# Prevalence and antimicrobial susceptibility profile of ESKAPE pathogens from the Federal District, Brazil

Prevalência e perfil de suscetibilidade aos antimicrobianos de bactérias do grupo ESKAPE no Distrito Federal, Brasil

Daniely M. Silva<sup>1</sup>; Eulina Maria N. Menezes<sup>2</sup>; Emerson V. Silva<sup>2</sup>; Thaís A. C. Lamounier<sup>1</sup>

1. Universidade de Brasília (UnB), Brasília, DF, Brazil. 2. Secretaria de Saúde do Distrito Federal, Brasília, DF, Brazil.

#### **ABSTRACT**

Introduction: The leading cause of hospital-acquired infections are the pathogens named by the acronym ESKAPE, which are the initials for the following bacterial: *Enterococcus faecium*, *Staphylococcus aureus*, *Klebsiella pneumoniae*, *Acinetobacter baumanni*, *Pseudomonas aeruginosa* and *Enterobacter* spp., which have high resistance rates by escaping the action of the antimicrobial. **Objective**: To trace the antimicrobial susceptibility profile of the ESKAPE pathogens in a primary public hospital in the Federal District, Brazil. **Methods**: A cross-sectional, retrospective and descriptive study was conducted by analyzing the corresponding data from January 2010 to December 2015 of samples considered positive to ESKAPE pathogens in order to generate an antimicrobial susceptibility profile. **Results**: Analyzing the Grampositive bacteria, almost 80% of *E. faecium* strains were vancomycin-resistant *enterococci* (VRE) and almost 40% of *S. aureus* strains were methicillin (oxacillin)-resistant *Staphylococcus aureus* (MRSA). It was observed that gram-negative strains (the ESKAPE group) examined in this study have a higher resistance rate to carbapenems than in other studies. In the molecular analysis, four *Klebsiella pneumoniae* strains were positive to *bla*<sub>KPC</sub> gene, three strains to *bla*<sub>NDM</sub> and one *Acinetobacter baumanni* strain was positive to *bla*<sub>OXA-23</sub> gene. **Conclusion**: Studies such as this should be performed periodically in order to evaluate the bacterial susceptibility profile. They demonstrate the importance of implementing strategies to prevent hospital-acquired infections, as well as greater antibiotic prescribing control.

Key words: microbial sensitivity tests; genes; bacteria.

#### INTRODUCTION

Bacteria may be intrinsically resistant to an antimicrobial, when it is an inherent characteristic of a species, related to chromosomal genes. Furthermore, they may acquire resistance to certain antimicrobials through chromosomal mutations or horizontal gene transfer by three mechanisms: 1) bacteriophage-mediated transduction; 2) transformation by chromatin incorporation of chromosomes, plasmids and deoxyribonucleic acid (DNA) from dead organisms; and 3) conjugation through plasmids and conjugative transposons<sup>(1, 2)</sup>. These transfers may occur in water, soil, food, and the digestive system of animals and humans<sup>(3)</sup>.

These resistances hindering patients' treatment, since the use of broad spectrum antimicrobials is required, but the development and approval of new drugs grow at a much slower rate than the emergence of bacterial resistance. Bacterial resistance mechanisms

usually occur two or three years after the introduction of a new antimicrobial into therapy<sup>(4)</sup> and a drug of this type can take 12 to 22 years to be available in the market<sup>(5)</sup>. This reduces the effectiveness of the antimicrobial treatment, making it difficult and costly, which can increase the length of hospital stay of infected patients and often leading to their death<sup>(6-8)</sup>. Bacteria from the ESKAPE group (*Enterococcus faecium*, *Staphylococcus aureus*, *Klebsiella pneumoniae*, *Acinetobacter baumanni*, *Pseudomonas aeruginosa* and *Enterobacter* spp.) are the main causes of hospital-acquired infections in the United States and they are able to escape from the antimicrobial actions due to the resistance profile<sup>(9)</sup>.

It is necessary that studies trace periodically the bacterial resistance profile, to contribute for both local and global epidemiological data. These data assist in therapeutic management, since they consider the prevalence of resistance

locally, adding it to the clinical effectiveness and cost of the antimicrobial. The present study aims to evaluate the susceptibility profile of ESKAPE pathogens in a primary hospital of the public network of the Federal District, Brazil.

## **METHODS**

# Characterization of the study

The present study is considered cross-sectional, retrospective and descriptive, in which were analyzed the data from antimicrobial susceptibility tests performed in the microbiology sector of the clinical analysis laboratory of a hospital in the public network of Brasília, Brazil, in the period from January 2010 to December 2015. The hospital analyzed is a primary hospital, and does not have an intensive care unit. This study was approved by the Research Ethics Committee of the Faculty of Health Sciences of the Universidade de Brasília (UnB) CAAE: 38856114.0.0000.0030.

# Data analysis

Using WHONET 5.6 software, 2,527 samples were analyzed, among which 577 were positive to the ESKAPE group pathogens. Data such as age, sex and type of sample were considered. The minimum inhibitory concentration (MIC) used to trace the susceptibility profile was interpreted from the cut-off points determined by the Clinical & Laboratory Standards Institute (CLSI) 2015.

WHONET is a program developed for the analysis and monitoring of microbiological data, especially for antimicrobial susceptibility test. It was developed in 1989 by the Collaborating Center for Surveillance of Antimicrobial Resistance of the World Health Organization (WHO), helping to understand epidemiological data for antimicrobial choice and outbreak detection<sup>(10)</sup>. It is a free program, used in more than 90 countries, and available in more than 20 languages, including Portuguese.

#### **Bacterial** isolates

The resistance genes were investigated in eight samples that were sent to the Central Public Health Laboratory of the Federal District [Laboratório Central de Saúde Pública do Distrito Federal (LACEN/DF)] for genetic identification, of which seven *Klebsiella pneumoniae* strains and one *Acinetobacter baumannii*. The criterion for sending the strains of these enterobacteria used by the laboratory of clinical analyzes of the public health network was the carbapenem-resistance profile

and also the modified Hodge test or enzymatic blockade test with positive ethylenediamine tetraacetic acid (EDTA)<sup>(11, 12)</sup>.

# Molecular detection of genes by polymerase chain reaction (PCR)

For molecular confirmation, the genes:  $bla_{\text{KPC}}$ ,  $bla_{\text{NDM}}$ ,  $bla_{\text{IMP}}$  e  $bla_{\text{VIM}}$ , were investigated, since they were the most prevalent in carbapenem-resistant enterobacteria, and the  $bla_{\text{OXA-23}}$ , gene in carbapenem-resistant *Acinetobacter baumannii* in Brazil (13,14). For this purpose, we used the primers indicated in **Table 1**.

TABLE 1 - Primers used

Primers	Sequence	Amplicon size (PB)	
NDM F	5' GGTTTGGCGATCTGGTTTTC	512	
NDM R	5' GGCCTTGCTGTCCTTGATC	514	
KPC F	5' TGTCACTGTATCGCCGTC	1011	
KPC R	5' CTCAGTGCTCTACAGAAAACC	1011	
VIM F	5' GATGGTGTTTGGTCGCATATC	332	
VIM R	5' CTCGATGAGAGTCCTTCTAGAG	334	
<i>IMP</i> F	5'AACACGGTTTGGTGGTTCTT	440	
IMP R	5 GGACTTTGGCCAAGCTTCTA	440	
OXA-23 F	5'GATCGGATTGGAGAACCAGA	501	
OXA-23 R	5' ATTTCTGACCGCATTTCCAT	501	

NDM: New Delbi metalo-betalactamase; F: forward; R: reverse; KPC: Klebsiella pneumoniae carbapenemase; OXA: oxacilinase.

Source: Faria Junior et al.  $(2016)^{(15)}$  and Woodford N et al.  $(2006)^{(16)}$ .

The reactions were subjected to the temperature cycles programming of 2 minutes at 94°C, followed by 40 cycles, denaturation at 94°C for 45 seconds, annealing at 58°C for 45 seconds and extension at 72°C for 1 minute. The PCR was concluded with the extension cycle at 72°C for 2 minutes. The PCR products, 15  $\mu$ l, were visualized on 2% agarose gel electrophoresis, prepared in 1× tris-acetate-EDTA (TAE) buffer, followed by gel staining in ethidium bromide solution; the amplified fragments were observed by ultraviolet light (UV) transilluminator using KODAK Gel Logic 200 Imaging System (Eastman Kodak Company).

## **RESULTS**

When analyzing the samples regarding age and gender, it was observed that 52% of the patients are male and 48.2% correspond to the age group of 60 years or older (**Table 2**).

From 577 samples, the most prevalent microorganism was *Klebsiella pneumoniae* (41%), followed by *Staphylococcus aureus* (22%), *Pseudomonas aeruginosa* (14%), *Enterobacter spp.* (11%),

TABLE 2 – Age group by sex of positive samples between 2010 and 2015

Age group (years)	Total	%	Female	%	Male	%
0-9	40	6.9	20	7.2	20	6.7
10-19	16	2.8	16	5.8	0	0
20-59	206	35.7	103	37.1	103	34.4
≥ 60	278	48.2	124	44.6	154	51.5
Not reported	37	6.4	15	5.4	22	7.4
Total	577	100	278	100	299	100

Acinetobacter baumannii (8%) and Enterococcus faecium (4%). The isolates analyzed were obtained mainly from urine (46%), retal swab (19%), nasal swab (11%) and blood (8%). In urine and rectal swab, the microorganism most commonly found Klebsiella pneumoniae, and in nasal swab and blood, was Staphylococcus aureus (Table 3).

When analyzing gram-positive bacteria, about 80% of the *Enterococcus faecium* strains were resistant to vancomycin — vancomycin-resistant enterococci (VRE) —, and also had a higher resistance to erythromycin (95.8%), followed by ciprofloxacin (91.7%), ampicillin (91.7%) and penicillin G (91.7%), and increased sensitivity to linezolid (87.5%) and daptomycin (83.3%) (**Figure**).

TABLE 3 – Microorganisms by sample type between 2010 and 2015

Microorganism	Urine	Rectal swab	Nasal swab	Blood	Other*	Total
Enterococcos faecium	3	20	-	1	-	24
Staphylococcus aureus	17	-	63	29	19	128
Klebsiella pneumoniae	165	54	-	5	10	234
Acinetobacter baumannii	8	16	1	5	15	45
Pseudomonas aeruginosa	30	9	-	3	39	81
Enterobacter spp.	46	9	-	2	8	65
Total	269	108	64	45	91	577

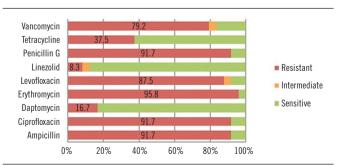


FIGURE - Susceptibility profile of Enterococcus faecium

The red bar represents the percentage of antimicrobial resistance, the orange bar indicates the intermediate profile and the green bar represents the percentage of sensitivity.

From the *Staphylococcus aureus*, 40% strains presented resistance to oxacillin (MRSA), 96.9% were resistant to penicillin G, 75.8% to erythromycin and 63.3% to ciproflxacin and levoflxacin. *S. aureus* strains resistant to linezolid (3.9%), daptomycin (4.7%) and vancomycin (2.3%) were also found.

When analyzing the susceptibility profile for *Klebsiella pneumoniae*, it was possible to observe that this strain presented the highest resistance rates to the following antimicrobials: aztreonam, cefepime and cefotaxime (75.2%) and ertapenem (69.7%); and the lowest rate of resistance was 2.6% to amikacin.

Acinetobacter baumannii presented the most worrying susceptibility profile, presenting a resistance frequency of 100% to imipenem; 91.1% to ciprofloxacin, ceftazidime and cefotaxime; 88.8% to cefepime; 86% to meropenem and 82.2% to levofloxacin.

The highest resistance rates to *Pseudomonas aeruginosa* were 54.3% to ciprofloxacin, 53.1% to levofloxacin and imipenem, and 47% to norfloxacin. While that to *Enterobacter* spp. 96% were resistant to cephalothin; 87.5% to amoxicillin/clavulanic acid; 79.2% to cefoxitin; 78.5% to ampicillin; and 73.9% to cefotaxime.

In molecular analysis, the  $bla_{\text{RPC}}$  gene was found in four  $Klebsiella\ pneumonieae$  strains, which were one in rectal swab specimens and three in urine samples; the  $bla_{\text{NDM}}$  gene was found in three  $Klebsiella\ pneumonieae$  positive retal swab samples, while the  $bla_{\text{OXA-23}}$  gene was found in one  $Acinetobacter\ baumannii$  strain isolated from a tissue fragment culture.

## **DISCUSSION**

The incidence of VRE among the *E. faecium* is worrisome, since these strains have become a worldwide problem due to infections caused in association with hospital morbidity and mortality<sup>(17)</sup>, as well as *S. aureus* strain resistant to linezolid and daptomycin, these antimicrobials are considered as the final option for the treatment of infections caused by MRSA<sup>(18)</sup>.

Compared to a study conducted in Latin America (Argentina, Brazil, Chile and Mexico) from 2008 to 2010, the susceptibility profile analyzed in this study showed a greater resistance to imipenem and meropenem, while in Latin America they showed resistance around 6% to these antimicrobials and 36% to cefepime. However, in this study the resistance found to amikacin and gentamicin was lower than that found in Latin America, 7.8% and 33%, respectively<sup>(19)</sup>. When comparing the results obtained

with a study conducted in 2011 in a university hospital in Londrina, Paraná, Brazil, the profile found in the present study demonstrated a higher frequency of resistance compared to the following antimicrobials: ertapenem (40%) and cefotaxime, cefepime and aztreonam  $(60\%)^{(20)}$ .

Acinetobacter baumannii showed the most worrying susceptibility profile, since this bacterium are easy to develop resistance. In the 1990s, most strains were sensitive to quinolones and carbapenems, and several outbreaks of multidrug resistant strains have been reported in recent years<sup>(21)</sup>. The decrease in susceptibility to carbapenems can be observed when comparing two studies conducted by the Sentry Antimicrobial Surveillance Program in Brazil in 2001 and 2010, especially imipenem, which reduced from 97.8% to 27%, and to meropenem, which previously was 96.7% and today is 27.3%<sup>(19, 22)</sup>. In a study that analyzed only the Gram-negative pathogens of the ESKAPE group in the Latin American countries *A. baumanni* was the pathogen with the highest resistance rate to amikacin, cefepime, ceftazidime, imipenem and levofloxacin<sup>(23)</sup>.

When comparing *Pseudomonas aeruginosa* with a study carried out in Bahia, by Assis *et al.* (2012)<sup>(24)</sup>, 66% of the strains were resistant to levofloxacin and 51%, to amikacin, that is, they showed greater resistance. However, this study showed less resistance to meropenem (34%)<sup>(24)</sup>. Comparing the results obtained with another study conducted in Latin America (Argentina, Brazil, Chile, Colombia, Costa Rica, Ecuador, Guatemala, Mexico, Panama, Peru and Venezuela), it was possible to observe a greater resistance to imipenem (44.9%) and to levofloxacine (38.2%), and a similar resistance rate to meropenem (38.4%) and Amikacin (20.5%)<sup>(25)</sup>.

Regarding *Enterobacter spp.*, this study presented a greater resistance to amoxicillin/clavulanic acid than the study carried out in Rondônia, Brazil, in which the resistance rate was 66.7%<sup>6</sup>.

Klebsiella pneumoniae carbapenemase (KPC) strain, New Delhi metalo-betalactamase (NDM-1) and oxacilinase-23 (OXA-23) are carbapenemases. KPC belongs to Amber class A and NDM-1, to Amber class B [metalo-β-lactamase (MBL)<sup>(26, 27)</sup>]. Class A hydrolyzes penicillins, cephalosporins, carbapenems and

aztreonam, but this hydrolysis is inhibited *in vitro* by the clavulanic acid and tazobactam. However, class B hydrolyzes all beta-lactams mentioned above, with the exception of aztreonam, and does not have its activity inhibited by beta-lactam inhibitors, since its hydrolysis depends on the interaction of betalactam with zinc ions at its catalytic site; its activity is inhibited *in vitro* by EDTA<sup>(14)</sup>. Oxacillinases are class D, and in Brazil OXA-23 stands out<sup>(13, 28)</sup>.

These genes have already been described in Brazil, and the first report of the presence of KPC in brazil was in 2009, after the discovery of the gene in strains isolated from a patient in an intensive care unit at a tertiary hospital in Recife, Brazil<sup>(29)</sup>. The first strain containing NDM-encoding gene in Brazil was isolated in Rio Grande do Sul and later a strain with this gene was also found in the Federal District<sup>(15, 30)</sup>. The first description of OXA-23 in Brazil was established from *A. baumannii* isolates found in 1999 in two tertiary hospitals in Curitiba, Paraná, Brazil<sup>(31)</sup>.

It is important to emphasize that the hospital studied is considered small and, although only few strains with resistance profile have been found, this is still worrying. This resistance is not specific to this hospital, since most of the patients come from other hospitals in the public health network of the Federal District, which arrive already colonized or infected by multidrug resistant (MDR) microorganisms.

#### **CONCLUSION**

Braziltookanimportantsteptowardsthe control of multi-drug resistant strains by prohibiting the sale of antimicrobials without prescription through the RDC 44/2010, which was replaced by the RDC 20/2011 of the National Sanitary Surveillance Agency [Agência Nacional de Vigilância Sanitária (Anvisa)], reducing its irrational use. However, many practitioners still prescribe antimicrobials without any need, or prescribe broad spectrum antimicrobials when the first-choice antimicrobials are effective. Thus, this work demonstrates important data that reflect an increase in the prevalence of MDR strains in hospitalized patients of the Federal District, Brazil.

#### **RESUMO**

Introdução: Os principais patógenos causadores de infecções nosocomiais foram resumidos pela sigla ESKAPE, que são as iniciais das seguintes bactérias: Enterococcus faecium, Staphylococcus aureus, Klebsiella pneumoniae, Acinetobacter baumanni, Pseudomonas aeruginosa e Enterobacter spp., as quais possuem altas taxas de resistência por conseguirem escapar das ações dos antimicrobianos. Objetivo: Traçar o perfil de suscetibilidade antimicrobiana do grupo ESKAPE em um bospital primário da rede pública do Distrito Federal, Brasil. Métodos: Foi realizado um estudo transversal, retrospectivo e descritivo, analisando os dados correspondentes de janeiro de 2010 a dezembro de 2015 para as amostras consideradas positivas para o grupo ESKAPE, com o intuito de gerar um perfil de sensibilidade aos antimicrobianos. Resultados: Ao analisar bactérias Gram positivas, quase 80% das cepas de Enterococcus faecium foram resistentes à vancomicina (VRE) e cerca de 40% das cepas de Staphylococcus aureus, resistentes à oxacilina (MRSA). Nas bactérias do grupo ESKAPE, observaram-se cepas com uma taxa de resistência maior aos carbapenens do que em outros estudos. Ao realizar uma análise molecular, quatro cepas de Klebsiella pneumoniae foram positivas para o gene bla<sub>RPC</sub> e três, para o bla<sub>RDM</sub>; uma de Acinetobacter baumanni foi positiva para o gene bla<sub>QCA-23</sub>. Conclusão: Estudos como este devem ser realizados periodicamente de modo a avaliar o perfil de suscetibilidade das bactérias. Eles demonstram a importância do uso de estratégias para evitar infecções nosocomiais, bem como um maior controle na prescrição de antimicrobianos.

*Unitermos: testes de sensibilidade microbiana; genes; bactérias.* 

#### REFERENCES

- 1. Alekshun MN, Levy SB. Molecular mechanisms of antibacterial. Multidrug resistance. Cell. 2007; 128(6): 1037-50.
- 2. Huycke MM, Sahm DF, Gilmore MS. Multiple-drug resistant enterococci: the nature of the problem and an agenda for the future. Emerg Infect Dis. 1998; 4(2): 239-49.
- 3. Verraes C, Van Boxstael S, Van Meervenne E, et al. Antimicrobial resistance in the food chain: a review. Int J Environ Res Public Health. 2013; 10(7): 2643-69.
- 4. Davies J. Where have all the antibiotics gone? Can J Infect Dis Med Microbiol. 2006; 17(5): 287-90.
- 5. Demain AL, Spizek J. The antibiotic crisis. In: Tegos G, Mylonakis E, editors. Antimicrobial drug discovery: emerging strategies. Wallingford, UK: CAB International; 2012. p. 26-43
- 6. Grillo VTRS, Gonçalves TG, Campos Júnior J, Paniágua NC, Teles CBG. Incidência bacteriana e perfil de resistência a antimicrobianos em pacientes pediátricos de um hospital público de Rondônia, Brasil. Rev Ciênc Farm Básica Apl. 2013; 34(1): 117-23.
- 7. Oliveira AC, Gonzaga C, Costa R, Damaceno QS, Garbaccio JL. Desafios e perspectivas para a contenção da resistência bacteriana na óptica dos profissionais de saúde. Rev Eletr Enf. 2013; 15(3): 747-54.
- $8. \ \ WHO. \ \ World \ \ Health \ \ Organization. \ Antimicrobial \ \ resistance: \ global \ \ report on surveillance; \ 2014.$
- 9. Rice LB. Federal funding for the study of antimicrobial resistance in nosocomial pathogens: no ESKAPE. J Infect Dis. 2008; 197: 1079-81.
- 10. Gosh AN, Bhatta DR, Ansari MT, et al. Application of WHONET in the antimicrobial resistance surveillance of uropathogens: a first user experience from nepal. J Clin Diagn Res. 2013; 7(5): 845-8.

- 11. CLSI. Clinical Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing; twenty-fifth informational supplement. CLSI document M100-S25. Wayne, PA: Clinical and Laboratory Standards Institute; 2015.
- 12. Giske CG, Gezelius L, Samuelsen O, Warner M, Sundsfjord A, Woodford N. A sensitive and specific phenotypic assay for detection of metalloblactamases and KPC in Klebsiella pneumoniae with the use of meropenem disks supplemented with aminophenylboronic acid, dipicolinic acid and cloxacillin. Clin Microbiol Infec. 2011; 17(4): 552-6.
- 13. Chagas TPG, Silveira MC, Albano RM, Carvalho-Assef APD, Asensi MD. Draft genome sequence of a multidrug-resistant Acinetobacter baumannii ST15 (CC15) isolated from Brazil. Mem Inst Oswaldo Cruz. 2015; 110(5): 691-2.
- 14. Nordmann P, Dortet L, Poirel L. Carbapenem resistance in Enterobacteriaceae: here is the storm! Trends Mol Med. 2012; 18(5): 263-72.
- 15. Faria Junior C, Rodrigues LO, Carvalho JO, Franco OL, Pereira AL; Brasilia Study Group On Bacterial Resistance. NDM producing enterobacteriaceae strains among hospitals in Brasília, Brazíl. J Microbiol Exp. 2016; 3(2).
- 16. Woodford N, Ellingtona MJ, Coelho JM, et al. Multiplex PCR for genes encoding prevalent OXA carbapenemases in Acinetobacter spp. Int J Antimicrob Agents. 2006; 27(4): 351-3.
- 17. Rosa RG, Schwarzbold AV, Santos RP, Turra EE, Machado DP, Goldani LZ. Vancomycin-resistant Enterococcus faecium bacteremia in a 50 tertiary care hospital: epidemiology, antimicrobial susceptibility, and outcome. Bio Med Res International. 2014; 1-6.
- 18. Monaco M, Araujo FP, Cruciani M, Coccia EM, Pantosti A. Worldwide epidemiology and antibiotic resistance of Staphylococcus aureus. Curr Top Microbiol Immunol. 2016; 1-36.

- 19. Gales AC, Castanheira M, Jones RN, Sader HS. Antimicrobial resistance among Gram-negative bacilli isolated from Latin America: results from SENTRY Antimicrobial Surveillance Program (Latin America, 2008-2010). Diagn Microbiol Infect Dis. 2012; 73: 354-60.
- 20. Rossi DJ, Rechenchoski DZ, Vivan ACP, et al. Evolução da resistência de Klebsiella pneumoniae no Hospital Universitário de Londrina no período de 2000 a 2011. Semina Cienc Biol Saude. 2015; 36(1): 267-74.
- 21. Ogutlu A, Guclu E, Karabay O, Utku AC, Tuna N, Yahyaoglu M. Effects of carbapenem consumption on the prevalence of Acinetobacter infection in intensive care unit patients. Ann Clin Microbiol Antimicrob. 2014: 13: 7.
- 22. Sader HS, Jones RN, Gales AC, Silva JB, Pignatari AC. SENTRY antimicrobial surveillance program report: latin american and brazilian results for 1997 through 2001. Braz J Infect Dis. 2004; 8(1): 25-79.
- 23. Karlowsky JA, Hoban DJ, Hackel MA, Lob SH, Sahm DF. Resistance among Gram-negative ESKAPE pathogens isolated from hospitalized patients with intra-abdominal and urinary tract infections in Latin American countries: SMART 2013-2015. Braz J Infect Dis. 2017; 21(3): 343-8.
- 24. Assis DAM, Oliveira FCS, Costa GB, et al. Prevalência e suscetibilidade de bactérias do grupo ESKAPE nas diversas infecções em pacientes do CTI de um hospital no sul da Bahia. Proceedings Congresso Brasileiro

- de Microbiologia; 2011; Natal. São Paulo: Sociedade Brasileira de Microbiologia; 2011. ref. 792-2.
- 25. Jones RN, Guzman-Blanco M, Gales AC, et al. Susceptibility rates in Latin American nations: report from a regional resistance surveillance program (2011). Braz J Infect Dis. 2013; 17(6): 672-81.
- 26. Echeverri Toro LM, Cataño Correa JC. Klebsiella pneumoniae como patógeno intrahospitalario: epidemiología y resistência. Iatreia. 2010; 23(3): 240-9.
- 27. Nordmann P. Carbapenemase-producing enterobacteriaceae: overview of a major public health challenge. Med Mal Infect. 2014; 44(2): 51-6.
- 28. Chagas TP, Carvalho KR, Santos ICO, Carvalho-Assef AP, Asensi MD. Characterization of carbapenem-resistant Acinetobacter baumannii in Brazil (2008-2011): countrywide spread of OXA-23-producing clones (CC15 and CC79). Diagn Microbiol Infect Dis. 2014; 79: 468-72.
- 29. Monteiro J, Santos AF, Asensi MD, Peirano G, Gales AC. First report of KPC-2-producing Klebsiella pneumoniae strains in Brazil. Antimicrob Agents Chemother. 2009; 53(1): 333-4.
- 30. Carvalho-Assef AP, Pereira OS, Albano RM, et al. Isolation of NDM-producing providencia rettgeri in Brazil. J Antimicrob Chemother. 2013; 68: 2956-7.
- 31. Dalla-Costa LM, Coelho JM, Souza HAPHM, et al. Outbreak of carbapenem-resistant Acinetobacter baumannii producing the OXA-23 enzyme in Curitiba, Brazil. J Clin Microbiol. 2003; 41(7): 3403-6.

#### CORRESPONDING AUTHOR

#### Thaís Alves da Costa Lamounier

Universidade de Brasília; Faculdade de Ceilândia; Centro Metropolitano, conjunto A, lote 01; CEP: 72220-275; Brasília-DF, Brasíl; e-mail: lamounier@unb.br.