

PHYTOPLANKTON DYNAMICS IN A HIGHLY EUTROPHIC ESTUARY IN TROPICAL BRAZIL

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ABSTRACT

The port of Recife in northeastern Brazil is an important ecological and social area but little is known about its environmental quality. Observations, sampling and measurements of phytoplankton were performed during spring and neap tides in the dry (January-February, 2005) and rainy (June, 2005) seasons to assess the environmental quality of the port of Recife. The area had chlorophyll-*a* concentrations ranging from 3.30 to 54.40 mg m⁻³, the highest values occurring at low tide during the dry season. A total of 129 species were identified: 53 of them in the dry season and 97 in the rainy season. Diatoms were the most diverse group and comprised 75.47% of the phytoplankton collected in the dry season and 60.82% of those collected in the rainy season. The dry season was characterized by *Coscinodiscus* sp. and *Helicotheca tamesis*; the rainy season by *Oscillatoria* sp. and *Coscinodiscus centralis*. Species diversity indices varied from 1.06 to 3.74 bits cel⁻¹. Low indices were related to the dominance of *Helicotheca tamesis*, *Coscinodiscus centralis*, *Coscinodiscus* sp. and *Aulacoseira granulata*. Seasonal forcing, rather than the estuarine flux, determined the phytoplankton community structure. The area is exposed to seasonally varying negative impacts due to metropolitan degradation and the high level of eutrophication.

RESUMO

Para avaliar a qualidade ambiental Porto do Recife (Nordeste do Brasil), uma área ecológica e socialmente importante mas pouco investigada, observações, medições e amostragem do fitoplâncton foram realizadas durante as marés de sizígia e quadratura no período de estiagem (janeiro-fevereiro / 2005) e chuvoso (junho/2005). A área apresentou concentração de clorofila *a* variando de 3,30 a 54,40 mg m⁻³, com maiores valores registrados durante a maré baixa no período de estiagem. Um total de 129 espécies foi identificada com 53 espécies durante o período de estiagem e 97 espécies no chuvoso. As diatomáceas formaram o grupo mais diverso, com 75,47% no período de estiagem e 60,82% no chuvoso. *Coscinodiscus* sp. e *Helicotheca tamesis* caracterizaram o período de estiagem e *Oscillatoria* sp. e *Coscinodiscus centralis* o período chuvoso. A diversidade de espécie variou de 1,06 a 3,74 bits cel⁻¹. Os baixos índices foram relacionados com a dominância de *Helicotheca tamesis*, *Coscinodiscus centralis*, *Coscinodiscus* sp. e *Aulacoseira granulata*. A sazonalidade determinou a estrutura da comunidade fitoplanctônica, em vez do fluxo estuarino. A área é exposta sazonalmente a vários impactos negativos, devido à degradação metropolitana e ao elevado nível de eutrofização.

Descriptors: Phytoplankton, Port, estuary, Diurnal variation, Tropical.
Descritores: Fitoplâncton, Porto, Estuário, Variação diurna, Tropical.

INTRODUCTION

The Pina Basin Estuaray is located in the metropolitan area of Recife in Pernambuco State, northeastern Brazil. As with many other tropical coastal areas, it is threatened by the diverse human uses of this body of water and the land areas that drain

into it. Consequently, the port basin presents a multitude of environmental problems. The most common problems are degraded natural habitats, declining plant and animal populations, diminishing fish and shellfish harvests and impaired water quality.

This coastal area is of great ecological, economic and social interest. It is also a highly variable system, where changes in the water

circulation patterns and fluctuations of input influences (e.g., Capibaribe River, Beberibe River, Tejipió River, sewage flow) induce high temporal variability on scales ranging from hours to seasons. This variability may be reflected in population dynamics, especially those of phytoplankton populations thriving in coastal systems.

Phytoplankton species can be used as a diagnostic tool in determining ecosystem quality (MAGUIRE, 1973; RESH, UNZICKER, 1975; DAY JR. et al., 1989). Previous research has identified biotic and abiotic factors that regulate primary productivity and has developed models that describe phytoplankton growth dynamics under specific environmental conditions (BAIRD et al., 2001).

Phytoplankton communities are highly complex multispecies communities in terms of their diversity and dynamics. Succession shifts in phytoplankton community structure are primarily due to changes in environmental variables (e.g., degree or type of nutrient limitation) and/or shifts in higher trophic levels (e.g., microzooplankton to mesozooplankton) (MILLER et al., 1995; GILBERT, 1998; LEWITUS et al., 1998; RIEGMAN; NOORDELOOS, 1998; RABOUILLE et al., 2001; FERNANDES; BRANDINI, 2004).

The influence of environmental variables and preferential grazing by herbivores on phytoplankton community composition is not well understood, particularly in harbor areas under extremely impacted influx. Identifying the ecological variables that regulate phytoplankton community structure is essential for facilitating the elaboration of broad hypotheses of pervasive environmental issues, such as eutrophication and harmful algal blooms (SMAYDA, 1997; RIEGMAN; FLAMELING; NOORDELOOS 1998; MAFRA JUNIOR et al., 2006). Since one of the most important factors affecting variability in the port of Recife and its basin is the tide, accurate estimates of the plankton requires sampling over a 24 h period (McLUSKY; ELLIOTT, 2004).

One question addressed in this study is whether the abiotic phenomena of the port basin of Recife (e.g., circulation patterns, tides) and water condition are related to identifiable phytoplankton assemblages or abundance patterns.

Despite the ecological and social importance of the port of Recife, few investigations of phytoplankton composition have been conducted in the harbor or in nearby areas (FEITOSA; PASSAVANTE, 1990; 1991/1993; FEITOSA et al., 1999; NASCIMENTO et al., 2003).

Many studies have been dedicated to the temporal variability of phytoplankton densities in marine systems (SOROKIN, 1995), but few data are

available on the dynamics of tropical harbor phytoplankton communities.

The main objective of the present study was to study temporal changes of phytoplankton and hydrographic parameters in the highly polluted port basin of Recife in northeastern Brazil.

STUDY AREA

The Port Basin Estuary is located in Recife, the capital of Pernambuco State, Brazil. Recife is the main urban center of northeastern Brazil and lies partly on the mainland and partly on the island. Dissected by waterways, it is often called the "Brazilian Venice" (Fig. 1).

This area has a hot and humid tropical climate and is categorized as Group A on the Köppen Scale. Annual average air and water temperatures are around 25°C, the annual minimum being 24°C and the maximum 31°C. Relative humidity varies from 80% to 90% and annual rainfall ranges from 1760 mm to 2270 mm, with 80% of it occurring between April and July (the rainy season). The dominant wind is from the southeast (ARAGÃO, 2004).

The maximum tidal height (during extreme spring tides) at the port is around 3 m, with an annual high tide average of 2.6 m during spring tides and 1.6 m during neap tides. There are two natural access channels to the port. The main access channel, South Channel, is approximately 260 m wide and 3.4 km long with a depth of 10.5 m. The North Channel, only used by smaller vessels, is narrower, being approximately 1.0 km long and 6.5 m deep. The basin constitutes a shallow, restricted environment that receives large quantities of untreated sewage and is, therefore, polluted, eutrophic and hypoxic.

MATERIAL AND METHODS

Sampling was carried out at one fixed station located at the confluence of the port of Recife, the Capibaribe River and the Pina Basin (08°04'01" S and 34°52'06" W), called the Port basin (Fig. 1). Sampling was conducted on two consecutive days during the dry season (January 25-26 and February 02-03, 2005) and the rainy season (June 07-08 and 14-15, 2005) at spring and neap tides during the diurnal high, low, flood and ebb tides.

The following hydrological data were collected concurrently with a Nansen bottle: pH (Hanna 8417 pHmeter); dissolved oxygen (Winkler method; GRASSHOFF et al., 1983); dissolved oxygen saturation (UNESCO, 1973); nutrients, including nitrites, nitrates, phosphates and silicate (STRICKLAND; PARSONS, 1972; GRASSHOFF et

al., 1983) and suspended particulate material (BAUMGARTEN et al., 1996). The following parameters were measured in situ : water temperature (Hanna digital thermometer), salinity (refractometer) and water transparency (Secchi disc). Chlorophyll-*a* was collected with a 1 liter Van Dorn bottle at the surface and measured using a Micronal B280 spectrophotometer (PARSONS; STRICKLAND, 1963; WETZEL; LINKENS, 1991).

Phytoplankton sampling was conducted with a plankton net (mesh size 64 μm) fitted with a flowmeter. Three-minute horizontal surface hauls were carried out. After collection, samples were preserved in a 4% buffered formaldehyde/seawater solution (NEWELL; NEWELL, 1963).

In the laboratory, 300 mL samples were homogenized and a 1 mL subsample was analyzed under a compound microscope. Each cell, colony or filamentous alga was counted as an individual and expressed as relative abundance (%). Identification was based on the phytoplankton literature (PÉRAGALLO; PÉRAGALLO, 1897-1908;

HUSTEDT, 1930-1966; CUPP, 1943; DESIKACHARY, 1959; SOURNIA, 1967; 1986; PRESCOTT, 1975; PARRA et al., 1982; TOMAS, 1996; KOMÁREK; ANAGNOSTIDIS, 2000).

The Shannon index was used to estimate the community diversity (SHANNON, 1948).

Cluster analysis of the sample-species data matrix was also performed, using the Bray & Curtis method. The Weighted Pair Group Method, using arithmetic averages, was the link method used for the dendrograms (WPGMA) (LEGENDRE; LEGENDRE, 1998). A cophenetic value matrix was applied to test the adequacy of fit of the cluster analysis (ROHLF; FISHER, 1968). A Principal Component Analysis was computed based on a matrix formed by richness, diversity, chlorophyll-*a* and hydrological data. Data analyses were carried out using the Numerical Taxonomy and Multivariate Analyses System (NTSYS ver. 1.30, Metagraphics Software Corporation, California, USA, 1987).



Fig. 1. Study area and sampling station (ST) in the Port Basin of Recife (Pernambuco, Brazil).

A non-parametric Kruskal-Wallis test was applied (BioEstat 3.0) to test significant differences ($p < 0.05$) between dry and rainy seasons and between tides, based on the physicochemical and biological parameters.

RESULTS

Physical Environment

No significant difference ($p > 0.5$) in the abiotic parameters (water temperature, salinity, transparency, particulate organic material, pH, dissolved oxygen, dissolved oxygen saturation tax,

nitrite+nitrate, phosphate and silicate) was found between spring and neap tides. Water temperature varied between 27°C and 32.6°C, the higher values being recorded in the late morning and early afternoon when the sunlight was more intense. Salinity varied between 21 and 36 during the dry season and ranged from 1 to 8 during the rainy season (Figs 2A, B; Figs 3A, B). Water transparency varied from 0.1 m to 1.0 m and higher values were recorded during the flood tide of the spring and neap tides in both seasons (Figs 2C, D; Figs 3C, D). The pH was > 7.0 the larger part of the time. In general, temperature, salinity, transparency and pH were significantly higher during the dry season ($p < 0.05$).

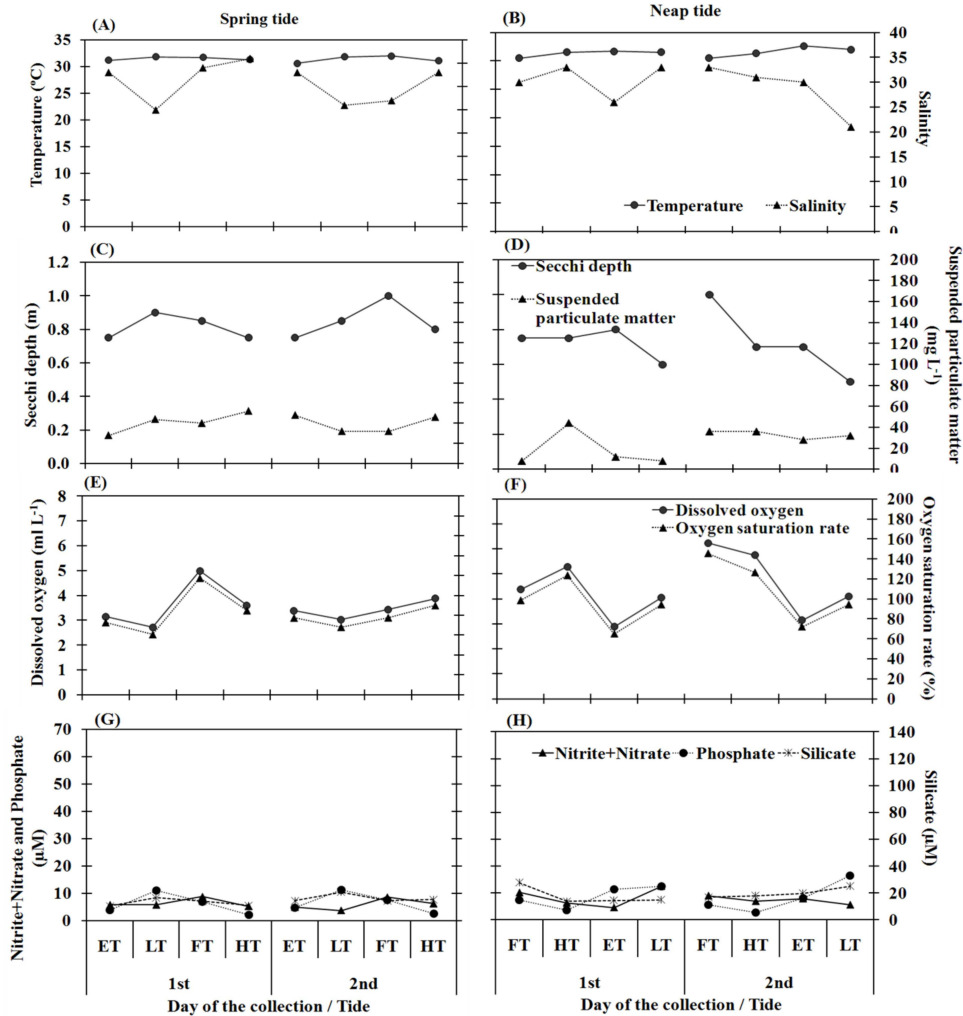


Fig. 2. Temporal variation of abiotic variables in the Port Basin of Recife (Pernambuco, Brazil) during the dry season (January 25-26 and February 02-03, 2005). Note: ST = spring tide, NT = neap tide, ET = ebb tide, LT = low tide, FT = flood tide and HT = high tide.

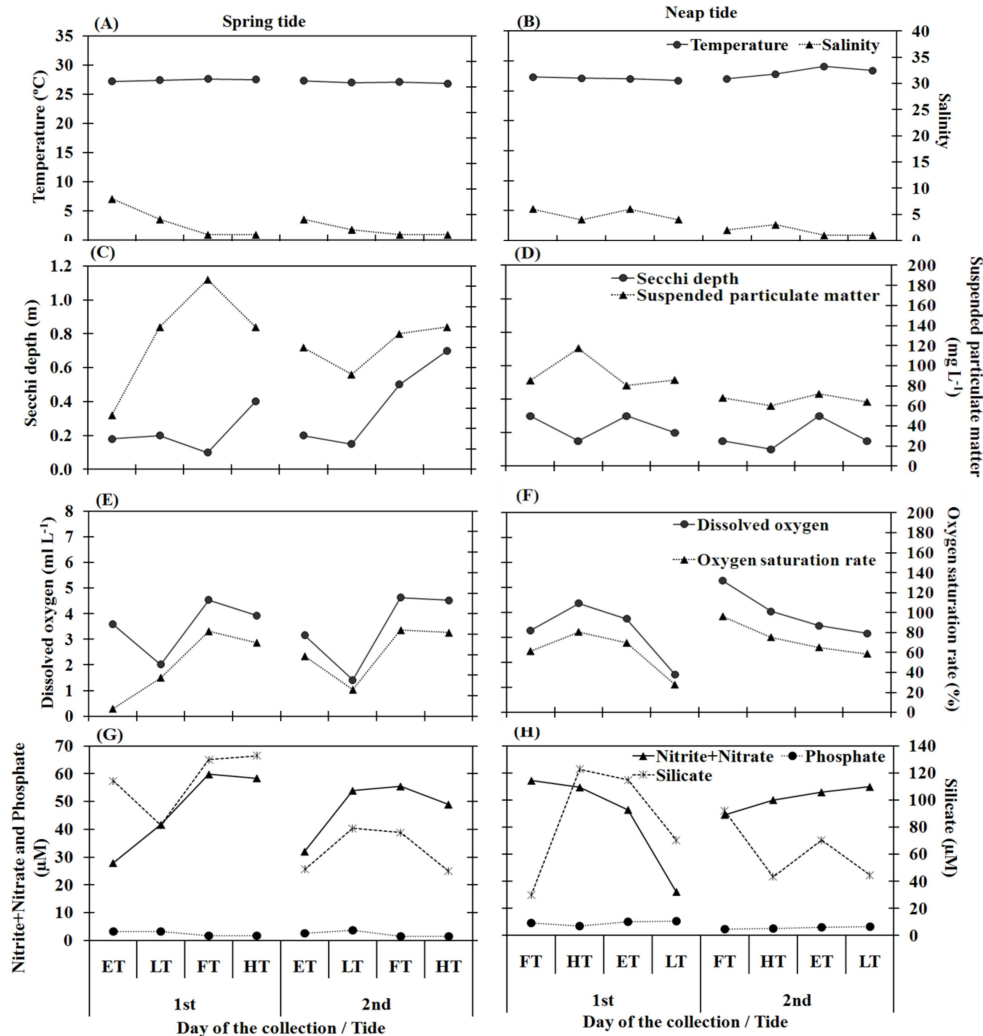


Fig. 3. Temporal variation of abiotic variables in the Port Basin of Recife Estuary (Pernambuco, Brazil) during the rainy season (June 07-08 and 14-15, 2005). Note: ST = spring tide, NT = neap tide, ET = ebb tide, LT = low tide, FT = flood tide and HT = high tide.

The suspended particulate material (SPM) varied from 8.0 mg L^{-1} to 187 mg L^{-1} , with higher values during flood and high tides in the rainy season (Figs 2C, D; Fig. 3C, D).

No significant difference ($p > 0.05$) was found between ebb tide (ET), low tide (LT), flood tide (FT) and high tide (HT) in terms of temperature, salinity, pH, transparency and suspended particulate material. However, significant differences were found between dry and rainy seasons ($p = 0.000$).

The dissolved oxygen was lower during low tide and higher during the spring and neap flood tides (Figs 2E, F; Fig. 3E, F). The minimum value was 2.72 ml L^{-1} and the maximum was 6.24 ml L^{-1} , with

significant differences between ET and LT ($p = 0.006$), ET and HT ($p = 0.010$), LT and FT ($p = 0.001$) and LT and HT ($p = 0.002$). However, no significant differences were observed between tidal stages (ET, FT, LT, and HT) or between dry and rainy seasons ($p > 0.05$). The dissolved oxygen saturation rate ranged from 6.93% to 145.45%, with lower values in the rainy season during low tide and higher values in the dry season during flood and high tide (Figs 2E, F; Figs 3E, F). Significant differences occurred between ET and FT ($p = 0.006$), ET and HT ($p = 0.013$), LT and FT ($p = 0.007$) and LT and HT ($p = 0.014$) and between seasons ($p = 0.004$).

Nitrite+nitrate ranged between 3.80 μM and 59.75 μM . Silicate ranged between 10.70 μM and 133.89 μM , with highest concentrations during the rainy season (Figs 2G, H; Figs 3G, H). No significant differences were found among tidal stages (ET, LT, FT, HT). Significant differences occurred between dry and rainy seasons ($p=0.000$).

In general, silicate content presented a similar distribution to nitrite + nitrate, with highest values in the rainy season (Figs 2G, H; Fig. 3G, H). However, concentrations higher than 10.70 μM in any season indicate that the environment has a great abundance of this nutrient. Significant differences were recorded between seasons ($p<0.05$) but no differences were found among tidal stages.

The phosphate varied from 1.50 μM to 16.40 μM and was abundant during the dry season (Figs 2G, H; Fig. 3G, H). Significant differences were observed between ET and HT ($p=0.035$), LT and HT ($p=0.001$) and between dry and rainy seasons ($p=0.000$).

Phytoplankton

Chlorophyll-*a* concentrations varied between 3.30 mg m^{-3} and 54.40 mg m^{-3} , with higher values at the low neap tide during the dry season (Figs 4A, B, C, D). There was no significant difference among tidal stages or between seasons.

A total of 127 taxa were identified: 52 in the dry season and 95 in the rainy season. In both seasons, diatoms were the group that most contributed to species richness. There were 40 diatom taxa (76.92%) in the dry season and 58 (61.05%) in the rainy season. There were 7 chlorophyte taxa (13.46 %) in the dry

season and 21 (22.11%) in the rainy season. Cyanobacteria were present with 4 taxa (7.69%) in the dry season and 13 taxa (13.68%) in the rainy season. Dinoflagellates were present only in the dry season with one taxon (1.92%) and the euglenophytes were present only in the rainy season with three taxa (3.16%) (Table 1).

The taxonomic richness ranged from 5 to 42, with highest values during the rainy season (Figs 5A, B, C, D) and significant seasonal differences ($p=0.000$).

The phytoplankton community was dominated by *Helicotheca tamesis* (Shrubsole) Ricard (80.72%) during the dry season at the spring tide and by *Coscinodiscus* sp. (60.87%) at neap tide (Table 2). In the rainy season, *Coscinodiscus centralis* Ehrenberg (53.54%) dominated in the spring tide and *Oscillatoria* sp. (56.16%) in the neap tide (Table 3).

Most taxa (62.59%) occurred sporadically, followed by those of low frequency of occurrence (28.78%), of frequent occurrence (7.19%) and of very frequent occurrence (1.44%). The species *Oscillatoria* sp. (84.38%) and *Aulacoseira granulata* (Ehrenberg) Ralfs (71.88%) were the most frequent (Table 1).

The specific diversity index ranged from 1.06 bits cel^{-1} (high tide) to 3.74 bits cel^{-1} (ebb tide) during spring tide in the dry season (Fig. 5A, B, C, D). Low species diversity was caused by the dominance of *Helicotheca tamesis*, *Coscinodiscus centralis*, *Coscinodiscus kutzingii* Schmidt and *Aulacoseira granulata* (Table 2; Table 3). There was no significant difference among tidal stages or between seasons.

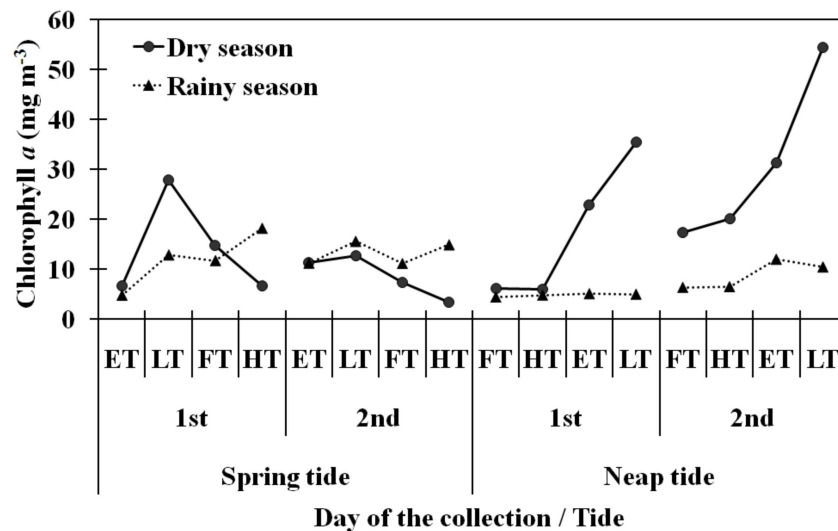


Fig. 4. Temporal variation of chlorophyll-*a* (mg m^{-3}) in the Port Basin of Recife Estuary (Pernambuco, Brazil) during the dry season (January 25-26 and February 02-03, 2005) and rainy season (June 07-08 and 14-15, 2005). Note: ST = spring tide, NT = neap tide, ET = ebb tide, LT = low tide, FT = flood tide and HT = high tide.

Table 1. Microphytoplankton species checklist for the Pina Basin Estuary, Recife, Brazil, during the dry season (January 25-26 and February 02-03, 2005) and rainy season (June 07-08 and 14-15, 2005). *^a Comments: Habitat and species typical or frequent in areas: plan = planktonic, tyco = tyco planktonic, fw = freshwater algal, est = estuarine, ner = neritic, oc = oceanic, vfre = very frequent species, fre = frequent species, lfre = low frequency species, espo = sporadic species. Note: ST= spring tide, NT = neap tide, ET = ebb tide, LT = low tide, FT = flood tide and HT = high tide.

TAXA	DROUGHT PERIOD				OBSERVATIONS* ^a
	DRY SEASON		RAINY SEASON		
	ST	NT	ST	NT	
CYANOPHYTA					
<i>Anabaena</i> sp.	ET		ET, LT, FT, HT	FT, HT, ET, LT	lfre
<i>Chroococcus dispersus</i> (Keissler) Lemmermann			HT	FT, HT, ET	plan, fw, vfre
<i>Chroococcus minor</i> (Kützing) Naegeli				LT	tyco, fw, spo
<i>Chroococcus</i> sp.				LT	spo
<i>Dactylococcosis acicularis</i> Lemmermann				ET	plan, fw, spo
<i>Geitlerinema</i> sp.			ET, LT, FT	LT, FT, HT	vfre
<i>Lyngbya versicolor</i> (Wartmann) Gomont			ET, LT, FT	FT, HT, ET, LT	tyco, fw, spo
<i>Lyngbya</i> sp.			FT	LT	fre
<i>Merismopedia tenuissima</i> Lemmermann				LT	tyco, fw, spo
<i>Oscillatoria princeps</i> Vaucher	ET, LT				tyco, fw, spo
<i>Oscillatoria tenuis</i> Agardh	ET		ET, LT, FT	FT, HT, ET, LT	tyco, fw, spo
<i>Oscillatoria</i> sp.	ET, LT, FT, HT	FT, HT, ET	ET, LT, FT	FT, HT, ET, LT	vfre
<i>Phormidium</i> sp.				ET	spo
<i>Planktolyngbya microspira</i> Kom. & Cronb.			ET, LT, FT	FT, HT, ET, LT	plan, fw, lfre
EUGLENOPHYTA					
<i>Euglena acus</i> Ehrenberg			ET	LT	plan, fw, spo
<i>Euglena spirogyra</i> Ehrenberg				LT	plan, fw, spo
<i>Euglena</i> sp.				LT	spo
DINOPHYTA					
<i>Ceratium</i> sp.	HT	FT, HT			spo
BACILLARIOPHYTA					
<i>Actinocyclus</i> sp.	ET, LT, EM				spo
<i>Amphiprora paludosa</i> Smith			ET		tyco, ner, spo
<i>Amphiprora</i> sp.			ET		spo
<i>Anomoeoneis serians</i> (Brébisson) Cleve			ET		tyco, fw, spo
<i>Asterionellopsis glacialis</i> (Castracane) Round		HT			tyco, ner, spo
<i>Aulacoseira granulata</i> (Ehrenberg) Ralfs	ET, LT, FT	HT, LT	ET, LT, FT, HT	FT, HT, ET, LT	plan, fw, vfre
<i>Auliscus</i> sp.	ET	FT, HT, ET, LT			lfre
<i>Biddulphia biddulphiana</i> Smith	ET	ET			tyco, ner, spo
<i>Biddulphia tridens</i> Ehrenberg				LT	tyco, ner, spo
<i>Biddulphia</i> sp.	ET, LT, FT, HT	FT, HT, ET, LT		LT	lfre
<i>Campylodiscus clypeus</i> Ehrenberg			LT, FT		tyco, ner, spo
<i>Campylodiscus</i> sp.			ET		spo
<i>Cerataulus turgidus</i> Ehrenberg	FT, HT			FT	tyco, ner, spo
<i>Cerataulus</i> sp.	FT	ET			spo
<i>Chaetoceros coarctatus</i> Lauder		HT			plan, oc, spo
<i>Chaetoceros</i> sp.	ET, LT, FT, HT	FT, HT, LT			lfre
<i>Climacospheia elongata</i> Bailey	ET				tyco, ner, spo
<i>Coscinodiscus centralis</i> Ehrenberg			ET, LT, FT, HT	ET, FT	plan, oc, lfre
<i>Coscinodiscus curvatulus</i> Grunow		LT			plan, oc, spo
<i>Coscinodiscus kutzingii</i> Schmidt			ET, LT, FT, HT	FT, HT, ET, LT	plan, oc, lfre
<i>Coscinodiscus</i> sp.	ET, LT, FT, HT	FT, HT, ET, LT	ET, FT	FT, HT, LT	fre
<i>Dimerogramma dubium</i> Grunow			ET, FT		tyco, ner, spo
<i>Diploneis smithii</i> (Brébisson) Cleve			FT		tyco, ner, spo
<i>Epithemia</i> sp.	LT	FT			spo
<i>Eunotia</i> sp.				ET	spo
<i>Fragilaria capucina</i> (Desmazière) Kützing			LT		tyco, fw, spo
<i>Fragilaria</i> sp.			ET		spo
<i>Grammatophora</i> sp.			ET	LT	spo
<i>Gyrosigma balticum</i> (Ehrenberg) Cleve	ET, LT	FT, HT, ET, LT	ET, LT	LT	plan, est, lfre
<i>Gyrosigma strigilis</i> (Smith) Griffith & Henfrey	ET				spo
<i>Helicotheca tamensis</i> (Shrubsole) Ricard	ET, LT, FT, HT	FT, HT, ET, LT			plan, ner, lfre
<i>Hyalodiscus</i> sp.			ET, LT, FT, HT	HT, ET, LT	lfre
<i>Licmophora flabellata</i> (Carmichael) Agardh	HT	LT			tyco, ner, spo
<i>Licmophora</i> sp.	LT, FT, HT	HT, ET, FT			lfre
<i>Lyrella lyra</i> (Ehrenberg) Karayeva	ET, LT, FT	LT			tyco, ner, lfre

Table 1. Continuation

TAXA	DROUGHT PERIOD				OBSERVATIONS* ^a
	DRY SEASON		RAINY SEASON		
	ST	NT	ST	NT	
CYANOPHYTA					
<i>Melosira</i> sp.			ET, LT	HT, LT	lfre
<i>Navicula gracilis</i> Ehrenberg			LT		tyco, ner, esp
<i>Navicula marina</i> Ralfs				LT	tyco, ner, spo
<i>Navicula</i> spp	HT	ET	ET, LT, FT, HT	FT, HT, ET, LT	spo
<i>Nitzschia angularis</i> Smith			ET, FT, HT	ET, LT	tyco, est, lfre
<i>Nitzschia constricta</i> (Kützing) Ralf				LT	tyco, ner, spo
<i>Nitzschia distans</i> Gregory			FT		plan, oc, spo
<i>Nitzschia fasciculata</i> Grunow			ET, LT, FT, HT		est, lfre
<i>Nitzschia insignis</i> Gregory			FT		tyco, est, spo
<i>Nitzschia linearis</i> (Agardh) Smith			ET		tyco, fw, spo
<i>Nitzschia longissima</i> (Brébisson) Grunow		LT		ET	tyco, fw, spo
<i>Nitzschia lorenziana</i> Grunow			ET, LT, FT		tyco, fw, spo
<i>Nitzschia máxima</i> Grunow			FT		est, spo
<i>Nitzschia sigma</i> (Kützing) Smith			ET, LT, FT, HT		tyco, fw, lfre
<i>Nitzschia sigmoidea</i> (Nitzsch) Smith			ET, LT, FT, HT	HT, LT	plan, oc, lfre
<i>Nitzschia thermalis</i> (Ehrenberg) Auerswald				ET	spo
<i>Nitzschia vermicularis</i> (Kützing) Hantsch			LT, FT, HT	ET, LT	ner, lfre
<i>Nitzschia vitrea</i> Norman			FT		fw, spo
<i>Nitzschia</i> sp.		FT, ET	ET, LT, FT, HT	FT, HT, ET, LT	spo
<i>Odontella aurita</i> (Lyngbye) Agardh	ET				tyco, ner, spo
<i>Odontella regia</i> (Shultz.) Hendeby	ET, FT				tyco, ner, spo
<i>Petrodictyon gemma</i> (Ehrenberg) Mann	ET, LT		ET, LT, FT, HT	LT	tyco, ner, lfre
<i>Pinnularia major</i> (Kützing) Ehrenberg			LT, FT		tyco, ner, spo
<i>Pinnularia tabellaria</i> Ehrenberg			LT		tyco, ner, spo
<i>Plagiogramma</i> sp.	ET, LT, FT, HT	HT			lfre
<i>Pleurosigma formosum</i> Smith	ET				tyco, est, spo
<i>Pleurosigma speciosum</i> Smith			ET, FT		tyco, ner, spo
<i>Pleurosigma</i> sp.	ET, LT, FT, HT	FT, HT, ET, LT	ET	HT, LT	fre
<i>Pleurosira laevis</i> (Ehrenberg) Campère	LT	LT	ET, LT, FT, HT	LT	tyco, ner, lfre
<i>Podocystis adriatica</i> Kützing				ET	tyco, ner, spo
<i>Psammodictyon panduriforme</i> (Gregory) Mann			ET, FT, HT		tyco, ner, spo
<i>Rhabdonema</i> sp.					spo
<i>Rhizosolenia styliiformis</i> Brightwell			HT	FT	plan, oc, spo
<i>Rhizosolenia</i> sp.	ET	LT			spo
<i>Sellaphora laevis</i> (Kützing) Grunow			HT		tyco, oc, spo
<i>Skeletonema costatum</i> (Greville) Cleve	LT, FT				plan, ner, spo
<i>Surirella spiralis</i> Kützing			FT		tyco, es, spo
<i>Surirella striatula</i> Jurpin			FT		tyco, es, spo
<i>Surirella</i> sp.	LT, FT, HT	FT, HT, ET, LT	FT, HT	FT, HT, ET, LT	spo
<i>Synedra gaillonii</i> (Bory) Ehrenberg	ET				tyco, est, spo
<i>Synedra ulna</i> (Nitzsch) Ehrenberg		LT			tyco, fw, spo
<i>Synedra</i> sp.	HT		FT		spo
<i>Tabellaria flocculosa</i> (Roth) Kützing	LT				tyco, fw, spo
<i>Terpsinoe americana</i> Bailey			LT		tyco, est, spo
<i>Terpsinoe musica</i> Ehrenberg			ET, LT	HT	tyco, est, spo
<i>Thalassiosira eccentrica</i> (Ehrenberg) Cleve	ET			LT	plan, oc, esp
<i>Thalassiosira</i> sp.	ET, LT, FT	FT, HT, ET, LT			fre
<i>Triceratium alternans</i> Bailey				LT	tyco, ner, spo
<i>Triceratium antedeluvianum</i> (Ehrenberg) Grunow			ET		tyco, ner, spo
<i>Triceratium pentacrinus</i> (Ehrenberg) Wallicia	ET	LT			tyco, ner, spo
CHLOROPHYTA					
<i>Actinastrum hantzschii</i> Lagerheim				FT, HT, ET	plan, fw, spo
<i>Ankistrodesmus falcatus</i> (Corda) Ralfs Wille			LT	FT, ET, LT	plan, fw, lfre
<i>Ankistrodesmus longissimus</i> Lemmerman			ET, LT, FT	FT, HT, ET, LT	plan, fw, lfre
<i>Closterium diana</i> Ehrenberg				ET	plan, fw, spo
<i>Closterium rostratum</i> Ehrenberg			ET	HT, LT	plan, fw, spo
<i>Closterium setaceum</i> Ehrenberg				FT	plan, fw, spo
<i>Closterium striolatum</i> Ehrenberg				FT	plan, fw, spo
<i>Closterium</i> sp.		ET		LT	spo
<i>Coelastrum microporum</i> Naegeli		LT			tyco, fw, spo
<i>Microspora</i> sp.	ET				spo
<i>Onychonema</i> sp.					spo

Table 2. Continuation.

	DROUGHT PERIOD																
	SPRING TIDE								NEAP TIDE								
	1 st				2 nd				1 st				2 nd				
	ET	LT	FT	HT	ET	LT	FT	HT	FT	HT	ET	LT	FT	HT	ET	LT	
<i>Tabellaria flocculosa</i> (Roth) Kützing	0.74																
<i>Thalassiosira eccentrica</i> (Ehr.) Cleve	5.08																
<i>Thalassiosira</i> sp.	37.04	21.31			4.83	23.59	21.57			38.89	21.74	26.98	2.50	9.38	17.75	2.17	5.00
<i>Triceratium pentacrinus</i> (Ehr.) Wal.	1.69												0.43				
CHLOROPHYTA																	
<i>Closterium</i> sp.													1.04				
<i>Coelastrum microporum</i> Naeg.													0.87				
<i>Microspora</i> sp.	1.69																
<i>Pediastrum duplex</i> Meyen					0.51												
<i>Scenedesmus</i> sp.													0.43				
<i>Staurastrum gracile</i> Ralfs	1.69	0.55															
<i>Staurastrum</i> sp.					3.45				1.30								

Table 3. Relative abundance (%) of microphytoplankton species in the Pina Basin Estuary, Recife, Brazil, during the rainy season (June 07-08 and 14-15, 2005). Note: ST = spring tide, NT = neap tide, ET = ebb tide, LT = low tide, FT = flood tide and HT = high tide.

	DROUGHT PERIOD																
	SPRING TIDE								NEAP TIDE								
	1 st				2 nd				1 st				2 nd				
	ET	LT	FT	HT	ET	LT	FT	HT	FT	HT	ET	LT	FT	HT	ET	LT	
CYANOPHYTA																	
<i>Anabaena</i> sp.	0.98	4.68		0.30		0.31	0.19		0.72		0.25	2.53		4.32			
<i>Chroococcus dispersus</i> (Keissler) Lemmermann									0.19		0.38		0.37	0.20		0.38	
<i>Chroococcus minor</i> (Kützing) Naegeli													0.48				
<i>Chroococcus</i> sp.													0.21				
Cyanophyceae	15.20	3.71		8.73	5.98	0.56		21.96	17.12	11.68	47.71	0.95	2.12	50.84	34.77		
<i>Dactylococcosis acicularis</i> Lemmermann	7.60												0.11				
<i>Geitlerinema</i> sp.	1.47	0.52		0.53						0.56	15.45	2.86			0.39	1.32	
<i>Lyngbya versicolor</i> (Wartmann) Gomont									0.14				0.11				
<i>Lyngbya</i> sp.	0.34	0.13		0.08	0.58	0.36			3.38	0.80	3.39	0.95	0.11	0.32	0.69	0.19	
<i>Merismopedia tenuissima</i> Lemmermann													0.88				
<i>Oscillatoria</i> sp.	5.74	21.77	2.86	9.86		27.48	4.84			21.96	56.16	25.88	30.92	49.47	27.60	10.56	13.72
Oscillatoriaceae	0.34								10.33		37.16		0.38		2.32		11.04
<i>Phormidium</i> sp.													0.34				
<i>Planktolingbya microspira</i> Komarek	1.37	1.44	0.55	1.51		0.23	1.00			0.40		1.51	1.72	3.11	0.99		3.95
EUGLENOPHYTA																	
<i>Euglena acus</i> Ehrenberg					0.08						0.24						
<i>Euglena spirogyra</i> Ehrenberg													0.08				
<i>Euglena</i> sp.													0.11				
BACILLARIOPHYTA																	
<i>Amphiprora paludosa</i> Smith	0.21				0.08												
<i>Amphiprora</i> sp.	0.05																
<i>Anomoeoneis serians</i> (Bréb.) Cl.	0.05																
<i>Aulacoseira granulata</i> (Ehrenberg) Ralfs	27.66	47.23		37.60	35.09	22.67	39.89	37.01	2.03	6.00	17.09	4.58	16.32	19.96	17.97	9.96	
Bacillariophyceae	6.52	0.62		2.64		1.28	0.21	1.31			3.12		1.34		0.21		2.44
<i>Biddulphia tridens</i> Ehrenberg													0.11				

Table 3. Continuation.

	DROUGHT PERIOD															
	SPRING TIDE								NEAP TIDE							
	1 st				2 nd				1 st				2 nd			
	ET	LT	FT	HT	ET	LT	FT	HT	FT	HT	ET	LT	FT	HT	ET	LT
<i>Surirella striatula</i> Jurpin	6.78				0.78											
<i>Surirella</i> sp.					4.27 0.19				0.25				0.42 0.74 0.19			
<i>Synedra</i> sp.					0.14											
<i>Terpsinoe americana</i> Bailey					0.04											
<i>Terpsinoe musica</i> Ehrenberg	0.25				0.12								0.19			
<i>Thalassiosira eccentrica</i> (Ehrenberg) Cleve													0.21			
<i>Triceratium alternans</i> Bailey									0.64				0.21			
<i>Triceratium antedeluvianum</i> (Ehrenberg) Grunow					0.08											
CHLOROPHYTA																
<i>Actinastrum hantzschii</i> Lag.													0.05 0.20 0.38			
<i>Ankistrodesmus falcatus</i> (Corda) Ralfs Wille					0.04				0.34				0.75 0.42 1.70 0.20			
<i>Ankistrodesmus longissimus</i> Lemm.	0.93 0.13				0.30 0.04				0.08 8.42 4.01 15.53				12.74 6.42 10.90			
Chlorophyceae	0.74 0.21 0.72				0.90 0.54 14.32 1.68				5.07 0.88 2.51 1.34				7.75 4.05 8.27			
<i>Closterium diana</i> Ehrenberg													0.11			
<i>Closterium rostratum</i> Ehr.					0.08				0.24				0.38			
<i>Closterium setaceum</i> Ehr.													0.13			
<i>Closterium striolatum</i> Ehr.													0.25			
<i>Closterium</i> sp.									0.34				0.21			
<i>Onychonema</i> sp.									0.68							
<i>Pediastrum biwae</i> Negoro	0.05 0.62				0.15 0.08				0.08 2.64				2.63 0.53 0.10 0.38			
<i>Pediastrum duplex</i> Meyen					0.10 0.08 0.04 0.07 0.19 0.68				2.76 0.38 1.32 2.23 0.49 1.13							
<i>Pediastrum simplex</i> Meyen									0.38				0.11			
<i>Pediastrum</i> sp.					0.03								0.53			
<i>Scenedesmus longispina</i> Chod					0.07 0.10 0.04				0.08 0.63 0.19 0.11				0.19			
<i>Scenedesmus</i> sp.	0.10												0.05 3.93			
<i>Spondylosium</i> sp.																
<i>Staurastrum gracile</i> Ralfs	0.05				0.12				0.08				0.19			
<i>Staurastrum leptocladum</i> Nor.	1.47 4.11 0.49 0.73 1.43 1.16								0.16 0.50				0.26 0.32 0.19			
<i>Staurastrum longiradiatum</i> West and West					0.04											
<i>Staurastrum subanchora</i> Gronbl.	0.05															
<i>Staurastrum subindentatum</i>					0.19											
<i>Staurastrum</i> sp.	0.41				0.15 0.31 0.28				0.13				0.21			

The first two PCA components accounted for 67.59% of the total variation. The first factor accounted for 53.39% and the second for 14.20%. The bi-dimensional projection shows two groups of variables with high internal correlation. Group A includes salinity, water temperature, water transparency and phosphate and characterizes the dry season. Group B brought together species richness, nitrogen (nitrate+nitrite), silicate and suspended particulate matter and characterized the rainy season. The dissolved oxygen and chlorophyll-*a* were related to the second axis and were inversely correlated to the species diversity (Fig. 6).

From the similarity analysis of the samples it was possible to distinguish two groups of species corresponding to the dry and rainy seasons (Fig. 7) when significant differences ($p=0.001$) were observed.

However, no significant differences were observed among the stages or between spring and neap tides.

DISCUSSION

A multitude of physical, chemical and biological processes affect organisms in marine areas (MANN, 1982). These processes operate over a range of spatial and temporal scales (TAGUSHI; LAWS, 1987) that must be considered when explaining variability in the structure, function and distribution of phytoplankton communities. In the tropics, this variability is highly affected by seasonal changes in rainfall (SOURNIA, 1969). Seasonal rainfall in tropical areas produces temporal and regional differences in river discharge. This induces fluctuations in salinity, nutrient concentrations, turbidity and biological productivity.

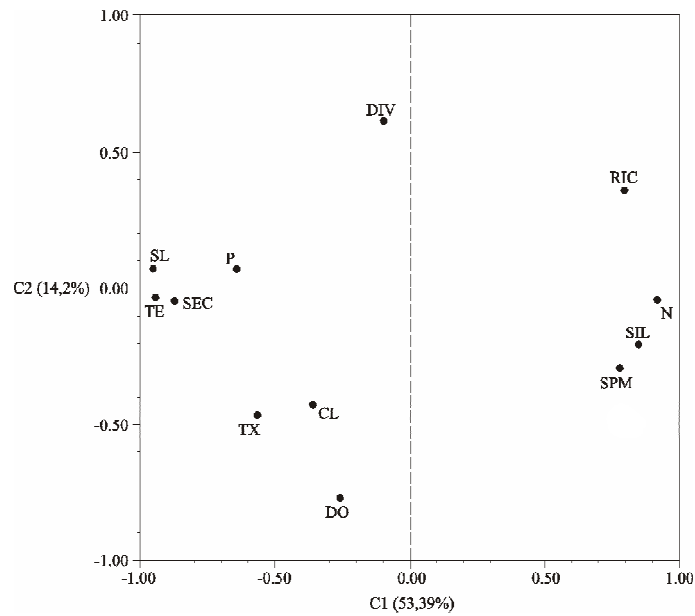


Fig. 6. Principal Component Analysis of the Port Basin of Recife Estuary (Pernambuco, Brazil) during the dry season (January 25-26 and February 02-03, 2005) and rainy season (June 07-08 and 14-15, 2005). Note: CL = chlorophyll-*a*, DIV = species diversity, P = phosphate, SPM = suspended particulate matter, N = nitrite+nitrate, RIC = number of species, DO = dissolved oxygen, SL = salinity, SIL = silicate, Tx = oxygen saturation rate, TE = water temperature and SEC = water transparency.

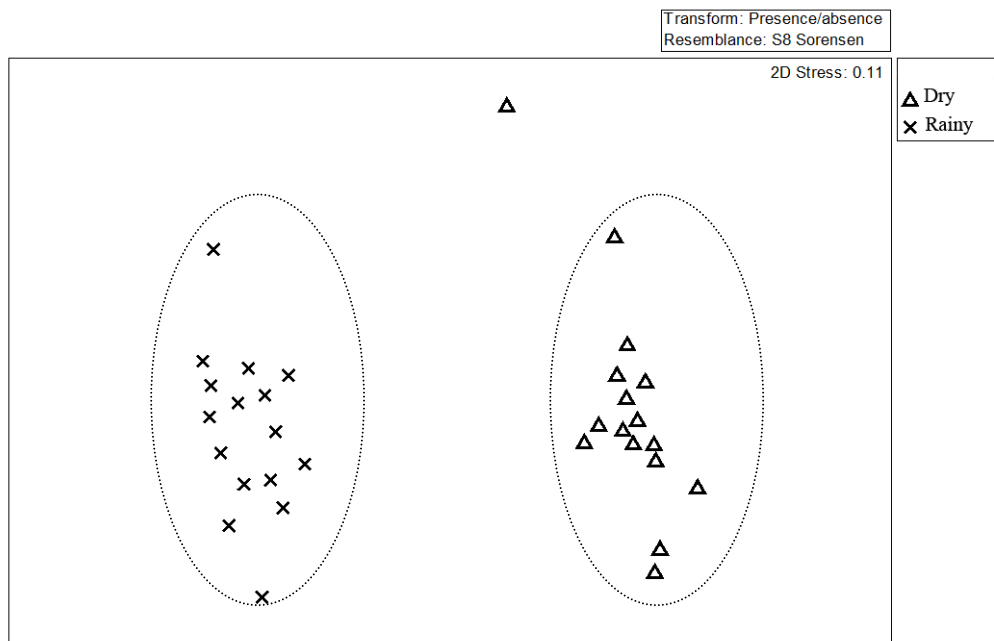


Fig. 7. Multidimensional scale of samples from the Port Basin of Recife Estuary (Pernambuco, Brazil) during the dry season (January 25-26 and February 02-03, 2005) and rainy season (June 07-08 and 14-15, 2005).

Previous research in the port of Recife and adjacent estuary shows that rainfall, Capibaribe and Beberibe Rivers and Pina Basin run-off affect the harbor area and produce temporal and spatial changes in the dynamics of coastal waters and phytoplankton standing crops (FEITOSA; PASSAVANTE, 1990; RESSURREIÇÃO et al., 1990; FEITOSA et al., 1999). The port of Recife presents features typical of estuarine waters during the rainy season when runoff is high and nutrients are abundant. Later, these nutrients fuel the phytoplankton blooms that commonly occur during the next dry season, when the river discharges decrease appreciably.

These blooms in the dry season bring about conditions of high productivity and create an unbalanced area. In our study, the high chlorophyll-*a* values were associated with high nutrient loads which enhance the eutrophication recorded in the adjacent Pina Basin (FEITOSA; PASSAVANTE, 1990; FEITOSA et al., 1999; NASCIMENTO et al., 2003) and other coastal environments (VARELA; PREGO, 2003; MELO-MAGALHÃES; KOENING; SANT'ANNA, 2004).

During this study, dissolved oxygen, saturation rate and phosphate showed significant differences with respect to changes in tidal stages, which suggests a great influence of the mixture between river flux and marine waters. Additionally, wind action can cause upwelling in shallow areas, causing resuspension of particulate matter and nutrients (McLUSKY; ELLIOT, 2004). The other physical, chemical and biological parameters showed no significant differences with low variation during the day, probably resulting from the mixing of these water masses.

Although nutrients appear to be available for the production of large quantities of phytoplankton in the study area, maximal production is apparently achieved only during neap tides in the dry season due to greater water transparency and duration of residence. In this time period, phytoplankton displayed an enhanced biomass with chlorophyll-*a* values up to 55 mg m⁻³.

According to McLusky and Elliot (2004), three factors limit phytoplankton production: turbidity can limit the penetration of light, shallowness means that blooms may not develop and the growth rate of phytoplankton may be less than the flushing rate of the estuary.

The estuarine area adjacent to Recife Harbor presents eutrophic to hyper-eutrophic conditions. This is typical of urban estuaries with strong anthropogenic impacts (e.g., dredging processes, dumping of municipal sewage), as documented for the Pina Basin (FEITOSA; PASSAVANTE, 1990; FEITOSA et al., 1999; NASCIMENTO et al., 2003) and the Capibaribe River (KOENING et al., 1995). This process has been

observed in many other urban areas (e.g., ALMEIDA et al., 2002; ODEBRECHT et al., 2005).

The higher phytoplankton richness in the rainy season was influenced by the input of freshwater species (chlorophytes and cyanobacteria). However, diatoms were the dominant phytoplankton group and some high density species (*Helicotheca tamesis*, *Coscinodiscus centralis*, *Coscinodiscus kutzingii*) were often responsible for the low diversity index. This group of species is predominant in other tropical estuaries under marine influence, probably due to their euryhaline characteristics (ESKINAZI-LEÇA et al., 2000; KOENING et al., 2003; LACERDA et al., 2004; MELO-MAGALHÃES; KOENING; SANT'ANNA, 2004; ROSEVEL DA SILVA et al., 2005; GAMEIRO et al., 2007).

The exceptionally high numbers of chlorophytes and cyanobacteria species (mainly *Anabaena*, *Oscillatoria*, *Pediastrum*, *Scenedesmus* and *Staurastrum*) in the rainy season were caused by the intense freshwater flux. This condition and the affinity of these groups with high nutrient concentrations favor their development, instead of the development of other photosynthesizing organisms (GRAHAM; WILCOX, 2000). Chlorophytes were the second group in terms of species richness. In general, this group is the most important component of oligotrophic and eutrophic continental waters (BICUDO; PARRA, 1995).

In our study we observed a great interplay of variables influencing the rate of phytoplankton photosynthesis (e.g., nutrient limitation, light limitation) and factors influencing species composition (e.g., tides, salinity). Rainfall influenced seasonal variability with high inputs of nutrients and sediments. Research into primary production in similar environmental conditions has shown that inputs stimulate phytoplankton growth when light conditions improve and that primary productivity decreases as turbidity increases (FEITOSA; PASSAVANTE, 1991, 1993; RESSURREIÇÃO; PASSAVANTE; MACEDO, 1996; ESKINAZI-LEÇA; KOENING; SILVA-CUNHA, 2000).

When primary production is high, eutrophication can lead to harmful algal blooms in the phytoplankton. Even in this case, total primary production will not necessarily change, but the changes in nutrient concentrations and ratios may influence the species composition of phytoplankton and have profound ecological implications (DAY JR. et al. 1989).

Studies carried out by Marone et al. (2005) close to a port in the Paranaguá Bay suggest that eutrophication processes and their consequences (e.g., oxygen depletion) tend to be more intense during the net heterotrophic rainy period and care should be taken to minimize the effect of the sewage load on the environment, especially during the rainy season.

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