

In vitro anthelmintic activity of *Lippia gracilis* Schauer essential oil against egg-hatching of goat gastrointestinal nematodes

Atividade anti-helmíntica *in vitro* do óleo essencial de *Lippia gracilis* Schauer contra a eclosão de nematódeos gastrintestinais de caprinos

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ABSTRACT: Since drug-resistant nematodes became a common problem in sheep and goat industries, alternative methods using natural products have emerged as a viable and sustainable anthelmintic treatment option. Here, the *in vitro* effect of essential oil extracted from *Lippia gracilis* Schauer was assessed on the hatching process of nematodes recovered from naturally infected goats. Essential oil at concentrations of 0.08% (0.008 µL/mL), 0.12% (0.012 µL/mL), and 0.16% (0.016 µL/mL) was able to induce an average inhibition of 74.7, 84 and 93%, respectively. The effective concentration required to inhibit egg hatching in 50% of eggs (EC50) was 0.03452%. Therefore, essential oil of *L. gracilis* showed promisor *in vitro* anthelmintic results against egg-hatching of goat gastrointestinal nematodes.

KEYWORDS: bioprospecting; ovicidal activity; small ruminants.

RESUMO: Como os nematóides resistentes a drogas se tornaram um problema comum nas indústrias de ovinos e caprinos, métodos alternativos que utilizam produtos naturais surgiram como uma opção de tratamento anti-helmíntico viável e sustentável. Aqui, o efeito *in vitro* do óleo essencial extraído de *Lippia gracilis* Schauer foi avaliado no processo de eclosão de nematóides recuperados de caprinos naturalmente infectados. O óleo essencial nas concentrações de 0,08% (0,008 µL/mL), 0,12% (0,012 µL/mL), e 0,16% (0,016 µL/mL) foi capaz de induzir uma inibição média de 74,7, 84 e 93%, respectivamente. A concentração efetiva necessária para inibir a eclosão de ovos em 50% dos ovos (CE50) foi de 0,03452%. Portanto, o óleo essencial de *L. gracilis* apresentou resultados anti-helmínticos *in vitro* promissores contra a eclosão de nematódeos gastrintestinais de caprinos.

PALAVRAS-CHAVE: bioprospecção; atividade ovicida; pequenos ruminantes.

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Goat farming is considered an activity of socioeconomic relevance, since it represents one of the main sources of protein for human consumption. Brazil is among the ten countries that have the largest number of goats in the world, with the largest population of this species concentrated in the Northeast region. According to data from the 2017 Census of Agriculture of the Brazilian Institute of Geography and Statistics, Brazilian goat farming reached 8.25 million head, a 16% increase in herds when compared to the previous census (IBGE, 2017).

For these small ruminants, the pasture is the most abundant option of feeding due to its lower cost of production. However, this food source provides a greater infection of these goats by gastrointestinal helminths, due to the presence of the infective larvae in the pasture (MELO et al., 2015). This occurs because of inadequate sanitary management, which in turn impedes the productive, nutritional and even reproductive management of these animals. Parasitic diseases are still considered the main cause of death of animals in herds (VERÍSSIMO et al., 2012).

Some of the genera of parasites of major clinical importance for small ruminants are *Haemonchus* spp., *Trichostrongylus* spp., *Oesophagostomum* spp. and *Strongyloides* spp., of which *Haemonchus* spp. is one of the most pathogenic (VERÍSSIMO et al., 2012). In an attempt to control widely disseminated parasitic infections, farmers began using anthelmintic drugs indiscriminately, which probably triggered a process of resistance, that is, the effectiveness of these chemical drugs against these parasites decreased (FORTES; MOLENTO, 2013).

Thus, alternative methods for control of gastrointestinal nematodes have been studied, with emphasis on phytotherapy. The essential oil of many medicinal plants has demonstrated some biological properties, such as antimicrobial, antiviral, antimutagenic, anticarcinogenic, antioxidant, immunomodulatory and anti-inflammatory (RIBEIRO et al., 2017).

Lippia gracilis Schauer is known in Brazil as “rosemary-of-Chapada” or “rosemary-board”, an herb commonly found in the Northeast region. It is a plant of semiarid climate that is found predominantly in the states of Bahia, Sergipe and Piauí, presenting as a shrub of about 2.5 cm high, with small leaves and white flowers (GOMES et al., 2011).

It is possible to extract essential oils of this species from stems, leaves, flowers and roots. The most common use of *L. gracilis* is the treatment of respiratory diseases, being used in colds, flu, bronchitis, coughs and asthma. Generally, members of the genus *Lippia* have similar chemical compositions, being among the most common components: p-cymene, carvacrol, humulene, thymol and 1,8-cineole; these compounds exhibit antimalarial, antiviral and cytostatic pharmacological activities, and may also be used to control insects and ticks (CRUZ et al., 2013; MELO et al., 2013).

The oil of *L. gracilis* has already been analyzed for its acaricidal (CRUZ et al., 2013), antidermatophytic and antileishmanial activities (MELO et al., 2013), but its possible action on gastrointestinal nematodes is still unknown. In this

context, the objective of this work was to evaluate the inhibitory effect of the essential oil extracted from *L. gracilis* in the process of hatching of goat gastrointestinal parasites.

The study was carried out in the municipality of Mossoró, which occupies an area of 2099 km² in the state of Rio Grande do Norte, with a mean temperature between 20 and 40 °C, and a mean humidity between 60 and 80%, sometimes being slightly higher or lower than these values (IBGE, 2012).

All experiments were performed in accordance with the recommended procedures of the National Council for the Control of Animal Experimentation. The study was approved by the Ethics Commission on Animal Experimentation of the Universidade Federal Rural do Semi-Árido (number 23091.009318/2016-40). All experiments were performed by the recommended procedures of the Ethics Commission on Animal Experimentation.

Essential oil was extracted from the leaves of *L. gracilis*. A voucher specimen was deposited under the designation 9317 at the Dárdaro de Andrade Lima Herbarium of the Universidade Federal Rural do Semi-Árido (UFERSA), Brazil.

Extraction of essential oil was performed as previously described by RASOOLI; MIRMOSTAFA (2003) (apud COSTA-JÚNIOR et al., 2016), where the leaves were macerated for 4 h, subsequently added in 500 ml of water, then hydrodistilled for 90 min. The oil was then isolated using a Clevenger apparatus for 2 h. The isolated oil was analyzed by gas chromatography–mass spectrometry for compounds identification.

Collection of feces was performed after at least 90 days from antiparasitic drugs administration to avoid residual effect. In parasitic resistance region (SANTOS et al., 2014), goats naturally infected were collected directly from the rectal ampulla to perform the fecal egg counting, and results were presented as number of eggs per gram of feces, according to the technique described by GORDON; WHITLOCK (1939) modified by CHAGAS et al. (2011). Eggs were recovered to accomplish the egg hatch test in compliance with the procedures of the World Association for the Advancement of Veterinary Parasitology (WAAVP). The recovery of the nematode eggs was performed according to the methodology of HUBERT; KERBOEUF (1992), where the feces pass through a sequence of sieves containing openings of different sizes 0.150, 0.100, 0.036 and 0.020 mm for egg retention (COLES et al., 1992).

In the first three sieves, the eggs passed through them, which retained the greatest impurities. However, in the last sieve, the eggs were retained and the water passed through the pores, causing concentration of eggs in a much-diminished volume. Thereafter, the resulting liquid was centrifuged for 5 min. Subsequently, the eggs were found in the pellet formed, then the supernatant was discarded and hypersaturated saline solution was added, aiming to allow the eggs to ascend to the supernatant by density and homogenized solution and centrifuged for another 5 min. Then, the supernatant was washed with distilled water so that the saline concentration was decreased

and a count of the eggs was made, when the calculation of how much solution should be placed in each well was performed (HUBERT; KERBOEUF, 1992; COLES et al., 1992)

Eggs were pooled and then submitted to treatment using concentrations of 0.08% (0.008 $\mu\text{L}/\text{mL}$), 0.12% (0.012 $\mu\text{L}/\text{mL}$), and 0.16% (0.016 $\mu\text{L}/\text{mL}$) essential oil diluted in 3% Tween 80 (CAMURÇA-VASCONCELOS et al., 2008). As negative control, just Tween 80 was used, and thiabendazole (32 $\mu\text{L}/\text{mL}$) (Sigma-Aldrich, St. Louis, MO, EUA) was used as positive control. The plates were placed in incubator of biochemical oxygen demand at 25 °C for a period of 48 h. At the end, a solution of lugol's iodine was added to each well and read under optical microscopy, where eggs and first-instar larvae were counted. All assays were repeated five times.

The collected data were analyzed through descriptive statistics, where the mean \pm standard deviation and coefficient of variation analyzed by the SigmaPlot program for Windows (SigmaPlot; Systat Software Inc) version 12.0. Differences among groups treated with different oil concentrations were analyzed using ANOVA ($\alpha = 0.05$), followed by Tukey's posttest ($\alpha = 0.05$), before that were verified the normality of data by Shapiro–Wilk test ($\alpha = 0.05$). Effective concentration capable to inhibit 50% of egg-hatching (EC50) was calculated with the software GraphPad Prism version 5.0 for Windows (GraphPad Software, USA).

The essential oil extracted from *L. gracilis* used in concentrations of 0.08% (0.008 $\mu\text{L}/\text{mL}$), 0.12% (0.012 $\mu\text{L}/\text{mL}$), and 0.16% (0.016 $\mu\text{L}/\text{mL}$) was able to induce inhibition of 74.7, 84 e 93% of egg-hatching on the egg hatch test, respectively, observing an increase in the inhibition rate in relation to the negative control (32%). An EC50 of 0.03452% was observed in this research. The genus *Lippia* also had its anthelmintic effect tested, such as *Lippia sidoides*, which showed 56.9% efficacy in sheep infected with nematodes resistant to

ivermectin (ALBUQUERQUE et al., 2006). Previous works have demonstrated the biological activity potential of the *L. gracilis* essential oil, since its antifungal, leishmanicidal and acaricide activity have been proven (MELO et al., 2013; COSTA-JR et al., 2016). The research in question shows another potential activity for this species, the anthelmintic action against gastrointestinal parasites of goats, with 93% inhibition of hatching eggs at their highest concentration.

MACEDO et al. (2011) demonstrated the anthelmintic properties of the essential oil of *Eucalyptus staigeriana* on *Haemonchus contortus*, with an impact on both egg hatching and larval development, reaching rates of 99.3 and 98.7% in both, respectively. The essential oil of *Piper aduncum* had also been tested on *H. contortus* (OLIVEIRA et al., 2014), also demonstrating action on egg hatching.

According to the results for the EC50, the essential oil showed a high activity, not only for the high inhibition rates demonstrated, but also for the fact that the EC50 was below the minimum value tested at work, demonstrating that even at low amounts, the oil has a significant effect.

Gas chromatography-mass spectrometry analysis revealed the predominant compounds of *L. gracilis* essential oil, being carvacrol (41.77%) and thymol (10.13%) the major ones. These compounds are generally related as the main active ingredients with antiparasitic action (CAMURÇA-VASCONCELOS et al., 2007). Carvacrol can be involved in acetylcholinesterase inhibition (SANTOS et al., 2014), and has an antioxidant action (KARKABOUNAS et al., 2006), whereas thymol is responsible for Ca^{2+} -ATPase inhibition at endoplasmic reticulum (SÁRKÖZI et al., 2007).

In conclusion, *L. gracilis* essential oil was capable to inhibit significantly egg hatching of goat gastrointestinal nematodes, and therefore could find application in anthelmintic therapy in veterinary practice.

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AVAILABILITY OF DATA AND MATERIAL: The datasets generated and/or analyzed during the current study are available in the Repositório da Biblioteca Orlando Teixeira da Universidade Federal Rural do Semi-Árido (<https://sigaa.ufersa.edu.br/sigaa/public/biblioteca/buscaPublicaAcervo.jsf>).

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