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Original Article

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■Keywords

⁴⁷Ca Distribution, quail chicks, biological half-life.



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Assessment of ⁴⁷Ca Distribution and Biological Half-Life in Japanese Quail Chicks

ABSTRACT

A study was conducted to assess ⁴⁷Calcium (⁴⁷Ca) distribution and biological half-life in different body organs of Japanese quail chicks. A total of 85, 4 week old chicks, were dosed with 1ml/chick volume of 1% (w/v) CaCl, solution containing 7.51x10⁻⁰⁸ Ci ⁴⁷Ca which is equivalent to 1.27x10⁻¹³ g. ⁴⁷Ca activity, in terms of decay per minute (DPM), was measured in different body organs at 12 hours time intervals, after 24 hours from ingestion for five days. The total activity of ⁴⁷Ca for each organ and activity per gram of organ (D.min⁻¹.g⁻¹) was calculated. Data were Statistically analyzed using completely randomized design (CRD), one way analysis of variance (ANOVA) as per the procedure given by SPSS (2002), 9.0 version for Windows. The results obtained indicated that following ⁴⁷Ca administration most of the total ⁴⁷Ca concentration was found in the bones with count rates of 110267±550 DPM. While in the fifth day following administration, most of the ⁴⁷Ca was found in the feathers with a total activity of 13322±760 DPM. The biological half-life time was found to be highest in the heart and kidneys, respectively compared to the other body organs. In conclusion, the current results suggest that the main excretory pathway for Ca is through the skin. Regarding organ dependency on Ca for the normal functioning, the results obtained in our study suggest that the heart is the most dependent organ on Ca.

INTRODUCTION

Calcium (Ca) has been recognized as an essential nutrient in poultry nutrition. It considers the highest abundant mineral in the body (Veum, 2010), required to sustain biological processes of various body functions such as: the ossification of bones and cardiac muscle activity, vascular contraction, activation of several enzymes, transmission of nerve impulses, intracellular signaling, and hormonal secretion, etc., (Pu et al., 2016). Adequate Ca intake is essential for the healthy functioning of bone, heart, muscles, blood and nerves, etc., while, Ca deficiency, Ca metabolism disorders (Weaver, 2014), factors that influence Ca absorption efficiency and thus influence requirements for Ca and also, any change in serum Ca homeostasis can affect one or more of these physiological functions (Shin & Kim, 2015). All these factors are subject to increase the risk of various bone diseases and lead to economic losses to farm holders, (Brini et al., 2013). In order for these processes to function properly, normal Ca ion levels in blood and extracellular Ca homeostasis of various body organs, such as the intestine, muscles, and bone tissue must be maintained within a narrow range (Brini et al., 2013; Brown, 2013). As reviewed above, it is necessary to have a detailed understanding of Ca absorption and metabolism of different body organs. Ca radioactive isotope (⁴⁷Ca), the most valuable tool for many Ca studies and biological investigations, has been widely used



as tracer dietary studies on the uptake, transport and absorption of Ca in biological materials, through the intestinal wall (Reynard et al., 2013; Melin et al., 2014; Heuser, 2016). The importance attributed to ⁴⁷Ca is due to the special properties of this isotope, it is naturally fractionated between different organs and body fluids (Moynier & Fujii, 2017) and eliminated from the body with a biological half-life of 4.5 days, which is long enough for numerous biological investigations and short enough to ensure that the body subjected to the investigation is not under prolonged irradiation. It emits gamma rays in addition to beta rays, and since the gamma rays are capable of penetrating many centimeters of tissue, determination of ⁴⁷Ca in the body is possible by external measurements. It is therefore, possible to monitor the Ca movements during different body organs and to investigate the metabolic path of Ca in the body: its absorption through the walls of the intestine into the blood, its deposition in the skeleton, its return to the blood and its excretion, through ⁴⁷Ca measurements. Since, no reference has been made to trace Ca throughout the body organs of Japanese quail chicks. Therefore, the aim of the present study. was to assess ⁴⁷Ca distribution and biological half-life in different body organs of Japanese quail chicks and define more closely the specific Ca requirements or the amount of Ca required for each organ using the ⁴⁷Ca.

MATERIALS AND METHODS

Production of ⁴⁷Ca

Five grams of Ca chloride, technical grade (Sigma-Aldrich), was placed in a polyethylene vial and irradiated in the thermal irradiation site of the second research reactor (ETRR-2) for 5 hours irradiation at thermal flux of ~ 2×10^{11} n.cm⁻².s⁻¹. After proper cooling time, ⁴⁷Ca activity was detected and measured using gamma ray spectrometer consisting of a high performance calibrated germanium (HPGe) detector associated with the necessary electronics for data acquisition and processing.

Birds' ethics and husbandry

All of the experiments were carried out according to the guidelines of the Institutional Animal Care and Use Committee (IACUC) for animal experiments, which is member in the Egyptian Network of Research Ethics Committee (ENERC). The scientific and ethics committee of the Biological Application Department, Nuclear Research Center, Egypt approved all procedures used in this experiment (protocol number 174; date of approval: 08-03-2018). The experimental study was conducted on Japanese quail chicks maintained at the Poultry experimental farm of the biological Application Department, Nuclear Research Center, Egypt. All procedures used in this experiment were approved by the Animal Ethics Committee of National Institute of Animal Health and complied with the "Guidelines for the Care and Use of Animals in Research".

Birds, Treatment, Samples and Measurements

A total number of 150 Japanese quail (Coturnix coturnix japonica) chicks, at 4 weeks of age with average body weight of $128\pm5g$ / chick, were used for this study. The chicks were reared in battery cages of $30 \times 60 \times 150$ cm in size and kept under the same environmental conditions of light 23:1 hrs Light: dark cycle, RH was $50\pm5\%$ and room temperature ($30\pm2^{\circ}C$).

The rearing conditions were $25\pm2^{\circ}$ C, $60\pm5^{\circ}$ relative humidity (RH), with a photoperiod of 14 L: 10 D hours

RH was $50\pm5\%$ and photoperiod was 14 L: 10 D hours.

Feed and water were provided ad libitum. Chicks, equal in body weight, were evenly distributed into 3 replications, each replicate pen containing 50 quails each. For one week (the treatment period), the quail chicks were fed the basal diet formulated to meet all nutrient requirements of Japanese quail according to (NRC, 1994), with protein level (22 %), metabolizable energy (3350 kcal/kg), Ca level (0.81%) and available phosphorus level (0.31%).

At the beginning of the treatment (time of ingestion), the activity of ⁴⁷Ca was measured using a gamma ray spectrometer consisting of a calibrated HPGe detector and found to be 7.51μ Ci/g CaCl₂ which is equivalent to 1.27×10^{-11} g ⁴⁷Ca since that the ⁴⁷Ca specific activity is 5.9×10^5 Ci/g. The radio activity half life time (t_{1/2}) of ⁴⁷Ca is 4.5 days (Reynard *et al.* 2013).

All chicks were administered orally 1ml volume/ chick of 1% (w/v) CaCl₂ solution containing 7.51x10⁻⁰⁸ Ci ⁴⁷Ca which was equivalent to 1.27x10⁻¹³g ⁴⁷Ca. After a 24-hour period following the administration, 6 quail chicks (2 chicks /pen) were randomly selected, weighed and slaughtered for carcass analysis: feather, skeletal muscles from breast and thigh, bones, liver, proventriculus, gizzard, intestine, kidney, and heart for each slaughtered chick were calculated and weighed individually. Excreta was also collected. Blood samples were collected from each slaughter chick into test tubes. For assay of the total Ca⁴⁷ content, each body organ



Calculated Activity of ⁴⁷Ca (Mean±SD) in different body organs and excreta at 24 hour time intervals measured after 24 hours from the ingestion.

and blood per chick was counted immediately for one-minute using gamma-counter in the term of decay per minute per gram (D/min/g) of organ. The same procedure was repeated at 12 hour intervals following the administration for five days. The data obtained were analyzed to estimate the bio-distribution and biological half life time of Ca for each organ.

⁴⁷Ca detection and amount of ⁴⁷Ca calculations per gram organ

The ⁴⁷Ca specific activity was employed to calculate both of the approximate total amount of ⁴⁷Ca (g) and the approximate amount of ⁴⁷Ca per gram organ at measuring time intervals.

The calculations of ⁴⁷Ca effective half life time $(t_{1/2e})$ and biological half-life time $(t_{1/2b})$ were performed according to (Segel, 1976) as follows:

t_{1/2e}, in hours, was calculated according to the following equation:

t_{1/2e}= 0.693x(A-A_o)/2.3logt_o/t

Where $A_{_{\rm o}}$ and A are any given activities of ${}^{_{47}}\text{Ca}$ at time $t_{_{\rm o}}$ and t

 $t_{1/2b}$, in minutes, of 47Ca in the chicks were calculated using the following equation:

 $t_{1/2b} = ((1/t_{1/2e}) - (1/(T_{1/2p} \times 24))) \times 60$

Where T_{1/2p} is the physical half life time of ⁴⁷Ca which equals 4.5 days.

Statistical analysis

Statistical analysis was done using completely randomized design (CRD), one way analysis of variance (ANOVA) as per the procedure given by SPSS (2002), 9.0 version for Windows. The test was employed to identify the significant differences amongst the different time, organs and feaces. P-value less than 0.05 is considered to be statistically significant.

RESULTS AND DISCUSSION

⁴⁷Ca distribution throughout the body

⁴⁷Ca total activity in DPM was tracked in various body organs of quail chick after 24 hours following administration and equivalent calculated amounts of ⁴⁷Ca in gram at 24-hour time intervals are summarized in Tables (1 and 2). As shown in Figure (1), the results obtained shows that, most of the total ⁴⁷Ca was found,

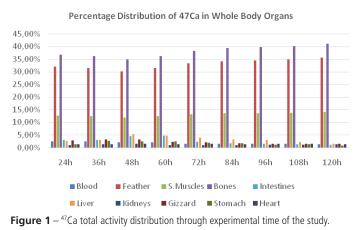
Drgan	24h	36h	48h	60h	72h	84h	96h	108h	120h
Blood	3848.7±103.9	3542.7±223.1	3034.7±151	2914.3±79.2	2167±30.1	2046±50.5	2054.7±138.5	2059±58.9	1861.6±120
	Adef	Bd	Cf	Cde	Dd	DEc	DEde	DEd	Ec
Feather	49167.7±1320	46464.8±1306	43653.3±1050	44190.3±1344	46066.3±864	46638.7±968	46670±802	46685.3±1213	47410±1301
	Ab	Bb	Cb	Cb	Bb	Bb	Bb	Bb	Bb
S.Muscles	19412±653.9	18344.7±845.6	17234.3±729.7	17446.3±912	18187.3±748.8	18413.4±868	18425.9±764.5	18432±825.8	18718.2±813
	Ac	Bc	Cc	Cc	Bc	Bb	Bc	Bc	Bc
Bones	56622.3±614.2	53509.3±789	50271.7±603.8	50890.4±760	53051.1±600	53709.6±660	53746.6±678.8	53763±600.4	54598.6±758.5
	Aa	Ba	Ca	Ca	Ba	Ba	Ba	Ba	Ba
Intestines	4498±166.9	4575±131.6	6541.7±58.1	6757.3±50.2	3055.3±173.7	2289±163.8	1923±46	1718.3±114.6	1527.3±130.2
	Cde	Cd	Bde	Ad	Dd	Ec	Fde	FGd	Gc
Liver	4121±335.3	4361±180.5	7503±394.5	6518.3±104.9	5553±150.1	4329.7±146.7	4034.6±124.6	3149±231.8	1900±360.5
	Ddef	Dd	Ad	Bd	Cd	Dc	Dd	Ed	Fc
Kidneys	1523.7±21.5	1814.7±30.7	2217.3±226.5	1259±45.2	1213.3±80.8	1385±147	1460.3±69.5	1628±63.3	1715.3±101.6
	CDEf	Be	Af	Fe	Fe	EFc	DEde	BCDd	BCc
Gizzard	4466±246.7	4635.7±126.3	4692.3±169	3289.3±48.8	2829.7±288.2	2428.3±111.4	2109.7±221.8	2052±93.8	1992.6±49.6
	Ade	Ad	Adef	Bde	Cd	Dc	Ede	Ed	Ec
Stomach	2076.7±107.8	3839.7 ±298.6	3604.7±611.7	3340.7±142	2556±76.1	2224±78.7	1648.3±45.2	1815.6±44.4	1010.3±38.5
	DEef	Ad	ABef	Bde	Cd	CDc	Ede	DEd	Fc
Heart	2010.3±91.9	1983.7±41	2160±144	1932.7±76.2	2000.7±96.7	1746.7±46.2	2049.3±66.5	1911.6±78.4	1664.6±61.2
	ABef	Bd	Af	Be	Bd	Cc	ABde	Bd	Cc
Excreta	6239.3±268	4674.7±281	3652±222	2374±24	1859±67.6	1470.3±61.1	1088.3±82.6	907.3±43.9	815.3±50
	Af	Bd	Cef	De	Ed	Fc	Ge	Gd	Gc
a, b, c,: Means wi A, B, C,: Means v	a, b, c,: Means with different superscripts in each row Differ significantly (p<0.05). A, B, C,: Means with different superscripts in each column Differ significantly (p<0.05)	in each row Differ signifi ts in each column Differ	cantly (p<0.05). significantly (p<0.05).						

Table 1 – Total



Table 2 – Approximate total amount of ⁴⁷Ca in different body organs and excreta at 24 hour time intervals measured after 24 hours from the ingestion.

Time Organ	24h	36h	48h	60h	72h	84h	96h	108h	120h
Blood	2.938E-15	2.705E-15	2.317E-15	2.225E-15	1.654E-15	1.562E-15	1.569E-15	1.572E-15	1.421E-15
Feather	3.754E-14	3.889E-14	3.908E-14	3.512E-14	1.930E-14	1.781E-14	1.682E-14	1.496E-14	1.725E-14
S.Muscles	1.482E-14	1.408E-14	1.392E-14	1.645E-14	1.490E-14	1.477E-14	1.183E-14	9.379E-15	3.286E-15
Bones	4.323E-14	9.034E-14	1.274E-13	1.595E-13	2.731E-13	2.503E-13	2.021E-13	1.575E-13	7.241E-14
Intestines	3.434E-15	3.493E-15	4.994E-15	5.159E-15	2.333E-15	1.748E-15	1.468E-15	1.312E-15	1.166E-15
Liver	3.146E-15	3.330E-15	5.728E-15	4.977E-15	4.240E-15	3.306E-15	3.080E-15	2.404E-15	1.451E-15
Kidneys	1.163E-15	1.385E-15	1.693E-15	9.612E-16	9.263E-16	1.057E-15	1.115E-15	1.243E-15	1.310E-15
Gizzard	3.410E-15	3.539E-15	3.582E-15	2.511E-15	2.160E-15	1.854E-15	1.611E-15	1.567E-15	1.521E-15
Stomach	1.586E-15	2.932E-15	2.752E-15	2.551E-15	1.951E-15	1.698E-15	1.258E-15	1.386E-15	7.713E-16
Heart	1.535E-15	1.515E-15	1.649E-15	1.476E-15	1.527E-15	1.334E-15	1.565E-15	1.459E-15	1.271E-15
Excreta	2.938E-15	2.705E-15	2.317E-15	2.225E-15	1.654E-15	1.562E-15	1.569E-15	1.572E-15	1.421E-15



as expected, in the bones which had ⁴⁷Ca count rates of 110267±550, 53747±2072, 60416±4188, and 46630±3232 DPM at 24, 48, 72, 96 hours following the administration, respectively. These count rates represented 67%, 48%, 56%, and 56%, in respect to the above mentioned count rates, of the total ⁴⁷Cadetected at mentioned times. At 120 hours after ingestion it was noticed that the count rate of ⁴⁷Ca was the highest in the feathers where ⁴⁷Ca activity recorded 13342±760 DPM and represented about 34% of the total detected activity, while in the bones, the ⁴⁷Ca recorded 11692±501 DPM and represented about 30% of the total detected activity. These results give obvious evidence that the main site of storage for Ca in terms of ⁴⁷Ca is in the bones which correlates with most of the studies and references (Taylor & Bushinsky, 2009; Lieben et al., 2010; Carmeli et et al., 2015; Pu et al., 2016). The above results also suggest that the main route of deposition and excretion of Ca may be through the skin and feathers, since the Ca amount, in terms of ⁴⁷Ca activity, was found to increase in the feathers as experimental time advanced. As a result, feathers can be a reliable indicator to detect the correlations between oral ingestion of ⁴⁷Ca and

its deposition in feathers and skin of quail chicks. Many investigators have used feathers as a simple method for biomonitoring to determine trace element levels, dietary exposure and external contamination in birds (Cardiel *et al.*, 2011; Brait & Antoniosi Filho, 2011; Jenni *et al.*, 2015). In skeletal muscles ⁴⁷Ca activity appears after 48 h of ⁴⁷Ca ingestion, recorded 12494±705 DPM and represented about 11% of the total detected activity, then a slight decrease occurred to 3619±190 DPM and represented about 9% of the total detected activity at 120 h of ⁴⁷Ca ingestion. This result of ⁴⁷Ca activity in skeletal muscles confirms the essential role of Ca in muscles contraction as shown in many studies (Sun *et al.*, 2009; Brini *et al.*, 2013; Brown, 2013; Weaver, 2014; Pu *et al.*, 2016).

⁴⁷Ca amount per gram tissue of each body organs

In this part of the study, the correlation between ⁴⁷Ca activity in DPM per gram organ and equivalent calculated amounts in gram per gram organ at 12 hr time intervals until 120 hours of the administration time are presented in Tables (3 and 4) and in Figures (2 and 3).

It was found that the picture changed when we studied the Ca requirements in terms of ⁴⁷Ca per gram in the studied organs, as shown in Figures (1 and 3) and detailed hereafter. The ⁴⁷Ca count rates in DPM per gram of organ at 24 hours after administration in the blood, feather, skeletal muscles, bones, intestine, liver, kidneys, gizzard, proventriculus, heart and excreta was found to be 661±46.24, 2891.3±241.79, 258±11.04, 1078.3±14.39, 460.3±11.57, 1317±48.7, 1364±55.8, 1326.7±80.38, 3651.7±134.1, 2101±188.5 and 556±16.4, respectively. As a result, the highest ⁴⁷Ca activities per gram organ were detectable at 24 h after ingestion in the proventriculus, feather and heart

Table 3 – Act	ivity of ⁴⁷ Ca per <u>c</u>	Jram (Mean±SD)	Table 3 – Activity of ⁴⁷ Ca per gram (Mean±SD) in different body organs and excreta at 12 hour time intervals measured after 24 hours from the ingestion.	organs and ex	creta at 12 hour	time intervals m	easured after 24	hours from the	ingestion.
Time Organ	24h	36h	48h	60h	72h	84h	96h	108h	120h
Blood	661±46.240	539±14.16	456.3±18.90	427±12.72	330±13.01	312.7±12.43	293.7±10.99D	278±7.23	234.3±10.35
	Ade	Bf	Cgh	Cf	Dg	Dg	Eh	Deg	Ef
Feather	2891.3±241.79 Ba	3273±80.92 Aa	3049±172.14ABa	2748±71.68 Bb	1591±29.83 Cd	1506.3±11.16 Cd	1383.3±25.90 Ce	1282.7±23.93 Cd	1433±97.28 Cd
S.Muscles	258±11.04 Abe	244.7±3.82 Bh	245±7.63 Bi	266.7±3.87 Ag	259.3±2.60ABgh	237.7±4.28 Bgh	208±7.28 Chi	158.3±7.28 Dh	58.3±7.69 Eg
Bones	1078.3±14.39	2043.3±66.07	3123±42.37	4044.7±84.57	7056±74.74	6270.6±89.88	5110.7±82.83	3673.3±90.35	2059.3±87.8
	Hcd	Gc	Fa	Da	Aa	Ba	Ca	Ea	Ga
Intestines	460.3±11.57	423±8.51	550.3±12.49	552±8.16	236.7±2.40	204.3±5.37	170.3±4.92	139±4.92	111±4.92
	Be	Cg	Ag	Af	Dgh	Ehi	Fi	Gh	Hg
Liver	1317±48.7	1184.7±85.5	1837.7±131.6	1667±58	1432.7±29.1	1184.7±29.9	976±35.6	863.3±29.1	523.7±8.2
	Dc	Ee	Ae	Bd	Ce	Ee	Ff	Ge	He
Kidneys	1364±55.8	1685.3±88.7	1965.7±53.6	1771±41.7	1605.3±93.1	1496±42.4	1554.7±36.2	1638±48.7	1759.3±50.3
	Fc	BCd	Ad	Bd	CDd	Ed	DEd	CDc	Bb
Gizzard	1326.7±80.38	1109.7±34.1	992.3±65.3	861.3±22.03	797.3±43	659±29	604.3±27.1	571±29	553±29
	Ac	Be	Cf	De	Df	Ef	EFg	Ff	Fe
Stomach	3651.7±134.1	3254.7±178.5	2833.7±143.3	2485±65	2162.3±76.1	1872±62.7	1759±37.3F	1662.3±49.6	1553±62.5
	Aa	Ba	Cb	Dc	Eb	Fc	Gc	GHc	Hc
Heart	2101±188.5	2247.3±51.1A	2257±197.7	2360.7±43.9	1876.7±36.1	2080±53.8	2082.7±53.1	2096±51.1	2116±52.8
	Bb	Bb	ABc	Ac	Cc	Bb	Bb	Bb	Ba

a, b, c, ..: Means with different superscripts in each row Differ significantly (p<0.05).

A, B, C, ...: Means with different superscripts in each column Differ significantly (p<0.05).

Table 4 – Approximate amount of ⁴⁷Ca in different body organs and excreta at 12 hour time intervals measured after 24 hours from the ingestion.

hours from					Amount of	Amount of 47Ca per gram organ (g)	organ (g)				
ingestion	Blood	Feather	Skeletal muscles	Bones	intestine	Liver	Kidneys	Gizzard	Proventriculus	Heart	Excreta
24	5.04657E-16	4.11513E-16	5.04657E-16 4.11513E-16 3.48374E-16 3.26004E-16 2.51947E-16 2.38739E-16 2.24233E-16 2.12246E-16 1.78882E-16	3.26004E-16	2.51947E-16	2.38739E-16	2.24233E-16	2.12246E-16	1.78882E-16	5.04657E-16 4.11513E-16	4.11513E-16
	2.20744E-15	2.49885E-15	2.20744E-15 2.49885E-15 2.32784E-15 2.09803E-15 1.21469E-15 1.15002E-15 1.05612E-15 9.7931E-16	2.09803E-15	1.21469E-15	1.15002E-15	1.05612E-15	9.7931E-16	1.09406E-15	1.09406E-15 2.20744E-15 2.49885E-15	2.49885E-15
48	1.96977E-16	1.86822E-16	1.96977E-16 1.86822E-16 1.87051E-16 2.03619E-16 1.97969E-16 1.81478E-16 1.58803E-16 1.20858E-16 4.45106E-17 1.96977E-16 1.86822E-16	2.03619E-16	1.97969E-16	1.81478E-16	1.58803E-16	1.20858E-16	4.45106E-17	1.96977E-16	1.86822E-16
60	8.23255E-16	1.56001E-15	8.23255E-16 1.56001E-15 2.38433E-15 3.08803E-15 5.38708E-15 4.78745E-15 3.90189E-15 2.80447E-15 1.57222E-15 8.23255E-16 1.56001E-15	3.08803E-15	5.38708E-15	4.78745E-15	3.90189E-15	2.80447E-15	1.57222E-15	8.23255E-16	1.56001E-15
72	3.51428E-16	3.2295E-16	3.51428E-16 3.2295E-16 4.2014E-16 4.21438E-16 1.80715E-16 1.55978E-16 1.3002E-16 1.06123E-16 8.47458E-17	4.21438E-16	1.80715E-16	1.55978E-16	1.3002E-16	1.06123E-16	8.47458E-17	3.51428E-16 3.2295E-16	3.2295E-16
84	1.0055E-15	9.04489E-16	1.0055E-15 9.04489E-16 1.40304E-15 1.27271E-15 1.09383E-15 9.04489E-16 7.45152E-16 6.59108E-16 3.99832E-16 1.0055E-15 9.04489E-16	1.27271E-15	1.09383E-15	9.04489E-16	7.45152E-16	6.59108E-16	3.99832E-16	1.0055E-15	9.04489E-16
96	1.04138E-15	1.28668E-15	1.04138E-15 1.28668E-15 1.50076E-15 1.35211E-15 1.22561E-15 1.14216E-15 1.18698E-15 1.25057E-15	1.35211E-15	1.22561E-15	1.14216E-15	1.18698E-15	1.25057E-15	1.34318E-15 1.04138E-15 1.28668E-15	1.04138E-15	1.28668E-15

75±5 Hq

81.7±2.9 Hh

98±2.6

135.3±6.1

185.7±6.6

242±6.24 Dg

340±10 Chi

439±11.6 Bg

556±16.4 Ae

Excreta

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8.47229E-16

4.22202E-16 1.18568E-15

4.35944E-16 1.26913E-15

4.61368E-16 1.34295E-15

6.08719E-16

6.57581E-16 1.89724E-15

7.57597E-16 2.16346E-15

8.47229E-16 2.48488E-15

1.0129E-15 2.78798E-15

108 120

5.0313E-16 1.42923E-15

1.65086E-15

2.48488E-15

1.0129E-15 2.78798E-15



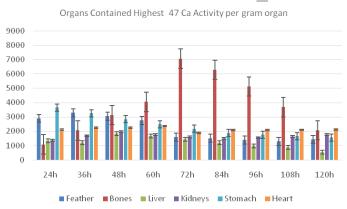


Figure 2 – distribution of 4^{7} Ca activity, per gram organ, in organs which found to have the highest per gram organ expressed as percentage from total 47 Ca activities throughout the time of the study.

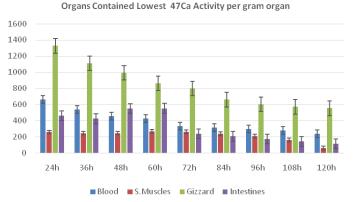


Figure 3 – distribution of 47 Ca activity, per gram organ, in organs which found to have the lowest expressed as percentage from total 47 Ca activities throughout time of the study.

(23.31, 18.46 and 13.41%), respectively, of total estimated ⁴⁷Ca activity, with intermediate activities in kidney, gizzard, liver and bones (8.71, 8.47, 8.41, and 6.88%), respectively, and low activities in intestine, blood, skeletal muscles and excreta (2.94, 4.22, 1.65 and 3.55%), respectively. With progress in time, starting from the 24th h until the 72nd h of the administration time, gradual decreases were recorded for the percentage of ⁴⁷Ca activity per gram in all organs and excreta, except the bones in which the ⁴⁷Ca reached 40% of total activity in the studied organs. The observed decreases in ⁴⁷Ca activity with progress of time, as observed in many studies (Khanal & Nemera, 2008; Geibel & Hebert, 2009; Christakos, 2012), confirmed the intestinal absorption and Ca metabolism, which refers to the movements and regulation of Ca into and out of various body compartments of quail chick, such as the intestine, the blood plasma, the extracellular and the intracellular fluid, and the bone tissue. The observed patterns of ⁴⁷Ca in bones bone tissue acts as a Ca storage center for deposits and withdrawals needed by the blood, via continual bone remodeling (Morgan et al., 2012; Brini

et al., 2013; Brown, 2013; Carmeli et et al., 2015). Most studies on the physiology of muscles showed that the muscles require Ca to contract, by increasing the amount of free intracellular Ca into the muscle cells (Ewing & Charlton, 2007; Pu et al., 2016; Shin & Kim, 2015; Heuser, 2016). In Skeletal muscles, liver, gizzard, intestine and excreta, it was noticed that, with progress of time starting from the 24th h towards the end of the measuring time at the 120th h of administration, ⁴⁷Ca activity per gram these organs decreased gradually and reached 0.56, 5, 5.28, 1.06, and 0.72%, respectively, of the total estimated ⁴⁷Ca activity. While, a slight increase was detected gradually in the behavior of ⁴⁷Ca activity per gram of kidney, starting from the 84th h towards the end of measuring time at the 120th h of administration time, and reached 16.79% of the total estimated ⁴⁷Ca activity. Similarly, a slight increase was detected gradually in the behavior of ⁴⁷Ca activity per gram of heart from 10% at the 84th h to 20.2% of the total estimated ⁴⁷Ca activity at the 120th h of administration time. Depending on the previous results, it can be concluded that the observed decrease in ⁴⁷Ca activity per gram of liver, gizzard, intestine and excreta after 24 h following the administration in this study is logically since the majority of the Ca, approximately 65%, is absorbed throughout the intestine but varies by region to provide a Ca pool to maintain serum levels (Pu et al., 2016; Heuser, 2016). Furthermore, most studies on Ca²⁺ homoeostasis in hepatic tissue have suggested a dynamic equilibrium between separate Ca²⁺ influx and Ca²⁺ efflux pathways (Reinhart et al., 1984). Moreover, these results may indicate that the quail's body reached the Ca balance state which is determined by the relationship between Ca intake and Ca absorption and excretion. Ewing & Charlton (2007) reported that, the Ca requirement is generally recognized to be the intake required to maintain Ca balance and therefore skeletal integrity. Examination of the data clearly detected a constant behavior of ⁴⁷Ca activity per gram of blood at approximately 3% of the total estimated ⁴⁷Ca activity which occur for 5 days following the administration. As a result, the regulation of Ca in the blood plasma is within narrow limits. As such, this may explain why an important aspect of Ca metabolism is plasma Ca homeostasis (Pu et al., 2016).

The biological half life time (t_{1/2b}) of ⁴⁷Ca

The calculated biological half life time $(t_{1/2b})$ in minutes of ⁴⁷Ca and the highest $t_{1/2b}$ for ⁴⁷Ca activity, are shown in Figure (4), and are found to be 101.1, 48.2, and 10.9, 6.2 minutes in the heart, kidney, bones and feathers respectively. The calculated biological half-life

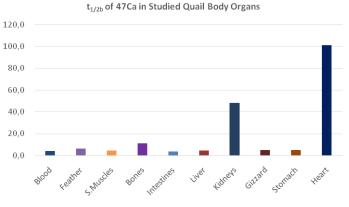
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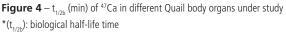


Table 5 – effective half life time (t_{1/2e}) and biological half life time (t_{1/2b}) of ⁴⁷Ca in Japanese quail body organs under study.

t _{1/2e} (hr)									
Blood	Feather	Skeletal muscles	Bones	intestine	Liver	Kidneys	Gizzard	Proventriculus	Heart
13.0	8.8	11.9	5.2	15.1	11.8	0.6	11.0	10.7	0.6
t _{1/2b} (min)									
4.0	6.2	4.5	10.9	3.4	4.5	48.2	4.9	5.0	101.1

values tended to increase with the increase in the Ca retention in the heart which may suggest that the heart muscle is the most dependent organ on Ca for proper functioning. In this respect, (Kranias et al., 2007; Bers, 2008; Sun et al., 2009) showed that Ca has an essential role in cardiac muscle physiology (Excitation contraction coupling), allowing the heart to function as a whole to contract properly and pump the blood efficiently into the arteries throughout the body. It was also showed that ⁴⁷Ca t_{1/2b} is higher in the kidneys than int he bones, this finding may be related to the primary mechanism of kidney for rapid release or absorption of Ca through the filtration and excretion functions. (Toka et al., 2015) showed that extra intestinal Ca can be processed through the kidneys and removed from the body through excretion, however, when correlated to the above total findings of total ⁴⁷Ca activity of feather, we found that the main secretion pathway of Ca is through the skin rather than the kidneys due to the feather's much greater size compared to the kidneys. Finally, the calculated t_{1/2b} suggest that, although Ca is apparently mainly deposited and utilized by the bones of quail chick, this is not the case on the specialized tissue levels of body organs where it was revealed using ⁴⁷Ca measurements and t_{1/2b} that the Ca retention in the heart is the highest, therefore there is minimal turnover and reduced exchange of calcium molecules between heart muscle cells and blood which may suggest that the heart muscle it the most dependent organ on Ca for proper functioning.





CONCLUSION

The results of the present study have provided a means for closely monitoring ⁴⁷Ca distribution throughout body organs which may occur following the administration in quail chicks and furthermore they allow a realistic estimation of the Ca content to be made and examine the ability of body organs to maintain a given ⁴⁷Ca load. Most of the total ⁴⁷Ca was found in the bones. While in the fifth day following the administration most the ⁴⁷Ca was found in the feathers. The highest biological half-life time was found to be in the heart and kidney, respectively. As a result, the heart is the most dependent organ on Ca requirements for the normal functioning and the main excretory pathway for Ca is through the skin.

CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

CONTRIBUTION OF AUTHOR'S

- Walaa F. Hassanin, hatamahmoud@yahoo.com, Principal Author, participated in the conception, design, planning methodology-of the research, performed analysis on all samples, helped in data interpretation and manuscript evaluation.
- Nashat Saeid Ibrahim, nashaat1977@yahoo. com, Co-Author, participated in the conception, participated in the design, data collection, statistical analysis, interpretation of the results, wrote the manuscript and acted as corresponding author.
- Emad. Eldien El-Barkouky, Emad1949@hotmail. com, Co-Author, participated in the planning methodology, biological materials, contributed substantially to the revising of the manuscript.
- Adel Mohamed. Abu-Taleb, adelabutaleb@ gmail.com, Co-Author, participated in the data collection, management and reporting, processing, materials (reagents).



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