



Effects of *Origanum Syriacum* Essential Oil on Blood Parameters of Broilers Reared at High Ambient Heat

<http://dx.doi.org/10.1590/1806-9061-2017-0511>

■ Author(s)

Tekce E^I
Gül M^{II}

^I Faculty of Applied Sciences, Bayburt University, Bayburt 69000, Turkey

^{II} Animal Nutrition and Nutrition Disease, Atatürk University Erzurum, Turkey

■ Mail Address

Corresponding author e-mail address
Emre Tekce
Organic Agriculture Management, Faculty
Of Applied Sciences, Bayburt University,
Bayburt 69000, Turkey.
Tel: +90 458-333-20-33
Email: Vet_emre_tekce@hotmail.com

■ Keywords

Broiler, *Origanum Syriacum* essential oil,
Biochemical and Hematological.

ABSTRACT

The biochemical and hematological blood values of broilers fed under heat stress and that were administered *Origanum syriacum* (OS) essential oil were investigated in this study. In total, 400 male broilers (Ross-308; age, 1 day) were used for the period of 42-days of the study. The experiment included ten groups (n = 50/group; each group divided into five sub-groups): normal heat, 22°C: basal feed (BF)-positive control group (PC), BF + 100 ppm OS; BF + 300 ppm OS; BF + 600 ppm OS). Heat stress, 36°C: BF-positive control group (PC), BF + 100 ppm OS; BF + 300 ppm OS; and BF + 600 ppm OS). Biochemical and hematological parameters were measured in blood collected into EDTA tubes. Total bilirubin, cholesterol, low-density cholesterol, Na⁺, Ca²⁺, and Mg²⁺ increased, whereas alanine aminotransferase, aspartate aminotransferase, creatine kinase (CK), CK-MB, urea, uric acid, Cl⁻, and K⁺ decreased ($p < 0.05$); however, no changes were detected in the other hematological values.

INTRODUCTION

Stress emerges from endogenous (nutrition and feeding problems, fast growth, sexual maturation period, and infection) and exogenous factors (climate, high density insufficient ventilation), and causes dire economic losses in poultry (Chenga & Muirb, 2004; Olanrewaju *et al.*, 2006; Hadımlı *et al.*, 2007). One stressor is high temperature which causes hyperventilation, blood pH > 7, and a decrease in plasma pyruvate kinase along with an increase in CO₂ density in the lungs; thus, causing livestock to perish within a short period (Hartlova *et al.*, 2002; Lin *et al.*, 2006; Al-fataftah & Abu-dieyh, 2007). Antibiotics have been used in the poultry industry to reduce the impact of stress, increase stock performance, and regulate gastrointestinal microflora. Nevertheless, the European Union (EU) banned antibiotics when the World Health Organization reported that microorganisms can become immune to specific antibiotics over time thus causing the emergence of resistant bacteria that pose a risk to human health (Gue *et al.*, 2004; Yörük *et al.*, 2008). In addition, growing sensitivity towards issues, such as human health, food safety, and environmental pollution, have emerged among consumers despite the progresses achieved in food production techniques and slaughterhouse hygiene in EU countries (Mazmanoğlu, 2008). Therefore, growing organic products by adding natural feed additives as an alternative to synthetic additives has begun in recent years. One of these alternative feed additives is thyme. Thyme is a herb in the Lamiaceae family with 40 species worldwide. Thyme contains carvacrol and thymol along with various phenolic monoterpenes. *Origanum syriacum* (OS) is an aromatic herb within the thyme family that grows primarily in Syria, Jordan, Lebanon, and the Sinai Peninsula, where it has different names, such as Syrian oregano, biblical hyssop,

Submitted: 03/March/2017

Approved: 15/August/2017



Za'atar, and marjoram (Alma *et al.*, 2003; IUCN, 2005; Lukas, 2010). While the fatty acids in the OS essential oil generally contains polymorphic compounds, such as thymol and carvacrol, compounds, such as γ -terpinene, P-cymene, and thymoquinene, can also be found depending on the growing area (Lukas *et al.*, 2009). Some studies have reported that essential fatty acids in OS have antioxidant (Luna *et al.*, 2010; Tavarez *et al.*, 2011), anti-microbial (Mitsch *et al.*, 2004; Gürakan *et al.*, 2008), anti-inflammatory (Yoshino *et al.*, 2006), anti-viral (Pilau *et al.*, 2011), anti-tumoral (El-desoukya *et al.*, 2009; Al-kalaldehy *et al.*, 2010), anti-ulcer (Afify *et al.*, 2012), anti-fungal (Manohar *et al.*, 2001), and anti-parasitic (Değerli *et al.*, 2012; Remmal *et al.*, 2013) properties, all of which have positive effects on stock performance (Jang *et al.*, 2007; Tekçe & Gül, 2016).

In this study, the effects of various doses of OS essential oil added to feed as an alternative to antibiotics in broilers fed under nominal heat (22°C) and heat stressed (36°C) conditions on biochemical and hematological parameters were studied.

MATERIALS AND METHODS

Animals, Experimental Design, Feeds

In this study, 400 1-day-old Ross-308 male broilers were used in the experiment. During the 7-day adaptation and a 35-day fattening period, the animals were kept indoors in 121 × 110 × 108 cm cages at

Atatürk University Veterinary Faculty Husbandry Research and Practice Unit; each group consisted of 10 animals. For each of the experimental periods, 5 treatments were prepared by supplementing the standard commercial ration with 100 mg/kg of Avilamycin antibiotic (Kartal chem, İstanbul, Turkey) or 0 mg/kg (PC), 100 mg/kg, 300 mg/kg or 600 mg/kg of OS. The animals were equal in live weight, and were separated into ten groups (PC, A-100, OS-100, OS-300, OS-600, NC, SA-100, SOS-100, SOS-300, and SOS-600) each of which contained 50 animals. Then, all groups were separated into five sub-groups each of which consisted of 10 animals. During the experimental period, the groups: positive control (PC, A-100, OS-100 ppm, OS-300 and OS-600) were exposed to a temperature of 22°C, while the NC, SA-100, SOS-100, SOS-300 and SOS-600 groups were exposed to 36°C. Basal broiler feed was given to the animals every day at the same hour (around 17:00). The PC and NC groups were provided with only the basal broiler feed, whereas the diets of the OS-100, OS-300, OS-600, SOS-100, SOS-300, and SOS-600 groups were supplemented with 100, 300, 600, 100, 300, and 600 ppm OS essential oil (Mahan cosmetic, Antakya, Turkey), respectively. Four different basal broiler feeds were given to the animals at certain intervals (Table 1). This study was approved by the ethics committee of Atatürk University Veterinary Faculty (25.10.2013/5/126).

Table 1 – Composition and analyses of the basal diet (g/kg).

| Raw Materials | Pre-Starter (0-14 d) | Starter (14-21 d) | Grower (21-28 d) | Finisher (28-42 d) |
|-------------------------------------|----------------------|-------------------|------------------|--------------------|
| Maize 7.6 | 52.11 | 53.77 | 47.73 | 50.12 |
| Wheat | - | - | 10 | 10 |
| Soybean meal 44% | 23.4 | 17.87 | 12.47 | 11 |
| Full Fat Soybean | 17 | 20 | 20 | 20 |
| Meat Bone's | - | 1 | 1 | - |
| Chicken Reputation 56 | 2.53 | 2.53 | 3.53 | 3.53 |
| Soy Oil | 1 | 1.53 | 1.27 | 0.6 |
| Animal Fat | - | - | 0.8 | 1.53 |
| Salt | 0.23 | 0.23 | 0.2 | 0.2 |
| L-Lysine | 0.17 | 0.15 | 0.15 | 0.15 |
| L-Threonine | 0.09 | 0.09 | 0.07 | 0.07 |
| Mono-calcium Phosphate | 0.87 | 0.6 | 0.47 | 0.53 |
| Vitamin-mineral premix ¹ | | | | |
| Chemical Analysis | | | | |
| ME (Mj/kg) | 12.64 | 13.08 | 13.40 | 13.44 |
| Crude Protein % | 23.2 | 23.4 | 23.3 | 22.5 |
| Crude Oil % | 8.2 | 8.8 | 8.9 | 9 |
| Dry Matter % | 89.3 | 90.5 | 90.7 | 89.6 |
| Crude Cellulose % | 3.4 | 3.7 | 4 | 4 |

The vitamin-mineral premix provided the following (per kg of diet): vitamin A, 12 000 IU; vitamin D3, 1500 IU; vitamin E, 50 mg; vitamin K3, 5 mg; vitamin B1, 3 mg; vitamin B2, 4 mg; vitamin B6, 4 mg; vitamin B12, 0.03 mg; calcium -D-pantothenate, 15 mg; folic acid, 1 mg; niacin, 25 mg; D-biotin, 0.115 mg; Co, 0.2 mg; Cu, 6 mg; Fe, 60 mg; K, 0.75 mg; Mn, 80 mg; Se, 0.15 mg; Zn, 60 mg.



Origanum Syriacum Essential Oil Composition

The OS essential oil contained 28.9% carvacrol, 19.9% γ -terpinene, 12.4% p-cymene, 7.1% thymol, and 6.5% α -terpinene.

Poultry House Heat Moisture and Illumination

The general temperature of the poultry house was 32–33°C during the first 2 days and 27–28°C during the next 5 days. On the other hand, a 36°C temperature and 75–85% relative humidity were applied to the groups subjected to heat stress, while 22°C temperature and 55–60% relative humidity were applied to the others. All groups were provided with illumination (60 W) for 24 hours.

Feed Analyses

A proximate analyses of the feeds used for this research was performed in accordance with the methods stated in A.O.A.C 2005.

Blood Analyses

On day 42 of the study, the chickens were fasted overnight for biochemical analyses. Then, 100 animals (10 from each group) were slaughtered randomly by cervical dislocation. Blood samples were collected into 10 ml biochemistry tubes and 5 ml EDTA hemogram tubes (Becton Dickinson Co. Brea, CA, USA), centrifuged at 4,100 rpm for 12 min at 4°C (NF 1200R, Nüve, Ankara Turkey), and the serum was transferred to Eppendorf tubes. Serum levels of glucose, albumin (ALB), creatinine, globulin, total protein, urea, alanine aminotransaminase (ALT), aspartate aminotransaminase (AST), glutamic pyruvate transaminase, total cholesterol, triglycerides (TG), high density lipoprotein cholesterol (HDL), low density lipoprotein cholesterol (LDL), very low density lipoprotein cholesterol (VLDL), Ca²⁺, P, Mg²⁺, Cl⁻, Fe²⁺, amylase, gamma-glutamyl transferase, alkaline phosphatase, and uric acid were assayed at the Erzurum Technical Chemistry and Medical Laboratories using a Cobas-8000 auto-analyzer, which is a closed spectrophotometric system, with Roche kits (Mannheim, Germany).

Statistical Analyses

The data were analyzed using SSPS 17.0. software (SPSS Inc., Chicago, IL, USA). Data are presented as mean \pm standard error. Repeated-measures analysis of variance was used to identify if differences existed

in the blood values between the groups. Duncan's multiple range test was applied to identify the group differences. A p-value < 0.05 was considered significant.

RESULTS

The effects of the OS essential oil added at different dosages (100, 300, and 600 ppm) on the metabolic, blood, and biochemical values are shown in Tables 2–5. Some of the serum values in broilers fed under heat stress and control conditions are as follows: glucose (22°C control: 270 vs. 36°C OS 300 ppm: 263.9 vs. 36°C O.S 600 ppm: 258.7 mM; $p < 0.01$; ALT (22°C control: 2.2 vs. 36°C OS 300 ppm: 1.3; $p < 0.00$); AST (22°C control: 295.8 vs. 36°C OS 300 ppm: 308.6; $p < 0.01$, ALB (22°C control: 1.3 vs. 36°C OS 300 and 600 ppm: 1.2; $p < 0.04$); CK-MB (22°C control: 0 vs. 36°C OS 600 ppm: 0; $p < 0.01$); Ca²⁺ (22°C control: 10.2 vs. 36°C OS 600 ppm: 11; $p < 0.01$); K⁺ (22°C control: 9.4 vs. 36°C OS 300 ppm: 6.6; $p < 0.01$). Adding the OS essential oil at various dosages had no effect on most hematological values compared to the values after administering the antibiotic or heat stress. However, the OS oil did have an effect on glucose ($p < 0.01$), liver enzymes (ALT and AST) and CK-MB ($p < 0.01$), which indicates heart and skeletal muscle degeneration. The 600 ppm OS essential oil dosage completely protected the muscles against various pathological defects ($p < 0.01$).

DISCUSSION

The levels of various biochemical parameters vary depending upon numerous biochemical reactions such as digestion and absorption of food. These parameters also vary with factors like strain, health, and environmental conditions (Gümüş, 2013). Blood glucose level is a biochemical indicator that increases in broilers under stress (Khaksar *et al.*, 2012). Due to increased gluconeogenesis and activation of muscle glycogen stores after corticosterone is released from the adrenal gland. While glucose and corticosterone concentrations increase in response to stress, the number of lymphocytes decreases, which diminishes the effectiveness of the humoral and cellular defense systems (Hayırlı *et al.*, 2005). Serum glucose level has been reported to increase in broilers as temperature rises over 30°C (Khan *et al.*, 2002; İsmail *et al.*, 2013; Toplu *et al.*, 2014). In our study, the OS essential oil had no effect on blood glucose level (Table 2), and the glucose levels after exposure to heat stress were

**Table 2** – Serum biochemical parameters in the experimental groups.

| Blood Biochemical | Glucose(mg/ dL) | | Triglycerides (mg/ dL) | | LDH U/L | | Uric acid mg/ dL | | Na mmol/L | | Cl mmol/L | |
|--------------------------------|---------------------|-----------|------------------------|----------|---------------------|--------------|-------------------|----------|---------------------|-----------|---------------------|-----------|
| | 22 °C | 36 °C | 22 °C | 36 °C | 22 °C | 36 °C | 22 °C | 36 °C | 22 °C | 36 °C | 22 °C | 36 °C |
| Control | 270,8±5,4 | 265,9±6,0 | 28,6±1,6 | 32,8±1,6 | 1399,7±160,8 | 2366,1±179,8 | 6,7±1,3 | 10,7±1,1 | 159,8±1,8 | 155,3±1,5 | 126,8±1,6 | 121,2±1,3 |
| Antibiotic 100 ppm | 245,9±5,4 | 252,6±6,0 | 29,6±1,6 | 38±1,6 | 1894,7±160,8 | 2206,7±179,8 | 6,4±3,3 | 10,3±1,2 | 158,0±4,5 | 154,7±1,6 | 122,0±4,0 | 119,1±1,4 |
| O.S 100 ppm | 237,7±5,4 | 280,2±6,0 | 32,8±1,6 | 37,3±1,6 | 1653,6±160,8 | 1999,1±169,5 | 6,0±1,9 | 7,3±1,1 | 157,3±2,6 | 161,8±1,5 | 123,0±2,3 | 122,7±1,3 |
| O.S 300 ppm | 242,0±5,4 | 263,9±6,0 | 33,7±1,6 | 34±1,6 | 2164,4±160,8 | 1596,0±169,5 | 4,0±1,9 | 5,3±1,1 | 155,0±2,6 | 156,8±1,5 | 120,3±2,3 | 121,7±1,3 |
| O.S 600 ppm | 258,3±5,4 | 258,7±6,0 | 30,6±1,6 | 38,3±1,6 | 1545,4±160,8 | 1868,4±160,8 | 5,6±1,2 | 6,5±1,4 | 160,9±1,6 | 162,5±1,8 | 125,0±1,4 | 124,0±1,6 |
| Source of variation (P values) | | | | | | | | | | | | |
| Diet | 0,00 | 0,00 | 0,00 | 0,00 | 0,01 | 0,01 | 0,04 | 0,04 | 0,98 | 0,98 | 0,20 | 0,20 |
| Temperature | 0,01 | 0,01 | 0,08 | 0,08 | 0,36 | 0,36 | 0,06 | 0,06 | 0,03 | 0,03 | 0,19 | 0,19 |
| Temperature x Diet | 0,00 | 0,00 | 0,10 | 0,10 | 0,00 | 0,00 | 0,71 | 0,71 | 0,15 | 0,15 | 0,29 | 0,29 |
| Main effect means diet | | | | | | | | | | | | |
| Control | 268,6 ^a | | 30,4 ^b | | 1829,2 ^a | | 9,1 ^a | | 157,1 ^{bc} | | 123,4 ^a | |
| Antibiotic 100 ppm | 248,9 ^b | | 33,3 ^{ab} | | 2033,4 ^a | | 9,8 ^a | | 155,1 ^c | | 119,4 ^b | |
| OS 100 ppm | 257,8 ^{ab} | | 33,8 ^a | | 1817,3 ^a | | 6,9 ^{ab} | | 160,6 ^{ab} | | 122,8 ^{ab} | |
| OS 300 ppm | 252,4 ^b | | 34,6 ^{ab} | | 1895,2 ^a | | 5,0 ^b | | 156,3 ^c | | 121,3 ^{ab} | |
| OS 600 ppm | 258,5 ^{ab} | | 34,9 ^a | | 1706,9 ^a | | 5,0 ^b | | 161,5 ^a | | 124,5 ^a | |
| Temperature | | | | | | | | | | | | |
| 22 °C | 250,9 | | 31,1 | | 1731,6 | | 5,74 | | 158,2 | | 123,4 | |
| 36 °C | 264,3 | | 36,1 | | 2007,3 | | 8,02 | | 158,2 | | 121,7 | |

Table 3 – Serum biochemical values in the experimental groups.

| Blood Biochemical | Urea mg/dL | | ALT U/L | | AST U/L | | Cholesterol mg/ dL | | HDL-C mg/ dL | | ALB g/ dL | |
|--------------------------------|-------------------|---------|-------------------|---------|---------------------|-------------|---------------------|-----------|--------------------|-----------|-------------------|----------|
| | 22 °C | 36 °C | 22 °C | 36 °C | 22 °C | 36 °C | 22 °C | 36 °C | 22 °C | 36 °C | 22 °C | 36 °C |
| Control | 4±0,8 | 5,8±0,6 | 2,2±0,4 | 4,0±0,4 | 295,8±25,86 | 478,3±25,86 | 150,6±5,7 | 135,7±5,7 | 98,7±3,7 | 85,6±3,7 | 1,3±0,06 | 1,2±0,06 |
| Antibiotic 100 ppm | 5±2,0 | 5,9±0,7 | 2,3±0,4 | 4±0,4 | 302,9±25,86 | 404,1±25,86 | 159,1±5,7 | 162,6±5,7 | 103,4±3,7 | 92,2±3,7 | 1,2±0,06 | 1,4±0,06 |
| O.S 100 ppm | 3,3±1,1 | 4,6±0,6 | 2±0,4 | 3±0,4 | 317,5±25,86 | 424,3±25,86 | 157,6±5,7 | 164,6±5,7 | 100,2±3,7 | 94±3,7 | 1,1±0,06 | 1,3±0,06 |
| O.S 300 ppm | 4,3±1,1 | 3,4±0,6 | 1,7±0,4 | 1,3±0,4 | 290,6±25,86 | 308,6±25,86 | 150,9±5,7 | 156±5,7 | 99,8±3,7 | 101,2±3,7 | 1±0,06 | 1,2±0,06 |
| O.S 600 ppm | 4,1±0,7 | 4,8±0,8 | 2,2±0,4 | 2,2±0,4 | 290,6±25,86 | 332±25,86 | 162,3±5,7 | 146,9±5,7 | 104,8±3,7 | 95,1±3,7 | 1,2±0,06 | 1,2±0,06 |
| Source of variation (P values) | | | | | | | | | | | | |
| Diet | 0,27 | 0,27 | 0,0 | 0,0 | 0,00 | 0,00 | 0,43 | 0,43 | 0,00 | 0,00 | 0,02 | 0,02 |
| Temperature | 0,62 | 0,62 | 0,0 | 0,0 | 0,00 | 0,00 | 0,02 | 0,02 | 0,21 | 0,21 | 0,05 | 0,05 |
| Temperature x Diet | 0,66 | 0,66 | 0,0 | 0,0 | 0,03 | 0,03 | 0,13 | 0,13 | 0,33 | 0,33 | 0,04 | 0,04 |
| Main effect means diet | | | | | | | | | | | | |
| Control | 5,1 ^{ab} | | 3 ^a | | 381,7 ^a | | 143,6 ^b | | 92,5 ^a | | 1,2 ^{ab} | |
| Antibiotic 100 ppm | 5,7 ^a | | 3,1 ^a | | 347,8 ^{ab} | | 160,6 ^a | | 98,4 ^a | | 1,3 ^a | |
| OS 100 ppm | 4,2 ^{ab} | | 2,5 ^{ab} | | 368,1 ^a | | 160,8 ^a | | 97,2 ^a | | 1,2 ^{ab} | |
| OS 300 ppm | 3,6 ^b | | 1,5 ^c | | 299,1 ^b | | 153,3 ^{ab} | | 100,4 ^a | | 1,1 ^b | |
| OS 600 ppm | 4,4 ^{ab} | | 2,2 ^{bc} | | 311,3 ^b | | 154,6 ^{ab} | | 99,9 ^a | | 1,2 ^{ab} | |
| Temperature | | | | | | | | | | | | |
| 22 °C | 4,1±0,5 | | 2,1 | | 299,4 | | 156,1 | | 101,3 | | 1,16 | |
| 36 °C | 4,8±0,3 | | 2,9 | | 389,4 | | 153,1 | | 93,6 | | 1,25 | |

**Table 4** – Serum biochemical values in the experimental groups.

| Blood Biochemical | ALP U/L | | CK-MB U/L | | Ca mg/dL | | Mg mg/dL | | K mmol/L | |
|--------------------------------|----------------------|--------------|----------------------|----------------|-------------------|----------|-------------------|---------|-------------------|---------|
| | 22 °C | 36 °C | 22 °C | 36 °C | 22 °C | 36 °C | 22 °C | 36 °C | 22 °C | 36 °C |
| Control | 1800,5±364,5 | 1663,8±364,5 | 0,0±2259,1 | 11895,6±2259,1 | 10,2±0,2 | 9,1±0,2 | 2,7±0,1 | 2,6±0,1 | 9,4±0,4 | 8,2±0,4 |
| Antibiotic 100 ppm | 1564,1±364,5 | 1511,8±364,5 | 0,0±2259,1 | 26065,7±2259,1 | 9,9±0,2 | 10,9±0,2 | 2,8±0,1 | 3±0,1 | 6,9±0,4 | 9,1±0,4 |
| O.S 100 ppm | 2712,8±364,5 | 2093,9±364,5 | 512,1±2259,1 | 17708,2±2259,1 | 10±0,2 | 10,9±0,2 | 2,7±0,1 | 2,9±0,1 | 8,4±0,4 | 6,6±0,4 |
| O.S 300 ppm | 2077,3±364,5 | 1966,2±364,5 | 1246,7±2259,1 | 12692,4±2259,1 | 10,4±0,2 | 10,3±0,2 | 2,5±0,1 | 2,9±0,1 | 7,8±0,4 | 8,1±0,4 |
| O.S 600 ppm | 1616,1±364,5 | 1870,3±364,5 | 0,0±2259,1 | 0,0±2259,1 | 10,8±0,2 | 11±0,2 | 2,9±0,1 | 3,1±0,1 | 8,4±0,4 | 6,7±0,4 |
| Source of variation (P values) | | | | | | | | | | |
| Diet | 0,58 | | 0,00 | | 0,24 | | 0,03 | | 0,12 | |
| Temperature | 0,18 | | 0,00 | | 0,00 | | 0,04 | | 0,03 | |
| Temperature x Diet | 0,83 | | 0,00 | | 0,00 | | 0,36 | | 0,00 | |
| Main effect means diet | | | | | | | | | | |
| Control | 1736,1 ^{ab} | | 5947,8 ^b | | 9,6 ^c | | 2,6 ^b | | 8,7 ^a | |
| Antibiotic 100 ppm | 1540,8 ^b | | 11584,7 ^a | | 10,3 ^b | | 2,9 ^{ab} | | 7,8 ^{ab} | |
| OS 100 ppm | 2419,6 ^a | | 8557,6 ^{bc} | | 10,4 ^b | | 2,8 ^{ab} | | 7,5 ^b | |
| OS 300 ppm | 2024,6 ^{ab} | | 6969,6 ^{bc} | | 10,3 ^b | | 2,7 ^b | | 7,9 ^{ab} | |
| OS 600 ppm | 1743,2 ^{ab} | | 0,0 ^c | | 10,9 ^a | | 3 ^a | | 7,5 ^b | |
| Temperature | | 1954,1±164,9 | | 351,7 | | 10,2 | | 2,7 | | 8,1 |
| 36 °C | | 1821,1±174,5 | | 13672,4 | | 10,4 | | 2,9 | | 7,7 |

Table 5 – Serum hematological in the experimental groups.

| Blood Hematology | WBC 10 ³ /μL | | RBC 10 ⁶ /μL | | HGB g/dL | | HCT % | | MCV fL | | MCH pg | | MCHC g/dL | |
|--------------------------------|-------------------------|------------|-------------------------|-----------|----------|-----------|----------|-----------|---------------------|------------|----------|-----------|--------------------|-----------|
| | 22 °C | 36 °C | 22 °C | 36 °C | 22 °C | 36 °C | 22 °C | 36 °C | 22 °C | 36 °C | 22 °C | 36 °C | 22 °C | 36 °C |
| Control | 279,2±7,4 | 298,3±7,4 | 2,4±0,8 | 2,7±0,8 | 10,8±0,3 | 12,1±0,3 | 37,9±1,2 | 42,6±1,2 | 155,8±1,4 | 156,9±1,4 | 44,6±0,4 | 44,8±0,4 | 28,6±0,2 | 28,5±0,2 |
| Antibiotic 100 ppm | 285,7±7,4 | 307,8±7,4 | 2,4±0,8 | 2,7±0,8 | 10,5±0,3 | 12,1±0,3 | 37,1±1,2 | 41,9±1,2 | 152,8±1,4 | 154,8±1,4 | 43,3±0,4 | 44,9±0,4 | 28,3±0,2 | 29,1±0,2 |
| O.S 100 ppm | 289,5±7,9 | 306,2±7,4 | 2,5±0,9 | 2,8±0,8 | 11,1±0,4 | 12,7±0,3 | 38,1±1,3 | 43,8±1,2 | 150,2±1,4 | 153,7±1,4 | 43,6±0,5 | 44,5±0,4 | 29,0±0,2 | 28,9±0,2 |
| O.S 300 ppm | 279,8±7,4 | 298,7±7,4 | 2,4±0,8 | 2,7±0,8 | 10,6±0,3 | 12,2±0,3 | 37,6±1,2 | 42,1±1,2 | 156,9±1,4 | 153,9±1,4 | 44,3±0,4 | 44,6±0,4 | 28,2±0,2 | 29,1±0,2 |
| O.S 600 ppm | 305,1±8,4 | 303,4±7,4 | 2,5±0,9 | 2,7±0,8 | 11,5±0,4 | 12,4±0,3 | 39,8±1,4 | 42,9±1,2 | 153,2±1,5 | 156,5±1,4 | 44,4±0,4 | 45,2±0,4 | 28,9±0,2 | 28,9±0,2 |
| Source of variation (P values) | | | | | | | | | | | | | | |
| Diet | 0,239 | | 0,392 | | 0,247 | | 0,579 | | 0,021 | | 0,323 | | 0,065 | |
| Temperature | 0,003 | | 0,000 | | 0,000 | | 0,000 | | 0,111 | | 0,009 | | 0,014 | |
| Temperature x Diet | 0,573 | | 0,823 | | 0,733 | | 0,898 | | 0,127 | | 0,500 | | 0,004 | |
| Main Effect Means Diet | | | | | | | | | | | | | | |
| Control | 288,7 | | 2,6 | | 11,5 | | 40,3 | | 156,4 ^a | | 44,7 | | 28,6 ^b | |
| Antibiotic 100 ppm | 296,8 | | 2,6 | | 11,3 | | 39,5 | | 153,8 ^{ab} | | 44,1 | | 28,7 ^{ab} | |
| OS 100 ppm | 298,4 | | 2,7 | | 11,9 | | 41,1 | | 152,0 ^b | | 44,1 | | 28,9 ^a | |
| OS 300 ppm | 289,3 | | 2,6 | | 11,4 | | 39,8 | | 155,4 ^a | | 44,5 | | 28,6 ^{ab} | |
| OS 600 ppm | 304,1 | | 2,7 | | 12,0 | | 41,6 | | 155,1 ^a | | 44,9 | | 28,9 ^a | |
| Temperature | | 287,9±3,47 | | 2,48±0,03 | | 10,9±0,15 | | 38,1±0,56 | | 153,7±0,62 | | 44,0±0,20 | | 28,6±0,07 |
| 36 °C | | 302,9±3,34 | | 2,75±0,03 | | 12,3±0,14 | | 42,0±0,54 | | 155,1±0,60 | | 44,8±0,20 | | 28,9±0,07 |



within the normal range for broilers (Reece, 2009). In our study, the effects of adding OS essential oil at different dosages to the diets of the heat stressed and unstressed groups were incompatible with some previous data (Khan *et al.*, 2002; Khaksar *et al.*, 2012; Ismail *et al.*, 2013).

Thymol and carvacrol, which are the main components of OS, have been reported to have a cholesterol lowering property by inhibiting hepatic 3-hydrox-3-methylglutaryl coenzyme A reductase, which is a cholesterol synthesis enzyme (Mazmanoğlu, 2008). Al-Kassie, (2009) reported that adding 200 ppm thyme oil to broiler feed significantly reduces cholesterol level. Khaksar *et al.* (2012) determined that 1 g/kg thyme oil in the feed of Japanese quail significantly reduces total cholesterol and TGs. Hong *et al.* (2012) reported that a mixture of 125 ppm thyme, anise seed, and citrus peel oils added to broiler feed significantly reduces cholesterol level. Sarica *et al.* (2005) stated that 1 g/kg thyme powder added to broiler feed significantly reduces total plasma cholesterol. In contrast, Demir *et al.* (2005) reported that 1 g/kg origanum powder added to broiler feed significantly increases total cholesterol and TG levels, and Bölükbaşı *et al.* (2006) obtained similar results with 200 ppm thyme oil. In the present study, 100, 300, and 600 ppm OS added to the feed of the heat stressed broiler group (Tables 2 and 3) significantly increased HDL, LDH, TG, and total cholesterol levels. This may be attributed to decreased hepatic 3-hydrox-3-methylglutaryl coenzyme A reductase enzyme activity. The high-fat diet also increased the capacity of the liver to store lipids and reduce liver function (Chawda *et al.*, 2014). OS also reduced CK-MB (Zhang *et al.*, 2014) and liver enzymes (ALT and AST) levels compared to the control and antibiotic groups.

Urea and uric acid are protein metabolites. Serum protein levels are important for preserving the immune system, and these values can increase under diseased or stressed conditions, such as toxicity (Arslan, 2012). Uric acid is an antioxidant that eliminates free radicals as a result of xanthine oxidase activity (Koizumi *et al.*, 1991). Al-kassie (2009) reported that 200 ppm origanum oil in boiler feed increases total serum protein, whereas Demir *et al.* (2005) found the same result with 1 g/kg origanum powder. In contrast, Köksal *et al.* (2012) reported a decrease in serum total protein in response to 0.75 g/kg essential oil mix (*Origanum vulgare*, *Thymus vulgare*, garlic, anise seed and raziyane). OS (300 ppm) significantly reduced urea and all dosages decreased uric acid levels in the heat stressed groups compared with those in the control and antibiotic

groups in the present study. The literature has few data on urea and uric acid in broilers.

In summary, the OS essential oil showed some protective effects against heat stress in broilers. We believe that it is necessary to better understand the effects of origanum syriacum essential oil by testing it under various stress conditions in further future research.

ACKNOWLEDGEMENTS

This study was funded and supported by the scientific research projects commission of Atatürk University, Erzurum, Turkey (Project code: 2013-34). This manuscript is a summary of a PhD thesis from Atatürk University Institute of Health Sciences and has presented a poster in the 3rd International Vetistanbul Group Congress, May 17- 20, 2016, Bosnia and Herzegovina.

REFERENCES

- Al-Fataftah AA, Abu-Dieyeh MHZ. Effect of chronic heat stress on broiler performance in Jordan. *International Journal of Poultry Science* 2007;6:64-70.
- Afify MRA, Esawy SH, El-Hadidy ME, Abdel-Salam ALM. Anti-ulcer activity of oregano (*Origanum syriacum* L.) against gastric ulcer in rats. *Advances in Food Sciences* 2012;34:145-149.
- Al-Kalaldeh JZ, Abu-Dahab R, Afifi FU. Volatile oil composition and antiproliferative activity of *Laurus nobilis*, *Origanum syriacum*, *Origanum vulgare*, and *Salvia triloba* against human breast adenocarcinoma cells. *Nutrition Research* 2010;30:271-278.
- Al-Kassie GAM. Influence of two plant extracts derived from thyme and cinnamon on broiler performance. *Pakistan Veterinary Journal* 2009;29:169-173.
- Alma MH, Mavi A, Yildirim A, Digrak M, Hirata T. Screening chemical composition and in vitro antioxidant and antimicrobial activities of the essential oils from *Origanum syriacum* L. growing in Turkey. *Biological and Pharmaceutical Bulletin* 2003;26:1725-1729.
- AOAC. Official methods of analysis of AOAC International. 18th ed. Rockville: Association of Official Analytical Chemists; 2005.
- Bölükbaşı SC, Erhan MK, Özkan A. Effect of dietary thyme oil and vitamin E on growth, lipid oxidation, meat fatty acid composition and serum lipoproteins of broilers. *South African Journal of Animal Science* 2006;36:189-196.
- Chawda HM, Mandavia DR, Parmar PH, Baxi SN, Tripathi CR. Hypolipidemic activity of a hydroalcoholic extract of *Cyperus scariosus* Linn. root in guinea pigs fed with a high cholesterol diet. *Chinese Journal of Natural Medicines* 2014;12:819-826.
- Chenga HW, Muirb WM. Chronic social stress differentially regulates neuroendocrine responses in laying hens: II. Genetic basis of adrenal responses under three different social conditions. *Psychoneuroendocrinology* 2004;29:961-971.
- Degerli S, Tepe B, Celiksoz A, Berk S, Malatyali E. In vitro amoebicidal activity of *Origanum syriacum* and *Origanum* on *Acanthamoeba castellanii* cysts and trophozoites. *Experimental Parasitology* 2012;131:20-24



- Demir E, Sarica Ş, Özcan MA, Suiçmez M. The use of natural feed additives as alternative to an antibiotic growth promoter in broiler diets. *Archiv für Geflügelkunde* 2005;69:110–116 .
- El-Desouky SK, Ibrahim FL, Kawashtya SA, El-Ansaria MA, Kimb YS, Chongb HS, *et al.* Phytochemical constituents and biological activities of *Origanum syriacum*. *Zeitschrift für Naturforschung* 2009;64:447-451.
- Gue FC, Kwakkel RP, Williams BA, Li WK, Li HS, Luo JY, Li XP, Wei YX, *et al.* Effects of mushroom and herb polysaccharides, as alternatives for an antibiotic, on growth performance of broilers. *British Poultry Science* 2004;45:684-694.
- Gümüş R, Imik H. The effect of *Yucca schidigera* powder added to lamb feed on fattening performance some blood parameters the immune system and the antioxidative metabolism of the hepatic tissue. *Turkish Journal of Veterinary and Animal Sciences* 2016;40:263-270.
- Gürakan GC, Aksoy C, Ögel ZB, Ören NG. Differentiation of *Salmonella Typhimurium* from *Salmonella Enteritidis* and other *Salmonella* Serotypes using random amplified polymorphic DNA analysis. *Poultry Science* 2008;87:1068–1074.
- Hadimli HH, Kav K, Erganiş O. Sıcaklık stresinin broiler piliçlerin humoral bağışıklıkları üzerine etkisi. *Veteriner Bilimler Dergisi* 2007;1:37-40.
- Hartlova H, Blaba J, Koubkova M, Draslrováz J, Fucftova A. Influence of heat stress on the metabolic response in broiler chickens. *Scientia Agriculturae Bohemica* 2002;33:145-149
- Hayirli A, Esenbuğa N, Macit M, Laçın E, Karaoğlu M, Karaca H, *et al.* Nutrition practice to alleviate the adverse effects of stress on laying performance, metabolic profile, egg quality in peak producing hens: I. The humate supplementation. *Asian-Australasian Journal of Animal Sciences* 2005;18(9):1310-1319.
- Hong JC, Steiner T, Aufy A, Lien TF. Effects of supplemental essential oil on growth performance, lipid metabolites and immunity, intestinal characteristics, microbiota and carcass traits in broilers. *Livestock Science* 2012;144:253–262.
- IUCN – International Union for Conservation Nature. A Guide to medicinal plants in north africa. 2nd ed. Glande: Centre For Mediterranean Cooperation; 2005.
- Ismail IB, Al-Busadah KA, El-Bahr SM. Oxidative stress biomarkers and biochemical profile in broilers chicken fed zinc bacitracin and ascorbic acid under hot climate, *American Journal of Biochemistry and Molecular Biology* 2013;3:202-214.
- Jang IS, Ko YH, Kang SY, Lee CY, Effect of a commercial essential oil on growth performance, digestive enzyme activity and intestinal microflora population in broiler chickens. *Animal Feed Science and Technology* 2007;134:304–315.
- Khaksar V, Van Krimpen M, Hashemipour H, Pilevar M, Effects of thyme essential oil on performance, some blood parameters and ileal microflora of Japanese quail. *Japan Poultry Science Association* 2012;49:106-110.
- Khan WA, Khan A, Anjum AD, Rehman ZU. Effects of induced heat stress on some biochemical values in broiler chicks, *International Journal of Agriculture & Biology* 2002;4(1):74–75.
- Koizumi L, Suzuki Y, Kaneko JJ, Studies on the fatty acid composition of intramuscular lipids of cattle, pigs and birds. *Journal of Nutritional Science Vitaminology* 1991;37:545-554.
- Köksal BH, Küçükersan MK. Effects of humate and vegetable extract mixture supplementation to diets on growth performance, some immunity and serum biochemistry parameters in broiler chickens . *Journal of the Faculty of Veterinary Medicine Kafkas University* 2012;18:103-108.
- Lin H, Decuyper E, Buyse J. Acute heat stress induces oxidative stress in broiler chickens. *Comparative Biochemistry and Physiology* 2006;144:11-17.
- Lukas BM. Molecular and phytochemical analyses of the genus *Origanum* (Lamiaceae) [thesis]. Wien (AUS): Universität Wien; 2010
- Lukas B, Schmiderer C, Franz C, Novak J, Composition of essential oil compounds from different Syrian populations of *Origanum syriacum* L. (Lamiaceae). *Journal of Agricultural and Food Chemistry* 2009;57:1362–1365.
- Luna A, Lábaque MC, Zygadlo JA, Marin RH. Effects of timol and carvacrol feed supplementation on lipid oxidation in broiler meat, *Poultry Science* 2010;89:366–370.
- Manohar V, Ingram C, Gray J, Talpur NA, Echard BW, Bagchi D, *et al.* Antifungal activities of origanum oil against *Candida albicans*. *Molecular and Cellular Biochemistry* 2001;228:111–117.
- Mazmanoğlu G. Effects of dietary antibiotic, essential oil mixture and organic acid supplementation; on performance, some organ weights and blood parameters in broilers [thesis]. Istanbul (TUR): Istanbul University, Institute of Health Science, Department of Animal Nutrition and Nutritional Diseases; 2008.
- Mitsch P, Zitterl-Eglseer K, Kohler B, Gabler C, Losa R, Zimpf I, The effect of two different blends of essential oil components on the proliferation of *Clostridium perfringens* in the intestines of broiler chickens. *Poultry Science* 2004;83:669–675.
- Olanrewaju HA, Wongpichet S, Nthaxton JP, Dozier WA, Branton SL. Stress and acid-base balance in chickens. *Poultry Science* 2006;85:1266–1274.
- Pilau MR, Alves SH, Weiblen R, Arenhart S, Cueto AP, Lovato LT. Antiviral activity of the *Lippia graveolens* (mexican oregano) essential oil and its main compound carvacrol against human and animal viruses. *Brazilian Journal of Microbiology* 2011;42:1616-1624.
- Reece OW. Functional anatomy and physiology of domestic animals. 4th ed. Hoboken: Wiley-Blackwell; 2009.
- Remmal A, Achahbar S, Bouddine L. Oocysticidal effect of essential oil components against chicken *Eimeria* oocysts. *International Journal of Veterinary Medicine* 2013;1-8.
- Sarica S, Ciftci A, Demir E, Kilinc K, Yildirim Y. Use of an antibiotic growth promoter and two herbal natural feed additives with and without exogenous enzymes in wheat based broiler diets. *South African Journal of Animal Science* 2005;35:61-72.
- Seven İ , Seven PT , Aslan AS , Yıldız N. The effects of dietary bee pollen on performance and some blood parameters in japanese quails (*Coturnix Coturnix Japonica*) breeding under different stocking densities. *Journal of Faculty of Veterinary Medicine Erciyes University* 2011;8(3):173-180.
- Tavarez MA, Boler DD, Bess KN, Zhao J, Yan Y, Dilger C, *et al.* Effect of antioxidant inclusion and oil quality on broiler performance, meat quality, and lipid oxidation. *Poultry Science* 2011;90:922–930.
- Tekce E, Gül M, Effects of *Origanum syriacum* essential oil added in different levels to the diet of broilers under heat stress on performance and intestinal histology. *European Poultry Science* 2016;80:1-11.
- Toplu HDO, Tunca R, Aypak SU, Coven F, Epikmen ET, Karaarslan S, *et al.* Effects of heat conditioning and dietary ascorbic acid supplementation on heat shock protein 70 expression, blood parameters and fear-related behavior in broilers subjected to heat stress. *Acta Scientiae Veterinariae* 2014;42:1-8.
- Yoshino K, Higashi N, Koga K. Antioxidant and anti-inflammatory activities of oregano extract. *Journal of Health Science* 2006;52:169-173.
- Yörük MA, Laçın E, Hayirli A, Yıldız A,. Effects of humate and prebiotic supplementation on laying performance, egg quality and blood parameters of Japanese quails reared in different cage densities, Faculty of Veterinary Medicine VYY University 2008;19:15-22.
- Zhang T, Yang S, Du J. Protective effects of berberine on isoproterenol-induced acute myocardial ischemia in rats through regulating HMGB1-TLR4 axis. *Evidence-Based Complementary and Alternative Medicine* 2014;13:1-8.

