COL6A3 promotes cellular malignancy of osteosarcoma by activating the PI3K/AKT pathway



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SUMMARY

OBJECTIVE: In this study, we aimed to investigate the role of COL6A3 on cell motility and the PI3K/AKT signaling pathway in osteosarcoma. **METHODS**: The relative expression of COL6A3 was achieved from a GEO dataset in osteosarcoma tissue. siRNA technology was applied to decrease the COL6A3 expression in cells, and cell counting kit-8 (CCK-8) assay and colony formation analysis were used to examine the cell proliferation potential. Knockdown COL6A3 made the proliferation and colony formation abilities worse than the COL6A3 without interference. Likewise, in contrast to the si-con group, cell invasion and migration were inhibited in the si-COL6A3 group. Moreover, the western blot results suggested that the PI3K/AKT signaling pathway was manipulated by measuring the protein expression of the PI3K/AKT pathway-related markers, due to the COL6A3 inhibition.

CONCLUSION: COL6A3 plays a crucial role in modulating various aspects of the progression of osteosarcoma, which would provide a potentially effective treatment for osteosarcoma.

KEYWORDS: Osteosarcoma. Neoplasms, bone tissue. Molecular targeted therapy. Collagen type VI.

INTRODUCTION

Osteosarcoma is a prevalent and aggressive tumor in the bones that has received widespread attention because of its particular features¹. Up to now, radical surgery, chemotherapy, and radiotherapy are the most commonly used treatments in osteosarcoma, but the cure rate is not significant². It is well-known that targeted therapy is a major and effective modality for cancer, and numerous genes have been proved to be

associated with the progression of tumors^{3,4}. Hence, novel biomarkers are urgently needed for the diagnosis and treatment of osteosarcoma.

Collagen VI (COL6) is a extracellular-matrix protein, which is related to the basement membrane and made up of three chains: alpha 1, alpha 2, and alpha 3⁵. Previous reports have illustrated that COL6 is involved in muscle regularity and cell membrane

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integrity⁶. There is evidence that the loss of COL6 can cause serious Bethlem myopathy and Ullrich congenital muscular dystrophy⁷.

However, there is no literature to interpret the functional characteristics of COL6A3 in osteosarcoma. The purpose of the study was to investigate the role of COL6A3 on cell motility and the PI3K/AKT signaling pathway in osteosarcoma.

ETHODSCELL LINES AND CULTURE CONDITION

Human osteosarcoma cell lines (U2OS, HOS, and MG-63) and normal cell lines hFOB1.19 were purchased from the cell bank of the Chinese Academy of Sciences (Shanghai, China), which were then incubated in RPMI-1640 medium. To maintain the normal viability of cells, fetal bovine serum (FBS; 10%), penicillin (100 U/mL), and streptomycin (100 U/mL) were used. All the cells were stored at 37°C and 5% CO2.

TRANSFECTION

To knockdown COL6A3, small interfering RNAs (siRNAs) including si-COL6A3 and si-con were designed by GenePharma (Shanghai, China). The sequences of two siRNAs are as follow: COL6A3 siRNA: 5'-GCTTTGCACATATTCGAGATT-3'; si-con: 5'-AATTCTCCGAACGGTCACGT-3'. The si-con was used as a negative control. For transfection, cells were treated with siRNAs using Lipofectamine2000 per the manufacture protocol. After 24 h transfection, the knockdown efficiency of COL6A3 can be observed for further detection.

Reverse transcription-quantitative polymerase chain reaction (qPCR)

To detect the mRNA expression level of a specific gene, TRIzol solution (Invitrogen, Carlsbad, CA, USA) was used to isolate total RNA, and then qRT-PCR analysis was carried out. Then RNA was converted into cDNA using the Fast Quant RT Kit (TaKaRa, Otsu, Shiga, Japan). qPCR was performed with the Applied Biosystems 7500 Real-Time PCR System (Thermo Fisher Scientific, Inc., Waltham, MA, USA) and, in the meantime, SYBR Green Master Mix was also utilized as a dyeing probe. The reaction procedure was the following: pre-heated at 95° for 5 min, denatured at 95°C for 30 s and 60° for 45 s with 40 cycles, 72° for 30 min. The forward primer of

COL6A3 was 5'-AACATCCTGGTCAGCTCTGC-3' and the reverse primer was 5'-TCCGGGATGAAGGAGAT-GGT-3'. In addition, the forward primer of GAPDH was 5'-TCCAAAATCAAGTGGGGCGA-3', and its reverse primer was 5'-TGATGACCCTTTTGGCTCCC-3'. GAPDH was assessed as internal control and all detections were conducted three times. The 2^{-ΔΔCT} method was used to examine relative expression.

Western blotting

Transfected cells for 24 h were placed on ice and lysed by RIPA reagent (Beyotime, Shanghai, China) supplemented with a protease inhibitor to extract proteins. Then, the isolated proteins were concentrated by the BCA method and boiled at 95° for 5 min. In the vertical electrophoresis tank, 20 µg protein was added into each well, separating by 12% SDS-PAGE at 110v for 1 h and transferred onto the PVDF membrane. Subsequently, the PVDF membrane was blocked using 5% skim milk powder for 1 h and antibodies. Primary antibodies (1:1,000; Abcam, Cambridge, MA, USA) were used to incubate the membrane at 4°C overnight, and a secondary antibody (1:2,000; Abcam, Cambridge, MA, USA) at room temperature for 1 h. Finally, ECL was added for development and the gray value of protein bands was scanned through the QUANTITY ONE software.

Cell proliferation and colony formation assays

Cell counting kit 8 (CCK-8) analysis and colony formation exploration were applied to evaluate the proliferative and clonal abilities. For the CCK-8 assay, the single-cell suspensions generated by the transfected cells were seeded into a 96-well plate with a density of 1, 000 cells per well, at 37°C for 0-72 h. Every 24 hours were considered as a detection time point and the cells should go through an additional culture 1.5 h after supplementing with 10 µL CCK-8 reagent. At last, the OD value was measured at 450 nm using a microplate reader. For colony formation exploration, transfected cells were inoculated in a 60 mm-depth dish, which was pre-filled with 5 mL warm medium; the density was 400 cells/dish. The culture was completed when macroscopic clones appeared in the culture dish (about two weeks), fixed then stained by 4% paraformaldehyde and 0.1% crystal violet, respectively, for 30 min. Finally, the clone was counted and captured under a microscope.

Transwell assay

Transwell assay was carried out to assess cell migration and invasion. Cells after 24 h transfection were turned into a cell suspension and put into the upper chamber. Meanwhile, the lower chamber was filled with 500 μ L complete medium. After waiting overnight, the residual cells on the surface of the upper chamber were wiped out and the chamber was washed using PBS for three times. The cells on the surface of the lower chamber were fixed using 4% paraformaldehyde, dyed via 0.1% crystal violet, and imaged through the microscope. Attentively, the upper chamber was pre-coated with Matrigel (BD Sciences, Franklin Lakes, NJ, USA) for invasion while migratory detection was not. Furthermore, the inoculated density was different: 5, 000 cells for migration and 1×10 5 cells for invasion.

Statistical analysis

Based on the SPSS 22.0 statistics software (SPSS Inc., Chicago, IL, USA) and GraphPad Prism 5.0 (GraphPad Software, San Diego, CA, USA), all data were analyzed and described as mean ± standard deviation (SD).

Differential analysis of the two groups was conducted with the student's t-test; the comparison of multiple groups was conducted by one-way ANOVA and Dunnett's post hoc test. The significance of this test was p < 0.05.

RESULTS

COL6A3 was associated with metastasis of osteosarcoma

Initially, we collected two arrays from the GEO dataset to examine the expression level of COL6A3 in osteosarcoma tissues and discovered that the expression level of COL6A3 was significantly increased in both GSE16088 (P = 0.0324) involving 14 osteosarcoma cases and 3 normal cases and GSE49003 (P = 0.0022) containing 6 non-metastasis cases and 6 metastasis cases. These findings indicate that the development of osteosarcoma was closely associated with the COL6A3

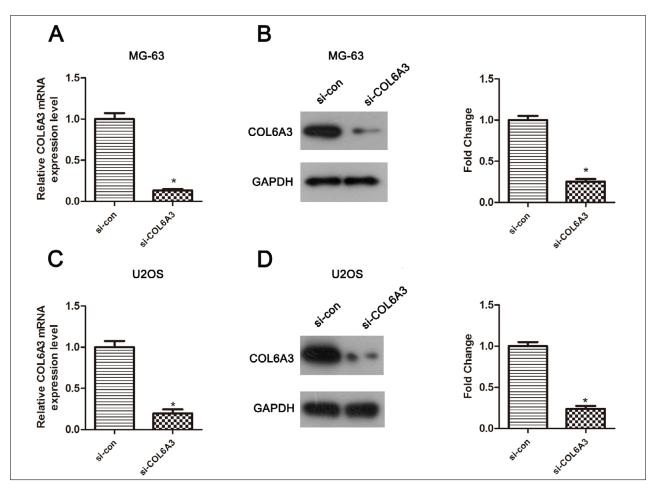


FIGURE 1. HIGH EXPRESSION OF COL6A3 WAS ASSOCIATED WITH METASTASIS IN OSTEOSARCOMA.

A) Relative expression of COL6A3 between normal tissue and osteosarcoma tissue, which was obtained from the GEO database, P = 0.0324. B) Relative expression of COL6A3 between non-metastasis tissue and metastasis tissue from the GEO database, P = 0.0022. C) The qRT-PCR analysis revealed a multiple expression tendency in three osteosarcoma cell lines (U2OS, HOS, and MG-63) and one in a normal cell line hFOB1.19, ***P < 0.001.

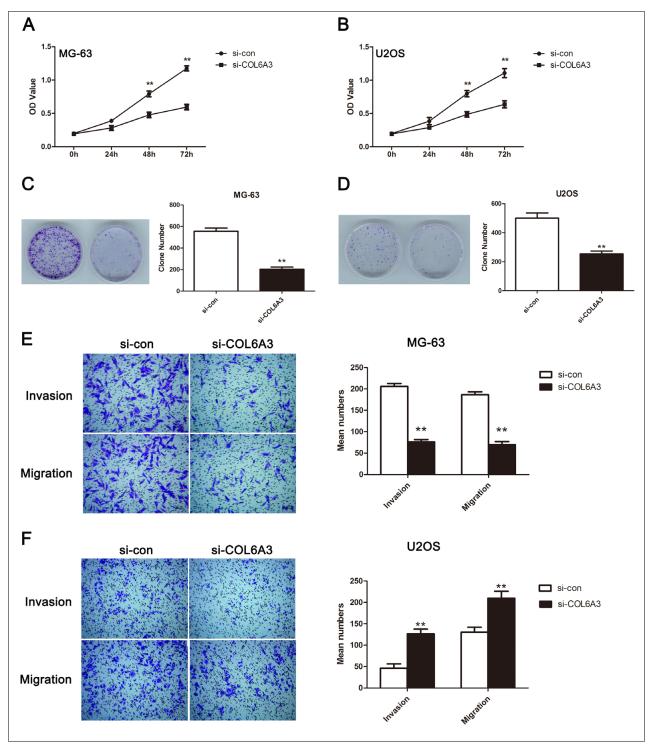


FIGURE 2. REDUCTION OF COL6A3 EXERTED NEGATIVE EFFECTS ON CELL MALIGNANT BEHAVIORS IN MG-63 AND U2OS CELLS. (A AND B) THE CELL VIABILITY OF OSTEOSARCOMA CELLS TRANSFECTED WITH SI-COL6A3 WAS HINDERED USING CCK-8 ASSAY, **P < 0.01. (C AND D), COLONY ACTIVITY WAS WEAKENED BY COLONY FORMATION ASSAY AFTER KNOCKDOWN OF COL6A3, **P < 0.01. (E AND F) SILENCING THE COL6A3 HAD NEGATIVE IMPACTS ON MIGRATORY AND INVASIVE ABILITIES IN OSTEOSARCOMA, **P < 0.01.

expression. Subsequently, we detected the COL6A3 expression in three osteosarcoma cell lines (U2OS, HOS, and MG-63) and one normal human cell line hFOB1.19. Compared with the normal human cell

line hFOB1.19, COL6A3 showed diverse expression patterns (Fig.1C, P < 0.001). Based on the expression level of COL6A3, we selected the two cell lines U2OS and MG-63 for future experiments.

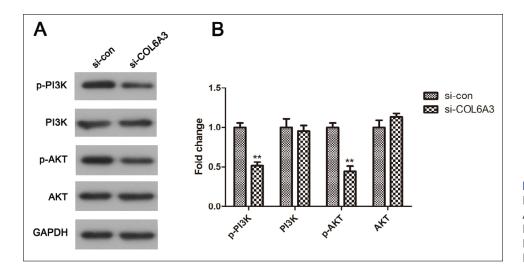


FIGURE 3. COL6A3 KNOCKDOWN WAS ASSOCIATED WITH THE INACTIVATION OF THE PI3K/AKT SIGNALING PATHWAY.

COL6A3 expression was down-regulated in U2OS and MG-63 cells

To verify the following results correctly, we used siRNA technology to knockdown the expression of COL6A3 and evaluate the silencing efficiency by qRT-PCR and western blot. As shown in Figure 1A and 2C, we observed that the interference of si-COL6A3 successfully decreased COL6A3 mRNA expression by qRT-PCR (P<0.05). Consistently, the western blotting results indicated that the COL6A3 protein expression level was inhibited in U2OS and MG-63 cells (Figure 1B and 2D, P < 0.05).

Depletion of COL6A3 inhibited cell proliferation, invasion, and migration in U2OS and MG-63

Next, we assessed the effects of knockdown COL6A3 on cell aggressive behaviors. The results of the CCK-8 assay revealed that the down-regulation of COL6A3 suppressed cell growth and an evident difference was observed at 48 h, 72 h between the si-COL6A3 group and si-con group in both MG-63 and U2OS cells (Figure 2A and 2B, P < 0.01). In the same way, the clonogenic ability of MG-63 and U2OS cells transfected with si-COL6A3 also showed a downward trend (Figure 2C and 2D, P < 0.01). To explore whether si-COL6A3 affected the capabilities of migration and invasion in osteosarcoma cells, we then performed a transwell assay. The representative images indicated that migratory and invasive cells in the si-COL6A3 group were fewer than in the si-con group in osteosarcoma cells (Figure 2E and 2F, P < 0.01).

COL6A3 knockdown was associated with the inactivation of PI3K/AKT signaling pathway

We determined the protein expression level of the PI3K/AKT pathway-related markers, including p-PI3K/PI3K and p-AKT/AKT, in osteosarcoma cells. As illustrated in Figure 3A, U2OS and MG-63 cells transfected with si-COL6A3 exhibited a more down-regulated expression of p-PI3K/p-AKT than cells in the si-con group. The quantified analysis of gray value encouraged the above-mentioned results (Figure 3B, P < 0.01).

DISCUSSION

We found that COL6A3 was increased in osteosarcoma tissues and cells when compared with normal control. COL6 is expected to generate a flourishing condition for the development of tumors by cooperating with the extracellular matrix (ECM) rebuilding. Cell anchoring has also been proven to correlate with COL6⁸⁻¹⁰. Considering the mentioned researches, we predicted that COL6A3, which acts as a component of COL6, would participate in tumor progression. Furthermore, its function has been evaluated in several cancers, such as pancreatic¹¹, giant cell tumors¹², and prostate¹³. However, the effects of COL6A3 on osteosarcoma are still unclear. Hence, we performed this study to determine the biological potential of COL6A3 in osteosarcoma.

Next, we paid much attention to the molecular mechanism of how COL6A3 manipulates cell activities. It is well-known that the PI3K/AKT signaling pathway is one of the most important cancer-related

pathways¹⁴. The activation of the PI3K/AKT signaling pathway is recognized to exert a pivotal role in cell malignant motility in cancer progress^{ion15,16}. In addition to that, COL6A3 has been reported to be involved in the progression of gastric cancer by regulating the PI3K/AKT signaling pathway¹⁷. The same function has also been indicated in esophageal cancer in the Chinese population¹⁸. Therefore, we conducted western blotting to demonstrate the expression level of the PI3K/AKT signaling pathway-related markers after knockdown of the COL6A3.

CONCLUSION

These findings propose a new perspective in the diagnosis and treatment of osteosarcoma, manifesting important clinical implications against osteosarcoma.

Conflicts of interest

They have no conflicts of interest.

Author Contributions

Conceptualization, Jia Sun; formal analysis, Ze-Long Song; writing (original draft preparation), Hong-Li Guo; writing (review and editing), Gang Chen; supervision, Xi-Hai Gao; funding acquisition, Yu-Xia Han

RESUMO

OBJETIVO: Neste estudo, investigamos a função do COL6A3 na mobilidade celular e na via PI3K/AKT em osteossarcomas.

METODOLOGIA: A expressão relativa do COL6A3 foi obtida a partir de dados GEO em tecidos de osteossarcoma. O RNA de interferência (siRNA) foi utilizado para reduzir a expressão do COL6A3 nas células, e o teste de contagem de células kit-8 (CCK-8) e a análise de formação de colônias foram realizados para examinar o potencial de proliferação celular. Além disso, o Transwell comprovou os efeitos do si-COL6A3 na invasão celular e migração em células de osteossarcoma. Para medir os níveis de expressão das proteínas e mRNAs, utilizamos transcriptase reversa quantitativa (qRT-PCR) e western blot.

RESULTADOS: O COL6A3 foi regulado nos tecidos e células do osteossarcoma quando comparado com o controle normal. A redução de COL6A3 reduziu a proliferação e a capacidades de formação de colônias em relação ao COL6A3 sem interferência. Do Mesmo modo, ao contrário do observado no grupo si-con, a invasão e migração celular foram inibidas no grupo si-COL6A3. Além disso, o resultado do western blot sugere que a via PI3K/AKT foi manipulada, medindo a expressão proteica dos marcadores relacionados à PI3K/AKT, devido à inibição do COL6A3.

CONCLUSÃO: O COL6A3 desempenha um papel crucial na modulação de vários aspectos da progressão do osteossarcoma, o que pode representar um possível tratamento eficaz para a doença.

PALAVRAS-CHAVE: Osteossarcoma. Neoplasias de tecido ósseo. Terapia de alvo molecular. Colágeno tipo VI.

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