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Row orientation effects on chemical composition and aromatic profile of Syrah winter wines

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Abstract

Sunlight and heating influence leaf and grape metabolism and therefore wine quality. As a recent management tool, no information exists on the effects of grapevine row orientation on the wine composition of Syrah vines within the context of double pruning management, a technique used to transfer the grape harvest from the wet summer to the dry winter season. This is a first attempt to investigate the wine composition from north-south- (NS) and east-west- (EW) oriented Syrah winter vines. EW wine samples had higher total acidity, residual sugars, alcohol and color hue, whereas NS wines exhibited higher content of color intensity, anthocyanins, total phenolics, total phenolic index, ashes and pH. The identification of volatile compounds was tentatively performed and demonstrated the presence of alkanes, volatile phenols and alkyl sulfide in NS wines, while butyrolactone and beta-damascenone were found mainly in EW wines. Row orientation contributed to wine composition and could be used as a management tool for obtaining individual wine styles.

Keywords: Vitis vinifera; double pruning management; phenolic compounds; aroma; quality.

Practical Application: The NS row orientation of vineyards is preferable to promote the sunlight exposure of bunches and therefore increase grapevine vigor and the phenolic maturity of the grapes. Knowledge about the impact of row orientation effects on winter wines may help viticulturists to determine the best orientation of their vineyards regarding topography limitations and desired wine styles.

1 Introduction

The concept of winter wines is relatively new in Brazil. The first vineyard managed with the double pruning technique was introduced in the coffee region of Três Corações in 2001 (Amorim et al., 2005). Under this management, the grapevines are first spur pruned at the end of winter (August or September) to develop the vegetative cycle with the removal of all clusters. The reproductive cycle then commences after the second spur pruning, realized in January (or February), to allow for grape harvesting during the dry season of winter (July or August), which improves wine grape quality (Mota et al., 2011; Favero et al., 2011; Regina et al., 2011; Pedro et al., 2017).

Vine growth, yield, and grape and wine quality attributes are affected by solar radiation and temperature regimes throughout the growing season (Bergqvist et al., 2001; Bertamini & Nedunchezhian, 2004; Jogaiah et al., 2012; Chaves et al., 2016). In general, most of the vineyards around the world are north-south (NS) oriented. NS rows, by receiving morning sun on one side and afternoon sun on the other, are better positioned to maximize light interception compared to east-west (EW) rows (Hunter et al., 2016; Campos et al., 2017). On the other hand, EW-oriented rows can capture the largest portion of total radiation in the cluster zone from soil-reflected radiation, and the leaves of EW-oriented vines can also display higher CO₂ assimilation, stomatal conductance and transpiration than those with a NW-SE orientation, as demonstrated by Grifoni et al. (2008) and Hunter et al. (2016). However, the reduced light interception in an EW row orientation may also have a negative impact on growth and yield compared to an NS direction (Chorti et al., 2018; Souza et al., 2019). Hunter & Volschenk (2018) observed that some wine sensory descriptors had lower scores for the EW row orientation in comparison with NS wines.

Although the choice of row orientation is mainly based on the best sunlight interception by the vine canopies, in some vineyard sites the topography and erosion potential should also be taken into account. To gain further knowledge on the double pruning management technique, this preliminary study aims to investigate the effects of row orientation on Syrah winter wine composition.

2 Materials and methods

The experiment was carried out in 2016 in a non-irrigated commercial vineyard located in Andradas (22°04' S 46°34' W, altitude of 920 m), south of Minas Gerais State, Brazil. Two adjacent vineyard blocks were north-south (NS) and east-west (EW) oriented and planted in 2007 using 'Syrah,' clone 174 ENTAV-INRA, grafted onto 1103 Paulsen. Each treatment consisted of 200 vines spaced 2.5×1.0 m apart, trained on a vertical shoot position and spur pruned with two spurs node (approximately 22 buds per vine) on a bilateral Royat Cordon. Double pruning management was applied to allow for grape harvesting during the winter, according to Favero et al. (2011). The first pruning to induce the vegetative cycle was performed in September 2015 in lignified shoots, and all bunches were removed at the bunch closure stage. In March 2016, the yield pruning was conducted in lignified shoots to promote the productive cycle during the autumn-winter season.

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Grapes were harvested with a mean soluble solids content of 21.5 °Brix, pH 3.50 and titratable acidity 5.50 g L⁻¹ tartaric acid. Harvested bunches from the two experimental sites were delivered at the winery and stored at 4 °C for 24 h. For each treatment, two replications of 10 kg of grape clusters were destemmed, crushed and placed in two 13.25 L Pyrex® glass carboys. The musts were inoculated with rehydrated wine yeast *Saccharomyces cerevisiae* × *S. kudriavzevii* (Maurivin®, AWRI 796, AB Biotek), and 80 mg SO, kg⁻¹ was added.

Wine density was determined daily during alcoholic fermentation at 21 °C. When the density reached approximately 990 g L¹, the wines were transferred to 5 L glass carboys for malolactic fermentation that was carried out at 21 °C, without lactic bacteria inoculation, until malic acid was not detected by the paper chromatography method (Amerine & Ough, 1980). The wines were racked to remove lees, treated with potassium metabisulfite (35 mg SO $_2$ L¹) and kept at 3 °C for 15 days to allow tartaric stabilization.

2.1 Wine composition

Physicochemical analyses consisted of alcohol, titratable acidity (g L $^{-1}$ tartaric acid), volatile acidity (g L $^{-1}$ acetic acid), pH, residual sugars (g L $^{-1}$), dry extract (g L $^{-1}$) and ashes (g L $^{-1}$) (Amerine & Ough, 1980). Furthermore, color intensity (CI) (A $_{420}$ +A $_{520}$ +A $_{620}$), color hue (A $_{420}$ /A $_{520}$), total phenolic index (TPI 280 nm) and polymerized pigments (A $_{420}$ and A $_{520}$ with water and SO $_2$) were evaluated by spectrophotometry, and total flavanoid content was evaluated by Bate-Smith reaction (Ribéreau-Gayon et al., 2006). Finally, anthocyanins and phenolics were measured by the pH differential method and Folin-Ciocalteau method, respectively (Amerine & Ough, 1980; Giusti & Wrolstad, 2000). Analyses were performed in triplicate of each glass carboy at bottling.

2.2 Volatile extraction and analysis

For the isolation and concentration of volatiles, the headspace solid-phase microextraction technique (HS-SPME) was used according with Gürbüz et al. (2006) with some modifications. All extractions were carried out using a DVB/CAR/PDMS fiber with a film thickness of $50/30~\mu m$ (Supelco, Bellefonte, PA, USA).

An aliquot of 10 g of wine was placed in 20 mL vials closed with a Teflon cap and stored at -20 °C. All samples were prepared in triplicate. Vials were unfrozen at room temperature and then heated to 30 °C under agitation with a magnetic stir bar for 10 min for headspace equilibrium. The adsorption time was 45 min at the same temperature. The SPME fiber was then injected directly into a gas chromatograph mass spectrometer (Agilent Technologies Inc., Santa Clara, USA) operating with ChemStation software. The SPME fiber was held for 10 min at 250 °C for desorption of volatile compounds, which were separated using a capillary column HP-5MS (30 m \times 0.25 mm \times 0.25 μ m) with helium as the carrier gas at a constant flow of 1 mL min⁻¹. The initial oven temperature was set to 40 °C, held for 5 min, then increased to 160 °C at 3 °C min⁻¹ and to 250 °C at 10 °C min⁻¹ and held for 10 min before returning to 40 °C, in a total cycle of 64 min with a transfer line temperature at 250 °C and the MS detector in SCAN mode 30-500 m/z.

Volatile compounds were tentatively identified by comparison with the National Institute of Standards and Technology (NIST) library (NIST 11, version 2.0, Gaithersburg, USA) considering a 70% similarity to the cut-off, further confirming the results with the retention indexes calculated according to the Kovats Index and compared to reported data on the Nist Webbook (National Institute of Standards and Technology, 2020), ChemSpider (2020) or PubChem (National Center for Biotechnology Information, 2020) websites. Relative odors were found on the Good Scents site (The Good Scents Company Information System, 2020). However, only aromatic compounds with a difference in Kovats retention indices lower than 50 units up or down were accepted.

2.3 Statistical analysis

All data sets were subjected to analyses of variance (ANOVAs). Tukey's HSD tests were carried out to determine differences between treatment means, using the SAEG software (ver. 9.1, UFV, Viçosa, Brazil). Moreover, a principal component analysis (PCA) was performed on the chemical composition of the wines to investigate the differences between NS and EW row orientations using the MetaboAnalyst program (MetaboAnalyst, 2020).

3 Results and discussion

Table 1 reports the results of chemical analysis carried out on the wines at bottling from NS- and EW-oriented vines.

Considering the means of each parameter for both treatments, data on winter wines' composition resemble that of Syrah wines from traditional regions such as Italy (Condurso et al., 2016), South Africa (Hunter & Volschenk, 2018), California (Pinnell & Kurtural, 2012; Brillante et al., 2018), Spain (Gutiérrez et al., 2005; Gil et al., 2013) and Australia (Antalick et al., 2015). This confirms the great potential of the double pruning technique for Brazilian viticulture.

Table 1. Physicochemical parameters of Syrah winter wine from north/south (NS) and east/west (EW) oriented vines.

Parameter	NS	EW	Tukey 5%*
Alcohol (% vol)	12.86 b	13.48 a	0.191
Residual sugar (g L-1)	2.60 b	2.94 a	0.312
Dry extract (g L-1)	27.65 b	28.16 a	0.432
Ashes (g L-1)	3.42 a	3.23 b	0.119
pH	4.03 a	3.98 b	0.020
Total acidity (g L-1)	5.90 b	6.09 a	0.164
Volatile acidity (g L-1)	0.55 a	0.51 b	0.012
Total phenolic index (TPI)	52.30 a	49.47 b	2.184
Total phenolics (mg L-1)	1,664.48	1,609.01	0.096 ns
total flavanols (mg L-1)	1,713.93	1,762.25	0.091 ns
Total anthocyanins (mg L-1)	458.31 a	440.60 b	17.65
Color intensity ($A_{420}+A_{520}+A_{620}$)	13.78 a	12.37 b	0.256
Color hue (A ₄₂₀ /A ₅₂₀)	0.66 b	0.68 a	0.010
Polymerized pigments (%)	37.76	37.42	1.031 ns

Average values of three replicates from each glass carboy. *Significant difference; ns = not statistically significant; same letter in row do not differ significantly between treatments as determined by Tukey's test (p < 0.05).

Furthermore, the PCA performed on the wine chemical parameters indicated a clear separation between treatments, with titratable acidity, residual sugars, alcohol and color hue with higher intensity on EW wine samples, whereas NS wines demonstrated higher content of color intensity, anthocyanins, total phenolics, total phenolic index, ashes and pH (Figure 1).

The influence of row orientation on alcoholic strength is not clear. Chorti et al. (2018) and Hunter & Volschenk (2018) did not find a significant difference between the alcohol content in NS and EW row orientations, although Jogaiah et al. (2012) and Souza et al. (2019) observed a slight increase in the soluble solids content in berries from an EW row orientation. Jogaiah et al. (2012) also found a higher potassium content in the must from an NS row, which could explain higher pH and ashes content in wines from NS-oriented vines.

In addition, Chorti et al. (2018) reported a lower anthocyanin content in wines from EW-oriented rows and higher color and phenolic content values from NS-oriented rows. However, as also observed by Jogaiah et al. (2012), the flavonol concentration did not exhibit differences between treatments.

In EW-oriented rows, the sun passes along the edges of the vine for most of the season, and bunches are less exposed to direct solar radiation. In contrast, in an NS orientation, bunches are exposed to sunlight in the morning on the east side of the canopy and in the afternoon on the west side, thereby ensuring direct sun exposure of the bunches at least in one period during the day. With this orientation, however, the east and west sides of the canopy do not receive the same quantity of radiation, and berries at the west side are exposed to higher temperatures (Hunter & Volschenk, 2018).

The wine aromatic profile of winter-harvested vines was also influenced by row orientation. The PCA plots demonstrate a clear separation between treatments (Figure 2) of Syrah vineyards at winter harvest, with NS samples located on the positive side of the PC1 and EW samples situated on the negative site.

Among the volatile compounds tentatively identified, esters represent the main class, followed by benzenes and alcohols (Table 2).

Wines were characterized by a greater amount of ester compounds. As mentioned by Ilc et al. (2016), esters are mainly secondary metabolites produced during fermentation from alcohol and acyl-CoA by yeast alcohol acyltransferase enzymes, and they contribute to fruity notes. Aliphatic acids, such as hexanoic acid, n-decanoic acid and octanoic acid, are also formed during fermentation as well as volatile phenols, stored in grapes as glycosides and hydrolyzed during winemaking (Dunlevy et al., 2009; González-Barreiro et al., 2015; Ilc et al., 2016).

The aromatic compounds that distinguished wines from an NS row orientation the most were hexadecane, ortho-cresol and guaiacol, ethyl salicylate and naphthalene, ethyl decanoate and hexanoic acid, methyl ester, while the EW row orientation pointed out with p-xylene, butyrolactone, propyl decanoate, nonanoic acid, ethyl ester and 2-ethylhexyl salicylate (Figure 3).

NS wines pointed out with the presence of alkanes, volatile phenols and alkyl sulfide, while butyrolactone and beta-damascenone

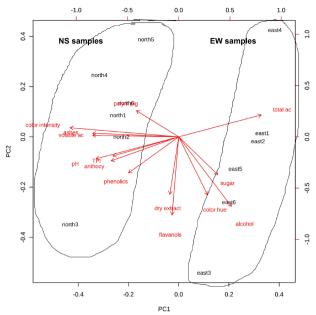


Figure 1. Principal component analysis (biplot graph) of chemical compounds of Syrah winter wines from north/south (NS) and east/west (EW) oriented vines. Principal component 1 (PC1) and PC2 account for 51.5% and 20.2% of the total variation in the dataset, respectively.

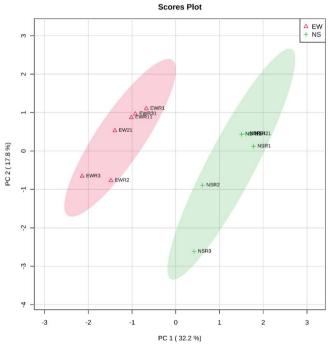


Figure 2. PCA plots for winter wine volatile compounds of Syrah vineyards under north/south (NS) and east/west (EW) orientation. Principal component 1 (PC1) and PC2 account for 32.2% and 17.8% of the total variation in the dataset, respectively.

were found mainly in EW wines. Despite the differences, both treatments displayed aromatic compounds with sweet and fruity flavors, with a possible phenolic, spicy and resinous note at the NS orientation and buttery and green nuances at the EW row. Future sensory evaluation, however, is necessary to evaluate consumers' perceptions of these differences.

Table 2. Aromatic volatile compounds tentatively identified in winter wines of the Syrah cultivar under north/south (NS) and east/west (EW) orientation.

compound	code	CASc	Kovatsa	Odor ^b
Acids				
Hexanoic acid	AJ	142-62-1	994	Rancid, sour, sharp, pungent, cheesy, fatty
n-Decanoic acid	AN	334-48-5	1375	Unpleasant rancid, sour, fatty, citrus, soapy
Octanoic acid	AH	124-07-2	1191	Fruity-acid
Alkanes				
Hexadecane	AP	544-76-3	1600	Mild wax
Alcohols				
1-Decanol	X	112-30-1	1275	Sweet, fat-like
1-Hexanol	U	111-27-3	875	Herbaceous, woody, sweet, green fruit, banana, flower, grass
1-Octanol	W	111-87-5	1068	Fresh, orange-rose, sweet, bitter almond, burnt matches, fat, floral
Cis-3-hexen-1-ol	AT	928-96-1	858	Grassy-green, herbaceous, leafy
Phenylethyl alcohol	A	60-12-8	1113	Floral, rose, dried rose, flower, rose water
Aldehydes				
Benzeneacetaldehyde	AB	122-78-1	1047/1046	Harsh, green
Decanal	Y	112-31-2	1209/1207	Sweet, waxy, flowery, citrus, fatty
Nonanal	AI	124-19-6	1106	Fatty, citrus-like
Benzenes				
Benzaldehyde	Н	100-52-7	964	Bitter almond
Benzyl alcohol	G	100-51-6	1039/1037	Fruity, pungent
Ethyl salicylate	AA	118-61-6	1271	Sweet, wintergreen, mint, floral, spicy, balsam
Naphthalene	С	91-20-3	1179	Pungent, resinous
p-Xylene	P	106-42-3	871	Sweet
Styrene	F	100-42-5	892	Sweet, balsam, floral, plastic
Ketones				
(E)-Beta-damascenone	BB	23726-93-4	1385	Apple, rose, honey, tobacco, sweet
Alkyl sulfide				
1-propanol, 3-(methylthio)-	AO	505-10-2	983	Sulfurous, onion, sweet, soup, vegetable
Ester				
1-Butanol, 3-methyl-, acetate	AF	123-92-2	881	Fruity – banana, pear, apple, glue
2-Ethylhexyl salicylate	Z	118-60-5	1805	Mild, orchid, sweet, balsam
2-Hexenoic acid, ethyl ester	AU	1552-67-6	1051	Fruity – pineapple, apple, green
Acetic acid, hexyl ester	AK	142-92-7	1019	Fruity – apple, cherry, pear
n-Caprylic acid isobutyl ester	AZ	5461-06-3	1351	Fruity, green, oily, floral
Diethyl succinate	AC	123-25-1	1186	Mild, fruity, cooked, apple, ylang
Ethyl butyrate	J	105-54-4	800	Fruity, juicy, pineapple, cognac
Ethyl crotonate	BA	6776-19-8	854	Found in alcoholic beverages. Component of strawberry aroma, guava
•				fruit, pineapple, yellow passion fruit
Ethyl decanoate	R	110-38-3	1400/1397	Sweet, waxy, fruity, apple, grape, oily, brandy
Ethyl 9-decenoate	BF	67233-91-4	1390	Fruity, fatty
Ethyl heptanoate	M	106-30-9	1101	Fruity, pineapple, cognac, rum, wine
Ethyl laurate	O	106-33-2	1596	Sweet, waxy, floral, soapy, clean
Ethyl palmitate	AS	628-97-7	1917	Mild, waxy, fruity, creamy, milky, balsam
Hexanoic acid, ethyl ester	AE	123-66-0	1003	Fruity – pineapple, banana
Hexanoic acid, methyl ester	Q	106-70-7	933	Ether-like
Hexanoic acid, 2-methylbutyl ester	AY	2601-13-0	1257	Ethereal
Isoamyl decanoate	AX	2306-91-4	1649	Waxy, banana, fruity, sweet, cognac, green
Isoamyl octanoate	AV	2035-99-6	1449	Sweet, oily, fruity, green, soapy, pineapple, coconut
Isobutyl hexanoate	K	105-79-3	1156	Sweet, fruity, pineapple, green, peach, tropical
Isopentyl hexanoate	AW	2198-61-0	1254	Fruity, banana, apple, pineapple, green
Methyl decanoate	S	110-42-9	1327	Oily, wine, fruity, floral
Methyl laurate	V	111-82-0	1526	Waxy, soapy, creamy, coconut, mushroom
Nonanoic acid, ethyl ester	AD	123-29-5	1298	Fruity, rose, waxy, rum, wine, natural, tropical

^aLinear retention indices calculated on capillary HP-5MS column according to Kovats equation. Data were considered within the mean ± 50 units respect to those reported on Nist, Chemspider or PubChem websites; ^bOdors extracted from Good Scents (The Good Scents Company Information System, 2020) or PubChem (National Center for Biotechnology Information, 2020) websites. ^cCAS number: unique numeric identifier of a chemical substance in the Chemical Abstracts Service (CAS), a division of the American Chemical Society.

Table 2. Continued...

compound	code	CASc	Kovatsa	Odor ^b
Octanoic acid, ethyl ester	N	106-32-1	1202/1199	Fruity, wine, waxy, sweet, apricot, banana, brandy, pear
Octanoic acid, methyl ester	T	111-11-5	1128	Winy, fruity – orange, oily
Pentadecanoic acid, ethyl ester	BE	41114-00-5	1872	Honey, sweet
Phenethyl acetate	I	103-45-7	1259	Floral, rose, sweet, honey, fruity, tropical
Propyl decanoate	BC	30673-60-0	1492	Waxy, fruity, fatty, green, vegetable, woody, oily
Propyl octanoate	AQ	624-13-5	1294	Coconut, caco, gin
Tetradecanoic acid, ethyl ester	AG	124-06-1	1796	Sweet, waxy, violet, orris
Undecanoic acid, ethyl ester	AR	627-90-7	1496	Soapy, waxy, fatty, cognac, coconut
Volatile phenols				
Ortho-cresol	D	95-48-7	1061	Musty, phenolic, plastic, medicinal, herbal, leathery
Phenol, 2-methoxy-	В	90-05-1	1090	Phenolic, smoke, spice, vanilla woody
Lactones				
Butyrolactone	E	96-48-0	918	Sweet, aromatic, buttery, creamy, oily, fatty, caramel
Monoterpenes				
Citronellol	L	106-22-9	1232	Flowery - rose
Citronellyl acetate	AM	150-84-5	1356	Floral, green, rose, fruity, citrus, wood, tropical fruit
Theaspirane	BD	36431-72-8	1296	Tea, herbal, green, wet, tobacco, leaf, metallic, woody, spicy

^aLinear retention indices calculated on capillary HP-5MS column according to Kovats equation. Data were considered within the mean ± 50 units respect to those reported on Nist, Chemspider or PubChem websites; ^bOdors extracted from Good Scents (The Good Scents Company Information System, 2020) or PubChem (National Center for Biotechnology Information, 2020) websites. ^cCAS number: unique numeric identifier of a chemical substance in the Chemical Abstracts Service (CAS), a division of the American Chemical Society.

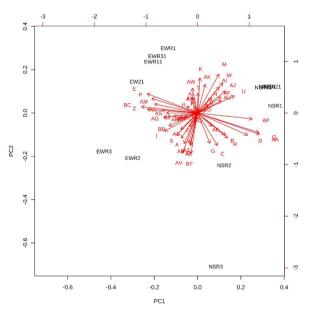


Figure 3. Principal component analysis (biplot graph) of volatile compounds for Syrah winter wines from vines under north/south (NS) and east/west (EW) orientation. Principal component 1 (PC1) and PC2 account for 32.2% and 17.8% of the total variation in the dataset, respectively.

4 Conclusion

Row orientation impacts grape and wine quality from winter harvests: NS-oriented vines resulted in wines with higher content of color intensity, anthocyanins, total phenolics, ashes and pH. Furthermore, the identification of volatile compounds also revealed differences between treatments. However, the composition of both treatments did not depreciate the quality of the wine, indicating that row orientation could be used as a management tool for obtaining different wine styles.

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