Ertapebem disk performance to predict *Klebsiella* pneumoniae carbapenemase produced by Gram-negative bacilli isolated in a São Paulo city public hospital

Desempenho do disco de ertapenem como preditor da produção de *Klebsiella pneumoniae* carbapenemase por bacilos Gram-negativos isolados de culturas em um hospital municipal de São Paulo

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ABSTRACT

Objective: To evaluate ertapenem disk performance to predict *Klebsiella* pneumonie carbapenemase production by Gram-negative bacilli. Methods: All Gram-negative bacilli isolated between January 2010 and June 2011 were tested by disk diffusion (Oxoid™) for sensitivity to ertapenem, meropenem and imipenem. Resistant or intermediate sensitivity strains (diameter ≤22 mm for ertapenem) were also tested for the blaked gene by polymerase chain reaction. Disk predictive positive value for Klebsiella pneumoniae carbapenemase and specificity were calculated. Results: Out of the 21839 cultures performed, 3010 (13.78%) were positive, and Gram-negative bacilli were isolated in 708 (23.52%) of them. Zone of inhibition diameter for ertapenem disk was ≤22 mm for 111 isolates, representing 15.7% of all Gram-negative isolates. The PCR assay for blaked detected 40 Klebsiella pneumoniae carbapenemase-producing strains. No strains intermediate or resistant to meropenem and imipenem were sensitive to ertapenem. The ertapenem disk presented a positive predictive value of 36% to predict blakec and 89% specificity. Conclusion: The resistance of Gram-negative bacilli detected by disk diffusion against ertapenem does not predict Klebsiella pneumoniae carbapenemase production. Other mechanisms, such as production of other betalactamases and porin loss, may be implicated. The need to confirm the presence of the blakec is suggested. Therefore, ertapenem was a weak predictor for discriminating strains that produce Klebsiella pneumoniae carbapenemase.

Keywords: Klebsiella; Carbapenems; Drug resistance, multiple, bacterial

RESUMO

Objetivo: Avaliar o desempenho do disco de ertapenem para predizer micro-organismos produtores de Klebsiella pneumoniae carbapenemase. Métodos: Bacilos Gram-negativos isolados em cultura entre janeiro de 2010 e junho de 2011 foram testados por disco-difusão (Oxoid™) para ertapenem, meropenem e imipenem. As cepas consideradas intermediárias ou resistentes (halo≤22mm) para ertapenem foram encaminhadas para a pesquisa do blakec por reação em cadeia da polimerase. Calcularam-se o valor preditivo positivo e a especificidade do disco. Resultados: Foram realizadas 21.839 culturas nesse período, sendo 3.010 (13,78%) positivas. Bacilos Gram-negativos foram isolados em 708 (23,52%) destas. A zona de inibição do disco de ertapenem foi ≤22mm para 111 (15,67%) dos isolados. A pesquisa do blakec caracterizou 40 cepas produtoras de Klebsiella peneumoniae carbapenemase. Não houve nenhum caso de disco intermediário ou resistente para meropenem ou imipenem com ertapenem sensível. O valor preditivo positivo foi de 36% e a especificidade calculada do disco de ertapenem para produção de Klebsiella pneumoniae carbapenemase foi de 89% em nosso serviço. Conclusão: A resistência ao disco de ertapenem não define bacilo produtor de Klebsiella pneumoniae carbapenemase. Mecanismos, como produção de outras betalactamases e perda de porinas, podem estar implicados. Sugerese a necessidade da confirmação da presença do gene blakec. O ertapenem, portanto, mostrou-se fraco preditor para discriminar cepas produtoras de Klebsiella pneumoniae carbapenemase.

Descritores: Klebsiella; Carbapenêmicos; Farmacorresistência bacteriana múltipla

Study carried out Hospital Municipal Dr. Moysés Deustch - M'boi Mirrim, São Paulo (SP), Brazil; Hospital Israelita Albert Einstein - HIAE, São Paulo (SP), Brazil

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Conflict of interest: none.

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INTRODUCTION

Ertapenem (ERT) is a beta-methyl-carbapenem that is active against Gram-negative bacteria producing extended-spectrum beta-lactamases (ESBL) or AmpC, and it is widely used in clinical practice since 2001. In the United States, the emergence of resistance against carbapenems is usually related to the production of carbapenemases, such as metallo-beta-lactamases and *Klebsiella pneumoniae* carbapenemase (KPC). However, other mechanisms may be involved in the resistance to this antibiotic class, including porin loss associated with ESBL or AmpC⁽¹⁾ production.

KPC is a class-A carbapenamase that inactivates all beta-lactam antibiotics. It was first described in *Klebsiella pneumonia*, but it has also been occasionally detected in other enterobacteria. This enzyme is coded by sequences related to transposons and are present in conjugative plasmids with high dissemination power⁽²⁾.

The identification of KPC-producing Gram-negative bacilli (GNB) is mandatory, as they may cause severe infections, and carbapenems (imipenem – IMP and meropenem – MER) are the therapy of choice in many nosocomial infections. In addition, the identification of carrier individuals allows controlling the dissemination of those agents. No precise phenotypic tool for their identification has yet been described, and the available tools are not able to differentiate resistance mechanisms^(3,4).

The Brazilian Health Surveillance Agency (ANVISA) currently recommends disk-diffusion with carbapenems for screening and determination of minimum inhibitory concentrations (MIC), followed by confirmation using molecular biology to identify the gene *bla*_{KPC}⁽⁵⁾. The ERT disc is considered a very sensitive marker, but data published in Brazilian and international literature on its specificity are still subjective and conflicting⁽⁶⁻⁸⁾.

OBJECTIVES

To evaluate the positive predictive value and the specificity of the ertapenem disk to predict the presence of KPC-producing Gram-negative bacilli.

METHODS

Gram-negative bacilli (GNBs) isolated in biological material cultures collected and processed at *Hospital Municipal Dr. Moiysés Deautch*, in M´Boi Mirim, SP, Brazil, between January 2010 and June 2011, were tested by disk-diffusion (Oxoid™) for ERT, MER, and IMP in Mueller-Hinton agar (BioMérieux®). The interpretation criteria applied were those suggested in the Technical Note published by ANVISA in 2010.

The strains that were considered intermediate or resistant to ERT (zone of inhibition diameter \leq 22 mm), MER (zone of inhibition diameter \leq 20mm) or IMP (zone of inhibition diameter \leq 20mm) were submitted to bla_{KPC} detection by polymerase chain reaction (PCR). DNA was extracted as previously described. Direct primer 5´TCGCTAACTCGAACAGG3´ and reverse primer 5´TTACTGCCCGTTGACGCCCAATCC3´ were used for amplification.

ERT disk positive predictive value (PPV) and specificity were calculated. PPV was calculated based on the true positive (TP)/TP+false positive (FP) ratio, where TP represents the strains positive for the $bla_{\rm KPC}$ gene and FP the number of ERT-resistant strains, but with negative PCR results. Assuming 100% ERT sensitivity, specificity was calculated by the d/b+d ratio, where "d" represents the ERT-sensitive strains, and "b" the ERT-resistant or intermediate strains negative for $bla_{\rm KPC}$.

RESULTS

Out of a total of 21839 cultures from different biological materials performed during that period, 3010 (13.78%) were tested positive, with GNB isolated in 708 (23.52%) of these positive cultures.

ERT disk zone of inhibition was \leq 22mm in 111 (15.67%) of the isolates, out of which 96 (86.48%) were considered resistant and 15 (13.51%) intermediate. Strains were submitted to molecular testing.

The blaked gene tests characterized 40 KPC-producing strains. Klebsiella pneumoniae was biochemically identified in 38 cases and Enterobacter cloacae in 2 cases. The bacteria were isolated in the urine in 16 cases, rectal/anal swab in 17 cases, tracheal secretion in 4 cases, blood in 2 cases and pancreatic abscess in one case. Eleven of the infected patients died. In all genepositive cases, the zone of inhibition diameter of the ERT disk was considered resistant. No MER or IMP intermediate or resistant samples were sensitive to ERT.

The results showed that there were 40 blakperpositive strains out of the 111 strains found to be resistant or intermediate to ERT among 708 isolated GNB. Calculated ERT disk PPV and specificity for KPC production were 36% and 89%, respectively.

DISCUSSION

The study of Anderson et al.⁽⁷⁾, considering the potential clinical impact, stresses the importance of evaluating the sensitivity to ERT because this is the most sensitive indicator of KPC, independently of the method applied.

However, Woodford et al.⁽⁸⁾ mentioned that the *in-vitro* resistance to that carbapenem is not specific for KPC production, particularly when considering centers where carbapenemase-producing bacteria are rare.

The present study shows objective data on PPV and specificity of the ERT disk to predict KPC in our center. It was observed that ERT-disk resistance does not determine that the isolated GNB produces KPC. Other resistance mechanisms, such as the production of other beta-lactamases and porin loss, may be implicated. Therefore, ERT was a weak predictor for the identification of KPC-producing strains in our cohort.

CONCLUSION

Despite the efforts to find an ideal phenotypic method to trace KPC production, there are still no results supporting the individual use of these tools. There are many groups working to find better methodologies, but no definite conclusions have been published yet. Consistent with ANVISA guidelines, the data obtained in the present study also indicate the need of confirming the presence of the *bla*_{KPC} gene to determine KPC production using molecular biology.

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