

Research Article

Linking dopamine neurotransmission and neurogenesis: The evolutionary history of the NTAD (*NCAM1-TTC12-ANKK1-DRD2*) gene cluster

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Abstract

Genetic studies have long suggested the important role of the *DRD2* gene in psychiatric disorders and behavior. Further research has shown a conjoined effect of genes in the Chr11q22-23 region, which includes the *NCAM1*, *TTC12*, *ANKK1* and *DRD2* genes, or NTAD cluster. Despite a growing need to unravel the role of this cluster, few studies have taken into account interspecies and evolutionary approaches. This study shows that behaviorally relevant SNPs from the NTAD cluster, such as rs1800497 (Taq1A) and rs6277, are ancient polymorphisms that date back to the common ancestor between modern humans and Neanderthals/Denisovans. Conserved synteny and neighborhood indicate the NTAD cluster seems to have been established at least 400 million years ago, when the first Sarcopterygians emerged. The NTAD genes are apparently co-regulated and this could be attributed to adaptive functional properties, including those that emerged when the central nervous system became more complex. Finally, our findings indicate that NTAD genes, which are related to neurogenesis and dopaminergic neurotransmission, should be approached as a unit in behavioral and psychiatric genetic studies.

Keywords: NTAD cluster, Taq1A SNP, shared synteny, co-regulation, psychiatric genetics.

Introduction

The role of the NTAD cluster in psychiatric disorders

The crucial role played by dopaminergic neurotransmission systems in psychiatric genetics has been of great interest for several years. Among the candidate genes, one of the main focuses of research has been the dopamine receptor D2 (DRD2), especially its neighboring singlenucleotide polymorphism (SNP) rs1800497 (Taq1A). The rs1800497 SNP was considered to be a silent mutation located about 10 kb from DRD2, in the 3' untranslated region of this gene. The identification of a novel gene in the neighboring forward-strand region of *DRD2*, named *ANKK1*, showed that the rs1800497 SNP is actually located in exon 8 of ANKK1 (Neville et al., 2004), where it causes an amino acid change (Glu713Lys) in its 11th ankyrin repeat. Although the rs1800497 polymorphism is localized in ANKK1, it seems to be in linkage disequilibrium with several DRD2 variants (Gelernter et al., 2006; Dubertret et al., 2010).

Despite the enormous number of studies regarding the role of the rs1800497 SNP in psychiatric disorders (es-

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pecially alcohol and nicotine dependence), it is still far from clear how and to what degree it affects psychiatric disorders. The three most recent meta-analyses support the link between the rs1800497 T allele and alcoholism (Munafò *et al.*, 2007; Smith *et al.*, 2008; Le Foll *et al.*, 2009). There are two meta-analysis studies focusing on the relationship between rs1800497 and smoking behavior, but with conflicting results (Li *et al.*, 2004; Munafò *et al.*, 2004). Strong heterogeneity has been a hallmark of several of these meta-analyses. Consequently, the strong divergence among such findings raises the need to identify the variables that might explain such heterogeneity.

It has been suggested that two other nearby genes, *NCAM1* and *TTC12*, are also good candidate genes for psychiatric disorders and could comprise causative variants to phenotypes previously attributed to *DRD2* polymorphisms (Gelernter *et al.*, 2006; Huang *et al.*, 2008; Dubertret *et al.*, 2010). For example, Yang *et al.* (2007) found an association of SNPs in the *NCAM1*, *TTC12* and *ANKK1* genes with alcohol dependence, but not so for SNPs in the *DRD2* gene. In another study, a single haplotype spanning *TTC12* and *ANKK1*, as well as multiple SNPs in these two genes, were associated with nicotine dependence (Gelernter *et al.*, 2006).

These genes are located on chromosome 11 (more precisely, the 11q22-23 region) and form a 521 kb gene

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cluster that comprises the NCAM1, TTC12, ANKK1 and *DRD2* genes, known as the NTAD gene cluster (Figure S1). The four genes that comprise the NTAD cluster all seem to act on the brain, although details of their specific functions in neural tissue have yet to be discovered. The neural cell adhesion molecule 1 (NCAM1) plays an important role in neurogenesis, specifically in axon and dendrite growth (McIntyre et al., 2010). TTC12 encodes the poorly understood tetratricopeptide repeat domain 12 protein, which seems to be involved in dopaminergic transmission and neurodevelopment via the Wnt signaling pathway (Kahto and Kahto, 2003; Castelo-Branco and Arenas, 2006). The ankyrin repeat and kinase domain containing 1 (ANKK1) gene encodes a signaling protein which takes part in indirect modulation of the expression of DRD2 (Huang et al., 2008), thus constituting the clearest currently known evidence of co-regulation in the NTAD cluster.

Gene cluster and genomic architecture: Evolutionary aspects

Gene order in eukaryotes cannot be attributed in its entirety to mere randomness. Multiple lines of evidence indicate that co-expressed, co-regulated and co-functional genes can be maintained as a gene cluster due to the pressure of natural selection (Hurst *et al.*, 2004; Sémon and Duret, 2006; Michalak, 2008).

Considering that polymorphisms of the NTAD gene cluster seem to have a conjoined effect, the question raised is how long these genes are being maintained with shared synteny (genes in the same chromosome) and conserved neighborhood (genes side-by-side in the same order) through the course of evolution. If the NTAD conformation is recent or human-specific it may reflect an adaptive novelty or stochastic clustering in Homo/Homo sapiens. Notwithstanding, the maintenance of a specific gene cluster such as NTAD over long evolutionary periods is much more difficult to be explained by actions of random processes only. An ancient clustering in this case might also reflect a functional benefit and/or an orchestrated evolution of genes involved in neurotransmission and neurogenesis, probably mediated by co-regulation, co-expression or molecular co-functionality.

Hominid comparative genomics has made great strides after the publication of the Neanderthal and Denisovan nuclear genomes since it has allowed researchers to build on knowledge about unique human phenotypes, including psychiatric disorder susceptibilities in present-day populations. Some of these psychiatric disorders have been repeatedly associated with variation in the whole NTAD cluster, but no previous study investigated it from a wider evolutionary perspective using comparative genomic approaches. Taking this information into account we herein performed the first *in silico* study addressing the genomic architecture and chromosomal dynamics of the NTAD cluster from an evolutionary perspective. Our results pro-

vide a more comprehensive view about this gene cluster and how its dynamics could shape future genetic studies of complex behavior phenotypes and psychiatric disorder in humans.

Material and Methods

Analysis of NTAD SNP status in primates

Seven human SNPs were chosen based on previous association with psychiatric disorders and/or with evidence of functionality: rs646558 from *NCAM1*; rs723077 and rs2303380 from *TTC12*; rs2734849 and rs1800497 from *ANKK1*; and rs6277 and rs2283265 from *DRD2* (Figure S1). These polymorphic sites were then compared with their counterparts in the genomes of two archaic hominids, *Homo neanderthalensis* (Green *et al.*, 2010) and a Denisovan specimen (Reich *et al.*, 2010), as well as nine non-human primates. The derived allele age was estimated for the rs1800497 and rs6277 SNPs according to the frequency based method, as proposed by Slatkin and Rannala (2000).

Three different approaches were used to predict whether the three non-synonymous mutations (rs1800497 and rs2734849 in *ANKK1*; rs723077 in *TTC12*) among the seven SNPs promote important functional changes in the proteins: PolyPhen2, SNAP and the assessment of Grantham scores of chemical distance (Grantham, 1974). For the latter we used the classification by Li *et al.* (1985) as: conservative (Grantham score = 0-50), moderately conservative (51-100), moderately radical (101-150) and radical (>151). These methods are used to predict the possible impact of amino acid substitutions on the structure and function of proteins by means of chemico-physical and comparative evidence.

Orthology, synteny and neighborhood status of the NTAD cluster in vertebrate genomes

The online databases Ensembl release 66, UCSC Genome Browser and UniProt were used as sources for the nucleotide and protein sequences of *NCAM1*, *TTC12*, *ANKK1* and *DRD2* genes in the human genome and their orthologues in 47 other vertebrate species (Table S1). Their synteny and neighborhood status were inferred from the available contigs. BLAST/BLAT searches in these databases were performed to find possible unannotated orthologues. BioEdit version 7.0.9.0 (Hall, 1999) was used to align the sequences of the orthologues, when necessary. The DECODE database was used to infer transcription factor binding sites in the NTAD cluster. DECODE is based on text mining applications by SABiosciences and gene annotations of regulatory binding sites available at the UCSC Genome Browser.

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Results and Discussion

Comparative analyses of seven SNPs belonging to the NTAD gene cluster in human and non-human primates

Our analyses based on the seven SNPs selected according to their association with psychiatric disorders and/or with evidence of functionality, revealed interesting results (Table 1). For instance, we were able to show that polymorphisms believed to be H. sapiens-specific turned out to be plausibly widespread in the Homo genus. Although the introgression of H. sapiens derived alleles to other hominids could not be disregarded, these polymorphisms could be traced back to at least 270,000-800,000 years when our lineage diverged from Neanderthals/Denisovans (Green et al., 2010; Reich et al., 2010). This seems to be the case for the rs1800497 SNP, where both ancestral and derived alleles (taking into account that a mutation event produces a new, mutant or derived allele, that is different from the "original" or ancestral one) are present in the Denisovan genome, while in Neanderthals only the derived allele could be found, denoting that the $A \rightarrow G$ (Glu713Lys) mutation occurred before the origin of *Homo* sapiens. Our estimate for the age of the derived allele (Slatkin and Rannala, 2000) is compatible with this hypothesis (379,000-447,000 years).

A similar situation was detected with the DRD2 rs6277 SNP, but in this case both alleles were found in Neanderthals (Table 1). Interestingly, the estimation of allele age for the derived allele of this polymorphism presented a greater discrepancy among populations. For Africans and Asians it was about 65,000 years, whereas for Europeans it was set at 359,000 years. These results raised some instigating hypotheses, including allele introgression from Neanderthals to *H. sapiens* in European populations, with subsequent dispersion to other continents. Selection processes and genetic drift, inflating the derived allele frequency in Europe, are other possible explanations. It seems however unlikely that stochastic processes would keep these polymorphisms unchanged for such a long time. Signals of positive selection were found in variant alleles of a related dopaminergic gene (DRD4) associated with modern psychiatric disorders. Some of these variants are associated with behavioral traits that could have had some adaptive advantage in the past, but today may have clinical implications (Ding et al., 2002; Wang et al., 2004; Tovo-Rodrigues et al., 2011). A similar scenario may be responsible for the maintenance of these polymorphic sites in the human lineage, although only further studies can test this hypothesis.

On the other hand, the *TTC12* rs2303380 and *DRD2* rs2283265 SNPs are likely to be *Homo sapiens*-specific. The *NCAM1* rs646558 derived allele (C) is also *H. sapiens*-specific, but curiously its ancestral allele (A) is exclusively found in Old World primates, while a third allele (G) is found in lemurs and New World monkeys (Table 1). This

Table 1 - Single nucleotide variation in the NTAD cluster across primate genomes.

Gene	Position SNP	SNP					Old Wo	Old World Primate						New World Primate	Lemurs		
					Human		Denisova	Denisova Neandertal	Chimp	Gorilla	Orangutan	Baboon	Rhesus	Marmoset	Tarsier	Mouse	Bushbaby
			Alleles	Alleles AFA M.A.F ASN M.A.F	ASN M.A.F	EUR M.A.F										lemur	
NCAMI	Intron 13	rs646558	A/C	0,575 (A)	0,196 (A)	0,252 (A)	A	¥	Ą	V	Ą	V	Ą	G	G	G	Ö
TTC12	Exon 3	rs723077*	A/C	0,097 (C)	0,308 (C)	0,487 (C)	∢	۶.	A	A	A	Ö	G	Ü	Ü	A	A
TTC12	Intron 7	rs2303380	A/G	0,338 (G)	0,39 (G)	0,381 (G)	∢	۶.	A	A	A	∢	A	A	A	A	A
ANKKI	Exon 8	rs2734849*	A/G	0,115 (G)	0,024 (G)	0,473 (G)	∢	A	A	II	A	∢	A	A	Ü	A	II
ANKKI	Exon 8	rs1800497* (Taq1A)	A/G	0,411 (A)	0,407 (A)	0,195 (A)	A/G	Ď	∢	4	⋖	A	∢	II	∢	∢	II
DRD2	Exon 7	rs6277	G/A	0,034 (A)	0,049 (A)	0,534 (A)	Ð	G/A	G	G	G	A	A	A	A	G	Ü
DRD2	Intron 5	rs2283265	C/A	0.092 (A)	0.422 (A)	0.167 (A)	Ö	ć	Ö	C	O	C	C	Ö	ć	Ö	II

Non-synonymous polymorphism. M.A.F = Minor Allele Frequency, indicated in parenthesis, obtained in HAPMAP. Mean frequencies were computed using European (CEU), Asian (CHB and JPT) and African (YRI) populations. Genomes included: Human (Homo sapiens), Denisova (Desinova cave specimen), Neanderthal (Homo neanderthalensis), Chimpanzee (Pan troglodytes), Gorilla (Gorilla gorilla gorilla gorilla gorilla) hamadryas), Rhesus (Macaca mulatta), Marmoset (Callithrix jacchus), Tarsier (Tarsier syrichta), Mouse Iemur (Microcebus murinus) and Bushbaby (Otolemur garnettii). (=): aligning species Orangutan (Pongo

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denotes that the $G \rightarrow A$ mutation possibly occurred on the phylogenetic branch leading to the Old World primates.

The additional analysis of the three non-synonymous SNPs rendered only the Met73Leu (rs723077) mutation in the *TTC12* gene as a non-neutral amino acid change, which the SNAP prediction tool indicated that the protein's chemico-physical properties may change in response to this substitution. The failure in detecting similar results for the other two mutations does not necessarily imply that they have no functional impact since the SNAP, PolyPhen2 and Grantham Score programs were designed to indicate only significant chemical changes in protein structure. Likewise, detecting a non-neutral signal does not necessarily imply direct association with relevant phenotypic changes.

Orthology, synteny and neighborhood status of the NTAD cluster in vertebrate genomes

Our comparative analysis involved human NTAD gene sequences as query for BLAST searches in the other 46 available vertebrate genome sequences (Table S1). Most of the orthologous genes were previously annotated in the databases used. Additionally, we were able to identify some new ANKK1 orthologues. This included an unannotated ~1.5kb long ANKK1 ortholog sequence in the porcine (Sus scrofa) genome (Table S1), comprising three exons (orthologous to the human exons 3, 4 and 5; 76% identity), from which a 119 amino acid-long protein sequence could be predicted (86% identity). This ANKK1 orthologue in pigs is located downstream from DRD2, changing the gene order in the cluster from NTAD to NTDA (NCAM1-TTC12-DRD2-ANKK1). This interesting result illustrates a unique chromosomal inversion in the porcine lineage (Figure 1). Two other probable new ANKK1 orthologues were identified in the alpaca (Vicugna pacos) and rabbit (Oryctolagus cuniculus) genomes (Table S1), but in both cases the expected NTAD gene order was retained.

Conserved synteny and neighborhood of the whole NTAD cluster was observed in 22 of the 48 vertebrate species studied (46%), other 4 species present conserved synteny but do not share full neighborhood (Teleostei: Danio rerio, Gasterosteus aculeatus, Tetraodon nigroviridis; Artiodactlyla: Sus scrofa; Figure 1). The synteny and neighborhood status of the NTAD cluster in the other genomes could not be ascertained due to low coverage and/or incomplete assembly.

One of the main features of the NTAD cluster (Figure S1) is that *DRD2* is located in the reverse strand (or minus strand) in relation to the other three genes in all vertebrate genomes, except in the porcine genome due to the inversion described above. The presence of more than one *NCAM1* and *DRD2* orthologue copies in teleost fishes (Figure 1B, Table S1) is in agreement with the known genome duplication in this taxon (Jaillon *et al.*, 2009). However, we did not observe conserved synteny in all copies of *NCAM1* and *DRD2*, since the extant copies not included in

the NTAD cluster are located in different chromosomes (Table S1). This raises the question of possible subfunctionalization or neofunctionalization of these gene copies.

It is worthy of note that NTAD cluster synteny is conserved in teleost fishes, but not the neighborhood of its genes. About three million bases, containing ~100 genes, separate *NCAM1* from the TAD cluster (*TTC12-ANKK1-DRD2*), which can be tracked back to at least ~525 Mya in the origin of vertebrates. The *NCAM1* neighborhood was apparently gained when the Sarcopterygians emerged ~400 Mya and seems to have been maintained since then (Figure 1C).

Although phenomena such as tandem duplications, inversions, rearrangements and indels may account for non-random patterns of the genome (Hurst *et al.*, 2004), none of these explain by themselves the origin and conservation of the NTAD cluster. Thus other hypotheses need to be considered.

Several authors have demonstrated that clustered genes are kept together for a long period of time to preserve intact their co-regulatory system and consequently phenotype integrity. For instance, several cis-regulatory sequences are preserved throughout all vertebrate genomes due to their role in development (Kikuta et al., 2007). One interesting fact about the NTAD gene cluster is that a polymorphism in one gene might indirectly affect the expression levels of a neighboring gene. Huang et al. (2008) demonstrated that the ANKK1 rs2734849 SNP alters the expression level of NF-κB-regulated genes and, since DRD2 gene expression is regulated by the transcription factor NF-κB (Fiorentini et al., 2002; Bontempi et al., 2007), it might be indirectly regulated by ANKK1. The search for regulatory transcription factors in the DECODE database showed that the NCAM1 gene also seems to be regulated by NF-κB, as well as TTC12 paralogues (Table S2), denoting a possible role of this transcription factor in the co-regulation of the NTAD genes.

Based on computer simulations, Yerushalmi and Teicher (2007) showed an extraordinary tendency for essential genes to cluster as a result of natural selection pressures. Our results illustrate, for the first time to our knowledge, such a tendency for genes with essential functions in neurogenesis and dopaminergic neurotransmission. Thus, it is likely that natural selection, through the formation of the NTAD cluster, has played a role in the emergence of an efficient mechanism of co-regulation, when the vertebrate central nervous system acquired novel traits and gained complexity. Equally important must have been the role of natural selection in maintaining the NTAD cluster practically intact for at least 400 million years.

Notwithstanding, certain limitations must be considered when interpreting the results of the present study. Several assumptions presented here rely on available genomic data and analyses that are still preliminary. This is especially true for tests considering alternative explanations for

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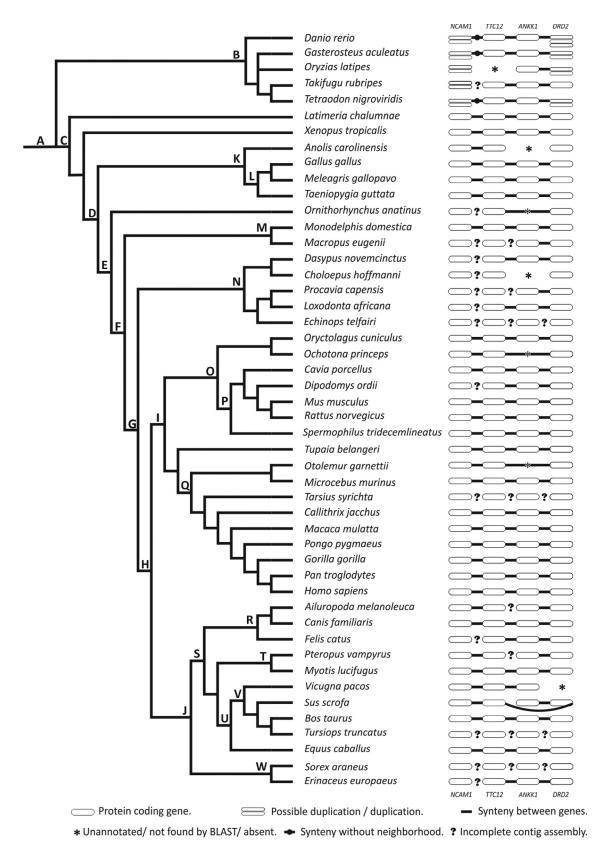


Figure 1 - Conservation of synteny and neighborhood in the NTAD cluster across 48 vertebrate genomes. The tree was compiled based on Agnarsson *et al.* 2010, Asher and Helgen 2010, Horner *et al.* 2007, Janes *et al.* 2010, Page and Goodman 2001, Pacheco *et al.* 2011. **A**: Vertebrata; **B**: Teleostei; **C**: Sarcopterygii; **D**: Amniota; **E**: Mammalia; **F**: Theria; **G**: Eutheria; **H**: Boreoeutheria; **I**: Euarchontoglires; **J**: Laurasiatheria; **K**: Sauria; **L**: Aves; **M**: Marsupialia; **N**: Atlantogenata; **O**: Glires; **P**: Rodentia; **Q**: Primates; **R**: Carnivora; **S**: Scrotifera; **T**: Chiroptera; **U**: Euungulata; **V**: Artiodactyla; **W**: Lipotyphla.

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specific SNPs that are widespread in the *Homo genus* and for the rate of maintained of four-gene clusters since or before the origin of vertebrates. We understand that a major goal of this study is to stimulate further research in the evolutionary history of gene clusters using the currently available genomic data and emerging bioinformatics tools.

Conclusion

Our results suggest that genes related to neurogenesis and dopaminergic neurotransmission may be interconnected in the course of the evolution of the complex vertebrate neural system via a common functional genomic architecture and chromosomal dynamics. Associated with due consideration of linkage disequilibrium patterns, this denotes the importance of approaching the NTAD cluster as a candidate functional unit, rather than its genes separately, in behavioral and psychiatric genetic studies.

Acknowledgments

We thank Brazilian funding agencies Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), Conselho de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), Fundação de Amparo a Pesquisa do Rio Grande do Sul (FAPERGS) and Programa de Apoio a Núcleos de Excelência (PRONEX) for financial support.

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Yerushalmi U and Teicher M (2007) Examining emergence of functional gene clustering in a simulated evolution. Bull Math Biol 69:2261-2280.

Internet Resources

- CisRED: Database of genome-wide regulatory module and element predictions, http://www.cisred.org/ (June 20, 2011).
- DECODE: Decipherment of DNA Elements, http://www.sabiosciences.com/chipqpcrsearch.php?app=T FBS (July 4, 2011).
- Ensembl Genome Browser, http://www.ensembl.org/index.html (July 5, 2011).
- GeneCards v.3: the Human Genome Compendium, http://www.genecards.org/ (May 17, 2011).
- UCSC Genome Browser, http://genome.ucsc.edu/ (July 10, 2011).
- UniProt Protein Database, http://www.uniprot.org/ (April 5, 2011).
- PolyPhen2: Prediction of Functional Effects of Human nsSNPs, http://genetics.bwh.harvard.edu/pph2/ (July 12, 2011).
- SNAP: Effects of Single Amino Acid Substitutions on Protein Function, http://rostlab.org/services/snap/, (July 12, 2011).

Supplementary Material

- The following online material is available for this article:
- Table S1 Orthologues of genes in the NTAD cluster in 48 available vertebrate genomes.
- Table S2 Transcription factors predicted by DECODE as regulators of the NTAD genes in humans.
 - Figure S1 NTAD cluster in the human genome.
- This material is available as part of the online article from http://www.scielo.br/gmb.

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Table S1. Orthologues of genes in the NTAD cluster in 47 available vertebrate genomes, showing accession number and location.

Taxon	Genome	Coverage	NCAM1	TTC12	ANKK1	DRD2
	Danio rerio (Zebrafish)		ncam1a	ENSDARG00000056896	ENSDARG00000056921	drd2a
			ENSDARG00000056181	<u>15:20908817-20942192:1</u>	<u>15:20975671-20986596:1</u>	ENSDARG00000056926
			<u>21:22615780-23024668:-1</u>			<u>15:21008646-21030749:-1</u>
			ncam1b			drd2b
		~7.5X	ENSDARG00000007220			ENSDARG00000011091
			<u>15:17312414-17518321:-1</u>			5:59424412-59461143:-1
						drd2l
						ENSDARG00000014858
						<u>16:57790203-57818030:1</u>
	Gasteroteus aculeatus (Stickleback)		1 of 2	ENSGACG00000009165	ENSGACG00000009158	drd2
			ENSGACG00000020773	groupl:8558661-8571918:-1	groupl:8533757-8538878:-1	ENSGACG00000009131
		6X	groupVII:24387904-24556179:-1			groupl:8509579-8522810:1
			2 of 2			prediction
TELEOSTEI			ENSGACG00000010697			ENSGACG00000012513
			groupl:11344900-11377049:1			groupXX:14002115-14008728:1
	Oryzias latipes (Medaka)		1 of 2	absent	ENSORLG00000007503	drd2
			ENSORLG00000014681		<u>13:15062437-15069570:1</u>	ENSORLG00000007515
		10.6X	<u>14:29382768-29464062:-1</u>			<u>13:15093829-15118346:-1</u>
			2 of 2			prediction
			ENSORLG00000005127			ENSORLG00000015196
			<u>13:10809441-10820108:-1</u>			<u>16:22530038-22544806:1</u>
	Takifugu rubripes (Fugu)	Takifugu rubripes (Fugu)	1 of 2	ENSTRUG00000014546	ENSTRUG00000014611	ENSTRUG00000014690
			ENSTRUG00000002966	scaffold 9:1673315-1683905:1	scaffold 9:1700087-1707491:1	scaffold 9:1719056-1732814:-1
	8.5X	8.5X	scaffold 454:35293-71171:-1			
			2 of 2			
			ENSTRUG00000007164			
			scaffold 156:454454-475510:-1			

Table S1 (cont). Orthologues of genes in the NTAD cluster in 47 available vertebrate genomes, showing accession number and location.

Taxon	Genome	Coverage	NCAM1	TTC12	ANKK1	DRD2
-	Tetraodon nigroviridis (Tetraodon)		1 of 2	ENSTNIG00000001367	ENSTNIG00000007778	drd2
			ENSTNIG00000003848	<u>16:938567-948048:-1</u>	<u>16:921218-925980:-1</u>	ENSTNIG00000007779
TELEOSTEI		7.9X	<u>Un_random:27638663-27656353:-1</u>			16:901459-913203:1
TELEOSTEI		7.98	2 of 2			prediction
			ENSTNIG00000016188			ENSTNIG00000004342
_			<u>16:3055810-3078332:1</u>			<u>Un_random:68441708-68447001:-1</u>
COELACANTHIMORPHA	Latimeria chalumnae	~7X	ENSLACG00000018069 JH126571.1:2687432-2874385	ENSLACG00000018234 JH126571.1:2926015-2964315	ENSLACG00000018279 JH126571.1:2974006-2990122	ENSLACG00000018381 JH126571.1:3151137-3199198
ANURA	Xenopus tropicalis	7.65X	ENSXETG00000011665	ENSXETG00000011664	ENSXETG00000011663	ENSXETG00000011662
71110101		7.03%	GL172954.1:1273359-1333515:-1	GL172954.1:1165658-1229867:-1	GL172954.1:1081038-1104804:-1	GL172954.1:921005-972728:1
	Anolis carolinensis (Anole Lizard)		ENSACAG00000008287	ENSACAG00000008658	absent	ENSACAG00000008768
		7.1X	GL343354.1:608176-677439:-1	GL343354.1:553247-588419:-1	-	GL343354.1:451491-460712:1
					_	
					-	_
	Gallus gallus (chicken)		ENSGALG00000007839	ENSGALG00000007833	ANKK1	ENSGALG00000007794
			24:5933204-6012150:-1	24:5913010-5927569:-1	ENSGALG00000007815	<u>24:5874347-5879422:1</u>
		6.6X			24:5895790-5901363:-1	
					LOC415708 (possible ortholog)	
SAURIA					ENSGALG00000021325	
					11:2632306-2645774:1	
	Meleagris gallopavo (Turkey)	17X	ENSMGAG0000004316	ENSMGAG00000004280	ENSMGAG00000004269	ENSMGAG00000004223
			<u>26:6151710-6193006:-1</u>	<u>26:6115795-6128289:-1</u>	<u>26:6094781-6100367:-1</u>	<u>26:6073432-6078559:1</u>
	Taeniopygia guttata (Zebra Finch)		ENSTGUG00000000275 24:1260906-1302411:-1	ENSTGUG00000000270 24:1238535-1250741:-1	ANKK1 ENSTGUG00000000267	ENSTGUG00000000255 24:1191855-1198991:1
		CV.			24:1217973-1221928:-1	
		6X			prediction	
					ENSTGUG00000006862	
					<u>11:6734086-6737862:1</u>	

Table S1 (cont). Orthologues of genes in the NTAD cluster in 47 available vertebrate genomes, showing accession number and location.

Taxon	Genome	Coverage	NCAM1	TTC12	ANKK1	DRD2
	Ornithorhynchus anatinus (Platypus)		NCAM1	ENSOANG00000010971	absent	ENSOANG00000010970
MONOTREMATA		6X	ENSOANG00000009247	Contig8462:8719-29891:-1		Contig8462:1327-6313:1
			Contig13035:10432-20417:1			
	Monodelphis domestica (Opossum)	6.8X	ENSMODG00000014314	ENSMODG00000014228	ENSMODG00000014205	ENSMODG00000014196
		0.07	<u>4:239164918-239249671:-1</u>	4:239070356-239121360:-1	4:239028762-239043233:-1	<u>4:238978598-239009228:1</u>
MARSUPIALIA	Macropus eugenii (Wallaby)		ENSMEUG00000012605	ENSMEUG00000009795	ENSMEUG00000000701	ENSMEUG00000000707
		2X	GeneScaffold_5807:10353-87553:1	GeneScaffold_5805:596-50221:1	GeneScaffold 5806:4947-18128:1	GeneScaffold_5806:34377-57516:-1
			-	_		_
			_		_	_
	Dasypus novemcinctus (Armadillo)	2X	ENSDNOG00000009390	ENSDNOG00000024773	ENSDNOG00000013438	ENSDNOG00000007008
XENARTHRA			GeneScaffold 4191:71042-134109:1	GeneScaffold 4190:3973-86588:1	GeneScaffold 4190:113028-126154:1	GeneScaffold 4190:138117-150433:-1
	Choloepus hoffmanni (Sloth)	2.05X	ENSCHOG00000002918	ENSCHOG00000010881	absent	ENSCHOG00000008200
			GeneScaffold 4607:4688-64734:1	GeneScaffold 4606:474-34240:1		GeneScaffold 4608:898-13027:-1
	Oryctolagus cuniculus (Rabbit)	7.48X	ENSOCUG00000006190	ENSOCUG00000011625	blat UCSC (85% id.)	ENSOCUG00000011657
LAGOMORPHA			1:103078226-103142725:-1	1:103004607-103049887:-1	chr1:102,979,282-102,990,588	<u>1:102961300-102971002:1</u>
	Ochotona princeps (Pika)	1.93X	ENSOPRG00000014977	ENSOPRG00000015703	absent	ENSOPRG00000015750
			GeneScaffold 2905:194352-257089:1	GeneScaffold 2905:281538-325574:1		GeneScaffold 2905:347086-356689:-1
	Cavia porcellus (Guinea pig)	6.79X	ENSCPOG00000004681	ENSCPOG00000004685	ENSCPOG00000001170	ENSCPOG00000009265
			scaffold 19:34540713-34601476:-1	scaffold 19:34455258-34511005:-1	scaffold 19:34420990-34430699:-1	scaffold 19:34398884-34411581:1
	Dipodomys ordii (Kangaroo rat)	1.85X	ENSDORG00000015038	ENSDORG00000013510	ENSDORG00000013515	ENSDORG00000013516
			scaffold 2068:5606-68849:-1	GeneScaffold 3885:3227-33111:1	GeneScaffold 3885:49442-58538:1	GeneScaffold 3885:66648-74803:-1
RODENTIA	Mus musculus (Mouse)		ENSMUSG00000039542	ENSMUSG00000040219	ENSMUSG00000032257	ENSMUSG00000032259
			<u>9:49310257-49607027:-1</u>	<u>9:49245068-49294330:-1</u>	<u>9:49223327-49235126:-1</u>	9:49148732-49216282:1
	Rattus norvegicus (Rat)	~2X	ENSRNOG00000031890	ENSRNOG00000008595	ENSRNOG00000025037	ENSRNOG00000008428
			<u>8:52822361-52885628:-1</u>	<u>8:52734796-52799135:-1</u>	<u>8:52714745-52722913:-1</u>	8:52641169-52707749:1
	Spermophilus tridecemlineatus (Squirrel)	1.90X	ENSSTOG00000003495	ENSSTOG00000003513	ENSSTOG00000003546	ENSSTOG00000003570
			GeneScaffold 3272:15324-20843:1	GeneScaffold 3272:98637-146451:1	GeneScaffold 3272:160571-171793:1	GeneScaffold 3272:179831-197728:-1

Table S1 (cont). Orthologues of genes in the NTAD cluster in 47 available vertebrate genomes, showing accession number and location.

	Genome	Coverage	NCAM1	TTC12	ANKK1	DRD2
Soil	prex araneus (Shrew)	1.9X	ENSSARG00000006760	ENSSARG0000005049	ENSSARG0000003877	ENSSARG0000001799
		1.98	scaffold 255145:60004-73832:-1	GeneScaffold 4137:32399-65702:1	GeneScaffold 5578:4431-103273:1	GeneScaffold 4138:3143-94291:-1
INSECTIVORA <i>Eci</i>	chinops telfairi (Lesser hedgehog tenrec)	2X	ENSETEG00000010392	ENSETEG00000009125	ENSETEG00000002513	ENSETEG00000012787
INSECTIVORA		2/	scaffold 319785:74303-79506:-1	GeneScaffold 4950:12296-130505:1	GeneScaffold 6711:8112-56359:1	scaffold 222770:3-9483:-1
Eri	rinaceus europaeus (Hedgehog)	1.86X	ENSEEUG00000000446	ENSEEUG00000004523	ENSEEUG00000004637	ENSEEUG00000004688
		1.00/	GeneScaffold 5083:215781-233965:1	GeneScaffold 5082:985-76059:1	GeneScaffold 5082:95143-106027:1	GeneScaffold 5082:114423-125550:-1
SCANDENTIA Tu	upaia belangeri (Tree Shrew)	2X	ENSTBEG00000002250	ENSTBEG00000003009	ENSTBEG00000004173	ENSTBEG00000005104
			GeneScaffold 3191:283798-297944:1	GeneScaffold 3191:385371-440831:1	GeneScaffold 3191: 463,541-476,825	GeneScaffold 3191:485260-525277:-1
Ta	arsius syrichta (Tarsier)	1.82X	ENSTSYG00000009955	ENSTSYG00000012003	ENSTSYG00000011390	ENSTSYG00000012280
			GeneScaffold 4816:549-67107:1	GeneScaffold 4815:897-48462:1	GeneScaffold 6508:3144-6436:1	GeneScaffold 4817:26-5539:-1
Ot	tolemur garnettii (Bushbaby)	1.5X	ENSOGAG00000006401	ENSOGAG00000006402	absent	ENSOGAG00000006407
			GeneScaffold 3006:89395-100429:1	GeneScaffold 3006:190790-254568:1		GeneScaffold 3006:308141-320255:-1
Mi	licrocebus murinus (Mouse Lemur)	1.93X	ENSMICG00000003994	ENSMICG0000004006	ENSMICG00000004027	ENSMICG00000004037
			scaffold_568:82754-94374:1	scaffold_568:171666-232202:1	scaffold_568:252261-258554:1	scaffold 568:267022-280232:-1
Са	allithrix jacchus (Marmoset)	6X	ENSCJAG00000012342	ENSCJAG00000012306	ENSCJAG00000012279	ENSCJAG00000012246
			<u>11:10710214-10782569:1</u>	<u>11:10812645-10872609:1</u>	11:10889021-10904331:1	11:10914111-10929399:-1
PRIMATES Ma	lacaca mulatta (Macaque)	5.1X	ENSMMUG00000004688	ENSMMUG00000014328	ENSMMUG00000014331	ENSMMUG00000014334
PRIIVIATES			14:111677926-111745516:1	14:111778513-111837307:1	14:111852166-111865392:1	14:111874406-111941325:-1
Po	ongo pygmaeus (Orangutan)	6X	ENSPPYG00000003883	ENSPPYG00000003884	UCSC	ENSPPYG00000003885
			11:109978390-110064543:1	11:110100792-110153360:1	chr11:110,175,216-110,187,450	<u>11:110197370-110212382:-1</u>
Go	orilla gorilla	2.1X	ENSGGOG00000013097	ENSGGOG00000003228	ENSGGOG00000003237	ENSGGOG00000003244
			11:111259892-111345449:1	<u>11:111381685-111435513:1</u>	11:111455141-111468472:1	<u>11:111477670-111542012:-1</u>
Pa	an troglodytes (Chimpanzee)	6X	ENSPTRG00000004291	ENSPTRG00000004292	ENSPTRG00000004293	ENSPTRG00000004295
			<u>11:111759836-112076094:1</u>	<u>11:112109519-112180130:1</u>	<u>11:112184597-112196060:1</u>	<u>11:112208040-112222288:-1</u>
Ho	omo sapiens		ENSG00000149294	ENSG00000149292	ENSG00000170209	ENSG00000149295
		high	11:112831997-113149158:1	11:113185251-113254266:1	11:113258513-113271140:1	11:113280318-113346001:-1

Table S1 (cont). Orthologues of genes in the NTAD cluster in 47 available vertebrate genomes, showing accession number and location.

Taxon	Genome	Coverage	NCAM1	TTC12	ANKK1	DRD2
	Pteropus vampyrus (Megabat)	2.63X	ENSPVAG00000007824	ENSPVAG00000007826	ENSPVAG00000012852	ENSPVAG00000012853
CHIROPTERA		2.03/	GeneScaffold 2166:404868-466481:1	GeneScaffold 2166:495074-539120:1	GeneScaffold 2977:967-5576:1	GeneScaffold 2977:14934-21721:-1
CHINOI TENV	Myotis lucifugus (Microbat)	7X	ENSMLUG00000011838	ENSMLUG00000011854	ENSMLUG00000011881	ENSMLUG00000011890
-		, , , , , , , , , , , , , , , , , , ,	GeneScaffold 3345:48657-61150:1	GeneScaffold 3345:144154-255563:1	GeneScaffold 3345:291777-293760:1	GeneScaffold 3345:302226-349693:-1
	Felis catus (Cat)	1.87X	ENSFCAG00000013891	ENSFCAG00000008291	ENSFCAG00000008292	ENSFCAG00000008293
			GeneScaffold 2791:232882-301289:1	GeneScaffold 2790:17645-63773:1	GeneScaffold 2790:84437-101130:1	GeneScaffold 2790:109558-121232:-1
CARNIVORA	Canis familiaris (Dog)	7.6X	ENSCAFG00000013844	ENSCAFG00000013902	ENSCAFG00000013898	ENSCAFG00000013890
			<u>5:22907424-22969273:-1</u>	<u>5:22838477-22881739:-1</u>	<u>5:22810681-22822862:-1</u>	<u>5:22791168-22802738:1</u>
	Ailuropoda melanoleuca (Panda)	2X	ENSAMEG00000016517	ENSAMEG00000016494	ENSAMEG00000006163	ENSAMEG00000006143
			GL193553.1:69580-131079:-1	GL193553.1:2181-47302:-1	GL193846.1:430758-441555:-1	GL193846.1:411593-423949:1
	Bos taurus (Cow)	7.1X	ENSBTAG00000005710	ENSBTAG0000010008	ENSBTAG00000010855	ENSBTAG00000010860
			<u>15:21855766-22058994:1</u>	<u>15:22152303-22197195:1</u>	<u>15:22225501-22235297:1</u>	15:22246344-22259339:-1
ARTIODACTYLA	Sus scrofa (Pig)	4X	ENSSSCG00000015045	ENSSSCG00000015047	BLAST (80% id)	ENSSSCG00000015048
			9:39876973-39943731:1	9:39974571-40018618:1	9:40074571-40076030:-1	9:40051843-40065065:1
	Vicugna pacos (Alpaca)	2.51X	ENSVPAG00000007274	ENSVPAG00000007275	blast (86% id.)	absent
			GeneScaffold 1967:356795-433389:1	GeneScaffold 1967:464956-509310:1	GeneScaffold_1967:536135-536217	
CETACEA	Tursiops truncatus (Dolphin)	2.59X	ENSTTRG00000000675	ENSTTRG00000010956	ENSTTRG00000014457	ENSTTRG00000007343
			GeneScaffold 1901:84476-148554:1	GeneScaffold 1900:5132-54722:1	scaffold 84548:9507-22715:-1	scaffold 111984:75732-88099:1
PERISSODACTYLA	Equus caballus (Horse)	6.8X	ENSECAG00000019193	ENSECAG00000017966	ENSECAG00000011781	ENSECAG00000013567
			<u>7:21647755-21713496:1</u>	<u>7:21745287-21793507:1</u>	7:21816607-21822636:1	<u>7:21833004-21893006:-1</u>
HYRACOIDEA	Procavia capensis (Hyrax)	2.19X	ENSPCAG00000005740	ENSPCAG00000016245	ENSPCAG00000006102	ENSPCAG00000006199
			scaffold 1490:10531-76838:-1	GeneScaffold 4301:8305-47937:1	GeneScaffold 5824:10185-18859:1	GeneScaffold 5824:29553-42938:-1
PROBOSCIDEA	Loxodonta africana (Elephant)	7X	ENSLAFG00000005371	ENSLAFG00000017571	ENSLAFG00000025747	ENSLAFG00000006523
			scaffold 147:1390575-1461424:1	scaffold 58:59908-112135:1	scaffold 58:151268-165444:1	scaffold 58:176422-189953:-1

Table S2. Transcription factors predicted by DECODE as regulators of the NTAD genes in humans.

Gene	Transcription factors
NCAM1	NF-κB , GR-α, GR-β, AP-1, c-Fos, c-Jun, p53, Sox9, POU2F1.
TTC12*	HOXA9B, Meis-1, RFX1, MIF-1, AREB6, Nkx3-1 v4, Nkx3-1, Nkx3-1 v1, Nkx3-1 v2, Nkx3-1 v3.
ANKK1	NF-кВ, CUTL1, AREB6, oct-B3, oct-B2, Oct-B1, POU2F2, POU2F2 (Oct-2.1), POU2F1, POU2F2B, POU2F2C.
DRD2	NF-κB , Egr-1, CREB, GR- α , GR- β , δCREB, c-Rel, p300, NRSF form 1.

^{*}Although NF-kB was not predicted as a regulator of *TTC12*, it regulates the *TTC12* paralogues *TOMM34* and *STIP1* as well as other genes encoding the tetratricopeptide repeat domain (not shown).

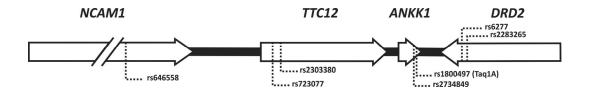


Figure S1- The NTAD cluster in the human genome, showing the single nucleotide polymorphisms (SNPs) included in this study. The sizes of the four genes are 317 kb, 69 kb, 13 kb and 66 kb, from left to right. Only *DRD2* is positioned in the reverse strand (arrow pointing to the left).