atherosclerotic disease is a ubiquitous cause of morbimortality worldwide, frequently treated with stent implantation. However, it is well known that re-narrowing of the lumen of the stent may occur within the first months following percutaneous intervention, a phenomenon referred to as restenosis.¹ Both entities (i.e. *de novo* atherosclerosis and in-stent restenosis) originate from markedly distinct pathogenetic mechanisms. Atherosclerotic plaque formation is a complex, multifactorial, long-lasting condition modulated by multiple systemic and local risk factors.² On the other hand, in-stent

restenosis is secondary to neointimal tissue growth, a vascular healing response triggered by vessel injury following device implantation.^{3,4} More recently, neoatherosclerosis has been described as another distinct cause of in-stent lumen narrowing. It is largely believed to be an accelerated form of atherosclerotic plaque formation probably induced by a sustained local tissular response to the stent metallic scaffold itself.⁵ The accumulation of inflammatory cells has been described as a central event for the development of *de novo* atherosclerosis^{2,6,7} and in-stent restenosis,⁸ as well as for neo-atherosclerosis.⁹ Local inflammation is believed to be an integral part of those conditions, functioning as the decisive step through which the vessel wall is dynamically modified as the pathologic process progresses. To date, however, it has been poorly described whether inflammatory profiles do vary according to the underlying type of condition, and if the potential differences can be assessed by clinical tools. Intravascular optical coherence tomography (OCT) provides *in vivo* near histology-level imaging,¹⁰ which has been largely used to investigate patients with coronary artery disease.¹¹⁻¹³ In addition to quantitatively measuring dimensional parameters, OCT has

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been validated as a tool to assess qualitative characteristics of the vessel wall, such as tissue type components, plaque accidents, and thrombus formation.^{11,12} Also importantly, OCT has been shown to accurately detect macrophage infiltration^{10,14} and intra-wall neovessel formation,¹⁵ two features associated with subjacent local inflammation. The present study aims at investigating whether atherosclerosis, in-stent restenosis, and neo-atherosclerosis differ in their inflammatory phenotype (i.e., macrophage and neovessel presence and quantity) as assessed by OCT imaging.

Methods

Patient selection

We conducted a search in our institution's database for patients who had undergone coronary OCT in native coronary arteries for clinical indications, between January of 2012 and December of 2019. All OCT runs from every patient were revised, and selected for final analysis if presenting: i) one or more *de novo* atherosclerotic lesions (defined as a plaque arc $\geq 180^\circ$), or ii) one or more lesions in a previously implanted stent (defined as at least $300 \mu\text{m}$ of in-stent tissue thickness). Lesions in the same vessel were considered discrete, and counted as such, if separated by a normal segment longer than 10 mm. Lesions at the stent edges (5-mm proximal or distal) were not included for analysis. Also, the present report only included lesions which OCT examination was performed prior to any intervention. This study was approved by the local ethics committee and is in accordance with the Declaration of Helsinki.

Image acquisition and analysis

Image acquisition was performed using standard techniques, during injection of contrast media as described elsewhere,¹⁶ using a frequency-domain OCT system (C7 or Ilumien OPTIS system, C7 DragonFly or DragonFly II imaging catheters, St. Jude Medical, St. Paul, MN).

Two independent reviewers blinded to any clinical information performed the evaluations of all OCT images. Any disagreement between the reviewers was resolved by consensus. Lesions were classified as *de novo*, in-stent restenosis, or in-stent neoatherosclerosis. The latter in-stent lesion was differentiated from the former by the presence of calcific or lipidic deposits in neoatherosclerotic lesions, as opposed to the homogenous appearance of the neointimal restenotic tissue (Figure 1).^{17,18}

Lesions were analyzed using standard definitions, as suggested elsewhere.¹⁸⁻²¹ Lipid tissue was defined as signal-poor regions with poorly defined, diffuse borders. Fibrous tissue was defined as a region with high backscattering and a relatively homogeneous signal. Calcific deposits were identified as signal-poor or heterogeneous structures with sharply delineated borders. The arc of calcium was measured at the frame with the largest extension of calcific deposit. Macrophages were identified by the presence of signal-rich, distinct, or confluent punctate images exceeding the intensity of background speckle noise (Figure 2A); macrophage accumulation was graded using a score from 0 to 4 in each frame and then summed the gradings for the whole lesion.²⁰ Neovascularization was defined as no-signal, intra-plaque structures without connection

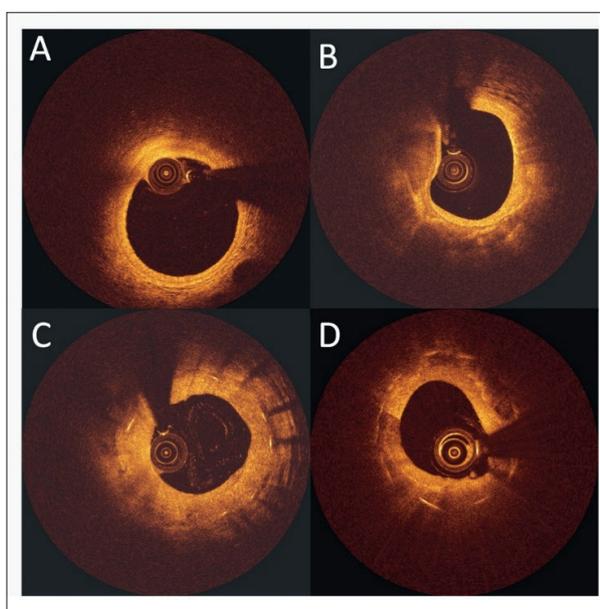


Figure 1 – OCT images of a normal coronary artery (A), *de novo* atherosclerosis (B), in-stent restenosis (C) and neoatherosclerosis (D). In-stent restenosis is characterized by the homogenous appearance of the neointimal restenotic tissue, while neoatherosclerosis presents in-stent lipidic and calcific deposits.

to the vessel lumen measuring between $50\text{-}300 \mu\text{m}$ and recognized in ≥ 3 consecutive frames (Figure 2B).^{18,21} The volume of neovascularization was calculated by summing the area of neovascularization in each frame and then applying Simpson's rule. Both macrophage accumulation and volume of neovascularization were indexed by plaque length, to allow for comparison among lesions. Thrombus was defined as a mass protruding into the vessel lumen, typically with irregular contours, discontinuous from the surface of the vessel wall (Figure 3A). Thin-cap fibroatheromas (TCFA) were defined as a region with maximal lipid arc more than 90° and cap thickness $< 65 \mu\text{m}$. Ruptured plaque was defined by the presence of intimal tearing, disruption, or dissection of the cap (Figure 3B).

Off-line quantitative OCT analyses utilized a dedicated software package (QIvus 3.0, Medis Medical, The Netherlands). Quantitative parameters included plaque length, minimal luminal cross-sectional area (CSA), and maximal luminal stenosis (minimal CSA \div mean [distal and proximal] reference lumen CSA). For *in-stent* lesions, neointimal area (stent CSA minus lumen CSA) and neointimal thickness (measured perpendicularly from stent strut to lumen) were also calculated.

Statistical analysis

Statistical analyses were performed using SPSS 26.0 (IBM Corp. Armonk, NY, USA). Categorical variables are presented as counts and frequencies and were analyzed using Chi-square or Fisher's exact test when appropriate. To test the normality of distribution, we performed a Shapiro-Wilks test. Continuous variables did not present normal distribution; therefore, their results are presented as median and interquartile range (IQR). We used the non-parametric Kruskal-Wallis test for multiple comparisons. When needed, pairwise comparisons

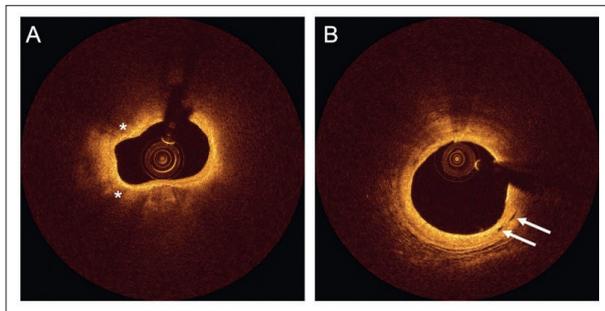


Figure 2 – OCT images of macrophage infiltration and neovascularization. The white asterisks in A indicate signal-rich, punctate images compatible with macrophage infiltration in OCT images. The white arrows in B indicate no-signal, intra-plaque images compatible with neovascularization.

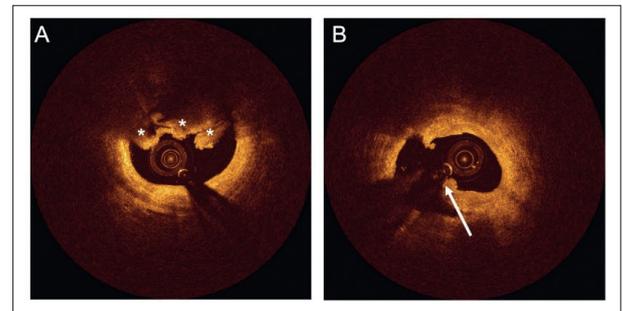


Figure 3 – OCT images of thrombus and ruptured plaque. The white asterisks indicate thrombus (A) and the white arrow indicates plaque rupture (B).

were performed using the Dunn-Bonferroni approach. A significance level of < 0.05 was adopted as statistically significant.

Results

Out of a total of 499 patients found in our database, 110 patients had at least one good-quality OCT run that imaged an entire lesion prior to any interventional manipulation, and comprised the present study population. Most patients were males, over 60 years of age, with multiple risk factors for coronary artery disease and presenting with acute coronary syndrome (ACS) upon hospital admission (Table 1).

In Table 2, we present the characteristics among the different types of plaque. Most characteristics were different

Table 1 – Demographic and clinical characteristics (n=110 patients)

Males	88 (80.0)
Age, years	63 (56 – 71)
Hypertension	75 (68.2)
Diabetes	33 (30.0)
Hyperlipidemia	90 (81.8)
Smoking (current or former)	61 (55.5)
Family history of CAD	60 (54.5)
Acute coronary syndrome	69 (62.7)

Numbers are counts (percentage) or median (interquartile range). CAD: coronary artery disease.

Table 2 – OCT characteristics of de novo, neointimal, and neoatherosclerotic lesions (n=121)

	De novo (n=74)	In-stent restenosis (n=29)	In-stent neoatherosclerosis (n=18)	p*
Calcification	56 (75.7)	-	10 (55.6)	< 0.01
TCFA	17 (23.3)	-	7 (38.9)	< 0.01
Plaque rupture	10 (13.9)	0	7 (38.9)	< 0.01
Thrombus	9 (12.5)	0	4 (22.2)	0.03
Neovascularization	48 (65.8)	3 (10.3)	17 (94.4)	< 0.01
Macrophage	71 (95.9)	2 (6.9)	18 (100)	< 0.01
Plaque length, in mm	24.1 (17.2-36.8)	25.8 (18.0-33.0)	23.5 (17.8-29.0)	0.9
Minimal luminal CSA, mm ²	2.42 (1.64-3.51)	2.72 (1.77-4.52)	1.85 (1.35-3.18)	0.07
Max. luminal stenosis, %	65.5 (54.8-74.6)	45.7 (33.1-63.0)	66.2 (53.9-76.2)	< 0.01
Max. IS tissue thickness, mm	-	0.74 (0.59-0.98)	1.13 (0.95-1.34)	< 0.01
Max. IS tissue CSA, mm ²	-	3.54 (2.87-4.69)	4.96 (4.22-6.21)	< 0.01
Neovasc. vol., 1000 $\mu\text{m}^3/\text{mm}$	950 (0-3400)	0 (0-0)	6220 (1250-13430)	< 0.01
Macrophage score	2.5 (0.9-4.9)	0.0 (0.0-0.0)	4.5 (3.1-7.3)	< 0.01

Numbers are counts (percentage) or median (interquartile range). CSA: cross sectional area; IS: in-stent; LAD: left anterior descending artery; LCx: left circumflex artery; Max: maximal; Neovasc: neovascularization; RCA: right coronary artery; TCFA: Thin-cap fibroatheroma; Vol: volume. *P-value for the overall comparison among the groups.

among the groups. Neovascularization was found in 65.8% of *de novo*, 10.3% in restenosis, and 94.4% in neoatherosclerosis ($p < 0.01$ for all) (Table 2). Accordingly, the volume of neovascularization was different among lesion types (950 vs. 0 vs. 6220, respectively [median values in $1000 \times \mu\text{m}^3/\text{mm}$]; $p < 0.01$ for all), being significantly higher in neoatherosclerosis and lower in restenosis (Figure 4).

The presence of macrophages differed among the lesions (95.9% in *de novo* vs. 6.9% in restenosis vs. 100% in neoatherosclerosis [$p < 0.01$ for all]). Also, the intensity of macrophagic infiltration was different among lesion types (2.5 vs. 0.0 vs. 4.5, respectively [median values of macrophage score]; $p < 0.01$ for all) (Figure 5), significantly higher in neoatherosclerosis and lower in restenosis (Figure 5).

When compared to stable patients, acute patients had more thrombus (16.2% versus 2.4%, $p = 0.029$) and lower intensity of macrophage infiltration (3.8 [1.2 – 5.9] versus 1.2 [0 – 3.6], $p = 0.008$). All other OCT features (plaque type; presence of neovascularization, macrophage, TCFA and plaque rupture; and volume of neovascularization) were not significantly different between the groups ($p > 0.05$ for all).

Discussion

Our study compared three different causes of coronary narrowing, namely *de novo*, restenotic and neoatherosclerotic lesions, and demonstrated marked differences among them in relation to their inflammatory phenotype by OCT, assessed by the presence and degree of macrophage accumulation and intra-lesion neovessels.

Inflammation is the cornerstone for understanding these three different processes that cause coronary stenosis. Pathogenesis of native coronary atherosclerosis has been extensively investigated in the last decades² and involves multiple inflammatory pathways. However, since coronary

stenting is a somewhat recent technique, in-stent restenosis is a pathological entity that did not exist previously and is not entirely understood yet. Following percutaneous intervention, blood flow disturbances, migration and proliferation of smooth muscle cells and fibroblasts into the intima ensues, causing deposition of extracellular matrix, collagen, lymphocytes and macrophages.^{4,8,22} Continuous inflammatory stimulus caused by the enduring metallic structures of the stent also leads to intra-plaque foreign-body reaction, accelerating atherosclerotic changes²³ and increasing the presence of neovascularization.²¹ Also, incomplete maturation of endothelial cells due to the anti-proliferative drugs eluted by the stents impairs the barrier function usually carried out by the normal endothelium.²⁴ Both increased neo-vessel presence and immature endothelium are likely responsible for allowing a steep inflow of inflammatory cells into the neointimal tissue. Unlike native vessel atherosclerosis, that develops over decades,² neoatherosclerosis is an accelerated atherosclerotic process set in an abnormally healed vessel wall that can occur in a few years or even months following stent implantation, particularly with drug-eluting stents.⁵ These differences are observed *in vivo* in our study, with neoatherosclerosis presenting significantly larger neovascularization volumes and macrophage density when compared to both *de novo* and restenotic lesions.

In-stent restenosis due to neointimal hyperplasia is believed to be limited to a certain timeframe following stent implantation²⁵ and was generally considered to be a somewhat benign, stable event, not frequently related to acute coronary events.²⁶ More recently, however, it has been observed that in-stent restenosis may present as acute coronary syndrome in more than 50% of the cases.²⁷ Neoatherosclerosis probably develops itself upon neointimal hyperplasia,²¹ following plaque modifications that infiltrate lipids and macrophages, which are associated with plaque rupture and acute coronary events. Our study population

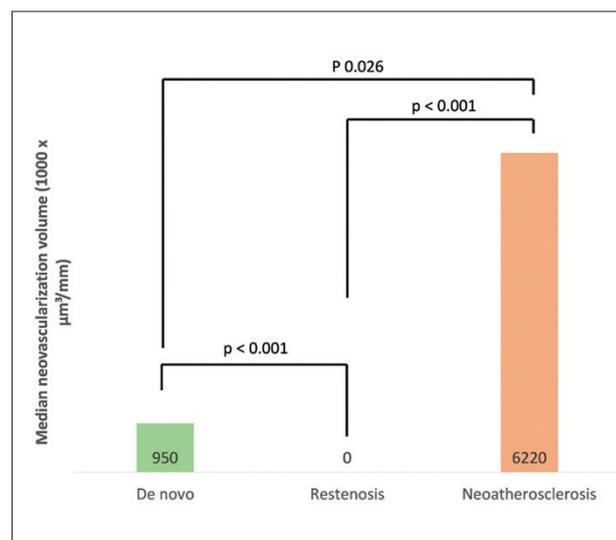


Figure 4 – Neovascularization volume by lesion type.

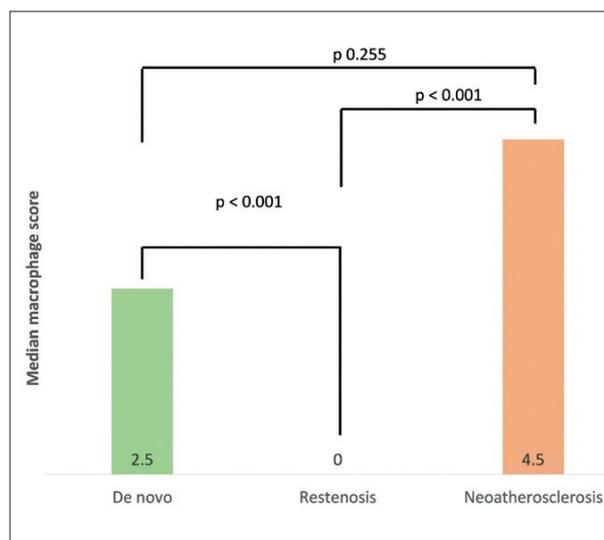


Figure 5 – Macrophage score by plaque type.

reflected such characteristics, with neoatherosclerotic plaques being significantly more prone to rupture than *de novo* and restenotic plaques, as well as presenting larger neointimal thickness and lumen sizes, which can be the result of such plaque modifications.

Our study has several limitations. It is an exploratory, observational, retrospective study, with a highly selected population of individuals with a high burden of cardiovascular risk factors and coronary artery disease, and most of our population (62.7%) was comprised of patients admitted to the hospital with acute coronary syndromes. Accordingly, it is not possible to extrapolate these findings to other clinical settings. Also, patients with acute coronary syndromes presented lower levels of macrophage infiltration in our sample. This finding may be explained by the fact that acute patients had significantly more thrombus when compared to stable patients, thus hindering the assessment of macrophage infiltration in these regions impossible in most cases. Although we had no information regarding the age or type of the stents implanted, in our opinion this was not detrimental to the interpretation of our findings, considering we were solely analyzing plaque characteristics.

Notwithstanding, this is supposed to be a hypothesis-generating study. Neoatherosclerosis is an important cause of late stent failure not reduced with the use of drug eluting stents and has a direct impact on the outcomes of coronary percutaneous interventions.²⁸ Risk factors such as hyperlipidemia, smoking and impaired renal function, all of which up-regulate systemic inflammation, have been associated with higher rates of neoatherosclerosis.^{29,30} Additionally, inflammation by itself has been linked with increased cardiovascular risk.² New evidence has emerged proving *in vivo* that modulation of the inflammatory response and risk factors control can reduce the rates of major cardiovascular events⁷ and reduce atherosclerotic plaque volume.³¹ Nonetheless, these effects are yet to be proven to reduce neointimal hyperplasia and neoatherosclerosis rates. In a recent article,³² Hashikata et al. demonstrated that the use of empaglifozin reduced neointimal hyperplasia at 12 months in diabetic patients when compared to standard glucose-lowering therapy. Mean neointimal thickness, volume and percentage were significantly lower in the empaglifozin group. Interestingly, this reduction was independent of lower glucose levels, suggesting a possible multi-factorial underlying mechanism. The current HUYGENS trial³³ included patients with non-ST segment elevation myocardial infarction who were treated with evolocumab or placebo in addition to intensive statin therapy for 52 weeks and underwent serial OCT and intravascular ultrasound imaging. The evolocumab group achieved lower LDL-C levels and imaging features that included a greater increase in minimum fibrous cap thickness, decrease in maximum lipid arc and plaque regression. More intensive lipid lowering with early addition of a PCSK9 inhibitor to statins after a NSTEMI produces stabilization and regression of coronary atherosclerosis. The improved clinical outcomes achieved with very low LDL-C levels associated with changes in plaque phenotype pave the way for these new lipid-lowering options become a perspective to prevent intra-stent neoatherosclerosis. Also, efforts are being made to

produce stents with novel absorbable scaffolds³⁴ and better drug delivery to modulate tissular response,³⁵ thus allowing a more physiologic endothelial regeneration and reducing the substrate that originates neoatherosclerosis.

To the best of our knowledge, this is the first study directly comparing plaque inflammation of native vessel atherosclerosis with neointimal hyperplasia and neoatherosclerosis using OCT. In our understanding, these findings stress the importance of inflammation in the pathogenesis of stent failure, suggesting that the future of PCI probably lies in fine-tuning tissular response and not leaving a metallic footprint behind.

Further prospective studies with aggressive lipid-lowering therapy, blood pressure and glucose control, smoking cessation, and control of inflammation may modify the evolution of neoatherosclerosis.

Conclusions

In summary, when compared using OCT, *de novo* atherosclerosis, in-stent restenosis and in-stent neoatherosclerosis presented markedly different inflammatory phenotypes (i.e., neovessel volume and macrophage quantification).

Author Contributions

Conception and design of the research: Pinheiro LF, Garzon S, Mariani J, Caixeta AM, Lemos PA; Acquisition of data and Analysis and interpretation of the data: Pinheiro LF, Garzon S, Mariani J, Prado GA, Caixeta AM, Almeida BO, Lemos PA; Statistical analysis: Pinheiro LF, Garzon S, Prado GA, Caixeta AM, Lemos PA; Writing of the manuscript and Critical revision of the manuscript for important intellectual content: Pinheiro LF, Garzon S, Lemos PA.

Potential Conflict of Interest

No potential conflict of interest relevant to this article was reported.

Sources of Funding

There were no external funding sources for this study.

Study Association

This article is part of the project of a postdoctoral submitted by Luiz Fernando Pinheiro, from Hospital Israelita Albert Einstein.

Ethics approval and consent to participate

This study was approved by the Ethics Committee of the Hospital Israelita Albert Einstein under the protocol number 3.722.061. All the procedures in this study were in accordance with the 1975 Helsinki Declaration, updated in 2013. Informed consent was obtained from all participants included in the study.

References

1. Dangas GD, Claessen BE, Caixeta A, Sanidas EA, Mintz GS, Mehran R. In-stent Restenosis in the Drug-Eluting Stent Era. *J Am Coll Cardiol.* 2010;56(23):1897-907. doi: 10.1016/j.jacc.2010.07.028. PMID: 21109112..
2. Libby P, Theroux P Pathophysiology of Coronary Artery Disease. *Circulation.* 2005;111(25):3481-8. doi: 10.1161/CIRCULATIONAHA.105.537878.
3. Weintraub WS. The Pathophysiology and Burden of Restenosis. *Am J Cardiol.* 2007;100(5A):3K-9K. doi: 10.1016/j.amjcard.2007.06.002.
4. Jukema JW, Verschuren JJ, Ahmed TA, Quax PH. Restenosis after PCI. Part 1: Pathophysiology and risk Factors. *Nat Rev Cardiol.* 2011;9(1):53-62. doi: 10.1038/nrcardio.2011.132.
5. Nakazawa G, Otsuka F, Nakano M, Vorpahl M, Yazdani SK, Ladich E, et al. The Pathology of Neointimal Hyperplasia in Human Coronary Implants Bare-Metal and Drug-Eluting Stents. *J Am Coll Cardiol.* 2011;57(11):1314-22. doi: 10.1016/j.jacc.2011.01.011.
6. Hansson GK. Inflammation, Atherosclerosis, and Coronary Artery Disease. *N Engl J Med.* 2005;352(16):1685-95. doi: 10.1056/NEJMra043430.
7. Ridker PM, Everett BM, Thuren T, MacFadyen JG, Chang WH, Ballantyne C, et al. Antiinflammatory Therapy with Canakinumab for Atherosclerotic Disease. *N Engl J Med.* 2017;377(12):1119-1131. doi: 10.1056/NEJMoa1707914.
8. Welt FG, Rogers C. Inflammation and restenosis in the Stent Era. *Arterioscler Thromb Vasc Biol.* 2002;22(11):1769-76. doi: 10.1161/01.atv.0000037100.44766.5b.
9. Romero ME, Yahagi K, Kolodgie FD, Virmani R. Neointimal Hyperplasia from a Pathologist's Point of View. *Arterioscler Thromb Vasc Biol.* 2015;35(10):e43-9. doi: 10.1161/ATVBAHA.115.306251.
10. Tearney CJ, Yabushita H, Houser SL, Aretz HT, Jang IK, Schlendorf KH, et al. Quantification of Macrophage Content in Atherosclerotic Plaques by Optical Coherence Tomography. *Circulation.* 2003;107(1):113-9. doi: 10.1161/01.cir.0000044384.41037.43.
11. Jang IK, Tearney CJ, MacNeill B, Takano M, Moselewski F, Iftima N, et al. In vivo Characterization of Coronary Atherosclerotic Plaque by use of Optical Coherence Tomography. *Circulation.* 2005;111(12):1551-5. doi: 10.1161/01.CIR.0000159354.43778.69.
12. Jang IK, Bouma BE, Kang DH, Park SJ, Park SW, Seung KB, et al. Visualization of Coronary Atherosclerotic Plaques in Patients Using Optical Coherence Tomography: Comparison with Intravascular Ultrasound. *J Am Coll Cardiol.* 2002;39(4):604-9. doi: 10.1016/s0735-1097(01)01799-5.
13. Yabushita H, Bouma BE, Houser SL, Aretz HT, Jang IK, Schlendorf KH, et al. Characterization of Human Atherosclerosis by Optical Coherence Tomography. *Circulation.* 2002;106(13):1640-5. doi: 10.1161/01.cir.0000029927.92825.f6.
14. Di Vito L, Agozzino M, Marco V, Ricciardi A, Concardi M, Romagnoli E, et al. Identification and Quantification of Macrophage Presence in Coronary Atherosclerotic Plaques by Optical Coherence Tomography. *Eur Heart J Cardiovasc Imaging.* 2015;16(7):807-13. doi: 10.1093/ehjci/jeu307.
15. Amano H, Koizumi M, Okubo R, Yabe T, Watanabe I, Saito D, et al. Comparison of Coronary Intimal Plaques by Optical Coherence Tomography in Arteries With Versus Without Internal Running Vasa Vasorum. *Am J Cardiol.* 2017;119:1512-17. doi: 10.1016/j.amjcard.2017.02.025.
16. Prati F, Jenkins MW, DiGiorgio A, Rollins AM. Intracoronary Optical Coherence Tomography, Basic Theory and Image Acquisition Techniques. *Int J Cardiovasc Imaging.* 2011;27(2):251-8. doi: 10.1007/s10554-011-9798-1.
17. Vergallo R, Yonetsu T, Uemura S, Park SJ, Lee S, Kato K, et al. Correlation between Degree of Neointimal Hyperplasia and Incidence and Characteristics of Neointimal Hyperplasia as Assessed by Optical Coherence Tomography. *Am J Cardiol.* 2013;112(9):1315-21. doi: 10.1016/j.amjcard.2013.05.076.
18. Yonetsu T, Kim JS, Kato K, Kim SJ, Xing L, Yeh RW, et al. Comparison of Incidence and Time Course of Neointimal Hyperplasia between Bare Metal Stents and Drug-Eluting Stents Using Optical Coherence Tomography. *Am J Cardiol.* 2012;110(7):933-9. doi: 10.1016/j.amjcard.2012.05.027.
19. Tearney CJ, Regar E, Akasaka T, Adriaenssens T, Barlis P, Bezerra HG, et al. Consensus Standards for Acquisition, Measurement, and Reporting of Intravascular Optical Coherence Tomography Studies: A Report from the International Working Group for Intravascular Optical Coherence Tomography Standardization and Validation. *J Am Coll Cardiol.* 2012;59(12):1058-72. doi: 10.1016/j.jacc.2011.09.079.
20. Tahara S, Morooka T, Wang Z, Bezerra HG, Rollins AM, Simon DI, et al. Intravascular Optical Coherence Tomography Detection of Atherosclerosis and Inflammation in Murine Aorta. *Arterioscler Thromb Vasc Biol.* 2012;32(5):1150-7. doi: 10.1161/ATVBAHA.111.243626.
21. Takano M, Yamamoto M, Inami S, Murakami D, Ohba T, Seino Y, et al. Appearance of Lipid-Laden Intima and Neovascularization after Implantation of Bare-Metal Stents Extended Late-Phase Observation by Intracoronary Optical Coherence Tomography. *J Am Coll Cardiol.* 2009;55(1):26-32. doi: 10.1016/j.jacc.2009.08.032.
22. Russo RJ, Silva PD, Teirstein PS, Attubato MJ, Davidson CJ, DeFranco AC, et al. A Randomized Controlled Trial of Angiography versus Intravascular Ultrasound-Directed Bare-Metal Coronary Stent Placement (the AVID Trial). *Circ Cardiovasc Interv.* 2009;2(2):113-23. doi: 10.1161/CIRCINTERVENTIONS.108.778647.
23. Inoue K, Abe K, Ando K, Shirai S, Nishiyama K, Nakanishi M, et al. Pathological Analyses of Long-Term Intracoronary Palmaz-Schatz Stenting: Is its Efficacy Permanent? *Cardiovasc Pathol.* 2004;13(2):109-15. doi: 10.1016/S1054-8807(03)00132-7.
24. Joner M, Nakazawa G, Finn AV, Quee SC, Coleman L, Acampado E, et al. Endothelial Cell Recovery between Comparator Polymer-Based Drug-Eluting Stents. *J Am Coll Cardiol.* 2008;52(5):333-42. doi: 10.1016/j.jacc.2008.04.030.
25. Kastrati A, Schömig A, Dietz R, Neumann FJ, Richardt G. Time Course of Restenosis During the First Year after Emergency Coronary Stenting. *Circulation.* 1993;87(5):1498-505. doi: 10.1161/01.cir.87.5.1498.
26. Levine GN, Chodos AP, Loscalzo J. Restenosis Following Coronary Angioplasty: Clinical Presentations and Therapeutic Options. *Clin Cardiol.* 1995;18(12):693-703. doi: 10.1002/clc.4960181203.
27. Walters DL, Harding SA, Walsh CR, Wong P, Pomerantsev E, Jang IK. Acute Coronary Syndrome is a Common Clinical Presentation of In-Stent Restenosis. *Am J Cardiol.* 2002;89(5):491-4. doi: 10.1016/s0002-9149(01)02285-8.
28. Park SJ, Kang SJ, Virmani R, Nakano M, Ueda Y. In-Stent Neointimal Hyperplasia: A Final Common Pathway of Late Stent Failure. *J Am Coll Cardiol.* 2012;59(23):2051-7. doi: 10.1016/j.jacc.2011.10.909.
29. Yonetsu T, Kato K, Kim SJ, Xing L, Jia H, McNulty I, et al. Predictors for Neointimal Hyperplasia: A Retrospective Observational Study from the Optical Coherence Tomography Registry. *Circ Cardiovasc Imaging.* 2012;5(5):660-6. doi: 10.1161/CIRCIMAGING.112.976167.
30. Lee SY, Hur SH, Lee SC, Kim SW, Shin DH, Kim JS, et al. Optical Coherence Tomographic Observation of In-Stent Neointimal Hyperplasia in Lesions with More than 50% Neointimal Area Stenosis after Second-Generation Drug-Eluting Stent Implantation. *Circ Cardiovasc Interv.* 2015;8(2):e001878. doi: 10.1161/CIRCINTERVENTIONS.114.001878.
31. Tsujita K, Sugiyama S, Sumida H, Shimomura H, Yamashita T, Yamanaga K, et al. Impact of Dual Lipid-Lowering Strategy with Ezetimibe and Atorvastatin on Coronary Plaque Regression in Patients with Percutaneous Coronary Intervention: The Multicenter Randomized Controlled PRECISE-IVUS Trial. *J Am Coll Cardiol.* 2015;66(5):495-507. doi: 10.1016/j.jacc.2015.05.065.

32. Hashikata T, Ikutomi M, Jimba T, Shindo A, Kakuda N, Katsushika S, et al. Empagliflozin Attenuates Neointimal Hyperplasia after Drug-Eluting-Stent Implantation in Patients with Type 2 Diabetes. *Heart Vessels*. 2020;35(10):1378-1389. doi: 10.1007/s00380-020-01621-0.
33. Nicholls SJ, Kataoka Y, Niessen SE, Prati F, Windecker S, Puri R et al. Effect of evolucumab on changes in coronary plaque phenotype and burden in statin-treated patients following myocardial infarction. *J Am Coll Cardiol Img*. Mar16,2022.Epublished DOI: 10.1016/j.jcmg.2022.03.002.
34. Nicol P, Bulin A, Castellanos MI, Stöger M, Obermeier S, Lewerich J et al. Preclinical investigation of neoatherosclerosis in magnesium-based bioresorbable scaffolds versus thick-strut drug-eluting stents. *EuroIntervention*. 2020;16:e922–29.
35. Santulli G, Wronska A, Uryu K, Diacovo TG, Gao M, Marx SO et al. A selective microRNA-based strategy inhibits restenosis while preserving endothelial function. *J Clin Invest*.2014;124:4102–114.



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