

Preparation and evaluation of azithromycin binary solid dispersions using various polyethylene glycols for the improvement of the drug solubility and dissolution rate

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Azithromycin is a water-insoluble drug, with a very low bioavailability. In order to increase the solubility and dissolution rate, and consequently increase the bioavailability of poorly-soluble drugs (such as azithromycin), various techniques can be applied. One of such techniques is "solid dispersion". This technique is frequently used to improve the dissolution rate of poorly water-soluble compounds. Owing to its low solubility and dissolution rate, azithromycin does not have a suitable bioavailability. Therefore, the main purpose of this investigation was to increase the solubility and dissolution rate of azithromycin by preparing its solid dispersion, using different Polyethylene glycols (PEG). Preparations of solid dispersions and physical mixtures of azithromycin were made using PEG 4000, 6000, 8000, 12000 and 20000 in various ratios, based on the solvent evaporation method. From the studied drug release profile, it was discovered that the dissolution rate of the physical mixture, as the well as the solid dispersions, were higher than those of the drug alone. There was no chemical incompatibility between the drug and polymer from the observed Infrared (IR) spectra. Drug-polymer interactions were also investigated using Differential Scanning Calorimetry (DSC), Powder X-Ray Diffraction (PXRD) and Scanning Election Microscopy (SEM). In conclusion, the dissolution rate and solubility of azithromycin were found to improve significantly, using hydrophilic carriers, especially PEG 6000.

Uniterms: Azithromycin/solubility. Azithromycin/dissolution. Solid dispersion. Polyethylene glycols.

A azitromicina é um fármaco insolúvel em água, com biodisponibilidade muito baixa. A fim de aumentar a taxa de solubilidade e dissolução e, consequentemente, aumentar a biodisponibilidade de fármacos fracamente solúveis (tais como azitromicina), várias técnicas podem ser aplicadas. Uma dessas técnicas é a "dispersão sólida", frequentemente usada para melhorar a taxa de dissolução de compostos fracamente solúveis em água. Devido à baixa taxa de solubilidade e de dissolução, este fármaco não tem biodisponibilidade adequada. Portanto, o principal objetivo desta pesquisa foi o de aumentar a taxa de solubilidade e dissolução da azitromicina, preparando a sua dispersão sólida, utilizando diferentes glicóis de polietileno (PEG). As dispersões sólidas e as misturas físicas de azitromicina foram preparadas utilizando PEG 4000, 6000, 8000, 12000 e 20000, em várias proporções, com base no método de evaporação do solvente. O perfil de liberação do fármaco foi estudado e verificou-se que tanto a taxa de dissolução da mistura física quanto as dispersões sólidas foram maiores do que as do fármaco sozinho. Espectros de IR não revelaram incompatibilidade química entre o fármaco e o polímero. Interações fármaco-polímero também foram investigadas usando calorimetria diferencial de varredura (DSC), Difração de Raios X (PXRD) e Microscopia Eletrônica de Varredura(SEM). Em conclusão, a taxa de dissolução e a solubilidade da azitromicina melhoraram, de forma significativa, utilizando suportes hidrofilicos, especialmente PEG 6000.

Unitermos: Azitromicina/solubilidade. Azitromicina/dissolução. Dispersão sólida. Polietilenoglicóis.

INTRODUCTION

Azithromycin (Figure 1) is a semi-synthetic antibiotic obtained from erythromycin. It is a Macrolide antibiotic. This drug is a nitrogen-containing macrolide (Azalide), with indications and usage similar to Erythromycin. All macrolides, such as azithromycin inhibit RNA dependent protein synthesis by binding reversibly to the 50S ribosomal subunits of susceptible microorganisms (Patil et al., 2011; Montejo-Bernardo et al., 2005; Beckers et al., 1995; Champney, Burdine, 1998). This antibiotic is one of the world's best-selling antibiotics, and is currently the world's most widely consumed antibiotic (WIPO, 2012). The dihydrate form of this drug is a white crystalline powder, with a molecular formula of C₃₈H₇₂N₂O₁₂.2H₂O and molecular weight of 748.984 g.mol⁻¹ (Moffat et al., 2011; USP, 2007; Amidon, 1995). It is used orally for the treatment of bronchitis, certain types of skin infections, sore throat, pharyngitis, tonsillitis, nosocomial infections and pneumonia. The excellent activity against Chlamydia trachomatis is its unique feature (Sweetman, 2009). It is prescribed for the treatment of respiratory-tract infections (including otitis media) and also for skin and soft-tissue infections. Azithromycin can also be used for prophylaxis, and act as a component of drug regimen in the treatment of Mycobacterium avium complex (MAC) infections. It has activity against gram-positive organisms and also offers an increased gram-negative coverage over erythromycin and clarithromycin (Moffat et al., 2011; Sweetman, 2009; Palanisamy, Khanam, 2011).

FIGURE 1 - Chemical structure of azithromycin (Adeli, 2014).

Poorly water-soluble drugs could create problems regarding decreased drug effectiveness. Majority of the drugs, which are soluble in small amounts of water, have low bioavailability. The effort to improve the dissolution and solubility of a poorly water-soluble drug remains one of the most challenging tasks in drug development. Many approaches are used to increase the dissolution rate of

drugs, including solid dispersion, chemical modification, micronization, co-solvency, complexation, pH adjustment, and micellar-solubilization.

The use of solid dispersions in improving the solubility of poorly water-soluble drugs dates back to 1961 (Sekiguchi, Obi, 1961; Patil et al., 2013; Damian et al., 2000). Solid dispersions could be prepared by different methods, including melting, solvent and melting-solvent techniques (Patil et al., 2013; Chiou, Riegelman, 1971). One common method in preparing solid dispersions is the melt-congealing or solvent evaporation method (Al-Hamidi et al., 2010; Han et al., 2011). The transformation of drugs in solid dispersions from amorphous form to crystalline form is a critical obstacle in making solid dispersions and a practical approach in the pharmaceutical industry. However, the utilization of surface adsorbents can be a useful approach to tackle this problem (Li et al., 2011; Patil, Gaikwad, 2011; Leuner, Dressman, 2000; Adeli, Mortazavi, 2014). It involves the formation of eutectic mixtures of drug with water-soluble carriers, by melting their physical mixtures. After dissolving the carrier, the drug precipitates in a finely divided state in water. Different mechanisms are considered for drug solubility and dissolution rate enhancement, using solid dispersions. Particle size reduction of drug to submicron size and the consequent increase in surface area seem to be the most important (Okonogi, Puttipipatkhachorn, 2006; Mura et al., 2005; Craig, 2002).

The solid dispersion technique has been used for a wide variety of poorly water-soluble drugs such as irbesartan (Adeli, 2015), nimesulide (Babu et al., 2003) and ursodeoxycholic acid (Okonogi et al., 1997). One major problem associated with azithromycin is its very poor solubility (belongs to BCS class II) in biological fluids which accounts for the poor bioavailability, upon its oral administration. In this study, the solid dispersion method was utilized in order to increase the dissolution rate and solubility of azithromycin. Although there have been previous studies on this technology for over half-a-century, its application in drug development has attained greater importance since early 1990s owing to the widespread application of combinatorial chemistry and high-throughput screening in drug discovery, which favors the selection of poorly water-soluble new chemical entities (NCEs) (Vasanthavada et al., 2008; Pudipeddi et al., 2006).

MATERIAL AND METHODS

Material

Azithromycin dihydrate, PEG 4000, PEG 6000,

PEG 8000, PEG 12000 and PEG 20000 were purchased from Fluka, USA. Sodium hydrogen phosphate, sodium hydroxide and ethanol 96% were purchased from Merck, USA. All the chemical reagents used were of analytical grade.

Methods

Preparation of physical mixtures

With great precision, 100 mg of pure azithromycin and appropriate amounts of PEG (4000, 6000, 8000, 12000 and 20000) in various drug-to-carrier weight ratios were weighed and thoroughly blended in a glass mortar for 5 min (as an example, PM1 (1:1) was prepared by 100 mg of azithromycin and 100 mg of PEG 4000, PM9 (1:7) was prepared by 100 mg of azithromycin and 700 mg of PEG 6000, similarly to PM25 (1:9), which was prepared by 100 mg of azithromycin and 900 mg of PEG 20000). The composition of various batches is shown in Table I. The physical mixtures were prepared by weighing the calculated amounts of azithromycin, followed by trituration mixture of the carrier (PEG) with the drug in a glass mortar. The samples prepared were kept for further studies.

Preparation of solid dispersions

Solid dispersions of azithromycin were prepared in drug: carrier weight ratios as 1:1, 1:3, 1:5, 1:7 and 1:9, using the solvent evaporation method. 100 mg of azithromycin dihydrate was dissolved in 10 mL of 96% ethanol and the carrier was then added and dissolved at 40 °C, following continuous mixing on a hot plate to obtain a clear solution. Afterwards, the solvent was allowed to evaporate. The prepared solid dispersions (Table I) were collected, sieved, and stored in desiccators until subsequent analysis. The process of evaporation was opted until a constant weight was obtained. Overall, the yield was approximately 98%.

Analysis of azithromycin dihydrate

Azithromycin was analyzed at 215 nm (USP, 2007; IP, 2007) using a UV spectrophotometer. The standard curve for the determination of drug was constructed by dissolving it in phosphate buffer pH = 6.0, in concentration range of 2 to 30 μ g/mL. In this concentration range, good linearity was observed (R² = 0.9998). The graph obeyed the Beer-Lambert's law in the selected concentration range.

Determination of drug content

Drug content was determined by dissolving accurately weighed quantities of binary systems of drug,

carrier, and adsorbent in pH = 6.0 buffer. The solutions were filtered and diluted appropriately followed by a spectrophotometrical measurement of samples at 215 nm using SHIMADZU-UVmini 1240 Spectrophotometer, Japan (USP, 2007; IP, 2007).

Saturation solubility studies

To evaluate the increase in drug solubility, solid dispersions and the physical mixtures, saturation solubility test was carried out. Excess amounts (approximately 100 mg) of the drug was added to 100 mL of phosphate buffer (pH = 6.0). Samples were stirred at 200 RPM at 25 °C for 24 h, filtered, suitably diluted and analysed by a UV spectrophotometer at 215 nm.

Dissolution studies

In vitro release studies of azithromycin, solid dispersions, and the physical mixtures were carried out in a USP Apparatus 2 (Erweka DT6R, Germany) at 100 rpm in 900 mL of phosphate buffer (pH = 6.0) maintained at $37.0\pm0.5\,^{\circ}\mathrm{C}$ (USP, 2007). Dissolution studies were carried out using 100 mg of the pure drug and an equivalent amount of individual sample preparations. Aliquots of 5 mL were withdrawn at specified time intervals of 5, 10, 15, 20, 30, 45 and 60 min and replaced with equal volumes of fresh medium. The resultant samples were filtered through Whatman filter paper and analyzed spectrophotometrically at 215 nm, for the determination of dissolved drug. The dissolution studies were carried out in triplicate (Deepti, Madan, 2007).

Dissolution Efficiency (DE)

Dissolution efficiency (DE) is a parameter for evaluation of *in vitro* dissolution data. It is the area under the dissolution curve divided by the rectangular area that shows 100% of the solute at a certain time. This parameter is equal to the total area under the curve of the dissolution curve, at time "t" (between two time-points, expressed as a percentage of the curve at maximum dissolution). DE can also be expressed as percentage of the area of the rectangle described by 100% dissolution at the same time. Dissolution efficiency (DE%) was calculated to compare the dissolution data (Anderson *et al.*, 1998). Where:"y" is the percent drug dissolved at the time "t".

Dissolution Efficiency (DE%) =
$$\left(\frac{\int_0^t y \cdot dt}{y_t^{100}}\right) \cdot 100$$

Infrared (IR) spectroscopy

The IR spectra of azithromycin, solid dispersion

TABLE I - Drug content and saturation solubility studies of pure Azithromycin, solid dispersions and physical mixtures (n = 3, mean \pm SD)

Carrier	Formulation	Preparation	Drug: Carrier (w/w)	Theoretical drug content		Assayed drug content		Saturation
	code				Expressed (%)	Amount (mg)	Expressed (%)	solubility (μg/mL)
	Azithromycin	Pure drug	1:0	100	100	99.92 ± 1.27	99.92	85 ± 1.08
	SD1	Solid dispersion	1:1	50	100	49.82 ± 0.33	99.64	185 ± 1.24
	SD2	Solid dispersion	1:3	25	100	24.77 ± 0.22	99.08	227 ± 2.04
	SD3	Solid dispersion	1:5	16.67	100	16.44 ± 0.13	98.62	235 ± 1.91
PEG 4000	SD4	Solid dispersion	1:7	12.5	100	12.26 ± 0.07	98.08	241 ± 1.45
	SD5	Solid dispersion	1:9	10	100	9.76 ± 0.07	97.57	243 ± 1.88
	PM4	Physical mixture	1:7	12.5	100	12.12 ± 0.16	96.96	126 ± 1.62
	SD6	Solid dispersion	1:1	50	100	49.81 ± 0.38	99.62	188 ± 1.43
	SD7	Solid dispersion	1:3	25	100	24.77 ± 0.10	99.08	229 ± 1.22
	SD8	Solid dispersion	1:5	16.67	100	16.43 ± 0.11	98.56	237 ± 1.60
PEG 6000	SD9	Solid dispersion	1:7	12.5	100	12.25 ± 0.04	98.00	253 ± 0.95
	SD10	Solid dispersion	1:9	10	100	9.75 ± 0.04	97.50	249 ± 1.14
	PM9	Physical mixture	1:7	12.5	100	12.37 ± 0.20	98.96	127 ± 2.06
	SD11	Solid dispersion	1:1	50	100	49.83 ± 0.37	99.66	170 ± 1.25
	SD12	Solid dispersion	1:3	25	100	24.81 ± 0.19	99.24	195 ± 1.50
	SD13	Solid dispersion	1:5	16.67	100	16.46 ± 0.09	98.74	209 ± 1.18
PEG 8000	SD14	Solid dispersion	1:7	12.5	100	12.27 ± 0.06	98.16	235 ± 1.56
	SD15	Solid dispersion	1:9	10	100	9.77 ± 0.06	97.70	228 ± 1.05
	PM14	Physical Mixture	1:7	12.5	100	12.37 ± 0.20	98.96	131 ± 2.08
PEG 12000	SD16	Solid dispersion	1:1	50	100	49.82 ± 0.52	99.64	181 ± 1.88
	SD17	Solid dispersion	1:3	25	100	24.80 ± 0.20	99.20	202 ± 1.63
	SD18	Solid dispersion	1:5	16.67	100	16.45 ± 0.09	98.68	217 ± 1.21
	SD19	Solid dispersion	1:7	12.5	100	12.27 ± 0.11	98.16	233 ± 1.70
	SD20	Solid dispersion	1:9	10	100	9.77 ± 0.07	97.70	230 ± 1.97
	PM19	Physical mixture	1:7	12.5	100	12.36 ± 0.17	98.88	141 ± 1.99
PEG 20000	SD21	Solid dispersion	1:1	50	100	49.85 ± 0.51	99.70	152 ± 1.55
	SD22	Solid dispersion	1:3	25	100	24.83 ± 0.30	99.33	167 ± 2.01
	SD23	Solid dispersion	1:5	16.67	100	16.48 ± 0.14	98.86	186 ± 1.60
	SD24	Solid dispersion	1:7	12.5	100	12.29 ± 0.07	98.32	207 ± 1.11
	SD25	Solid dispersion	1:9	10	100	9.77 ± 0.06	97.70	228 ± 1.49
	PM25	Physical mixture	1:9	10	100	9.85 ± 0.12	98.50	149 ± 1.87

and the physical mixture were obtained using an IR spectrometer (PerkinElmer 843 System, USA). About 2 to 3 mg of the test sample was mixed with dry KBr and the spectra were obtained over a wavenumber range of 4000 to 200 cm⁻¹. An average of 20 scans was taken.

Differential scanning calorimetry (DSC)

Differential scanning calorimetry thermograms of pure the pure drug, PEG 6000, solid dispersions and all the other preparations were obtained using a SHIMADZU DSC-60instrument (Japan). The calorimeter was calibrated using Indium as standard at 156.6 ± 1 °C.

Accurately weighed samples (6.00 mg) were crimped in aluminum pans and heated from 20.0 to 200.0 °C at a heating rate of 10 °C/min in nitrogen atmosphere. An empty sealed aluminum pan was used as the reference.

Powder X-Ray Diffraction (PXRD)

Powder X-ray diffraction patterns of plain azithromycin, pure PEG 6000, solid dispersions of Azithromycin and the physical mixtures were recorded using a powder X-ray diffract meter (PHILIPS PW 1800 X-Ray Diffraction Machine, The Netherlands) with a copper tube anode over the interval 5-90° $2\theta^{-1}$. The

operational parameters were as follows: generational tension (voltage) of 45 kV; generator current of 40 mA; scans step time of 9 sec⁻¹ and scan step size of 0.008° (2θ).

Scanning Electron Microscopy (SEM)

The surface morphology of samples was determined using an analytical scanning electron microscope (VEGA\\TESCAN-LMU, Czech Republic, equipped with thermo-emission cathode [Balzers Union Ltd, Balzers, Lichtenstein]). The samples were placed on a sample disc carrier carbon stub (10 mm diameter, 3 mm height) and coated with gold under vacuum (0.25 Torr). The images were generated using a 30 kV electron beam.

Statistical analysis

The data obtained from dissolution and solubility studies were statistically compared using the analysis of variance [ANOVA] (Hinkelmann, Kempthorne, 2008).

RESULTS AND DISCUSSION

Drug content and saturation solubility studies

Table I presents the results of drug content and saturation solubility test of samples. The calculated percentage of drug content for drug-PEG 6000 was between the ranges of 97.50% to 99.62%, also the drug content of the drug-PEG 4000SDs was in the range of 97.57% to 99.64%. All determinations were presented as mean \pm SD of three measurements. The maximum solubility of azithromycin in phosphate buffer medium (pH = 6.0) was observed as $253 \pm 0.95 \mu g/mL$ for the SD9 (azithromycin:PEG 6000; ratio weight of 1:7). Also, saturation solubility of SD4 (drug:PEG 4000) with a weight ratio of 1:7 was about 241 \pm 1.45 μ g/mL (Table I). Further studies showed that SD9 had the maximum drug solubility. The SD21 (azithromycin:PEG 20000) demonstrated a minimum drug solubility. Consequently, the drug-PEG 6000 or the drug-PEG 4000 solid dispersions, both with the same ratio of 1:7 (w/w)were considered as the best formulations. The probable reason for the saturation solubility improvement in their formulations was the locally imposed solubilizing activity of the carriers used (Bashiri-Shahroodi et al., 2008; Chaud et al., 2010). Reduced drug particle size, decreased particle aggregation and higher wettability of drug in the presence of hydrophilic carrier could also be considered as the most probable mechanisms for enhanced dissolution rate of azithromycin from solid dispersion systems. The physical mixtures (PMs) were prepared as controls of solid dispersion (SDs) formulations. Based on the Table I, SD25 (of PEG 20000) with a further saturation solubility compared to other formulations of this group, was selected, thus PM25 was prepared. In addition, prepared physical mixtures of drug-polymer having higher molecular weights, such as PEG 20000 (PM25) due to high hydrophilicity and having further -OH functional group, demonstrated higher saturation solubility (Adeli, 2014, Valizadeh *et al.*, 2004).

In vitro dissolution studies

Table II and Figure 2 showed the results of in vitro azithromycin dissolution. The dissolution of pure azithromycin was rather low. Hence, the cumulative amount of dissolved drug after 60 min was about 22%. The dissolution rate of the azithromycin physical mixture was improved, but to a lesser extent than that of the solid dispersions. Figure 2B illustrates the comparative drug release profile of various solid dispersions and the physical mixtures of azithromycin with PEG 6000. The best result was observed by PEG 6000 group. The SD9 formulation showed the greatest amount of drug release (49.10% at 60 min) among the investigated solid dispersion samples (Figures 2A-2E). The SD21 formulation showed the least amount of drug release (30.50% at 60 min) among studied SD formulations. The formulations containing PEG 4000 (SD4) and PEG 6000 (SD9) dissolved well after 30 min and showed higher dissolution efficiencies, in comparison with pure azithromycin (DE₃₀ for SD4 = 30.91and for SD9 = 31.44, which were the highest DE values observed). However, SD9 was selected as the best formulation because of the slightly better results obtained. Additional tests were performed on this formulation. For PEG 20000 polymer, SD24 was selected (35.3% at 60 min), therefore PM24 was prepared. Perhaps, due to establish sink condition and for reasons that will be mentioned, SD24 (and consequently PM24) has further dissolution compared to other formulations of this group. Generally, there seems to be an increase in the solubility of azithromycin in the presence of hydrophilic carriers surrounding the drug particles. Actually, increasing the PEG (4000, 6000, 8000, 12000 and 20000) concentration in solid dispersions, up to an optimum amount, resulted in an improved drug dissolution. However, further increase in polymer content beyond this optimum amount showed no additional improvement on drug dissolution. This might be attributed to the viscous layer formed around the solid particles due to the presence of higher polymer concentrations, resulting in a decrease in the diffusion coefficient and dissolution of the drug (Giorgetti et al., 2014; Narasaiah et al., 2011).

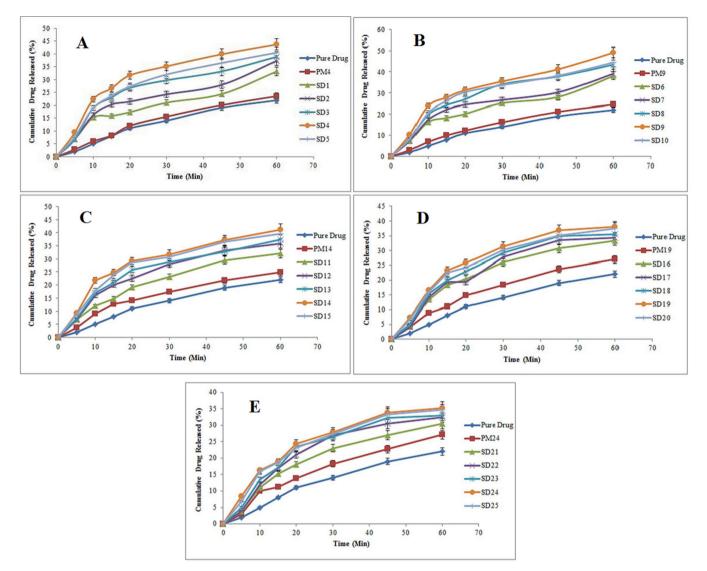


FIGURE 2 - *In vitro* dissolution release profiles of pure azithromycin, solid dispersions and the physical mixtures containing: (A) PEG 4000, (B) PEG 6000, (C) PEG 8000, (D) PEG 12000, (E) PEG 20000; (n = 3, mean ± SD).

Based on these results, there was significant improvement in the dissolution profile of Azithromycin in all the prepared solid dispersions (p < 0.05). A similar trend was observed with physical mixtures, although in a smaller extent. The probable reason for the improvement in the dissolution rate in the physical mixtures was the local solubilizing effect of PEG 6000 (Patil et al., 2013). Overall, the solubility studies showed that the solid dispersion method could help to increase the solubility of Azithromycin, particularly when used alongside PEG 6000. Consequently, the dissolution of Azithromycin solid dispersion using PEG 6000 (at a drug:polymer ratio of 1:7) experienced up to a 100% increase, in comparison to the pure drug (2.23 times more). Comparison of the effect of increasing the drug solubility, when using different PEGs in this study is as follows:

PEG 6000 > PEG 4000 > PEG 8000 > PEG 12000 > PEG 20000

The dissolution rate of poorly water soluble drugs included within solid dispersions is influenced by various factors, including the carrier type, viscosity of carriers around drug particles within the dissolution medium and the drug: carrier ratio. The chemical nature of the drug is also important. Therefore, it is not possible to propose a specific process for prediction of the increase in the dissolution rate of solid dispersion samples. The increase in solubility or dissolution can be explained by three reasons: the high hydrophilicity of the polymer, the chemical properties of drugs and a reduction in the particle size of the drug.

TABLE II - Dissolution efficiency of pure azithromycin, solid dispersion samples and physical mixtures (n = 3, mean \pm SD)

Formulation code	Preparation	Drug:Polymer (w/w)	DE ₁₀ (%)	DE ₃₀ (%)
Azithromycin	Pure Drug	1:0	4.50 ± 1.12	14.32 ± 0.95
SD1	Solid Dispersion	1:1	14.22 ± 0.93	23.07 ± 0.94
SD2	Solid Dispersion	1:3	15.45 ± 0.84	23.89 ± 0.76
SD3	Solid Dispersion	1:5	15.80 ± 0.59	27.43 ± 0.76
SD4	Solid Dispersion	1:7	20.76 ± 0.48	30.91 ± 1.38
SD5	Solid Dispersion	1:9	16.16 ± 0.83	29.22 ± 1.04
PM4	Physical Mixture	1:7	14.73 ± 0.92	14.47 ± 1.52
SD6	Solid Dispersion	1:1	14.28 ± 1.37	23.24 ± 0.93
SD7	Solid Dispersion	1:3	15.59 ± 0.39	25.03 ± 0.87
SD8	Solid Dispersion	1:5	15.65 ± 2.19	28.07 ± 0.46
SD9	Solid Dispersion	1:7	18.90 ± 1.48	31.44 ± 0.49
SD10	Solid Dispersion	1:9	17.72 ± 1.38	29.44 ± 1.04
PM9	Physical Mixture	1:7	5.03 ± 2.10	15.18 ± 1.39
SD11	Solid Dispersion	1:1	9.89 ± 1.10	20.63 ± 1.47
SD12	Solid Dispersion	1:3	13.57 ± 0.49	23.35 ± 0.69
SD13	Solid Dispersion	1:5	14.61 ± 0.38	26.78 ± 037
SD14	Solid Dispersion	1:7	19.26 ± 1.69	27.12 ± 0.36
SD15	Solid Dispersion	1:9	15.85 ± 1.46	26.39 ± 2.51
PM14	Physical Mixture	1:7	7.87 ± 1.11	15.80 ± 1.04
SD16	Solid Dispersion	1:1	11.07 ± 1.17	21.64 ± 1.36
SD17	Solid Dispersion	1:3	13.08 ± 0.84	24.17 ± 1.32
SD18	Solid Dispersion	1:5	14.26 ± 1.05	25.09 ± 2.41
SD19	Solid Dispersion	1:7	14.67 ± 1.73	26.87 ± 1.39
SD20	Solid Dispersion	1:9	14.53 ± 0.47	26.12 ± 0.59
PM19	Physical Mixture	1:7	6.48 ± 2.08	16.65 ± 0.34
SD21	Solid Dispersion	1:1	9.70 ± 1.74	20.46 ± 0.68
SD22	Solid Dispersion	1:3	9.88 ± 1.04	22.42 ± 0.37
SD23	Solid Dispersion	1:5	11.10 ± 0.49	23.08 ± 0.22
SD24	Solid Dispersion	1:7	12.84 ± 1.94	24.37 ± 0.50
SD25	Solid Dispersion	1:9	12.68 ± 1.43	25.15 ± 1.23
PM24	Physical Mixture	1:7	8.46 ± 2.01	17.07 ± 1.47

IR studies

The IR spectra showed that a particular change or interaction does not occur between carriers and the drug. Figure 3 depicts the IR spectra of pure azithromycin, PEG 6000 (as the selected polymer), the physical mixture and solid dispersions. The spectrum of azithromycin (Figure 3A) shows characteristic peaks at 1724 cm⁻¹ (C=O stretch), 1190 cm⁻¹ (C-O-C asymmetrical stretching) and 1049 cm⁻¹ (C-O-C symmetrical stretching). IR spectrum of the PEG 6000 also displayed characteristic peaks at 1108 cm⁻¹

(for C-O-C stretch), 2907 cm⁻¹ (for C-H stretch), and 3244 cm⁻¹ (O-H stretch) (Figure 3B). On the other hand, the peak related to C-O-C stretching bond of the drug in 1049 cm⁻¹ for samples PM9, SD9 and SD4 shifted to higher wave numbers. This can be due to the formation of *Van der Waals* forces between azithromycin and carrier. There was no substantial shifting in the position of functional groups (Stuart, 2002). According to the IR spectra of solid dispersions (such as SD9, Figure 3C) and physical mixtures (such as PM9, Figure 3D). Figures 3E and 3F illustrates the IR spectra of SD4 and PM4 (as selected

samples of PEG 4000), respectively. From the figures, there was no undesirable chemical interaction between the drug and polymer. The main peaks of the drug and polymer were clearly distinguished in solid dispersion samples and physical mixtures.

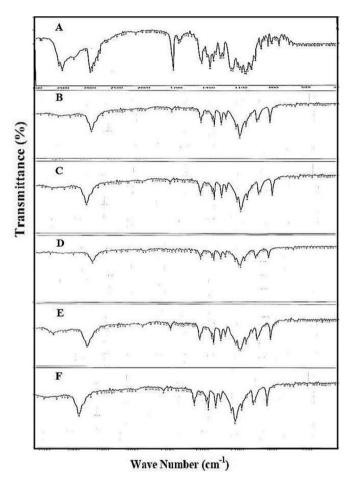


FIGURE 3 - IR Spectra of: (A) azithromycin, (B) PEG 6000, (C) SD9, (D) PM9, (E) SD4 and (F) PM4.

DSC studies

DSC as important thermal analyses techniques were used to determine the thermodynamic characteristics of SD samples. This test was performed for the thermodynamic behavior of anhydrous polymorphs of Azithromycin and carriers in the present study. The DSC thermogram of azithromycin, PEG 6000, SD9 (as the selected solid dispersion system) and PM9 (as the physical mixture) have been presented in Figure 4. The differential scanning calorimetry shows a sharp endothermic fusion peak between 109.45 °C to 127.77 °C, which corresponds to the melting point of azithromycin (Figure 4A and Table III). The sharp endothermic peak of PEG 6000 was exhibited at 62.14°C (Figure 4B). A melting endothermic peak

with decomposition at 144.28 °C can also be depicted. In addition, there was no interaction between the drug and the polymer according to the observed endothermic peaks. The melting peak of pure azithromycin disappeared in solid dispersion and physical mixtures, presumably due to a transformation in drug structure, from a crystalline to an amorphous form (Figure 4).

PXRD studies

Azithromycin crystals display various diffraction peaks (Figure 5A) due to their crystalline structure. The PXRD pattern revealed sharp peaks at diffraction angles (2θ) of 9.56°, 7.72°, 20.67°, 19.59°, 23.80°, 17.31°, 28.68° and 26.55°. For PEG 6000, the 2θ value was 19.09° (Figure 5B). In the solid dispersion samples studied, there were no azithromycin peaks, and this may be attributed to structural transformation of the drug molecule from the crystalline from into an amorphous form (Figures 5C and 5E). However, some of the smaller peaks of PXRD of the major peaks of pure azithromycin can be detected in SD9, PM9 and other samples at the same 2θ value, but with much lower intensities, which affirms that azithromycin crystal do not completely change into the amorphous form in solid dispersion and physical mixtures, and some crystalline structure still remains in these samples during the preparation process (Figures 5C and 5E). Based on these results, the enhancement in azithromycin dissolution in the form of solid dispersions can be partly due to the structural changes, alongside the influence of the carrier itself as well as a reduction in particle size.

SEM studies

Surface morphological studies of the samples were carried out using SEM and illustrated in Figure 6. Cubic-shaped crystals with sharp edges constitute the morphology of crystalline Azithromycin powder. The morphology of PEG 6000 was needle-shaped. The solid dispersion process resulted in the formation of a significantly shorter and amorphous length of the PEG needles compared to the unaltered azithromycin alone (Figure 6). The SEM of solid dispersions reveals the existence of irregular particles with several microscopic cracks and crevices, which provides additional surface for deposition of drug particles. Drug molecules seem to be dispersed within the carrier matrix of the solid dispersions. Furthermore, a particle size reduction in drug molecules is also proposed. These events could help to enhance the release profile and dissolution rate of the drug molecules.

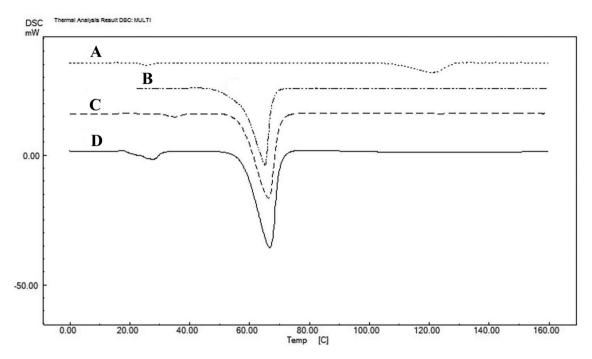


FIGURE 4 - DSC thermograms of: (A) azithromycin, (B) SD9, (C) PM9 and (D) PEG 6000.

TABLE III - Data obtained from the DSC thermograms of azithromycin, PEG 6000 solid dispersion, SD9 and physical mixture PM9 samples

Sample:	Azithromycin	PEG 6000	SD9	PM9
T _{onset} (°C):	22.34	21.46	57.82	31.17
	109.45	59.38	-	57.14

Physical stability studies

The selected formulations underwent accelerated stability studies, based on ICH guidelines. Study parameters were: 40 ± 2 °C and $75 \pm 5\%$ relative humidity for 120 days (Shel-Lab SMI2 Forced-Air Incubator, USA, Sheldon Manufacturing, Inc.). One hundred milligram samples were individually stored, kept in glass vials sealed with rubber plugs, followed by analysis of the drug content and physical changes of the stored samples at time intervals: 15, 30, 60, 90 and 120 days. During these time periods, the saturation solubility and dissolution rates of Azithromycin were re-examined. No significant change in the test samples was observed. Hence, the samples appeared to remain stable over the study period (Table IV).

Statistical analysis studies

The Pearson correlation test in order to evaluation of the relationship between used polymer percentage and solubility using SPSS software was carried out. Pearson's

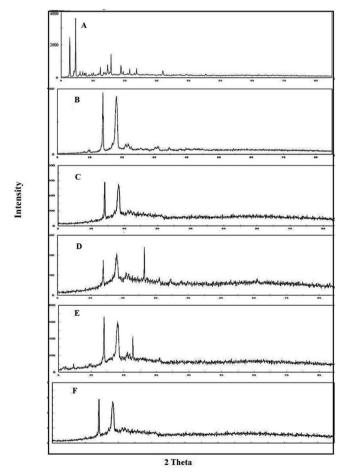


FIGURE 5 - PXRD Diffractograms of: (A) azithromycin, (B) PEG 6000, (C) SD9, (D) PM9, (E) SD4 and (F) PM4.

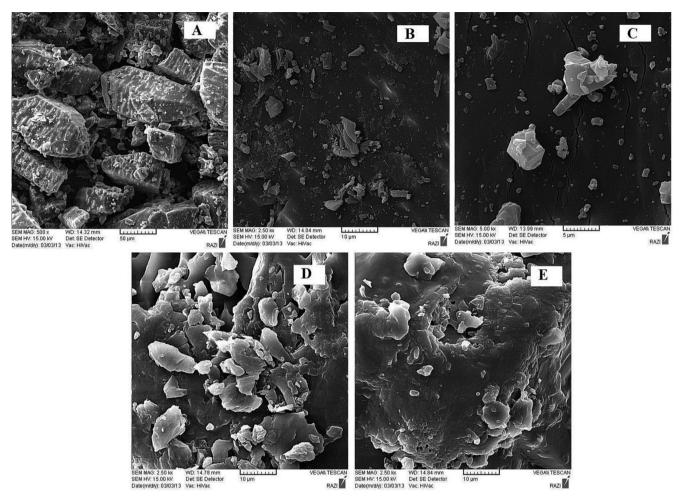


FIGURE 6 - SEM of: (A) azithromycin [magnification: 500×], (B) azithromycin [magnification: 2500×], (C) azithromycin [magnification: 5000×], (D) SD9 [magnification: 2500×] and (E) SD9 [magnification: 2500×].

TABLE IV - Drug content of selected formulations after undergoing stability studies based on the **ICH** guidelines for storage conditions, with was 40 ± 2 °C at $75 \pm 5\%$ RH (n = 3, mean \pm SD)

Time	Pure azithromycin (%)	SD4 (%)	SD9 (%)
0 days	99.92 ± 1.27	98.08 ± 2.17	98.00 ± 1.57
15 days	99.92 ± 1.02	98.06 ± 1.98	97.95 ± 1.09
30 days	99.91 ± 2.34	97.88 ± 0.95	97.91 ± 1.95
60 days	99.90 ± 0.14	97.75 ± 1.63	97.84 ± 1.03
90 days	99.89 ± 1.22	97.53 ± 2.00	97.44 ± 2.07
120 days	99.88 ± 0.78	97.29 ± 2.67	97.37 ± 1.97

r with alpha level of 0.05 for PEG 4000, PEG 6000, PEG 8000, PEG 12000 and PEG 20000 was calculated, the results are as follows: 0.7913, 0.8068, 0.8541, 0.8154 and 0.9309, respectively. In all the groups, solubility of the selected samples as SD4, SD9, SD14, SD19 and SD25 demonstrated significant differences when compared with other samples of the same group (p < 0.05).

CONCLUSIONS

In this investigation, the solvent evaporation approach was used for the preparation of Azithromycin solid dispersions. The solubility and the dissolution rate of the drug were significantly increased in all of the solid dispersion prepared with PEG. The physical

mixture samples had lower dissolution rates than the solid dispersion systems. The best result was obtained from solid dispersions containing azithromycin:PEG 6000 in a ratio of 1:7. The amount of azithromycin released from this solid dispersion sample (SD9) was more than 49% in 60 min. Greater wettability of the drug by this hydrophilic carrier, as well as an increase in surface area (Adeli, 2014) seems to be the main reasons for the increase in drug dissolution rate. Hence, the solid dispersion method, using a hydrophilic carrier such as PEG 6000, could be considered as an appropriate technique for dissolution enhancement of azithromycin, which is a poorly-soluble drug.

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