

Activity of powdered methylxanthine applied to poultry litter on adults of *Alphitobius diaperinus* (Panzer, 1797) (Coleoptera: Tenebrionidae)

*Atividade da Metilxantina em pó aplicada na cama de frango sobre adultos de *Alphitobius diaperinus* (Panzer, 1797) (Coleoptera: Tenebrionidae)*

Masaio Mizuno Ishizuka¹ , Nadja Susana Mogyca Leandro² , Julia Marixara Sousa da Silva² , Raphael Rodrigues dos Santos² , Helder Freitas de Oliveira² , Marcos Barcellos Café*² 

¹Universidade de São Paulo (USP), São Paulo, São Paulo, Brazil

²Universidade Federal de Goiás(UFG), Goiânia, Goiás, Brazil

*correspondent: mcafe@ufg.br

Abstract

One of the pests that most affect and compromise poultry production worldwide is the insect *Alphitobius diaperinus*, known as the lesser mealworm. This insect is a vector of diseases that compromise not only chicken production but also human health. This study proposes to examine the efficacy and determine the appropriate rate of methylxanthine (MTX), a natural insecticide extracted from caffeine, for the control of an adult population of lesser mealworms in poultry litter. A total of 2,500 adult mealworms were distributed into five treatments in a completely randomized design using 10 replications with 50 insects per replication. The treatments consisted of a control group and four concentrations of MTX (14, 16, 18, and 20 g/m²) spread in plastic boxes containing reused poultry litter and feed, allocated in a broiler shed, to simulate the farm condition. The experimental period was 18 days, and five readings were performed on days 2, 4, 6, 10, and 18. Methylxanthine affected ($P<0.05$) the mealworms' cumulative mortality rate, with the groups of insects housed in boxes treated with 16 g/m² MTX showing the highest cumulative mortality (86.6%) at the end of the experimental period. In conclusion, MTX has insecticidal action on adults of lesser mealworm and can be used on chicken litter to control the population of this insect in poultry sheds. The MTX concentration of 16 g/m² showed the greatest effectiveness.

Keywords: chicken bed; insecticide; lesser mealworm; methylxanthine

Resumo

Uma das pragas que mais afetam e comprometem a produção avícola no mundo é o inseto *Alphitobius diaperinus*, conhecido como cascudinho. Este inseto é vetor de doenças que comprometem não só a produção de frangos como também a saúde humana. Objetivou-se com esse trabalho avaliar a eficácia e determinar a dose adequada de Metilxantina (MTX), inseticida natural extraído da cafeína, para o controle da população adulta de cascudinhos em cama de frango. Foram utilizados 2.500 cascudinhos adultos distribuídos em delineamento inteiramente casualizado, cinco tratamentos, 10 repetições com 50 insetos por repetição. Os tratamentos consistiram de grupo controle e quatro concentrações 14 g/m², 16 g/m², 18 g/m², 20 g/m² de MTX espalhadas em recipientes plásticos contendo cama de frango reutilizada e ração, alocados em um galpão de frangos de corte a fim de simular a condição de granja. Período experimental foi de 18 dias e realizadas cinco leituras nos dias dois, quatro, seis, 10 e 18. A MTX afetou ($P<0,05$) a taxa de mortalidade acumulada de cascudinhos, grupos de insetos alojados em caixas tratadas com 16 g/m² de MTX apresentaram maior mortalidade acumulada (86,6%) ao final do período experimental. Conclui-se que MTX tem ação inseticida sobre adultos de cascudinho, podendo ser utilizada sobre a cama de frango para o controle da população deste inseto em galpões de criação de frangos, a concentração 16 g/MTX/m² demonstrou maior efetividade.

Palavras-chave: Metilxantina; inseticida; cama de frango; cascudinho

Received: April 28, 2022. Accepted: September 2, 2022. Published: September 30, 2022.



This is an Open Access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Introduction

Alphitobius diaperinus, a beetle commonly known as the lesser mealworm, is one of the most abundant insects in chicken production facilities. The species stands out in the poultry industry for having the status of a pest, as it is a reservoir and vector of pathogens and is difficult to control. The lesser mealworm breeds in litter used on chicken farms and feeds on poultry waste, feed leftovers, broken eggs, dead birds, and other organic materials^(1,2,3). The greatest economic losses in poultry production establishments are due to: i) the preference of birds to consume this insect instead of feed, which reduces their weight gain^(4,5); ii) damage to the structures of the facilities, as mealworm larvae usually dig tunnels into the walls, insulating materials, and on the floor of the shed in search of places to pupate and escape from the enemies present in the litter^(2, 6, 7, 8, 9, 10, 11, 12, 13, 14).

The birds' health is also compromised because *A. diaperinus* act as transmission routes for etiological agents of diseases such as: i) *Salmonella* ^(15, 16, 17), *Campylobacter* sp.⁽¹⁸⁾, *Escherichia coli*^(19,20,21); ii) fungi⁽²⁰⁾; iii) Marek's disease virus⁽²²⁾, Newcastle disease and yaws⁽²³⁾, enteritis⁽²⁴⁾, and Gumboro disease^(25,26); iv) chicken tapeworm⁽²⁷⁾, and *Ascaridia galli*⁽²⁷⁾. The chemical products most widely used are pyrethroids (bifenthrin, deltamethrin, fenitrothion, pirimiphos-methyl), which are highly toxic to birds and whose accumulation in muscle tissue renders the meat of these chickens unsuitable for human consumption^(1,28). The number of reports of lesser mealworm populations resistant to these compounds is increasing^(29, 30, 31), including in Brazil^(32,33).

Chemical control of the lesser mealworm through the application of pyrethroid and organophosphate insecticides is commonly used as a preventive measure⁽³⁴⁾, and this approach during downtime is more efficient for producers in the short term. However, research has already shown that this type of chemical control with the use of pyrethroids in litters during downtime no longer yields effective results due to the return of the presence of this pest during housing⁽³⁵⁾. In this scenario, the use of natural products is another method that has stood out in the control of several pathogenic microorganisms and pests. These products can be an efficient and viable alternative for the control of the lesser mealworm, since this method does not depend on the absence of birds in the poultry shed. Some essential oils have already been tested and their insecticidal effect proven, e.g. *Melaleuca alternifolia* oils^(1,36).

Caffeine (1,3,7 trimethylxanthine) is a fat-soluble alkaloid belonging to the class of compounds called methylxanthine⁽³⁷⁾ that acts on the human central nervous system and is also known for its antioxidant

properties⁽³⁸⁾. Together with theophylline, it is found in wild fruits, seeds, and leaves of numerous plant species including tea, coffee, cocoa, and nuts⁽³⁹⁾. Studies have shown that caffeine causes toxic effects on *Aedes aegypti* larvae, interfering with their development and consequently preventing them from reaching the adult stage^(40, 41), which leads to a decrease in oviposition rate⁽⁴²⁾.

The objective of this study was to examine the efficacy and determine the effective concentration of methylxanthine extracted from caffeine against adults of lesser mealworm (*Alphitobius diaperinus*) in poultry litter.

Material and methods

The experiment was carried out in the Experimental Poultry House in the Poultry Section of the Animal Science Department of the Veterinary and Animal Science School at the Federal University of Goiás, located in Goiânia - GO, Brazil. A total of 2,500 lesser mealworm (*Alphitobius diaperinus*) adults captured in several broiler farms in the region of Itaberaí - GO were used in a completely randomized experimental design with five treatments and 10 replications with 50 insects per experimental unit. The treatments consisted of four concentrations of methylxanthine (MTX) (14, 16, 18, and 20 g/m²) spread in plastic boxes (41 × 27 × 12.5 cm) containing a 10-cm-high layer of reused chicken litter with a portion of chicken feed, plus a control group without the use of any product.

After the mealworms and MTX were placed, the boxes were sealed with tulle fabric to prevent the entry and exit of insects. Subsequently, the boxes were transferred to a broiler shed to simulate the farm condition. The experiment lasted 18 days and five readings were performed, on days 2, 4, 6, 10, and 18. At each elapsed period, the seals were removed to count dead and live insects with the aid of surgical tweezers and data were recorded in collection forms. Afterwards, the boxes were sealed again, and this process was repeated until the end of the experimental period. Data were evaluated by analysis of variance (ANOVA) and Tukey's test using the R computer package and adopting $\alpha=0.05$.

Results

The methylxanthine (MTX) concentration affected ($P<0.05$) the cumulative mortality rate of lesser mealworm. The groups of insects housed in boxes treated with 16 g MTX/m² showed the highest cumulative mortality (86.6%) at the end of the experimental period (Table 1).

Table 1. Cumulative mortality rate of lesser mealworm (*Alphitobius diaperinus*) in chicken litter treated with methylxanthine (MTX)

Observation day	Treatment																	
	12 g MTX/m ²		14 g MTX/m ²		16 g MTX/m ²		18 g MTX/m ²		Control									
	Live	Dead	Live	Dead	Live	Dead	Live	Dead	Live	Dead								
2nd	354	146	352	148	198	302	322	178	497	3								
4th	287	67	285	67	138	60	270	52	495	2								
6th	269	18	255	30	117	21	241	29	493	2								
10th	233	36	219	36	103	14	208	33	491	2								
18th	183	50	167	52	67	36	158	50	486	5								
Cumulative mortality (%)	63.4 b		66.6 b		86.6 a		68.4 b		2.8 c									
P-value	<0.0001																	
SEM ¹	4.288																	

Means followed by distinct letters differ from each other by Tukey's test (P<0.05); ¹Standard error of the mean.

Discussion

The primary effect of methylxanthine (MTX) in the mealworm was found to be due to the inhibition of phosphodiesterase activity and intracellular increase of cyclic adenosine monophosphate (cyclic AMP). At low concentrations, they are potent synergists of other insecticides known to activate adenylate cyclase in insects. These data suggest the use of MTX in the control of arthropods as a natural insecticide, by inhibiting phosphodiesterase alone with the involvement of AMP cyclase; or in combination with other compounds⁽⁴³⁾. Nathanson⁽⁴³⁾ demonstrated the pesticidal and peststatic activity of MTX in goliath worm (*Manduca sexta*) larvae and observed lethality within 24 h. Polo⁽⁴⁴⁾ showed that MTX alters the synthesis pattern of esterases, which are important in several physiological processes and are even involved in blocking insect metamorphosis. Esterases are also involved in several physiological processes, including neuronal activity⁽⁴⁵⁾, insect juvenile hormone metabolism⁽⁴⁶⁾, and insecticide resistance⁽⁴⁷⁾.

In neuronal activity, acetylcholine is released into the synaptic cleft, binds to transmembrane receptors, and generates signal transmission. Later, acetylcholinesterase hydrolyzes acetylcholine, triggering the stimulus⁽⁴⁸⁾. Nishi et al.⁽⁴⁹⁾ confirmed that caffeine inhibits the pattern of esterase gene expression. Juvenile hormone esterase controls the concentration of juvenile hormone and, therefore, caffeine alters the gene expression pattern of esterases. Juvenile hormone is a class of sesquiterpenoids produced in the insect *Corpora Allata* and distributed throughout the hemolymph, being directly involved in several metabolic activities of insects such as metamorphosis and oogenesis. During metamorphosis, it modulates ecdysteroid activity (20E), preventing molt from occurring during the larval stage⁽⁵⁰⁾.

Among the different plant extracts with insecticidal effect demonstrated in laboratory studies, caffeine has been the one that acts in the intoxication of larvae, interrupting their development and leading them

to death^(40,42,51). Caffeine proved to be potentially effective in controlling adult red flour beetles through repellent, fumigant, and contact effects, and its action is believed to be linked to the inhibition of certain enzymes such as carboxylesterase⁽⁵²⁾. In adult insects, caffeine causes motor immobility by increasing the activity of the dopamine receptor, an antagonist of adenosine receptors, inhibiting their action^(49,53,54). Caffeine acts in a dose-dependent manner, with the concentration of 1 mg/mL of water being lethal to larvae⁽⁴⁰⁾.

Herbal products are recognized as natural insecticides, as is the case of tea extract-based emulsion, which can be used as an insecticide to control the green peach aphid⁽⁵⁸⁾. Some formulations of insecticides based on medicinal herbs were effective in combating several pests, e.g. rosemary-pepper⁽⁵⁹⁾, neem (*Azadirachta indica*)^(60, 61), garlic⁽⁶²⁾, and several species of eucalyptus⁽⁶³⁾, whose main deleterious effects in the fight against insects are related to methylxanthine.

In a similar study, Ananenka⁽⁶⁴⁾ tested the efficacy of a natural insecticide composed of caffeine on the house cricket (*Acheta domesticus*) and concluded that caffeine results in increased neuronal activity and consequently the death of the insect due to depolarization of neurons in the brain membrane, constituting an option to control this species. In the present study, the MTX concentration of 16 g/m² had positive results for cumulative mortality rate in the total observation period, which was 80% higher than in the control treatment, and ensured a consecutive increase in mortality from the 2nd to the 18th day of observation.

The control of *Alphitobius diaperinus* is essential due to its many detrimental effects on the productivity of these animals, as this species can cause superficial wounds and negatively affect the birds' growth. In addition, the ingestion of large amounts of *A. diaperinus* compromises the nutritional quality of diets, since adults and larvae can cause intestinal obstruction in broilers due to the lack of chitinase for the digestion of chitin, which is widely found in the exoskeleton of the mealworm. This

results in necrotic enteritis, reduced nutrient absorption, and, ultimately, considerable economic losses for the poultry industry^(65, 66).

Several studies have demonstrated that cypermethrin, dichlorvos, and triflumuron are efficient in controlling ectoparasites in poultry production in Brazil^(67, 68). However, resistance of pest populations to these compounds has already been reported in various countries^(69, 70). Therefore, the use of plant extracts such as citronella, caffeine, among others that have an insecticidal action on the central nervous system of insects, is encouraged. These compounds have been proven to harm the development of insects and have repellent and larvicidal activity^(71, 72), thereby constituting efficient ways to control the damage caused by *Alphitobius diaperinus* in poultry production.

Conclusion

Methylxanthine, extracted from caffeine, has an insecticidal action on lesser mealworm (*Alphitobius diaperinus*) adults and can be used on chicken litter to control the population of this insect in poultry sheds. For this purpose, the methylxanthine concentration of 16 g/m² is the most effective.

Conflicts of interest

The authors declare they have no conflicts of interest.

Author contributions

Conceptualization: M.M. Ishizuka, N.S.M. Leandro and M.B. Café. *Data curation:* J.M.S. Silva, R.R. Santos and H.F. Oliveira. *Formal Analysis:* M.B. Café. *Resources:* M.M. Ishizuka and N.S.M. Leandro. *Writing (original draft):* M.M. Ishizuka, M.B. Café and H.F. Oliveira. *Writing (review & editing):* H.F. Oliveira

References

- Volpato A, Galli GM, Campigotto G, Glombowsky P, Santos RCV, Silva AS, Vaucher RA. Avaliação *in vitro* dos efeitos inseticida e larvícola de oito óleos essenciais sobre o casquinho aviário (*Alphitobius diaperinus*). Arch. Vet. Sci. 2018; 23(2):84-90. Available from: <http://dx.doi.org/10.5380/avs.v23i2.46127>
- Axtell RC, Arends JJ. Ecology and management of arthropod pests of poultry. Annu. Rev. Entomol. 1990; 35(1990):101-126. Available from: <https://doi.org/10.1146/annurev.en.35.010190.000533>
- Rueda LM, Axtell RC. Arthropods in litter of poultry (broiler chicken and turkey) houses. J. Agri. Entomol. 1997; 14(1):81-91. Available from: <http://www.lib.ncsu.edu/resolver/1840.2/2110>
- Santos JC, Alves LFA, Opazo MAU, Mertz NR, Marcomini AM, Oliveira DGP, Bonini AK. Eficiência da aplicação de inseticida químico no solo para o controle de *Alphitobius diaperinus* Panzer (Coleoptera: Tenebrionidae) em avíario de frango de corte. Arq. Inst. Biol. 2009; 76(3):417-425. Available from: <http://dx.doi.org/10.1590/1808-1657v76p4172009>
- Wojciechowski P, Pedrassani D, Fedalto LM. Terra de diatomáceas para controle do *Alphitobius diaperinus* em granjas de frango de corte. Saúde Meio Ambiente. 2015; 4(1):66-78. Available from: <https://doi.org/10.24302/sma.v4i1.695>
- Somerfield KG. Recent aspects of stored product entomology in New Zealand. New Zealand J. Agric. Res. 1981; 24(0):403-408. Available from: <http://hbs.bishopmuseum.org/fiji/pdf/somerfield1981.pdf>
- Vaughan JA, Turner EC. Studies on the infestation into polystyrene insulation by the lesser mealworm (*Alphitobius diaperinus* Panz.), a common inhabitant of deep-pit caged layer houses. Va. J. Sci. 1982; 33(3):91-91.
- Despins JL. Investigations of the destructive behavior, and methods for control of the lesser mealworm, *Alphitobius diaperinus* (Panzer) (Coleoptera: Tenebrionidae). Ph.D. Tesisn, Blacksburg, Virgínia, 1987.
- Despins JL, Turner EC, Ruszler PR. Effects of poultry manure moisture and poultry house construction materials on movements of the lesser mealworm, *Alphitobius diaperinus* (Panzer) (Coleoptera: Tenebrionidae), a structural insect pest in high-rise layer houses. Poult. Sci. 1989; 68(10):1326-1331. Available from: <https://doi.org/10.3382/ps.0681326>
- Despins JL, Turner EC, Pfeifer DG. Evaluation of methods to protect poultry house insulation from infestation by lesser mealworm (Coleoptera: Tenebrionidae). J. Agric. Entomol. 1991; 8(3):209-217. Available from: <http://hdl.handle.net/10919/73829>
- O'Connor JP. *Alphitobius diaperinus* (Panzer) (Coleoptera: Tenebrionidae) damaging polystyrene insulation on an Irish pigery. Entomol. Monthly Magaz. 1987; 123(0):1472-1475.
- Geden CJ, Axtell RC. Factors affecting climbing and tunneling behavior of the lesser mealworm, *Alphitobius diaperinus* (Coleoptera: Tenebrionidae). J. Econom. Entomol. 1987; 80(6):1197-1204. Available from: <https://doi.org/10.1093/jee/80.6.1197>
- Axtell RC. Biology and economic importance of the darkling beetle in poultry houses. Proceedings of the North Carolina State University Poultry Supervisors' Short Course. 1994; 8-17. Available from: <http://www.lib.ncsu.edu/resolver/1840.2/2108>
- Salin C, Delettre YR, Cannavaccioulo M, Vernon P. Spatial distribution of *Alphitobius diaperinus* (Panzer) (Coleoptera: Tenebrionidae) in the soil of a poultry house along a breeding cycle. Eur. J. Soil. Biol. 2000; 36(2):107-115. Available from: [https://doi.org/10.1016/S1164-5563\(00\)01054-2](https://doi.org/10.1016/S1164-5563(00)01054-2)
- McAllister JC, Steelman CD, Skeels JK. Reservoir competence of the lesser mealworm (Coleoptera: Tenebrionidae) for *Salmonella typhimurium* (Eubacteriales: Enterobacteriaceae). J. Med. Entomol. 1994; 31(3):369-372. Available from: <https://doi.org/10.1093/jmedent/31.3.369>
- Crippen TL, Zheng L, Sheffield CL, Tomberlin JK, Beier RC, Yu Z. Transient gut retention and persistence of *Salmonella* through metamorphosis in the lesser mealworm, *Alphitobius diaperinus* (Coleoptera: Tenebrionidae). J. Appl. Microbiol. 2012; 112(5):920-926. Available from: <https://dx.doi.org/10.1111/j.1365-2672.2012.05265.x>
- Roche AJ, Cox NA, Richardson LJ, Buhr RJ, Cason JA, Fairchild BD, Hinkle NC. Transmission of *Salmonella* to broilers by contaminated larval and adult lesser mealworms, *Alphitobius diaperinus* (Coleoptera: Tenebrionidae). Poult. Sci. 2009; 88(1):44-48. Available from: <https://doi.org/10.3382/ps.2008-00235>
- Strother KO, Steelman CD, Gbur EE. Reservoir competence of lesser mealworm (Coleoptera: Tenebrionidae) for *Campy-*

- lobacter jejuni* (Campylobacterales: Campylobacteraceae). J. Med. Entomol. 2005; 42(1):42-47. Available from: <https://doi.org/10.1093/jmedent/42.1.42>
19. De Las Casas E, Pomeroy BS, Harein PK. Infection and quantitative recovery of *Salmonella typhimurium* and *Escherichia coli* from within the lesser mealworm, *Alphitobius diaperinus* (Panzer). Poult. Sci. 1968; 47(6):1871-1875. Available from: <https://doi.org/10.3382/ps.0471871>
20. De Las Casas E, Harein PK, Pomeroy BS. Bacteria and fungi within the lesser mealworm collected from poultry brooder houses. Environ. Entomol. 1972; 1(1):27-30. Available from: <https://doi.org/10.1093/ee/1.1.27>
21. McAllister JC, Steelman CD, Skeels JK, Newberry LA, Gbur EE. Reservoir competence of the *Alphitobius diaperinus* (Coleoptera: Tenebrionidae) for *Escherichia coli* (Eubacteriales: Enterobacteriaceae). J. Med. Entomol. 1996; 33(6):983-987. Available from: <https://doi.org/10.1093/jmedent/33.6.983>
22. Eidson CS, Schmitt SC, Goode RB, Lal JB. The role of the darkling beetle (*Alphitobius diaperinus*) in the transmission of acute leukosis in chickens. Poult. Sci. 1965; 44(5):1366-1367. Available from: <https://doi.org/10.3382/ps.0441347>
23. De Las Casas E, Harein PK, Deshmukh DR, Pomeroy BS. Relationship between the lesser mealworm, fowl pox and Newcastle disease virus in poultry. J. Econ. Entomol. 1976; 69(6):775-779. Available from: <https://doi.org/10.1093/jee/69.6.775>
24. Despins JL, Axtell RC, Rives DV, Guy JS, Ficken MD. Transmission of enteric pathogens of turkeys by darkling beetle larva (*Alphitobius diaperinus*). J. Appl. Poult. Res. 1994; 3(1):61-65. Available from: <https://doi.org/10.1093/japr/3.1.61>
25. Watson DW, Guy JS, Stringham SM. Limited transmission of turkey coronavirus in young turkeys by adult *Alphitobius diaperinus* (Coleoptera: Tenebrionidae). J. Med. Entomol. 2000; 37(3):480-483. Available from: <https://doi.org/10.1093/jmedent/37.3.480>
26. Mullen G, Durden L. Medical and veterinary entomology. 3 rd ed. San Diego, United States, Academic Press, 2019;794p.
27. Elowni EE, Elbahi S. Natural and experimental infection on the beetle, *Alphitobius diaperinus* (Coleoptera: Tenebrionidae) with *Choanotaenia infundibulum* and other chicken tape-worms. Vet. Sci. Commun. 1979; 3:171-173. Available from: <https://doi.org/10.1007/BF02268965>
28. Galli A, Souza D, Garbellini GS, Coutinho CFB, Mazo LH, Avaca LA, Machado SAS. Utilização de técnicas eletroanalíticas na determinação de pesticidas em alimentos. Quím. Nova. 2006; 29(1):105-112. Available from: <https://doi.org/10.1590/S0100-40422006000100020>
29. Tomberlin JK, Richman D, Myers HM. Susceptibility of *Alphitobius diaperinus* (Coleoptera: Tenebrionidae) from broiler facilities in Texas to four insecticides. J. Econ. Entomol. 2008; 101(2):480-483. Available from: [https://doi.org/10.1603/0022-0493\(2008\)101\[480:sadct\]2.0.co;2](https://doi.org/10.1603/0022-0493(2008)101[480:sadct]2.0.co;2)
30. Chernaki-Leffer AM, Sosagómez DR, Almeida LM, Lopes ION. Susceptibility of *Alphitobius diaperinus* (Panzer) (Coleoptera, Tenebrionidae) to cypermethrin, dichlorvos and triflumuron in southern Brazil. Rev. Bras. Entomol. 2011; 220(1):125-128. Available from: <https://doi.org/10.1590/S0085-56262011000100020>
31. Fogaca I, Ferreira E, Saturnino KC, Santos TR, Cavali J, Porto MO. Álcool para controle de cascadinho em cama de frangos de corte. Arch. Zootec. 2017; 66(256):509-514. Available from: <https://doi.org/10.21071/az.v66i256.2766>
32. Chernaki-Leffer AM, Sosa-Gomez DR, Almeida LM. Susceptibilidade de *Alphitobius diaperinus* (Panzer, 1797) (Coleoptera: Tenebrionidae) a reguladores de crescimento de insetos (RCI). Arq. Inst. Biol. 2006; 73(1):51-55. Available from: https://www.researchgate.net/publication/284549911_Susceptibility_of_Alphitobius_diaperinus_Panzer_1797_Coleoptera_Tenebrionidae_to_insect_growth_regulators_IGR
33. Gazoni FL, Flores F, Bampi RA, Silveira F, Boufleur R, Lovato. Avaliação da resistência do cascadinho (*Alphitobius diaperinus*) (Panzer) (Coleoptera: Tenebrionidae) a diferentes temperaturas. Arq. Inst. Biol. 2012; 79(1):69-74. Available from: <https://www.scielo.br/j/aiab/a/QDGt4PYpcqL76Yt3XtPH5bD/?format=pdf&lang=pt>
34. Alves LFA, Uemura-Lima DH, Oliveira DGP, Godinho RPV. Eficiência de um novo inseticida comercial para o controle do cascadinho dos aviários (*Alphitobius diaperinus*) (Panzer) (Coleoptera: Tenebrionidae). Arq. Inst. Biol. 2010; 77(4):693-700. Available from: <https://doi.org/10.1590/1808-1657v77p6932010>
35. Dias DA, Vargas AB, Almeida FS. Efeitos de dosagem mais concentrada de cipermetrina no controle de cascadinho. Rev. Acad. Ciênc. Agrár. Ambient. 2013; 11(4):437-442. Available from: <https://doi.org/10.7213/academico.011.004.AO11>
36. Volpato A, Lorenzetti WR, Zortea T, Giombelli LCDD, Barretta D, Santos RCV, Vaucher RA, Raffin RP, Souza ME, Stefani LM, Boligon AA, Athayde ML, Silva AS. *Melaleuca alternifolia* essential oil against the lesser mealworm (*Alphitobius diaperinus*) and its possible effect on the soil fauna. Rev. Bras. Ciênc. Avíc. 2016; 18(1):41-46. Available from: <https://doi.org/10.1590/1516-635X1801041-046>
37. Maria CAB, Moreira RFA. Caféina: revisão sobre métodos de análise. Quim. Nova. 2007; 30(1):99-105. Available from: http://static.sites.saq.org.br/quimicanova.saq.org.br/pdf/Vol30-No1_99_20-RV05372.pdf
38. Krisko A, Kveder M, Pifat G. Effect of caffeine on oxidation susceptibility of human plasma low density lipoproteins. Clin. Chim. Acta. 2005; 355(1-2):47-53. Available from: <https://doi.org/10.1016/j.cccn.2004.12.001>
39. Varago FC, Silva LP, Ribeiro JR, Fernandes CA, Carvalho BC, Gioso MM, Moustacas VS. Teofilina como agente capacitante do semen bovino. Arq. Bras. Med. Vet. Zootec. 2017; 69(6):1670-1614. Available from: <https://doi.org/10.1590/1678-4162-9173>
40. Laranja AT, Manzato AJ, Bicudo HEMC. Effects of caffeine and used coffee grounds on biological features of *Aedes aegypti* (Diptera: Culicidae) and their possible use in alternative control. Genet. Mol. Biol. 2003; 26(4):419-429. Available from: <https://doi.org/10.1590/S1415-47572003000400004>
41. Guirado MM, Bicudo HEMMC. Attractiveness of bioinsecticides caffeine and used coffee grounds in the choice of oviposition site by *Aedes aegypti* (Diptera: Culicidae). Int. J. Mosquito Res. 2016; 47(3):47-51. Available from: <https://docs.bvsalud.org/biblioref/ses-sp/2016/ses-38077/ses-38077-7041.pdf>
42. Laranja AT, Manzato AJ, Bicudo HEMC. Caffeine effect on mortality and oviposition in successive generations of *Aedes aegypti*. Rev. Saúde Pública. 2006; 40(6):1112-1117. Available from: <https://doi.org/10.1590/S0034-89102006000700022>
43. Nathanson JA. Caffeine and related methylxanthines: possible naturally occurring pesticides. Science. 1984; 226(4671):184-187. Available from: <https://doi.org/10.1126/science.6207592>
44. Pólo AM. Efeito da cafeína no desenvolvimento de *Aedes aegypti* (Diptera: Culicidae): o significado biológico das alterações

- ções do padrão de síntese de esterases. Dissertação Mestrado. UNESP, 2014. Available from: <http://www.sbicafe.ufv.br/handle/123456789/8993>
45. Harel M, Kryger G, Rosenberry TL, Mallender WD, Lewis T, Fletcher RJ, Guss JM, Silman I, Sussman JL. Three-dimensional structures of *Drosophila melanogaster* acetylcholinesterase and of its complexes. *Protein Sci.* 2000; 9(6):1063-1072. Available from: <https://dx.doi.org/10.1110%2Fps.9.6.1063>
46. Davey K. The interaction of feeding and mating in the hormonal control of egg production in *Rhodnius prolixus*. *J. Insect Physiol.* 2007; 53(3): 208-215. Available from: <https://doi.org/10.1016/j.jinsphys.2006.10.002>
47. Lucena ALM, Gigliolli AAS, Lapenta AS. Análise das esterases durante as fases do desenvolvimento em *Sitophilus oryzae* (Coleoptera: Curculionidae) e sua relação com a resistência ao inseticida malathion. *SaBios: Rev. Saúde Biol.* 2012; 7(3):36-44. Available from: <https://revista2.grupointegrado.br/revista/index.php/sabios/article/view/771>
48. Pohanka M. Alpha-7-nicotinic acetylcholine receptor is a target in pharmacology and toxicology. *Int. J. Mol. Sci.* 2012; 13(2):2219-2238. Available from: <https://dx.doi.org/10.3390%2Fijms1302219>
49. Nishi Y, Sasaki K, Miyatake T. Biogenic amines, caffeine and tonic immobility in *Tribolium castaneum*. *J. Insect Physiol.* 2010; 56(6):622-628. Available from: <https://doi.org/10.1016/j.jinsphys.2010.01.002>
50. Mansur JF, Figueira-Mansur J, Santos AS, Santos-Junior H, Ramos IB, Medeiros MN, Machado EA, Kaiser CR, Muthukrishnan S, Masuda H, Melo ACA, Moreira MF. The effect of lufenuron, a chitin synthesis inhibitor, on oogenesis of *Rhodnius prolixus*. *Pestic. Biochem. Physiol.* 2010; 98(1):59-67. Available from: <https://dx.doi.org/10.1016/j.pestbp.2010.04.013>
51. Guirado MM, Bicudo HEMC. Effect of used coffee grounds on larval mortality of *Aedes aegypti* L. (Diptera: culicidae): Suspension concentration and age versus efficacy. *BioAssay.* 2007; 2(5):1-7. Available from: <http://www.seb.org.br/biosay/arquivos/journals/1/articles/52/public/52-254-1-PB.pdf>
52. Phankaen Y, Manaprasertsak A, Pluempanupat W, Koul O, Kainoh Y, Bullangpoti V. Toxicity and repellent action of *Coffea arabica* against *Tribolium castaneum* (Herbst) adults under laboratory conditions. *J. Stored Prod. Res.* 2017; 71(2017):112-118. Available from: <https://doi.org/10.1016/j.jspr.2017.01.006>
53. Fredholm BB, Bättig K, Holmén J, Nehlig A, Zvartau EZ. Actions of caffeine in the brain with special reference to factors that contribute to its widespread use. *Pharmacol. Rev.* 1999; 51(1):83-133. Available from: <https://pharmrev.aspetjournals.org/content/51/1/83>
54. Zahniser N, Simosky JK, Mayfield RD, Negri CA, Hanania T, Larson GA, Kelly MA, Grandy DK, Rubinstein M, Low MJ, Fredholm BB. Functional uncoupling of adenosine A(2A) receptors and reduced response to caffeine in mice lacking dopamine D2 receptors. *J. Neurosci.* 2000; 20(16):5949-5957. Available from: <https://doi.org/10.1523/jneurosci.20-16-05949.2000>
55. Crippen TL, Sheffield CL, Esquivel SV. The acquisition and internalization of *Salmonella* by the lesser mealworm, *Alphitobius diaperinus* (Coleoptera: Tenebrionidae). Vector Borne Zoonotic Dis. 2009; 9(1):65-71. Available from: <https://doi.org/10.1089/vbz.2008.0103>
56. Lambkin TA, Rice SJ. Baseline responses of *Alphitobius diaperinus* (Coleoptera: Tenebrionidae) to cyfluthrin and detection of strong resistance in field populations in eastern Australia. *J. Econ. Entomol.* 2006; 99(3):908-913. Available from: <https://doi.org/10.1603/0022-0493-99.3.908>
57. Kaufman PE, Strong C, Rutz DA. Susceptibility of lesser mealworm (Coleoptera: Tenebrionidae) adults and larvae exposed to two commercial insecticides on unpainted plywood panels. *Pest Manag. Sci.* 2008; 64(2):108-111. Available from: <https://doi.org/10.1002/ps.1475>
58. Khoshraftar Z, Shamel A, Safekordi AA, Zaefizadeh M. Chemical composition of an insecticidal hydroalcoholic extract from tea leaves against green peach aphid. *Int. J. Environ. Sci. Technol.* 2019; 16(11):7583-7590. Available from: <http://dx.doi.org/10.1007/s13762-018-2177-x>
59. Gomes GA, Monteiro CMO, Julião LS, Maturano R, Senra TOS, Zeringóta V, Calmon F, Matos RS, Daemon E, Carvalho MG. Acaricidal activity of essential oil from *Lippia sidooides* on unengorged larvae and nymphs of *Rhipicephalus sanguineus* (Acari: Ixodidae) and *Amblyomma cajennense* (Acari: Ixodidae). *Exp. Parasitol.* 2014; 137(0):41-45. Available from: <https://doi.org/10.1016/j.exppara.2013.12.003>
60. Boursier CM, Bosco D, Coulibaly A, Negre M. Are traditional neem extract preparations as efficient as a commercial formulation of azadirachtin A?. *Crop Prot.* 2011; 30(3):318-322. Available from: <https://doi.org/10.1016/j.croppro.2010.11.022>
61. Anjali CH, Sharma Y, Mukherjee A, Chandrasekaran N. Neem oil (*Azadirachta indica*) nanoemulsion--a potent larvical agent against *Culex quinquefasciatus*. *Pest Manag. Sci.* 2012; 68(2):158-163. Available from: <https://doi.org/10.1002/ps.2233>
62. Yang Z, Baldermann S, Watanabe N. Recent studies of the volatile compounds in tea. *Food Res. Int.* 2013; 53(2):585-599. Available from: <https://doi.org/10.1016/j.foodres.2013.02.011>
63. Filomeno CA, Barbosa LCA, Teixeira RR, Pinheiro AL, Farias ES, Silva EMP, Picanço MC. *Corymbia* spp. and *Eucalyptus* spp. essential oils have insecticidal activity against *Plutella xylostella*. *Ind. Crops Prod.* 2017; 109(0):374-383. Available from: <https://dx.doi.org/10.1016/j.indcrop.2017.08.033>
64. Ananenka A. Insecticides and invertebrate neurophysiology: testing the efficacy of caffeine as an insecticide. 2018. Thesis (PhD in Biology). Montreal: Concordia University, 2018. Available from: https://digitalcommons.csp.edu/cup_commons_undergrad/124
65. Tamburro M, Sammarco ML, Trematerra P, Colacci M, Ripabelli G. Potential role of *Alphitobius diaperinus* Panzer (Insecta, Coleoptera) in poultry farm as transmission vector of bacterial pathogens in broilers and humans. *J. Appl. Microb.* 2022; 74(6):883-892. Available from: <https://doi.org/10.1111/lam.13679>
66. Rumbos C, Pantazis I, Athanassiou C. Population growth of the lesser mealworm, *Alphitobius diaperinus* (Panzer) (Coleoptera: Tenebrionidae), on various commodities. *J. Econ. Entomol.* 2020; 113:1001-1007. Available from: <https://doi.org/10.1093/jee/toz313>
67. Oliveira DGP, Cardoso RR, Mamprim AP, Angeli LF. Laboratory and field evaluation of a cypermethrin-based insecticide for the control of *Alphitobius diaperinus* Panzer (Coleoptera: Tenebrionidae) and its in-vitro effects on *Baeouveria bassiana* bals. vuill. (Hypocreales: Cordycipitaceae). *Braz. J. Poult. Sci.* 2016; 18(3):371-380. Available from: <https://dx.doi.org/10.1590/1806-9061-2015-0115>
68. Souza CJ, Barbosa FM, Marujo MM, Santos ET, Domingues CH, Oliveira D, Sgavioli S. Effect of cypermethrin on the control of lesser mealworm (*Alphitobius diaperinus*) and broiler performance. *Pesq. Vet. Bras.* 2021; 41:e06859. Available from:

- <https://doi.org/10.1590/1678-5150-PVB-6859>
69. Tomberlin JK, Richman D, Myers H. Susceptibility of *Alphitobius diaperinus* (Coleoptera: Tenebrionidae) from broiler facilities in Texas to four insecticides. J. Econ. Entomol. 2008; 101(2):480-483. Available from: <https://dx.doi.org/10.1603/0022-0493>
70. Hickmann F, Morais AF, Bronzatto ES, Giacomelli T, Guedes JVC, Bernardi O. Susceptibility of the lesser mealworm, *Alphitobius diaperinus* (Coleoptera: Tenebrionidae), from broiler farms of southern Brazil to insecticides. J. Econ. Entomol. 2018; 111(2):980-985. Available from: <https://dx.doi.org/10.1093/jee/toy059>
71. Gurib-Fakim A. Medicinal plants: traditions on yesterday and drugs of tomorrow. Mol. Aspects Med. 2006; 27(1):1-93. Available from: <https://dx.doi.org/10.1016/j.mam.2005.07.008>
72. López MD, Pascual-Villalobos MJ. Mode of inhibition of acetylcholinesterase by monoterpenoids and implications for pest control. Indust. Crops. Products. 2010; 31(2):284-288. Available from: <https://dx.doi.org/10.1016/j.indcrop.2009.11.005>