

Suzuki-Miyaura Coupling between 3-Iodolawsone and Arylboronic Acids. Synthesis of Lapachol Analogues with Antineoplastic and Antileishmanial Activities

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A series of 2-hydroxy-3-arylnaphthalene-1,4-diones (3-aryllawsones) were synthesized by Suzuki-Miyaura cross coupling reaction of 3-iodolawsone with arylboronic acids/esters. The hydroxylated resulting products were transformed into their corresponding *N,N*-diethyl carbamates. The antineoplastic and antileishmanial activities of the compounds were evaluated and compared with lapachol and its carbamate, providing promising results.

Keywords: Suzuki-Miyaura coupling, lapachol analogues, pd-catalysis

Introduction

Quinones are present in many naturally occurring compounds, and are responsible for taking part in the life cycle of many living organisms.¹ They are well known for being significant source of biologically active compounds.² For example, the benzoquinone mitomycin C³ and the anthraquinone doxorubicin⁴ are relevant members of this family, being used in clinic as antineoplastic agents (Figure 1).

Among the quinone family, naphthalene-1,4-diones is the most important class with many representatives such as synthetic atovaquone,⁵ that is used in anti-malarial therapy and chemoprophylaxis of malaria; the naturally occurring 2-hydroxy-3-prenylnaphthalene-1,4-dione lapachol (**1**), first isolated by E. Paterno from *Tabebuia avellanedae* in 1882 exhibiting antimalarial, antifungal, antitumor,

leishmanicidal, bactericidal and antiparasitic activities;⁶ and the antibiotic WS-5995-C,⁷ a functionalized naphthalene-1,4-dione isolated from *Streptomyces auranticolor* species, that possess chemoprotective activity against *Eimeria tenella* (Figure 1). *Eimeria tenella* is a protozoan disease that causes poultry coccidiosis impacting directly in chicken industry and global food supply due to the high mortality.⁸

Lapachol (**1**) has been used as starting point to obtain new bioactive quinones showing interesting pharmacological profiles.^{9,10} Herein, we would like to report the synthesis of 2-hydroxy-3-arylnaphthalene-1,4-diones, analogues of WS-5995-C that can also be considered analogues of **1** in which the prenyl side chain is substituted by an aromatic ring (Figure 1). We rationalized the use of the aromatic ring in a way to mimic the π -system of **1**, knowing that prenyl and phenyl can be considered isosteres.¹¹

Carbamates are widely used in organic synthesis as protecting groups and they have been found in the

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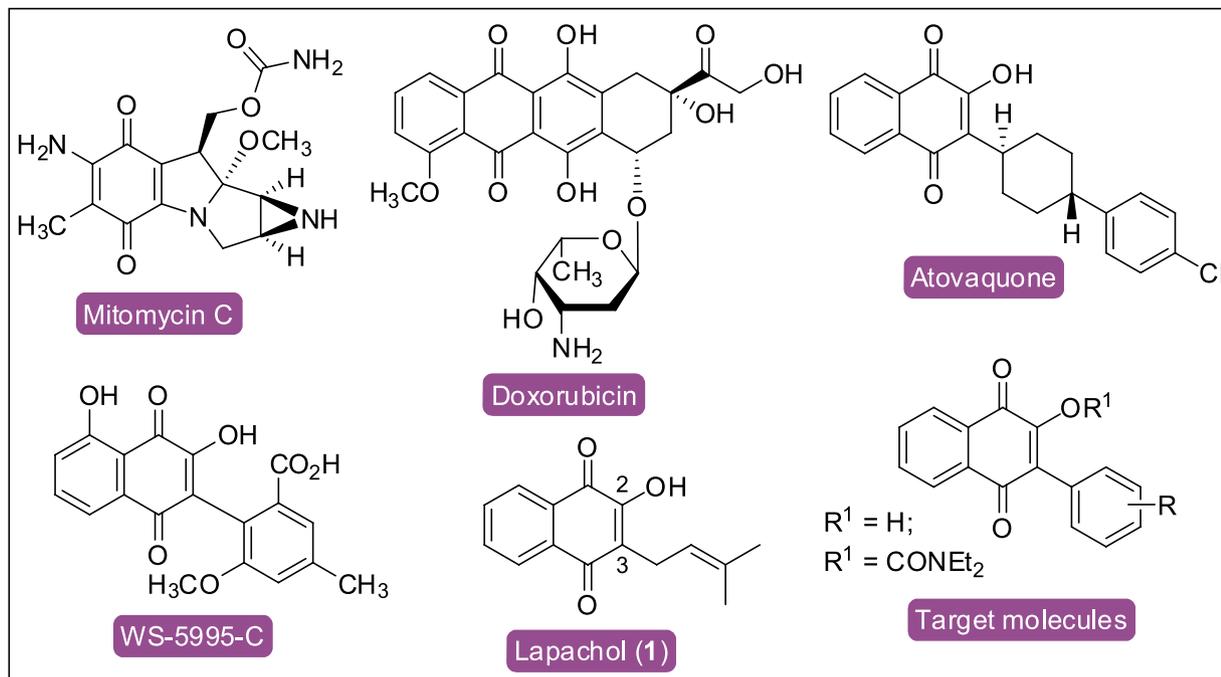


Figure 1. Natural and synthetic compounds from quinone family.

structure of promising antiviral, antifungal, antibacterial, antimicrobial and anticancer molecules.¹² Organic carbamates can act as an electrophilic site, alkylating biological nucleophiles,¹³ having their use also correlated to prodrug design.¹⁴ These precedents prompted us to synthesize new potentially active analogues combining the 2-hydroxy-3-arylnaphthalene-1,4-dione scaffold and the carbamate function, aiming to obtain compounds with dual action.

The antineoplastic and antileishmanial activities of these compounds were evaluated and the results compared with **1**. Cancer cells and protozoan parasites are known to possess several biochemical similarities and to share mutual features such as highly proliferative behavior and resistance to the programmed cell death. Therefore, antineoplastic and antileishmanial agents have common biological targets that act on enzymes inhibition of both parasite and human cells enabling apoptosis.¹⁵

Results and Discussion

Chemistry

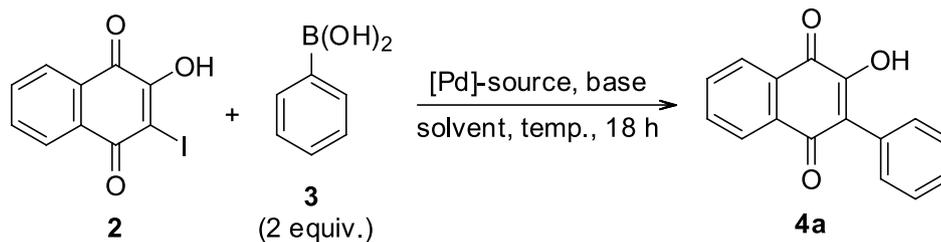
Few general methods are available in literature to prepare 2-hydroxy-3-arylnaphthalene-1,4-diones such as: (i) the palladium catalyzed Stille-type coupling involving phenyliodonium ylides of hydroxyquinones and naphthopyranyl stannanes;¹⁶ (ii) the palladium catalyzed Suzuki-type coupling of phenyliodonium ylides

of hydroxyquinones with arylboronic acids;¹⁷ (iii) the metal free arylation of 2-hydroxy-naphthalene-1,4-diones with phenyliodonium ylides mediated by BF_3 ;¹⁸ (iv) the oxidative arylation of 2-hydroxy-naphthalene-1,4-diones using *o*-iodoxybenzoic acid and phenylhydrazines;¹⁹ and (v) $\text{K}_2\text{S}_2\text{O}_8$ -catalyzed direct C–H functionalization of 2-hydroxy-naphthalene-1,4-diones with arylboronic acids.²⁰ The main disadvantages associated with the described methods involve the use of hypervalent iodines, that confront the atom economy principle of the green chemistry; employment of toxic hydrazines and scope limitation. In this paper we take advantage of the Suzuki-Miyaura cross coupling reaction²¹ to achieve 2-hydroxy-3-arylnaphthalene-1,4-diones structurally related to **1**. After the submission of this paper, Louvis *et al.*²² reported a similar method to synthesize 2-hydroxy-3-arylnaphthalene-1,4-diones.

We began with the preparation of the starting material according to the phenol iodination method previously described by our group, in which 2-hydroxynaphthalene-1,4-dione was easily iodinated by using a morpholine-iodine complex as the iodinating agent.²³

With the starting material in hands, we first examined the reaction of 2-hydroxy-3-iodonaphthalene-1,4-dione **2** with phenylboronic acid **3a** leading to **4a** under the classical Suzuki-Miyaura conditions,²¹ but moderate yield was obtained (Table 1, entry 1).

Taking into consideration that the steric hindrance present in **2** may be a problem, we looked for protocols

Table 1. Optimization of the reaction conditions

entry ^a	[Pd]-source	Base	Solvent	Temperature / °C	Yield ^b / %
1	Pd(PPh ₃) ₄ (5 mol%)	Na ₂ CO ₃	DME	85	54
2	Pd(PPh ₃) ₄ (2 mol%)	K ₃ PO ₄	DMF	110	0
3	10% Pd/C (5 mol%)	K ₂ CO ₃	Dioxane-H ₂ O	95	96
4 ^c	10% Pd/C (5 mol%)	K ₂ CO ₃	Dioxane-H ₂ O	95	80
5	10% Pd/C (10 mol%)	K ₂ CO ₃	Dioxane-H ₂ O	95	89
6	10% Pd/C (1 mol%)	K ₂ CO ₃	Dioxane-H ₂ O	95	85
7 ^d	10% Pd/C (5 mol%)	K ₂ CO ₃	Dioxane-H ₂ O	95	85

^aAll the reactions were carried out under a nitrogen atmosphere and heated overnight; ^bisolated yields; ^creaction carried out using 5,5-dimethyl-2-phenyl-1,3,2-dioxaborinane as the coupling partner; ^dexperiment done with 1.4 equivalents of **3**. DME: 1,2-dimethoxyethane; DMF: *N,N*-dimethylformamide.

that could be better suited on this system, for example, the one reported by Suzuki and Miyaura applying K₃PO₄ as base and DMF (*N,N*-dimethylformamide) as solvent.²⁴ However, no product was detected under these conditions (Table 1, entry 2). When conditions described by Blanchet and co-workers²⁵ were tested, which involves the use of the cheap catalyst 10% Pd/C, **4a** was achieved in almost quantitative yield (Table 1, entry 3). The evaluation of the performance of arylboronic ester 5,5-dimethyl-2-phenyl-1,3,2-dioxaborinane under Blanchet's conditions was also accomplished (Table 1, entry 4).

Further optimization was performed. Increasing the catalyst load from 5 to 10 mol% resulted in a drop of the chemical yield (Table 1, entry 5). When diminishing the catalyst amount to 1 mol%, the yield also suffered a decrease (Table 1, entry 6). Excess of **3** was reduced from 2 equivalents to 1.4 equivalents, but a lower yield was obtained (Table 1, entry 7). In this way, optimal conditions were assumed as being the one described in entry 3.

Aiming to exploit the reactivity of the naphthoquinone system, we protected the free hydroxyl of 2-hydroxy-3-iodonaphthalene-1,4-dione (**2**) using groups with two different electronic demands. Therefore, 2-methoxy-3-iodonaphthalene-1,4-dione (**5**) and 3-iodo-1,4-dioxo-1,4-dihydronaphthalen-2-yl acetate (**6**), were prepared from **2** by the reaction with dimethyl sulfate²⁶ and acetic anhydride,²⁷ respectively. These compounds were then submitted to the optimized conditions described above and on both reactions we observed that a loss of the protecting group had occurred. Performing the reaction with **5**,

compound **4a** was provided in 47% yield; while the reaction with **6** led to 35% yield of **4a**.

Next, we examined the scope of this transformation and the results are presented in Table 2.

Moderate yields were obtained when (3,4-dimethoxyphenyl)boronic acid and (4-formylphenyl) boronic acid were employed (**4b** and **4e**). Aryl boronates containing fluoro substituents furnished good yields (**4c** and **4d**), however, aromatic boronates *ortho*-substituted were not tolerated on this process, probably due to steric effects (**4f-g**). From these results we found that the best substrate was the one with no substitution pattern (**4a**).

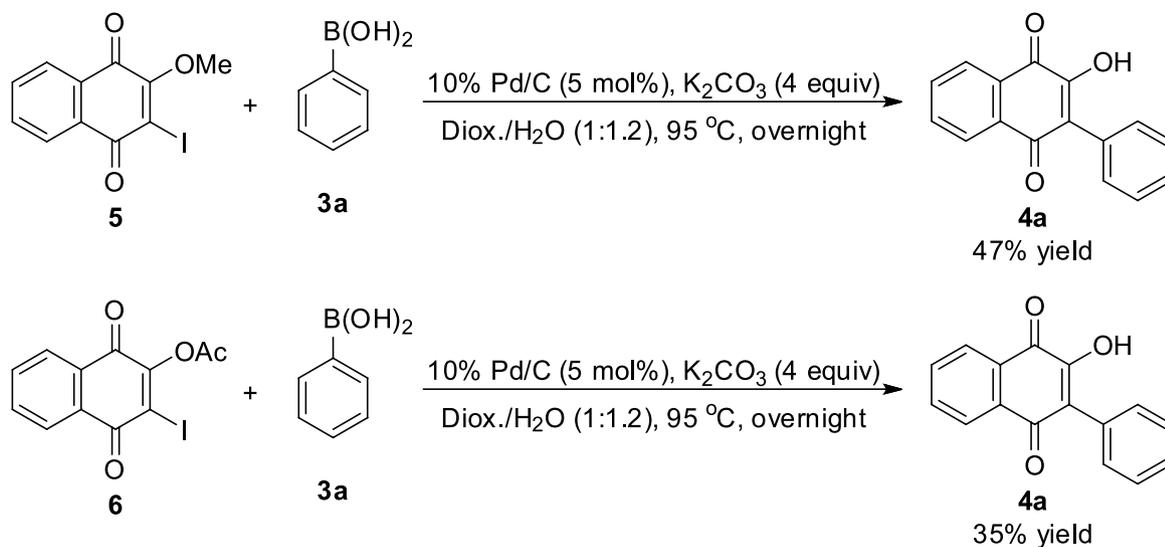
Continuing the work, lapachol (**1**) and analogs **4a-e** were transformed into their corresponding *N,N*-diethyl carbamates **8** and **7a-e**, respectively, according to the well-known protocol described in literature,²⁸ furnishing moderate yields (Table 3).

With lapachol analogs in hands, we evaluated their pharmacological profiles in relation to antineoplastic and antileishmanial activities.

Antineoplastic activity

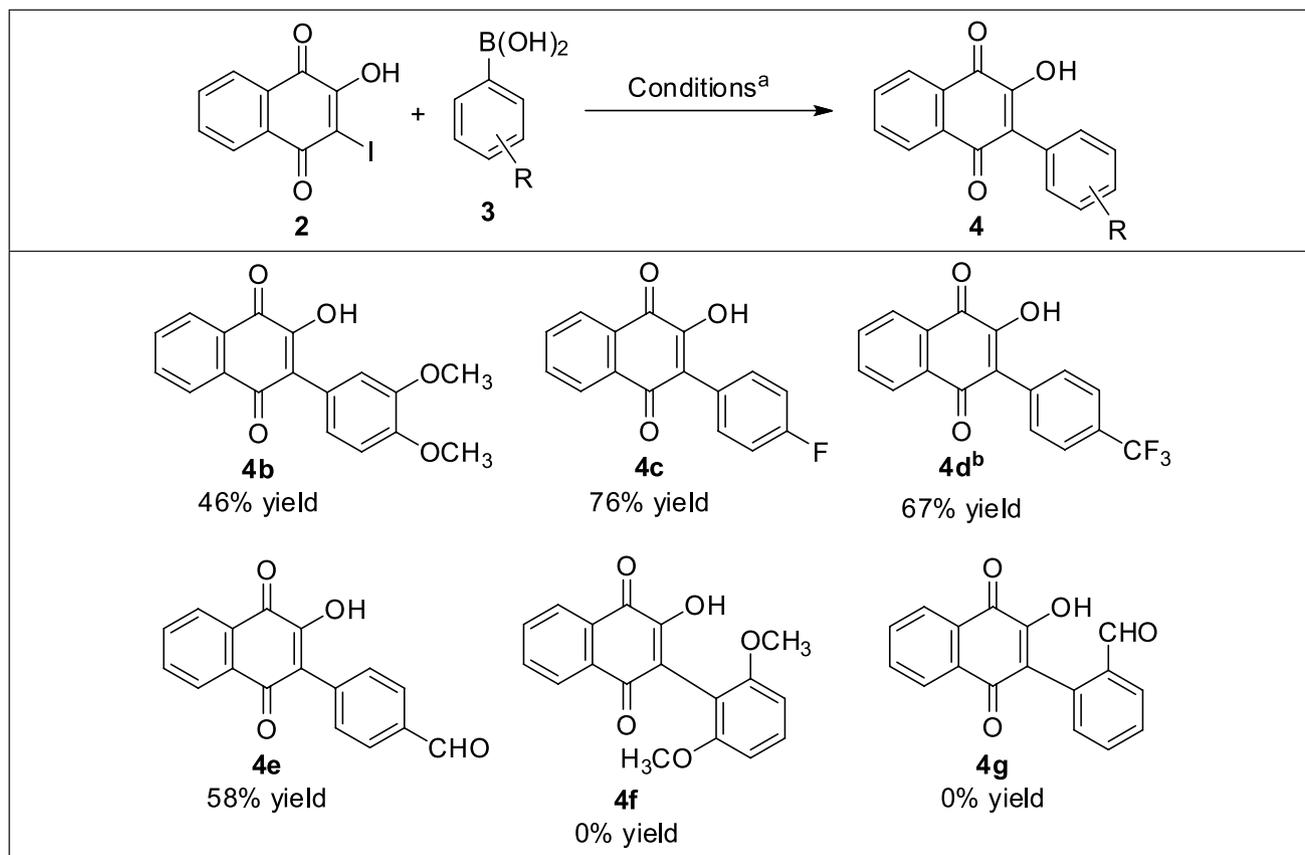
The antineoplastic activity of the prepared naphthoquinones was studied in two human leukemia cell lines, HL-60 (promyelocytic leukemia) and K562 (chronic myelogenous leukemia),²⁹ and the results are shown in Table 4.

Lapachol (**1**) presented moderate potency against these cell lines (entry 1) and the exchange of the allyl group



Scheme 1. Analyzing the reactivity of the naphthoquinone system.

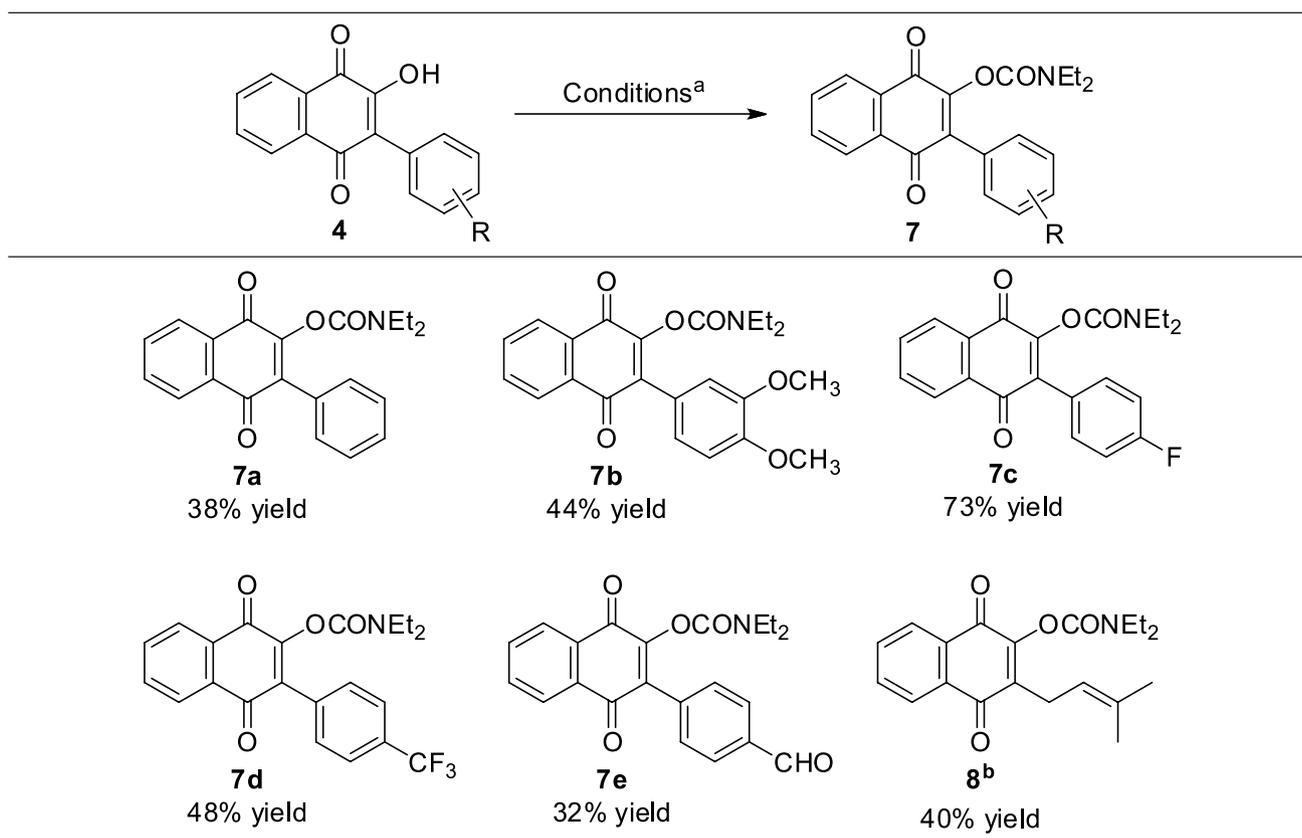
Table 2. Scope of the reaction



^aReaction conditions: **3** (2 equiv.), 10% Pd/C (5 mol%), K₂CO₃ (4 equiv.), dioxane/H₂O (1:1.2), 95 °C, 18 h, carried under a nitrogen atmosphere; ^b5,5-dimethyl-2-(4-(trifluoromethyl)phenyl)-1,3,2-dioxaborinane was used as the coupling partner.

in **1** by aryl groups, as in **4a-c** and **4e**, led to products still less active (entries 2-4 and 6). Interestingly, **4d** bearing a *p*-CF₃Ph substituent, was six to seven times more potent than **1** for both cell lines (entry 5).

The results obtained with carbamates **8** and **7a-e** are shown in entries 7-12. For HL-60, **8** was still less potent while in K562 the carbamate of lapachol exhibited moderate potency (entry 7). The corresponding carbamates

Table 3. Carbamates synthesis

^aReaction conditions: ClCONEt₂ (20 equiv.), Py (5 equiv.), CHCl₃, 60 °C, 18 h; ^blapachol was used as the starting material. Py: pyridine.

Table 4. Antineoplastic activity of compounds **1**, **4a-e**, **7a-e** and **8** in HL-60 and K562 (IC₅₀, μM). Doxorubicin (D) was used as reference

entry ^a	Compound	HL-60	K-562
1	1	12.0	16.0
2	4a	> 80	> 80
3	4b	> 80	> 80
4	4c	46.3	> 80
5	4d	2.5	2.2
6	4e	> 80	> 80
7	8	35.7	9.9
8	7a	2.3	5.4
9	7b	4.3	6.1
10	7c	1.9	1.4
11	7d	6.9	11.4
12	7e	2.6	1.4
13	D	0.04	0.14

^aData represent the means of three independent experiments, with each concentration tested in duplicate.

of **4a-c**, compounds **7a-c** (entries 8-10), showed enhanced cytotoxic activity when compared to lapachol **1**. In contrast, for **4d**, the corresponding carbamate **7d** presented a slight lower potency (entry 11), while **7e** was much more potent than **4e** (entry 12).

PBMC (peripheral blood mononuclear cell) tests were performed only with compounds which presented IC₅₀ < 10 μM for tumor cells HL-60 and K562 (Table 5).

The best selectivity indexes were found for compound **4d** (entry 2) followed by compound **7c** (entry 5). In contrast, compound **7d** showed no selectivity toward HL-60 or K562 (entry 6).

Compounds **4d** and **7c** are promising. Studies aiming to establish the probable mechanism of action of these compounds are under investigation.

Antileishmanial activity

In the antileishmanial assay, **1** showed to be moderately active against promastigotes of *L. amazonensis*, but the IC₅₀ in amastigotes was above the highest concentration tested (Table 6, entry 1). The IC₅₀ in intracellular amastigote, which is the most clinically relevant form for treatment, is the criteria to select hit and lead compounds in drug discovery for leishmaniasis.³⁰

The substitution of prenyl for an aromatic ring in **4a-b** led to increased potency against amastigote forms and reduction of toxicity to the macrophages. No significant

Table 5. Cytotoxic effect of compounds **1**, **4d** and **7a-e** tested in PBMC. Doxorubicin (D) was used as positive control

entry	Compound	PBMC	PBMC/HL60 (IC ₅₀ / μM) ^a	PBMC/K562 (IC ₅₀ / μM) ^b
1	1	> 80	ND	ND
2	4d	78	31.2	35.4
3	7a	13.7	5.9	2.5
4	7b	34.0	7.9	5.6
5	7c	24.0	12.6	17.1
6	7d	11.7	1.7	1.02
7	7e	9.4	3.6	6.7
8	D	1.78	44.5	12.7

^aData represent the ratio between PBMC and HL-60 IC₅₀ values. The IC₅₀ values for HL-60 are described in Table 4; ^bdata represent the ratio between PBMC and K562 IC₅₀ values. The IC₅₀ values for K562 are described in Table 4. ND: not determined.

Table 6. Antileishmanial activity of compounds **1**, **4a-e**, **7a-d** and **8** in promastigotes and intracellular amastigotes of *Leishmania amazonensis* and toxicity to murine macrophages. Pentamidine was used as reference

entry ^a	Compound	Promastigote IC ₅₀ / μM	Amastigote IC ₅₀ / μM	Macrophage LD ₅₀ / μM	Selectivity index (SI) ^a
1	1	18.5	> 50	74.3	ND
2	4a	85.8	25.0	115.2	4.6
3	4b	> 100	21.3	152.3	7.15
4	4c	70.5	> 50	90.5	ND
5	4d	25.5	17.2	37.5	2.1
6	4e	> 100	21.5	57.5	2.6
7	8	2.90	4.5	15.1	3.3
8	7a	1.98	3.6	10.2	2.8
9	7b	3.0	1.5	8.0	5.3
10	7c	4.5	1.9	5.2	2.7
11	7d	3.3	2.6	4.9	1.9
12	pentamidine	1.3	0.9	41.5	46.1

^aData represent the ratio between macrophage LD₅₀ and IC₅₀ in amastigote values. ND: not determined.

alteration was observed for promastigote forms (entries 2 and 3). Interestingly, this modification improved the antiamastigote activity, making **4b** the compound with the most favorable selectivity index (SI) (i.e., many times more selective against intracellular amastigotes than host cells; calculated using the LD₅₀/IC₅₀ ratios),³⁰ although presenting moderate potency. Compounds **4c-e** showed moderate activity against amastigotes and increased toxicity to the macrophages (entries 4-6). The addition of the carbamate function in the structure of **1**, originating **8**, potentiated the antileishmanial activity; however the toxicity to macrophages was also increased (entry 7). Carbamates **7a-d**, in which both carbamate and aromatic ring were added to lapachol core, were much more active (entries 8-11). Compound **7b** was the most potent against intracellular amastigotes, with IC₅₀ as low as 1.5 μM and selectivity index of 5.3 (entry 9). These data suggest that

replacing the prenyl side chain of **1** by an aromatic ring is important to enhance the selectivity, while the introduction of the carbamate group is important to enhance the antileishmanial activity.

Conclusions

Despite the number of methods already existing to produce 3-aryllawsones, all of them have considerable drawbacks making it necessary the search for new synthetic tools. We developed a simple approach to achieve 2-hydroxy-3-arylnaphthalene-1,4-dione (**4a-e**) in moderate to good yields, involving the palladium-catalyzed Suzuki-Miyaura cross coupling reaction of arylboronic acids/esters (**3a-e**) and 2-hydroxy-3-iodonaphthalene-1,4-dione (**2**). Unfortunately, *ortho*-substituted boronates were not tolerated.

The corresponding carbamates **7a-e** were also obtained in moderate yields and all synthesized products were submitted to anticancer and antileishmanial evaluation, for the first time, providing promising results. Molecules **4d** and **7c** showed good activity against cancer cell lines and compound **4b** exhibited good antiplasmodial performance. The introduction of the carbamate moiety was important to potentiate the anticancer and antileishmanial activities but also increased the toxicity of the molecules to healthy cells.

Experimental

Chemistry

All solvents and chemicals were used as received from commercial sources. Flash column chromatography was carried out using SiliaFlash® F60 silica gel (particle size: 40-63 μm , 230-400 mesh, Silicycle, Quebec City). Infrared (IR) spectra were performed on a Shimadzu IRPrestige-21 FT-IR instrument via ATR unit or as KBr pellets. ^1H and ^{13}C nuclear magnetic resonance (NMR) spectra were recorded either on Varian 400 MHz or 500 MHz NMR spectrometer using tetramethylsilane (TMS) as reference. The high resolution mass spectra were acquired by LC-MS (liquid chromatography mass spectrometry) in the ESI (+) mode.

General procedure for the preparation of 2-hydroxy-3-iodonaphthalene-1,4-dione (**2**)²³

A mixture of 2-hydroxy-naphthalene-1,4-dione (1 equiv, 5.74 mmol) and K_2CO_3 (3.05 equiv, 17.5 mmol) in H_2O (58 mL) was stirred at room temperature for 10 minutes. After this time, the morpholine-iodine complex²³ (1.95 equiv, 11.2 mmol) was slowly added, in similar portions, every 15 minutes during 2 hours and then the reaction was allowed to stir for 3 more days. The reaction mixture was cooled to 0 $^\circ\text{C}$, acidified with 25% phosphoric acid solution until pH ca. 2 and placed on refrigerator for 24 hours. The resulting precipitate was filtered under vacuum, washed with H_2O and purified by recrystallization from glacial acetic acid. This compound was obtained as a yellow solid in 55% yield, mp 178-180 $^\circ\text{C}$; IR (KBr) ν / cm^{-1} 3200, 1666, 1620, 1581, 1354, 1259, 1116, 721; ^1H NMR (400 MHz, CDCl_3) δ 8.21 (dd, 1H, J 7.0, 1.7 Hz, Ar-H), 8.15 (dd, 1H, J 7.0, 1.7 Hz, Ar-H), 7.76 (m, 2H, 2Ar-H); ^{13}C NMR (101 MHz, $(\text{CD}_3)_2\text{CO}$) δ 180.09, 178.02, 162.29, 135.51, 134.34, 132.18, 130.47, 128.05, 127.37, 92.59.

General procedure for the preparation of 2-hydroxy-3-arylnaphthalene-1,4-diones (**4a-e**)

A stirred solution of **2** (1 equiv, 0.5 mmol), arylboronic acids or esters **3** (2 equiv, 1 mmol), K_2CO_3 (4 equiv, 2 mmol) and Pd/C 10 mol% (5 mol%) in dioxane/ H_2O in proportion of 1:1.2 (11.9 mL) was heated at 95 $^\circ\text{C}$ for 18 hours under nitrogen atmosphere. Dichloromethane (50 mL) was added to the reaction mixture and the organic layer was first washed with a solution of hydrochloric acid 1N (50 mL) and then with brine (3×50 mL), dried over anhydrous Na_2SO_4 and concentrated *in vacuo*. The crude product was purified by flash column chromatography on silica gel.

2-Hydroxy-3-phenylnaphthalene-1,4-dione (**4a**)³¹

After column chromatography using hexanes/EtOAc (95:5) as eluent, this compound was obtained as an orange solid in 96% yield, mp 135-137 $^\circ\text{C}$; IR (KBr) ν / cm^{-1} 3346, 3049, 1655, 1587, 1365, 1331, 1274, 1000; ^1H NMR (500 MHz, CDCl_3) δ 8.21 (dd, 1H, J 7.6, 0.8 Hz, Ar-H), 8.16 (dd, 1H, J 7.6, 0.8 Hz, Ar-H), 7.82 (td, 1H, J 7.5, 1.3 Hz, Ar-H), 7.75 (td, 1H, J 7.5, 1.3 Hz, Ar-H), 7.58 (s, 1H, OH), 7.54-7.50 (m, 2H, 2Ar-H), 7.50-7.45 (m, 2H, 2Ar-H), 7.43-7.39 (m, 1H, Ar-H); ^{13}C NMR (126 MHz, CDCl_3) δ 183.82, 181.92, 152.33, 135.39, 133.25, 132.87, 130.73, 130.03, 129.37, 128.76, 128.02, 127.37, 126.23, 122.22.

2-(3,4-Dimethoxyphenyl)-3-hydroxynaphthalene-1,4-dione (**4b**)³¹

After column chromatography using hexanes/EtOAc (75:25) as eluent, this compound was obtained as a brown solid in 46% yield, mp 174-176 $^\circ\text{C}$; IR (KBr) ν / cm^{-1} 3368, 1653, 1630, 1585, 1518, 1369, 1259, 1148, 1015; ^1H NMR (500 MHz, CDCl_3) δ 8.10 (dd, 1H, J 7.7, 0.8 Hz, Ar-H), 8.13 (dd, 1H, J 7.6, 1.0 Hz, Ar-H), 7.80 (td, 1H, J 7.6, 1.3 Hz, Ar-H), 7.72 (td, 1H, J 7.5, 1.2 Hz, Ar-H), 7.65 (s, 1H, OH), 7.15 (dd, 1H, J 8.3, 2.0 Hz, Ar-H), 7.10 (d, 1H, J 1.9 Hz, Ar-H), 6.96 (d, 1H, J 8.4 Hz, Ar-H), 3.92 (s, 3H, OCH_3), 3.90 (s, 3H, OCH_3); ^{13}C NMR (126 MHz, CDCl_3) δ 184.10, 181.82, 152.05, 149.49, 148.40, 135.30, 133.25, 132.93, 129.41, 127.36, 126.17, 124.07, 122.44, 122.04, 114.06, 110.68, 56.04, 55.98.

2-(4-Fluorophenyl)-3-hydroxynaphthalene-1,4-dione (**4c**)³²

After column chromatography using hexanes/EtOAc (90:10) as eluent, this compound was obtained as a yellow solid in 76% yield, mp 183-186 $^\circ\text{C}$; IR (KBr) ν / cm^{-1} 3329, 1665, 1645, 1597, 1510, 1356, 1233; ^1H NMR (400 MHz, CDCl_3) δ 8.21 (dd, 1H, J 7.7, 1.0 Hz, Ar-H), 8.16 (dd, 1H, J 7.7, 1.0 Hz, Ar-H), 7.83 (td, 1H, J 7.5, 1.3 Hz, Ar-H),

7.75 (td, 1H, J 7.5, 1.3 Hz, Ar-H), 7.57-7.48 (m, 2H, 2Ar-H), 7.21-7.10 (m, 2H, 2Ar-H); ^{13}C NMR (101 MHz, CDCl_3) δ 183.79, 181.84, 162.87 (d, $^1J_{\text{C-F}}$ 249 Hz), 152.36, 135.49, 133.37, 132.90, 132.83 (d, $^3J_{\text{C-F}}$ 8 Hz), 129.40, 127.45, 126.32, 125.96 (d, $^4J_{\text{C-F}}$ 4 Hz), 121.25, 115.17 (d, $^2J_{\text{C-F}}$ 22 Hz).

2-Hydroxy-3-(4-(trifluoromethyl)phenyl)naphthalene-1,4-dione (**4d**)

After column chromatography using hexanes/EtOAc (85:15) as eluent, this compound was obtained as a yellow solid in 67% yield, mp 224-227 °C; IR (KBr) ν / cm^{-1} 3333, 1668, 1634, 1589, 1361, 1335, 1277, 1159, 1111; ^1H NMR (400 MHz, CDCl_3) δ 8.22 (d, 1H, J 7.6 Hz, Ar-H), 8.18 (d, 1H, J 7.6 Hz, Ar-H), 7.85 (t, 1H, J 7.5 Hz, Ar-H), 7.77 (t, 1H, J 7.5 Hz, Ar-H), 7.72 (d, 2H, J 8.2 Hz, 2Ar-H), 7.64 (d, 2H, J 8.2 Hz, 2Ar-H); ^{13}C NMR (101 MHz, $(\text{CD}_3)_2\text{CO}$) δ 183.00, 181.40, 154.67, 135.75, 134.91, 133.21, 132.60, 131.62, 130.02, 128.97 (q, $^2J_{\text{C-F}}$ 32 Hz), 126.43, 125.70, 124.50 (q, $^1J_{\text{C-F}}$ 271 Hz), 124.23 (q, $^3J_{\text{C-F}}$ 4 Hz), 120.55. HRMS (ESI) m/z , calcd. for $\text{C}_{17}\text{H}_9\text{F}_3\text{O}_3$ [$\text{M} - \text{H} + 2\text{Na}$] $^+$: 363.0215, found: 363.0245.

4-(3-Hydroxy-1,4-dioxo-1,4-dihydronaphthalen-2-yl)benzaldehyde (**4e**)

After column chromatography using hexanes/EtOAc (75:25) as eluent, this compound was obtained as a yellow solid in 58% yield, mp 216-220 °C; IR (KBr) ν / cm^{-1} 3200, 1688, 1670, 1643, 1605 1359, 1258; ^1H NMR (400 MHz, CDCl_3) δ 10.07 (s, 1H, CHO), 8.22 (d, 1H, J 7.7 Hz, Ar-H), 8.19 (d, 1H, J 7.5 Hz, Ar-H), 7.98 (d, 2H, J 8.2 Hz, 2Ar-H), 7.85 (t, 1H, J 7.6 Hz, Ar-H), 7.78 (t, 1H, J 7.6 Hz, Ar-H), 7.75 (s, 1H, OH), 7.70 (d, 2H, J 8.1 Hz, 2Ar-H); ^{13}C NMR (101 MHz, DMSO) δ 192.94, 183.18, 181.31, 155.47, 138.06, 135.16, 134.88, 133.41, 131.98, 131.59, 130.05, 128.59, 126.16, 125.77, 120.98. HRMS (ESI) m/z , calcd. for $\text{C}_{17}\text{H}_{10}\text{O}_4$ [$\text{M} - \text{H} + 2\text{Na}$] $^+$: 323.0291, found: 323.0290.

General procedure for the preparation of 2-methoxy-3-iodonaphthalene-1,4-dione (**5**)³³

To a stirred suspension of K_2CO_3 (5 equiv, 2.5 mmol) in acetone (12 mL) at room temperature, **2** (150 mg, 0.5 mmol) was added. After 5 minutes, dimethyl sulfate (2.5 equiv, 1.25 mmol) was slowly added and then the reaction was heated under reflux overnight. The reaction mixture was cooled to room temperature and ethyl acetate (25 mL) was added. The organic layer was first washed with brine (3 \times 25 mL) and then with H_2O (3 \times 25 mL), dried over anhydrous Na_2SO_4 and concentrated *in vacuo*. The crude product was purified by flash column chromatography on

silica gel using hexanes/EtOAc (90:10) as eluent. This compound was obtained as yellow needles in 74% yield, mp 160-161 °C; IR (KBr) ν / cm^{-1} 1668, 1587, 1562, 1437, 1329, 1306, 1248, 1211, 1041, 1007, 914, 719; ^1H NMR (500 MHz, CDCl_3) δ 8.17-8.12 (m, 1H, 1Ar-H), 8.12-8.06 (m, 1H, 1Ar-H), 7.77-7.64 (m, 2H, 2Ar-H), 4.31 (s, 3H, OCH_3); ^{13}C NMR (126 MHz, CDCl_3) δ 179.83, 177.84, 163.48, 134.23, 133.83, 131.02, 130.18, 127.62, 127.04, 105.58, 62.00.

General procedure for the preparation of 3-iodo-1,4-dioxo-1,4-dihydronaphthalen-2-yl acetate (**6**)

A mixture of **2** (150 mg, 0.5 mmol) and 4-dimethylaminopyridine (20 mol%) in acetic anhydride (2.5 mL) was stirred at room temperature for 4 hours. After this time, ethyl acetate (25 mL) was added and the organic layer was washed with brine (3 \times 25 mL), dried over anhydrous Na_2SO_4 and concentrated *in vacuo*. The crude product was purified by flash column chromatography on silica gel using hexanes/EtOAc (80:20) as eluent. This compound was obtained as a yellow solid in 70% yield, mp 153-154 °C; IR (KBr) ν / cm^{-1} 1778, 1674, 1597, 1371, 1327, 1302, 1275, 1244, 1165, 721; ^1H NMR (500 MHz, CDCl_3) δ 8.22-8.18 (m, 1H, Ar-H), 8.16-8.12 (m, 1H, Ar-H), 7.81-7.73 (m, 2H, 2Ar-H), 2.46 (s, 3H, OAc); ^{13}C NMR (126 MHz, CDCl_3) δ 179.09, 174.97, 166.63, 158.90, 134.55, 134.50, 130.51, 130.47, 128.30, 127.53, 113.88, 20.75. HRMS (ESI) m/z , calcd. for $\text{C}_{12}\text{H}_7\text{IO}_4$ [$\text{M} + \text{Na}$] $^+$: 364.9281, found: 364.9276.

General procedure for the preparation of 1,4-dioxo-3-aryl-1,4-dihydronaphthalen-2-yl diethylcarbamates (**7a-e**) and 3-(3-methylbut-2-en-1-yl)-1,4-dioxo-1,4-dihydronaphthalen-2-yl diethylcarbamate (**8**)

A stirred solution of **1** (0.1 mmol) or **4** (0.1 mmol), pyridine (5 equiv), *N,N*-diethylcarbamoyl chloride (20 equiv), in CHCl_3 (10 mL) was heated at 60 °C for 18 hours. Dichloromethane (30 mL) was added to the reaction mixture and the organic layer was first washed with a solution of hydrochloric acid 1N (30 mL) and then with brine (3 \times 30 mL), dried over anhydrous Na_2SO_4 and concentrated *in vacuo*. The crude product was purified by flash column chromatography on silica gel.

1,4-Dioxo-3-phenyl-1,4-dihydronaphthalen-2-yl diethylcarbamate (**7a**)

After column chromatography using hexanes/EtOAc (95:5) as eluent, this compound was obtained as an orange solid in 38% yield, mp 90-93 °C; IR (neat) ν / cm^{-1} 1730,

1666, 1595, 1471, 1257, 1184, 1146; ^1H NMR (500 MHz, CDCl_3) δ 8.19-8.13 (m, 2H, 2Ar-H), 7.79-7.73 (m, 2H, 2Ar-H), 7.48-7.37 (m, 5H, 5Ar-H), 3.37-3.24 (m, 4H, 2 CH_2), 1.17 (t, 3H, J 7.1 Hz, CH_3), 1.06 (t, 3H, J 7.1 Hz, CH_3); ^{13}C NMR (126 MHz, CDCl_3) δ 184.36, 179.91, 152.67, 150.67, 136.81, 134.33, 133.94, 132.18, 131.15, 130.16, 129.56, 129.24, 128.02, 127.08, 126.60, 42.54, 42.50, 13.85, 13.29. HRMS (ESI) m/z , calcd. for $\text{C}_{21}\text{H}_{19}\text{NO}_4$ $[\text{M} + \text{Na}]^+$: 372.1206, found: 372.1209.

3-(3,4-Dimethoxyphenyl)-1,4-dioxo-1,4-dihydronaphthalen-2-yl diethylcarbamate (**7b**)

After column chromatography using hexanes/EtOAc (95:5) as eluent, this compound was obtained as a red solid in 44% yield, mp 115-118 °C; IR (neat) ν / cm^{-1} 1730, 1666, 1595, 1514, 1454, 1259, 1184, 1145, 1026; ^1H NMR (500 MHz, CDCl_3) δ 8.20-8.12 (m, 2H, 2Ar-H), 7.81-7.73 (m, 2H, 2Ar-H), 7.04 (dd, 1H, J 8.3, 2.0 Hz, Ar-H), 7.00 (d, 1H, J 2.0 Hz, Ar-H), 6.95 (d, 1H, J 8.3 Hz, Ar-H), 3.94 (s, 3H, OCH_3), 3.88 (s, 3H, OCH_3), 3.44-3.24 (m, 4H, 2 CH_2), 1.20 (t, 3H, J 7.1 Hz, CH_3), 1.12 (t, 3H, J 7.1 Hz, CH_3); ^{13}C NMR (126 MHz, CDCl_3) δ 184.59, 179.88, 152.92, 150.34, 150.01, 148.48, 136.61, 134.26, 133.93, 132.27, 131.20, 127.11, 126.56, 123.67, 121.95, 113.45, 110.72, 55.05, 55.99, 42.58, 14.01, 13.37. HRMS (ESI) m/z , calcd. for $\text{C}_{23}\text{H}_{23}\text{NO}_6$ $[\text{M} + \text{Na}]^+$: 432.1418, found: 432.1422.

3-(4-Fluorophenyl)-1,4-dioxo-1,4-dihydronaphthalen-2-yl diethylcarbamate (**7c**)

After column chromatography using hexanes/EtOAc (95:5) as eluent, this compound was obtained as a yellow oil in 73% yield. IR (neat) ν / cm^{-1} 1730, 1666, 1593, 1504, 1259, 1225, 1184, 1145; ^1H NMR (500 MHz, CDCl_3) δ 8.20-8.12 (m, 2H, 2Ar-H), 7.81-7.74 (m, 2H, 2Ar-H), 7.44-7.39 (m, 2H, 2Ar-H), 7.17-7.11 (m, 2H, 2Ar-H), 3.38-3.28 (m, 4H, 2 CH_2), 1.19 (t, 3H, J 7.1 Hz, CH_3), 1.10 (t, 3H, J 7.1 Hz, CH_3); ^{13}C NMR (126 MHz, CDCl_3) δ 184.25, 179.73, 163.26 (d, $^1J_{\text{C-F}}$ 249 Hz), 152.59, 150.70, 135.74, 134.41, 134.04, 132.28 (d, $^3J_{\text{C-F}}$ 8 Hz), 132.01, 131.04, 127.09, 126.63, 125.37 (d, $^4J_{\text{C-F}}$ 3 Hz), 115.22 (d, $^2J_{\text{C-F}}$ 22 Hz), 42.56, 42.51, 13.92, 13.29. HRMS (ESI) m/z , calcd. for $\text{C}_{21}\text{H}_{18}\text{FNO}_4$ $[\text{M} + \text{Na}]^+$: 390.1112, found: 390.1116.

1,4-Dioxo-3-(4-(trifluoromethyl)phenyl)-1,4-dihydronaphthalen-2-yl diethylcarbamate (**7d**)

After column chromatography using hexanes/EtOAc (95:5) as eluent, this compound was obtained as a yellow oil in 48% yield. IR (neat) ν / cm^{-1} 1730, 1666, 1595, 1330, 1257, 1184, 1144, 1067, 1018; ^1H NMR (500 MHz, CDCl_3) δ 8.21-8.14 (m, 2H, 2Ar-H), 7.82-7.76 (m, 2H, 2Ar-H),

7.71 (d, 2H, J 8.0 Hz, 2Ar-H), 7.54 (d, 2H, J 8.0 Hz, 2Ar-H), 3.38-3.26 (m, 4H, 2 CH_2), 1.18 (t, 3H, J 7.1 Hz, CH_3), 1.08 (t, 3H, J 7.1 Hz, CH_3); ^{13}C NMR (126 MHz, CDCl_3) δ 183.84, 179.58, 152.42, 151.23, 135.46, 134.54, 134.17, 133.37, 132.00, 131.14 (q, $^2J_{\text{C-F}}$ 33 Hz), 131.10, 130.65, 127.15, 126.75, 124.98 (q, $^3J_{\text{C-F}}$ 4 Hz), 124.09 (q, $^1J_{\text{C-F}}$ 272 Hz), 42.66, 42.61, 13.84, 13.22. HRMS (ESI) m/z , calcd. for $\text{C}_{22}\text{H}_{18}\text{F}_3\text{NO}_4$ $[\text{M} + \text{Na}]^+$: 440.1080, found: 440.1079.

3-(4-Formylphenyl)-1,4-dioxo-1,4-dihydronaphthalen-2-yl diethylcarbamate (**7e**)

After column chromatography using hexanes/EtOAc (95:5) as eluent, this compound was obtained as a yellow oil in 32% yield. IR (neat) ν / cm^{-1} 1730, 1694, 1666, 1605, 1259, 1209, 1184, 1145; ^1H NMR (500 MHz, CDCl_3) δ 10.08 (s, 1H, CHO), 8.21-8.14 (m, 2H, 2Ar-H), 7.96 (d, 2H, J 8.2 Hz, 2Ar-H), 7.83-7.77 (m, 2H, 2Ar-H), 7.60 (d, 2H, J 8.2 Hz, 2Ar-H), 3.40-3.24 (m, 4H, 2 CH_2), 1.18 (t, 3H, J 7.1 Hz, CH_3), 1.07 (t, 3H, J 7.1 Hz, CH_3); ^{13}C NMR (126 MHz, CDCl_3) δ 191.98, 183.76, 179.53, 152.40, 151.16, 136.52, 135.86, 135.59, 134.56, 134.18, 131.98, 131.07, 130.98, 129.21, 127.14, 126.76, 42.67, 42.60, 13.91, 13.22. HRMS (ESI) m/z , calcd. for $\text{C}_{22}\text{H}_{19}\text{NO}_5$ $[\text{M} + \text{Na}]^+$: 400.1155, found: 400.1164.

3-(3-Methylbut-2-en-1-yl)-1,4-dioxo-1,4-dihydronaphthalen-2-yl diethylcarbamate (**8**)

After column chromatography using hexanes/EtOAc (95:5) as eluent, this compound was obtained as a brown oil in 40% yield. IR (neat) ν / cm^{-1} 1730, 1694, 1666, 1593, 1454, 1360, 1337, 1259, 1186, 1145, 1051; ^1H NMR (500 MHz, CDCl_3) δ 8.13-8.03 (m, 2H, 2Ar-H), 7.77-7.66 (m, 2H, 2Ar-H), 5.17-5.11 (m, 1H, CH), 3.48 (q, 2H, J 7.1 Hz, CH_2), 3.41 (q, 2H, J 7.1 Hz, CH_2), 3.32 (d, 2H, J 7.3 Hz, CH_2), 1.76 (s, 3H, CH_3), 1.68 (s, 3H, CH_3), 1.32 (t, 3H, J 7.1 Hz, CH_3), 1.24 (t, 3H, J 7.1 Hz, CH_3); ^{13}C NMR (126 MHz, CDCl_3) δ 184.96, 179.67, 152.53, 151.27, 137.81, 134.50, 134.02, 133.71, 132.33, 131.19, 126.70, 126.57, 119.07, 42.61, 25.89, 23.72, 18.04, 14.17, 13.34. HRMS (ESI) m/z , calcd. for $\text{C}_{20}\text{H}_{23}\text{NO}_4$ $[\text{M} + \text{Na}]^+$: 364.1519, found: 364.1520.

Antineoplastic activity

The cell lines HL-60 and K562 were obtained from the National Cancer Institute, Bethesda, MD, USA. All cancer cells were maintained in RPMI 1640 (Roswell Park Memorial Institute 1640) medium supplemented with 10% fetal bovine serum, 2 mM glutamine, 100 U mL^{-1} penicillin, 100 μg mL^{-1} streptomycin at 37 °C with 5%

CO₂. Heparinized blood (from healthy, non-smoker donors who had not taken any drug at least 15 days prior to sampling) was collected and PBMC were isolated by a standard method of density-gradient centrifugation over Ficoll-Hypaque. PBMC were washed, resuspended at a concentration of 3×10^5 cells mL⁻¹ and plated in a 96-well plate with RPMI 1640 medium supplemented with 20% fetal bovine serum, 2 mM glutamine, 100 U mL⁻¹ penicillin, 100 µg mL⁻¹ streptomycin at 37 °C with 5% CO₂. Phytohemagglutinin (3%) was added at the beginning of culture. After 24 h, tested compounds (0.4-80 µM) dissolved in RPMI 1640 medium with 1% of DMSO (dimethyl sulfoxide) were added to each well and incubated for 72 h.

The cytotoxicity of all compounds were tested using the 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide (MTT)³⁴ (Sigma-Aldrich Co., St. Louis, MO, USA) reduction assay. For all experiments, tumor cells were plated in 96-well plates (3×10^5 cells mL⁻¹). Tested compounds (0.4-80 µM) dissolved in DMSO 1% were added to each well and incubated for 72 h. Control groups received the same amount of DMSO. At the end of the incubation, the plates were centrifuged and the medium was replaced by fresh medium (150 µL) containing 0.5 mg mL⁻¹ MTT. After 3 h, the formazan product was dissolved in 150 µL of DMSO, and the absorbance was measured using a multiplate reader (DTX 880 Multimode Detector, Beckman Coulter, Inc., Fullerton, California, USA). The IC₅₀ values and their 95% confidence intervals for two different experiments were obtained by nonlinear regression using GraphPad Prism version 5.0 for Windows (GraphPad Software, San Diego, California, USA).

Antileishmanial activity

The compounds were dissolved in DMSO, Sigma®. Assay concentrations were prepared in culture medium used in the experiments, as indicated in each case. For antipromastigote activity,¹⁰ promastigotes of *Leishmania amazonensis* (MHOM/BR/75/LTB0016) were adjusted to a concentration of 1×10^6 cells mL⁻¹ in Schneider medium supplemented with 10% de fetal bovine serum, penicillin (100 U mL⁻¹) and streptomycin (100 µg mL⁻¹), and incubated at 26 °C for 72 h with the compounds (0-100 µM). The antileishmanial activity was evaluated by adding in each well 22 µL of MTT at 5 mg mL⁻¹ (Sigma®). After 2 h, 80 µL of DMSO was added. The optical density was determined at a wavelength of 570 nm in microplate reader (µQuant Bio-Tek Instruments®, Winooski, Vermont, USA). The inhibition percentage was estimated by the comparison with non-treated control

cultures. The assays were carried out in triplicate in 96-well plates (Costar®, New York, NY, USA) and repeated at least three times. For intracellular amastigote assays, BALB/c mice macrophages were obtained by peritoneal lavage with 5 mL of cold RPMI medium (Sigma®). The cell suspension (2×10^6 macrophages mL⁻¹) was applied in Labtek chambers (Nunc®, New York, NY, USA) and incubated for 1 h at 37 °C, 5% CO₂. Then, the cultures were washed with phosphate buffer saline (PBS) at 37 °C for removal of non-adherent cells. The remaining cells were incubated at 37 °C, 5% CO₂ with stationary phase promastigotes of *L. amazonensis* at a ratio of 3:1. After 3 h, the chambers were washed again to remove free parasites and incubated with compounds (0-50 µM) at 37 °C, 5% CO₂ for 72 h. The anti-amastigote activity was analyzed microscopically by counting at least 100 macrophages *per* sample, after staining cells with hematological system Instant Prov (New Prov®, Curitiba, Brazil).³⁵ The experiments were performed in duplicate and repeated twice. Results were expressed as infection index (II) using the following formula: $II = (\% \text{ infected cells}) \times (\text{number of amastigotes} / \text{total macrophages number})$. For toxicity assays, BALB/c mice macrophages were obtained by peritoneal lavage with 5 mL of cold RPMI medium (Sigma®). The macrophages at 2×10^6 cells well⁻¹ in RPMI culture medium (pH 7.2, supplemented with 10% fetal bovine serum) were incubated with the compounds (0-200 µM) for 72 h at 37 °C under 5% CO₂ in 96-well plates. After removing the supernatant, viable cells were quantified by adding MTT (200 µL, 0.5 mg mL⁻¹) in PBS. After 2 h, the supernatant was removed and DMSO (100 µL) was added in each well. The optical density was determined at wavelength of 570 nm in the microplate reader. The percentage of viable cells was calculated relative to the control cells. The tests were carried out in triplicate and repeated twice. Logarithm regression analysis was performed using GraphPad Prism 5.0 (San Diego, CA, USA) in order to obtain the values of IC₅₀ and LD₅₀. The selectivity index was determined as macrophage LD₅₀/intracellular amastigote IC₅₀. This study was approved by the Animal Ethics Committee of Oswaldo Cruz Foundation (license number LW7/2010).

Supplementary Information

Supplementary data (¹H and ¹³C NMR spectra) are available free of charge at <http://jbcbs.sbj.org.br> as PDF file.

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