Long-lasting endothelium-dependent relaxation of isolated arteries caused by an extract from the bark of *Combretum leprosum*

Extrato das cascas de *Combretum leprosum* causa relaxamento dependente de endotélio de longa duração em artérias isoladas

Francisco das Chagas Alves Filho¹, Paulo Marques da Silva Cavalcanti², Rita de Cassia Aleixo Tostes Passaglia¹, Gustavo Ballejo¹

ABSTRACT

Objective: To describe and to characterize the relaxing effect of an extract of the bark of Combretum leprosum on isolated arterial rings from different animals. Methods: Rings (3 to 4mm) from rabbit, rat, or porcine arteries rings were suspended in an organ bath (Krebs, 37°C, 95%0,/5%C0₂) to record isometric contractions. After the stabilization period (2 to 3 hours) contractions were induced by the addition of phenylephrine (0.1 to $0.3\mu M$) or U46619 (10 to 100nM), and Combretum leprosum extract was added on the plateau of the contractions. Experiments were performed to determine the potency, duration, reversibility, and to get insights on the potential mechanism involved in extract-induced relaxations. Results: In all rings tested, Combretum leprosum extract (1.5µg/mL) was able to cause relaxations, which were strictly endothelium-dependent. In rabbit or rat thoracic aorta rings, the relaxations were reversed by vitamin B_{12a} or L-N^Gnitroarginine. In porcine right coronary arteries and rabbit abdominal aorta, extract caused both L-NG-nitroarginine-sensitive and L-NGnitroarginine-resistant relaxations. In rabbit thoracic aorta, the extract was relatively potent (EC₅₀=0.20 μ g/mL) and caused relaxations; intriguingly the endothelium continued to produce relaxing factors for a long period after removing the extract. The magnitude of extract-induced relaxations was significantly reduced in the absence of extracellular Ca^{2+} ; in addition, the TRPs channels blocker ruthenium red (10 μ M) was able to revert extract-induced relaxations. Phytochemical analyses indicated that the extract was rich in polyphenol-like reacting substances. **Conclusions:** Combretum leprosum extract contains bioactive compounds capable of promoting Ca2+-dependent stimulation of endothelial cells which results in a prolonged production of relaxing factors.

Keywords: *Combretum*; Arteries/drug effects; Endothelium/drug effects; Nitric oxide/pharmacology; Polyphenols/pharmacology

RESUMO

Objetivo: Descrever e caracterizar os relaxamentos induzidos por um extrato das cascas de Combretum leprosum em anéis de artérias de diferentes espécies de animais. Métodos: Anéis (3 a 4mm) de artérias de coelho, rato e porco foram montados em cubas para órgão isolado (Krebs, 37°C, 95%0,/5%CO,) para registro das contrações isométricas. Após um período de estabilização (2 a 3 horas), as contrações foram induzidas com fenilefrina (0,1 a 0,3 μ M) ou U46619 (10 a 100nM); no platô dessas contrações, adicionamos o extrato Combretum leprosum. Diferentes protocolos foram realizados para determinar potência, duração, reversibilidade e mecanismo dos relaxamentos induzidos pelo extrato. Resultados: Em todas as preparações testadas, o extrato de Combretum leprosum (1,5µg/mL) provocou relaxamentos dependentes de endotélio. Em aorta torácica de coelho ou rato, os relaxamentos foram revertidos pela vitamina B₁₂a ou L-NG-nitro-arginina. Em anéis de aorta abdominal de coelho e de artérias coronárias de porco, o extrato causou relaxamentos sensíveis e resistentes à L-NG-nitro-arginina. Em aorta torácica de coelho, o extrato foi relativamente muito potente $(EC_{50}=0,20\mu g/mL)$ e quando causou relaxamentos; intrigantemente o endotélio continuou a produzir fatores relaxantes por um longo período após remoção do extrato. A magnitude dos relaxamentos induzidos pelo extrato foi significativamente reduzida em ausência Ca2+ extracelular; ademais, o vermelho de rutênio (10µM), um bloqueador de canais TRPs, foi capaz de reverter os relaxamentos induzidos pelo extrato. Análises preliminares indicaram que o extrato continha compostos com reatividade química semelhante à polifenóis. Conclusão: 0 extrato de Combretum leprosum contem compostos bioativos capazes de promover estimulação dependente de Ca2+ das células endoteliais a qual resulta numa produção prolongada de fatores relaxantes.

Descritores: *Combretum*; Artérias/efeitos de drogas; Endotélio/efeitos de drogas; Óxido nítrico/farmacologia; Polifenóis/farmacologia

Corresponding author: Francisco das Chagas Alves Filho – Avenida Bandeirantes, 3,900 – Monte Alegre – Zip code: 14049-900 – Ribeirão Preto, SP, Brazil – Phone: (55 16) 3206-3326 E-mail: semprechycoman@yahoo.com.br

Received on: Jan 14, 2015 - Accepted on: June 24, 2015

Conflict of interest: none.

DOI: 10.1590/S1679-45082015AO3242

¹ Faculdade de Medicina de Ribeirão Preto, Universidade de São Paulo, Ribeirão Preto, SP, Brazil,

² Universidade Federal do Piauí, Teresina, PI, Brazil.

INTRODUCTION

Combretum leprosum Mart. (Combretaceae) is a shrub or small tree that grows in northeastern Brazil where it is known as *mufumbo*, *mofumbo*, *cipoaba* and *pente-de-macaco*. In folk medicine, extracts from different parts of the plant are being used for alleged expectorant, hemostatic, sedative, and aphrodisiac properties. (1,2)

In a bioactivity-based screening of the potential effects of the lyophilized hydroalcoholic extract of *Combretum leprosum* (ECL) bark on visceral and vascular smooth muscles, it was observed that the extract caused a potent endothelium-dependent relaxation (EDR) of rabbit thoracic aorta rings. Although some substances isolated from related species of the *Combretum* genus have been shown to cause EDR in rat aorta (mollic acid glucoside, isolated from *Combretum molle*) as well as methanolic fractions rich in flavonoids from *Combretum celastroides* and *Combretum racemosum*, ^(3,4) all previously reported effects of flower, leaf and/or root ECL did not include any cardiovascular bioactivity.

Endothelial cells exhibit remarkable heterogeneity in responsiveness to agents that induce EDR either of homologous arteries from different animals species or of arteries from different vascular beds from the same animal; *e.g* acetylcholine (ACh), but not bradykinin (BK), induces EDR in the rabbit and rat aorta; ^(5,6) similarly, histamine causes EDR in rat aorta but not in rabbit aorta. Therefore, it is highly recommended when describing the EDR of a new drug or natural product to examine its effects in same artery from more than one animal species and in different arteries from the same animal.

OBJECTIVE

To describe and interpret the results of experiments conceived to answer the following questions regarding the observed novel effect of extract of *Combretum leprosum*: (1) Which is the site of action, the potency, and the duration of the effect of the extract of *Combretum leprosum* in provoking relaxations of the rabbit thoracic aorta rings? (2) Is the extract of *Combretum leprosum* capable of causing endothelium-dependent relaxation in isolated arterial rings from other vessels of the rabbit or from vessels of rat, mice, guinea-pig and pig? (3) Which are the possible mechanisms involved in the extract of *Combretum leprosum*-induced relaxations of rabbit thoracic aorta rings?

METHODS

Plant material

The stem bark of *C. leprosum* was collected in the morning (10 to 11:00am) on July 15, 2005, at the

Agrarian Science Center, *Universidade Federal do Piau*í, Teresina (PI), Brazil. A voucher specimen (number 10,557) was deposited in the Graziela Barroso Herbarium (TEPB, at the same institution). The plant material was dried in the shade at 40±1°C, and the stem bark powder (500g) was extracted (three times) with 1L of 70% ethanol, evaporated in a vacuum at 50°C and lyophilized to obtain a dry extract, which was stored under refrigeration (4°C) until further use. The extract was freshly diluted in distilled water for experiments.

Isolated organ experiments

Thirty male and female New Zealand rabbits (2.5 to 3.5kg) and 20 Wistar rats (200 to 250g), 5 male guinea-pigs (350g), and 10 male mice (24 to 30g) were employed for all the experiments. The animals were obtained from the vivarium of the Ribeirão Preto Campus of the Universidade de São Paulo. Hearts from ten pigs were obtained from a local slaughterhouse; they were removed immediately after the death of the animal, washed with cold Krebs solution to remove most of the blood, and transported to the laboratory in sealed plastic bags in a cooler box containing crushed ice. Within 2 to 3 hours after heart removal from the animals, the right and/or left coronary arteries were dissected out for preparation of the arterial rings. All experimental protocols were in accordance with the ethical principles for animal experimentation recommended by the Colégio Brasileiro de Experimentação Animal [Brazilian College of Animal Experimentation] and were approved by the Animal Experimentation Ethics Committee (number 084/2011) of Faculdade de Medicina de Ribeirão Preto of Universidade de São Paulo.

Rats, mice, guinea-pigs, and rabbits were euthanized under sodium pentobarbital anesthesia (50mg/kg i.p. for rats, mice, and guinea-pigs and/or by marginal ear vein injection for rabbits). The thoracic descending aorta (from all animals) as well as the abdominal aorta, the superior mesenteric artery, and the common carotid artery (from rabbits) were quickly removed and placed in Krebs solution (116mM of sodium chloride -NaCl; 4.5nM potassium chloride - KCl; 1.14mM of monobasic sodium phosphate - NaH₂PO₄; 1.16nM of magnesium chloride - MgCl₂; 2.5nM of Calcium chloride - CaCl₂; 25nM od Sodium bicarbonate -NaHCO₃; and 11.1 of D-glucose) containing diclofenac (10µM) to inhibit synthesis of cyclooxygenase derivedproducts.⁽⁶⁾ After all adherent tissues were removed, the arteries were cut into rings (3 to 4mm) taking care to preserve the endothelium; when required, the intima of some rings was rubbed with a metal stick to destroy the endothelium. The rings were suspended between thin metal brackets and mounted under an initial tension of 2 to 4g in isolated organ chambers (volume: 5 to 10mL) containing Krebs solution at $37\pm1^{\circ}\text{C}$ bubbling continuously with 5% of carbon dioxide (CO₂) and 95% of oxygen (O₂). The isometric tension of each ring was continuously recorded and stored using the data acquisition system Lab Chart software.

Experimental protocols

To test the viability of the rings, submaximal concentrations of phenylephrine (PE; 0.1 to 1μ mol/L) or U46619 (10 to 30nmol/L) were added twice at hourly intervals. During the contraction plateau induced by the second addition of PE (or U46619), the functional integrity of the endothelium was tested by addition of ACh (1μ mol/L) or BK (0.1μ mol/L, pig coronary artery). Those rings in which ACh (or BK) caused a decrease of at least 80% of PE (or U46619)-induced tension were considered as containing intact functional endothelium. One hour after determining endothelium functionality, PE (or U46619) was again added to contract the rings and at the plateau of the contraction, and a supramaximal concentration of ECL (1.5μ g/mL) was added.

To determine whether nitric oxide (NO) synthesis and/or release were required for ECL-induced relaxations, L-N^G-nitro-L-arginine (L-NNA, 300μ mol/L) or hydroxocobalamin (B_{12a}; 30 to 100μ M) were added when the relaxation response induced by ECL attained a steady state. To determine whether ECL-induced relaxations required the presence of extracellular Ca²⁺, the effect of ECL was tested in preparations incubated in Krebs solution containing nominally zero Ca²⁺. To determine whether Ca²⁺ influx into endothelial cells was involved in ECL-induced relaxations, the effect of the addition of ruthenium red (RR; 10μ mol/L, a non-selective blocker of Ca²⁺ permeable channels) when the relaxation response induced by ECL attained a steady state was also examined.

The potency of ECL was calculated from concentration-effect curves obtained in a non-cumulative manner using intact endothelium rings from rabbit thoracic aortas, *i.e.* by adding single concentrations of ECL (0.1, 0.3, and $1.0\mu g/mL$) on the plateau of PE-induced contractions at 45 to 60 minutes intervals. All drugs and the extract solutions were added in the medium chamber through a pipette in a volume of 1 to $30\mu L$.

Measurements of polyphenols

To estimate the total phenolic OH groups, the Folin-Ciocalteu (FC) method was utilized. (7) The ECL (0.5 mg/mL) solutions were diluted in aqueous sodium hydroxide (NaOH; 1%) to produce solutions with following concentrations: 1.0, 2.5, 5.0, 7.5, 10, and $12.5\mu g/mL$. To these solutions, $500\mu L$ of FC reagent were added followed by vigorous shaking for 2 minutes. Later, 500µL of sodium carbonate (Na₂CO₃; 75g/mL) were added and these solutions were incubated at room temperature or at 50°C, for 20 minutes. Incubations were ended by cooling the solutions in ice water bath for 5 minutes. The specific absorbance at 700nm was determined using an Enzyme Linked Immuno Sorbent Assay (ELISA) plate reader; determinations were performed in triplicate. Solutions of gallic acid (GA) and quercetin (dissolved in 10% aqueous methanol) were utilized as standard polyphenol compounds.

Statistical analyses

The ECL-induced relaxations of arterial rings were expressed as percentage of reduction (% tension) of the tension developed by PE- or U46619-induced contractions; the reversion of these relaxations by inhibitors was expressed as percentage of the relaxation magnitude (% relaxation). Values are presented as mean \pm standard error of the mean (SEM). The ECL concentration causing half-maximal relaxation (EC₅₀) was determined by fitting the original concentration-response curve to a sigmoidal curve using GraphPad Prism® Software, version 5.0. Values (in percentage) of changes in the tonus were analyzed by the *t* Student test (two-tailed), paired (in the same arterial ring) or unpaired (in the different arterial rings). Values of p<0.05 were considered statistically significant.

Chemicals

ACh, BK, FC reagent, GA, B_{12a}, PE, quercetin, reagent grade ethanol, RR, and U46619 were obtained from Sigma (United States). Diclofenac was obtained from Calbiochem™ (United States). L-NNA was obtained from Research Biochemicals International (United States).

RESULTS

Initial experiments using rings from rabbit thoracic aorta with intact endothelium showed that an aqueous suspension of ECL at the concentration of 1.0 to $1.5\mu g/mL$ caused relaxations of magnitude similar to or even

greater than those caused by ACh (1 μ M) (Figure 1A). While relaxations elicited by ACh were initiated almost immediately after its addition, those elicited by ECL required between 60 to 90 seconds to become apparent; furthermore while the relaxing effect of ACh reached a plateau in about 2 minutes, ECL induced effects took about 10 minutes to reach the plateau. The maximal velocity of relaxations, measured from the first derivative equation (dT/dt) were -90.33 \pm 5.41mg/s (n=8) and -39.83 \pm 2.28mg/s (n=8) for ACh (1 μ M) and ECL (1.0 μ g/mL), respectively (Figure 1B).

ECL-induced relaxation could be due to a direct action on the smooth muscle cells or to an indirect action mediated by the endothelium as is now well established for ACh and other agents. (5) To distinguish between these alternatives, the effect of ECL was determined in rabbit thoracic aorta rings containing endothelium or in rings in which the endothelium had been removed,

as can be observed in figure 1C. ECL did not cause relaxations in rings without endothelium.

In order to determine whether ECL caused EDR in other arteries from the rabbit or in arteries from other animal species, the effect of ECL was determined in rings from rabbit abdominal aorta, rabbit superior mesenteric artery, rabbit common carotid artery, rat thoracic aorta, guinea-pig thoracic aorta, mouse thoracic aorta, and porcine coronary arteries. As shown in figure 1D and 1E, ECL at $1.5\mu g/mL$ caused relaxations which were also strictly endothelium-dependent in rat thoracic aorta and in porcine right coronary artery rings. Similar findings were observed in all the other arteries tested.

In rabbit and rat thoracic aorta, EDR are entirely mediated by NO; thus, the effect of a nitric oxide synthase (NOS) inhibitor (L-NNA) and of a NO scavenger (B_{12a}) upon ECL- induced EDR was examined. As shown in figures 2A, 2B, 2C, 2D, and 2E the addition of L-NNA

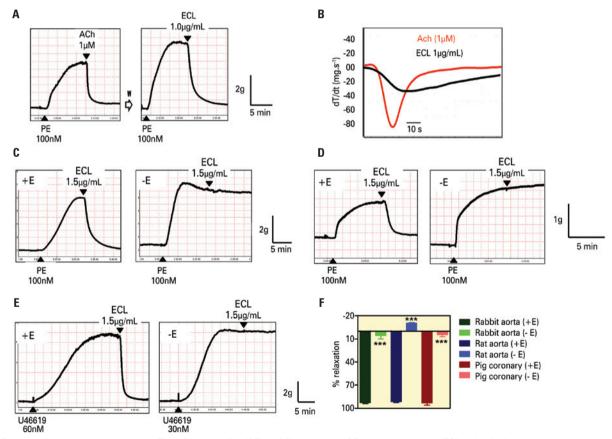


Figure 1. Representative tracings showing the effect of acetylcholine (ACh; 1μ M) and extract of *Combretum leprosum* (ECL; 1.0μ g/mL) on the isometric tension developed by rings of rabbit thoracic aorta pre-constricted with Phenylephrine (PE; 100nM) (A). The first derivative (dT/ dt, mg/s) of relaxing effects induced by ECL (1.0μ g/mL) and ACh (1μ M) (B). The effect of ECL (1.5μ g/mL) on the isometric tension developed by rings, with (+E) or without (-E) endothelium, of rabbit thoracic aorta (C), rat thoracic aorta (D) pre-constricted with phenylephrine (PE), and porcine right coronary artery pre-constricted with U46619 (E). In (F), mean \pm standard error of the mean (SEM) values of the magnitude of the relaxations observed in 4-20 similar experiments (relaxation magnitude is expressed as a percentage of reduction of PE-or U46619-induced contraction). ***Two-tailed p < 0.0001 (unpaired t-test). W: washout of the preparation for 60-45 minutes

or B_{12a} on the plateau of ECL-induced relaxations of rabbit and rat aortic rings completely reversed the relaxations. ECL (1.5 μ g/mL) was also able to induce relaxations in rabbit abdominal aorta in the presence of L-NNA (100 μ M, 20 minutes) (Figure 2F). The magnitude of these relaxations was smaller than the ones observed in the absence of L-NNA.

In experiments designed to calculate the potency of the extract by the single concentration method using the same rabbit thoracic aorta ring, it was observed that the magnitude of the relaxation induced by low concentrations (0.1 to $0.3\mu g/mL$) of ECL increased with successive additions and required at least three additions of the same concentration at hourly intervals in order to become stabilized (Figures 3A and 3B); furthermore it was observed that an initial priming of the rings with $1\mu g/mL$ was also

required. In these conditions the calculated EC₅₀ was $0.2\mu g/mL$ (95% confidence interval – 95%CI=0.17-0.25) (Figure 3C).

Interestingly, the magnitude of contractions elicited by PE after washing out the extract, especially at the higher concentrations (1 to $1.5\mu g/mL$), did not return to that observed before adding the extract. For example, the magnitude of PE-induced contractions 1 hour after washing out the ECL ($1.5\mu g/mL$) was $0.62\pm0.11g$ (n=4), which is significantly lower than $2.22\pm0.15g$ (n=4) observed before adding ECL. Furthermore, the magnitude of PE-induce contractions remained reduced ($1.17\pm0.14g$; n=4) even 4 hours after washing out ECL, whereas the addition of B_{12a} increased the magnitude of PE-induced contraction (which was decreased 1 hour after ECL washout) from $0.30\pm0.04g$ (n=4) to $3.70\pm0.21g$ (n=4) (Figure 3D).

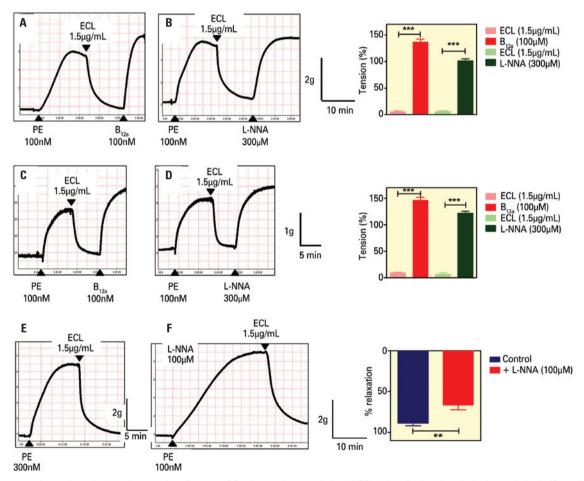


Figure 2. Representative tracings showing the reversion of extract of Combretum leprosum-induced (ECL-induced) relaxations by hydroxocobalamin ($B_{12a'}$: 100 μ M,) or L-N⁶-nitro-L-arginine (L-NNA; 300 μ M) on rings of rabbit thoracic aorta (A, B) or rat thoracic aorta (C, D). Representative tracings showing ECL-induced relaxations in the absence (E; n=8) and in the presence of L-NNA (F; 100 μ M, n=7) in rabbit abdominal aortic rings. Right panels (A-D) show the mean \pm standard error of the mean (SEM) values of tension of 4-8 similar experiments (tension expressed as a percentage of phenylephrine - PE-induced contraction). **Two-tailed p=0.0041 (unpaired *t*-test); ***two-tailed p<0.0001 (paired *t*-test). Right panel (E-F) shows the mean \pm SEM values of the magnitude of ECL-induced relaxation (relaxation magnitude, expressed as a percentage of reduction of PE-induced contraction)

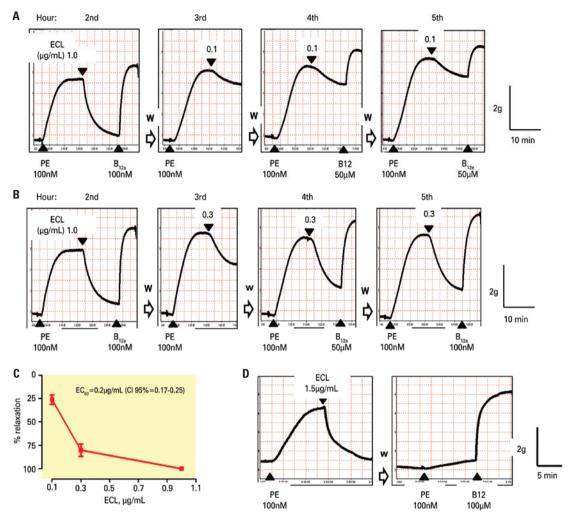


Figure 3. Representative tracings showing the effect of low concentrations of extract of *Combretum* leprosum (ECL) on rings from rabbit thoracic aorta pre-constricted with phenylephrine (PE). The small concentrations of ECL were added in a non-cumulative manner after "priming" of preparations with ECL $(1.0\mu g/mL)$ and reversion of ECL-induced relaxations by hydroxocobalamin (B_{12a}, 100 μ M). It can be observed that the magnitude of ECL $(0.1-0.3\mu g/mL)$ -induced relaxations increased in the first additions and became stabilized only after the 4th or 5th hour (A and B). Panel C shows the concentration-effect curve (mean \pm standard error of the mean - SEM values) observed after the 4 hours (**two-tailed p<0.0067, paired t-test). Panel D shows that the magnitude of PE-induced contraction was reduced even removing ECL $(1.5\mu g/mL)$ from the chamber and the recovery of contraction by B_{12a} addition. W= washout of the preparation for 60-45 minutes; EC₅₀: half-maximal relaxation; 95%CI: 95% confidence interval

Considering that endothelial cells require the presence of Ca^{2+} in the extracellular medium in order to produce NO and EDHF (endothelium-derived hyperpolarization factor), we determined whether the effect of ECL was affected by absence of extracellular Ca^{2+} . As shown in figures 4A and 4B, when rabbit thoracic aorta, rat thoracic aorta, and pig right coronary artery rings were incubated in Krebs solution without Ca^{2+} (- Ca^{2+}), the magnitude of ECL-induced relaxations was significantly reduced. In order to identify pharmacologically the putative Ca^{2+} influx pathways activated by ECL, the effect of RR was next examined. In rabbit thoracic aorta rings, the addition of RR ($10\mu M$) on the

plateau of ECL-induced relaxations reversed almost completely such relaxations (Figures 4C and 4D). Similar results were observed in rings from rat and mice thoracic aorta.

Because tree barks are rich in polyphenol compounds, the amount of total polyphenols present in the ECL was estimated using the FC method using GA as standard of polyphenol-like reactive compounds. From the linear regression equation relating absorbance and gallic acid concentration (y=0.0578 x -0.0348), it was calculated that a solution of 12.5mg/mL of ECL contained 4.94mg/mL (approximatively 39.5%) of gallic acid equivalents.

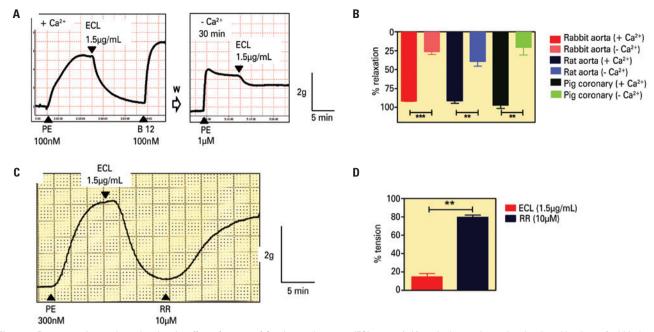


Figure 4. Representative tracings showing the effect of extract of *Combretum* leprosum (ECL; $1.5\mu g/mL$) on the isometric tension developed by rings of rabbit thoracic aorta pre-constricted with phenylephrine (PE), in A, in the presence (+ Ca²+) or absence (- Ca²+) of extracellular Ca²+. In B, the mean \pm standard error of the mean - SEM values of the magnitude of relaxation (relaxation magnitude expressed as a percentage reduction of PE- or U46619-induced contraction) of four similar experiments in rings from thoracic rabbit aorta, thoracic rat aorta, and right coronary pig artery. In C, the reversion of ECL-induced relaxations by ruthenium red (RR, $10\mu M$) and D shows the mean \pm SEM values of tension of three similar experiments (tension expressed as a percentage PE-induced contraction). **Two-tailed p<0.001 or ****p<0.0001 (paired *t*-test). W: washout of the preparation for 60 minutes; -Ca²+: washing and incubation with Krebs solution for 30 minutes

DISCUSSION

The present study showed for the first time that ECL produced relaxations of isolated arterial rings from different animal species, which indicated that the bark of *C. leprosum* contains bioactive compounds capable of influencing the function of blood vessels. Extracts or isolated compounds of *C. leprosum* have been previously shown to exhibit antinociceptive,⁽⁸⁻¹¹⁾ anticholinesterase,⁽¹²⁾ antiulcerogenic,⁽¹³⁾ antileishmanial,⁽¹⁴⁾ anti-inflammatory, and antiproliferative effects;^(15,16) however, to our knowledge, the vascular effects described in the present work have not been mentioned previously.

The fact that the effect was observed at relative low concentrations (0.1 to $1.0\mu g/mL$), and, considering that it is crude extract that may contain hundreds of different compounds, indicates that the bioactive compound responsible for the effect is either the most abundant, or if present in small amounts, is extremely potent; alternatively the effect may be caused by the synergistic action of different compounds. Such synergism has been described for polyphenols present in red wine extract (RWP), for instance, the EC₅₀ of raw RWP to induce EDR in rat aorta is 0.53 to $0.63\mu g/mL$, while the EC₅₀ of pure delphinidin or leucocyanidol, polyphenol compounds isolated from RWP, are $8.91\mu g/mL$ and

1.77 μ g/mL, respectively.^(17,18) Substances identified in other *Combretum* genera members are also capable of inducing EDR in rat aortas; however the concentrations required are higher than those of ECL; for example, 5 to 80μ g/mL (to mollic acid glucoside isolated from *C. molle* leaf) and EC₅₀ of 3.9 μ g/mL and 9.5 μ g/mL for methanolic extract of *C. celastroides* leaves, and *C. racemosum* roots respectively.^(3,4) As in the rat aorta, 1.5 μ g/mL of ECL causes complete relaxation of PE-induced contractions the vasodilator in the extract of compounds present in ECL are unlikely to be related to those already described in other members of the genus *Combretum*.

The endothelial cells might constitute the main site of action of the bioactive compounds present in the ECL, this suggestion is based on the observation that it requires an intact endothelium to cause relaxation in all the arteries examined. Alternatively, the ECL could be acting on the vascular smooth cells to enhance their responsiveness to relaxing factors produced basally by the endothelium like as the phosphodiesterase inhibitors but the relaxations caused by these inhibitors is still present in endothelium denuded rings from rat or rabbit thoracic aorta. (19,20) In fact, such mechanism has been recently proposed for the relaxant effect of a dichloromethane fraction from *Anogeissus leiocarpus*, a combretaceae. (21)

Considering that ECL-relaxed thoracic aorta rings from both rabbits and rats in which BK has no effect(6) and that histamine relaxes rat thoracic aorta but not rabbit thoracic aorta,(22) together with the fact that it relaxes rings from pig coronary arteries in which ACh causes contraction would suggest that the mechanisms involved in its action are different from those involved in the action of classical receptor agonists which cause EDR. In addition, the kinetic characteristics of ECL-induced relaxation such as slower initiation and longer time for attaining the steady state clearly distinguish its effects from those caused by ACh and BK and histamine. Furthermore, the fact that the effect remained for at least 4 hours after removing the extract contrasts with the transient relaxation caused by ACh or BK whose effects are not already evident 0.5 to 1 hour after removing them from the organ bath.

As in some arteries (rabbit superior mesenteric and common carotid arteries, as well as from pig coronary artery and from guinea-pig thoracic aorta) ECL is able to cause EDR in the presence of NOS inhibitors, we suggest that its effect is apparently related to the activation of endothelial cells to produce whatever relaxing factor is being predominantly produced. It has been thoroughly described that endothelial cells are heterogeneous in terms of responsiveness to agonists and relaxing factors being produced; for instance, endothelial cells from small arteries and arterioles produce mainly EDHF, whereas endothelial cells from the aorta produce almost exclusively NO.⁽²³⁾

Our results showing that a major part of ECL-induced EDR requires the presence of extracellular Ca²⁺ indicate that ECL action is related to the activation of Ca²⁺ influx in endothelial cells. Although the pathways mediating Ca²⁺ influx in endothelial cells have not been yet entirely elucidated, the fact that RR was able to revert ECL-induced relaxations would suggest that one or more of the various RR-sensitive Ca²⁺ permeable channels⁽²⁴⁾ is activated directly or indirectly by bioactive compounds in the extract.

There are good reasons to suggestion that polyphenol compounds could constitute the active principles accounting for the ECL effect. First, substances with polyphenol-like chemical reactivity are present in ECL in substantial amounts (approximately 40% w/w). Second, plant-derived polyphenols have been shown to cause EDR in several arterial segments. Third, previous studies have shown the presence of a variety of compounds, including polyphenols, in the ethanol extracts of flowers, leaves, roots, fruits, and bark from *C. leprosum*, such as flavonoids, 12,26,27) triterpenes, 26-28) mono- and oligosaccharides, fatty acids, sterols, 12,27,28) and stilbenes.

Finally the description of long-lasting vasorelaxing effect of ECL might be clinically relevant since it could lead to the development of potential new therapeutic approaches for pathological conditions in which blood flow is reduced or impaired due to increased vascular tonus such as hypertension and vasoespastic disorders. More experiments are necessary to establish the therapeutic potential of ECL, for instance we need to answer the questions: What are effects of ECL upon blood circulation in intact animals? What are the active compounds present in the extract? What are the mechanisms by which the ECL promotes synthesis and release of endothelial factors?

CONCLUSION

The present findings showed for the first time that an aqueous suspension of a lyophilized hydroalcoholic extract of Combretum leprosum causes relaxation of arterial rings isolated from different animal species. These Combretum leprosum extract-induced relaxations were completely dependent on a functional endothelium and appear to be mediated by the release of endothelial relaxing factors such as nitric oxide in the rat or rabbit thoracic aorta or by nitric oxide and others factors nonderived from nitric oxide synthases or cyclooxygenase activity in rabbit abdominal aorta and pig coronary arteries. Finally, since Combretum leprosum extract requires Ca2+ influx through ruthenium red-sensitive pathways to cause relaxation the elucidation of its mechanism of action, this could reveal a new approach to activate endothelial cells to produce and release relaxing factors.

ACKNOWLEDGMENTS

Francisco das Chagas Alves Filho was a recipient of PhD fellowships from *Fundação de Amparo à Pesquisa do Estado de São Paulo* (FAPESP, 2011/ 02866-0) and from *Conselho Nacional de Desenvolvimento Científico e Tecnológico* (CNPq, 870307/ 1997-5).

REFERENCES

- Agra MF, Nurit-Silva K, Basílio IJ, Freitas PF, Barbosa-Filho JM. Survey of medicinal plants used in the region Northeast of Brazil. Rev Bras Farmacogn. 2008;18(3):472-508. Review.
- Lorenzi H, Matos FJ. Plantas medicinais do Brasil: nativas e exóticas. 2a ed. Nova Odessa, SP: Plantarum; 2008.
- Ojewole JA. Cardiovascular effects of mollic acid glucoside, a 1alphahydroxycycloartenoid saponin extractive from Combretum molle R Br ex G Don (Combretaceae) leaf. Cardiovasc J Afr. 2008;19(3):128-33.
- Nsaudi Manga F, El Khattabi C, Fontaine J, Berkenboom G, Duez P, Lami Nzunzu J, et al. Vascular effects and antioxidant activity of two Combretum species from Democratic Republic of Congo. J Ethnopharmacol. 2012;142(1):194-200.

- Furchgott RF. Role of endothelium in responses of vascular smooth muscle. Circ Res. 1983;53(5):557-73. Review.
- Cherry PD, Furchgott RF, Zawadzki JV, Jothianandian D. Role of endothelial cells in relaxation of isolated arteries by bradykinin. Proc Natl Acad Sci USA. 1982;79(6):2106-10.
- Singleton VL, Rossi Jr JA. Colorimetry of total phenolics with phosphomolybdicphosphotungstic acid reagents. Am J Enol Vitic. 1965;16(3):144-58.
- Lira SR, Almeida RN, Almeida FR, Oliveira FS, Duarte JC. Preliminary Studies on the Analgesic Properties of the Ethanol Extract of Combretum leprosum. Pharm Biol. 2002;40(3):213-5.
- Pietrovski EF, Rosa KA, Facundo VA, Rios K, Marques MC, Santos AR. Antinociceptive properties of the ethanolic extract and of the triterpene 3beta, 6beta,16beta-trihidroxilup-20(29)-ene obtained from the flowers of Combretum leprosum in mice. Pharmacol Biochem Behav. 2006;83(1):90-9.
- Lopes LS, Marques RB, Pereira SS, Ayres MC, Chaves MH, Cavalheiro AJ, et al. Antinociceptive effect on mice of the hydroalcoholic fraction and (-) epicatechin obtained from Combretum leprosum Mart & Eich. Braz J Med Biol Res. 2010;43(12):1184-92.
- Lopes Lda S, Marques RB, Pereira Sda S, Ayres MC, Chaves MH, Almeida FR. Mechanisms of the antinociceptive action of (—) epicatechin obtained from the hydroalcoholic fraction of Combretum leprosum Mart & Eic in rodents. J Biomed Sci. 2012;19(1):1-6.
- Facundo VA, Rios KA, Medeiros CM, Militão JS, Miranda AL, Epifanio RA, et al. Arjunolic acid in the ethanolic extract of Combretum leprosum root and its use as a potential multi-functional phytomedicine and drug for neurodegenerative disorders: anti-inflammatory and anticholinesterasic activities. J Braz Chem Soc. 2005;16(6b):1309-12.
- Nunes PH, Cavalcanti PM, Galvão SM, Martins MC. Antiulcerogenic activity of Combretum leprosum. Pharmazie. 2009;64(1):58-62.
- Teles CB, Moreira LS, Silva AA, Facundo VA, Zuliani JP, Stábelia RG, et al. Activity of the Lupane isolated from Combretum leprosum against Leishmania amazonensis Promastigotes. J Braz Chem Soc. 2011;22(5):936-42.
- Longhi-Balbinot DT, Lanznaster D, Baggio CH, Silva MD, Cabrera CH, Facundo VA, et al. Anti-inflammatory effect of triterpene 3b, 6b, 16b trihydroxylup-20 (29) -ene obtained from Combretum leprosum Mart & Eich in mice. J Ethnopharmacol. 2012;142(1):59-64.
- Horinouchi CD, Mendes DA, Soley Bda S, Pietrovski EF, Facundo VA, Santos AR, et al. Combretum leprosum Mart. (Combretaceae): potential as an antiproliferative and anti-inflammatory agent. J Ethnopharmacol. 2013;145(1):311-9.
- 17. Andriambeloson E, Kleschyov AL, Muller B, Beretz A, Stoclet JC, Andriantsitohaina

- R. Nitric oxide production and endothelium-dependent vasorelaxation induced by wine polyphenols in rat aorta. Br J Pharmacol. 1997;120(6):1053-8.
- Andriambeloson E, Magnier C, Haan-Archipoff G, Lobstein A, Anton R, Beretz A, et al. Natural dietary polyphenolic compounds cause endothelium-dependent vasorelaxation in rat thoracic aorta. J Nutr. 1998;128(12):2324-33.
- Martin W, Furchgott RF, Villani GM, Jothianandan D. Depression of contractile responses in rat aorta by spontaneously released endothelium-derived relaxing factor. J Pharmacol Exp Ther. 1986;237(2):529-38.
- Martin W, Furchgott RF, Villani GM, Jothianandan D. Phosphodiesterase inhibitors induce endothelium-dependent relaxation of rat and rabbit aorta by potentiating the effects of spontaneously released endothelium-derived relaxing factor. J Pharmacol Exp Ther. 1986;237(2):539-47.
- Belemnaba L, Ouédraogo S, Auger C, Chataigneau T, Traore A, Guissou IP, et al. Endothelium-independent and endothelium-dependent vasorelaxation by a dichloromethane fraction from Anogeissus Leiocarpus (DC) Guill. Et Perr. (Combretaceae): possible involvement of cyclic nucleotide phosphodiesterase inhibition. Afr J Tradit Complement Altern Med. 2012;10(2):173-9.
- 22. Van de Voorde J, Leusen I. Role of the endothelium in the vasodilator response of rat thoracic aorta to histamine. Eur J Pharmacol. 1983;87(1):113-20.
- Garland CJ, Plane F, Kemp BK, Cocks TM. Endothelium-dependent hyperpolarization: a role in the control of vascular tone. Trends Pharmacol Sci. 1995;16(1):23-30. Review. Erratum in: Trends Pharmacol Sci. 1995;16(5):177.
- Vriens J, Appendino G, Nilius B. Pharmacology of vanilloid transient receptor potential cation channels. Mol Pharmacol. 2009;75(6):1262-79. Review.
- Shini-Kerth VB, Auger C, Etienne-Selloum N, Chataigneau T. Polyphenol-induced endothelium-dependent relaxations role of NO and EDHF. Adv Pharmacol. 2010;60:133-75. Review.
- 26. Facundo VA, Andrade CH, Silveira ER, Braz-Filho R, Hufford CD. Triterpenes and flavonoids from Combretum leprosum. Phytochemistry. 1993;32(2):411-5.
- 27. Ayres MC, Almeida BC, Chaves MH. Constituintes químicos e atividade antioxidante da espécie Combretum leprosum Mart. et Eicher. In: 32ª Reunião Anual da Sociedade Brasileira de Química [Internet]. Fortaleza (CE): Sociedade Brasileira de Química. Fortaleza, CE; 2009. [citado 2015 Jun 23]. Disponível em: http://sec.sbq.org.br/cdrom/32ra/resumos/T1190-1.pdf
- Facundo VA, Rios KA, Moreira LS, Sancho L J, Militão JS, Stabelli RG, et al. Two new cycloartanes from Combretum leprosum MART. (combretacea). Rev Latinoam Quím. 2008;36(3):76-82.
- Queiroz SC, Assalin MR, Nobre S, Melo IS, Moraes RM, Ferracini VL, et al. Determination of Combretastatin A-4 in Combretum leprosum. Planta Med. 2010;76(10):53.