

Assessment of an anti-ageing structured cosmetic formulation containing goji berry

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Based on previous studies, it has been found that goji berry (GB), popularly known as a ‘miracle fruit’, has excellent antioxidant potential and can be used in the treatment of skin disorders associated with ageing. This study aimed to incorporate GB into a structured cosmetic in order to optimise its penetration. Stability studies of the formulation, determination of the antioxidant activity of the extract and of the formulation, rheological measurements, SAXS, polarised light microscopy and bioadhesion analyses were performed. The results indicated the antioxidant activity of the extract, which can be incorporated into an emulsified cosmetic formulation. The emulsified formulation containing the extract remained stable, even after being submitted to thermal and luminous stresses for 30 days. In addition, rheological tests revealed that the addition of the GB soft extract reduced the viscosity of the formulation, increasing thixotropy and deformation. These systems were characterised by SAXS as a lamellar phase, which was confirmed by polarised light microscopy. These highly organised structures indicate their excellent stability. *In vitro* bioadhesion experiments revealed that these formulations exhibited skin adhesion strength statistically similar to commercial anti-ageing formulation. These results suggest that this formulation has excellent potential to be used as a topical treatment for ageing.

Keywords: Goji Berry. Skin. Formulation. Lycium/drug effects. Skin Aging/immunology. Cosmetics/pharmacokinetics. Antioxidants/pharmacokinetics. Cosmetic Technology.

INTRODUCTION

The skin is a complex covering organ and is the largest organ in the human body, with functions such as pigmentation, keratogenesis, thermoregulation, sweating, defence and absorption. It consists of two layers: the outer epidermis outermost and the inner dermis. The hypodermis, or subcutaneous tissue, lies just below the dermis. This tissue is not part of the skin, but represents a region of union between the skin and other organs (Batistela, Chorilli, Leonardi, 2007; Alchorne, Abreu, 2008).

Over time, some changes occur in tissues due to ageing. In the skin, these changes are more visible. The most apparent signs of senile skin are atrophy, wrinkling, ptosis, lassitude, dyschromia, among others (Oriá *et al.*,

2003). The skin ageing process can occur due to various factors, including intrinsic or chronological ageing and extrinsic ageing (Velasco *et al.*, 2004). Changes in the genetic material of cells, including those that compose the skin, may occur during the skin ageing process. This phenomenon generates protein alterations and decreases cellular proliferation, with a consequent loss of elasticity and vigour. This process is accelerated by chemical and enzymatic oxidation in which there is the formation of free radicals (Hirata, Sato, Santos, 2004).

Free radicals are organic and inorganic molecules that contain one or more unpaired electrons, making them highly unstable and chemically reactive. The presence of free radicals is critical to the maintenance of normal physiological functions (Bianchi, Antunes, 1999). Conversely, antioxidants are compounds that inhibit or retard the oxidation of molecules by preventing chain oxidation reactions from starting or spreading. The antioxidant activity of a compound may be due to its

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oxide-reducing properties, with a role in the neutralisation of free radicals. Antioxidant agents can be both naturally occurring and synthetic (Degáspari, Waszczynskyj, 2004).

The main sources of antioxidants to the human body are food or dietary supplementation. However, other ways of affecting body homeostasis through antioxidants have been used and studied. Flavonoids, found in red fruits, have present phenolic groups in their chemical structure, with iron chelating power and action against free radicals (Silva, Degáspari, 2014). Among the red fruits, goji berry (*Lycium barbarum* L.) (GB), belonging to the Solanaceae family, has received attention in recent years, and is widely disseminated worldwide. It exhibits excellent antioxidant and immunomodulatory potential and has been known for a long time by the Chinese. Due to its characteristics, it has been denominated as a 'miracle fruit', 'super fruit' or 'super food' because it is part of a group of red fruits that have in their composition several secondary metabolites, among them flavonoids, along with bioactive effects. Flavonoids are one of the compounds responsible for the characteristic pigmentation found in fruits and vegetables and protect the body against oxidative damage (Jamin, 2009; Reeve *et al.*, 2010; Karp, 2012; Silva, Degáspari, 2014; Donno *et al.*, 2015).

Reeve *et al.* (2010) evaluated the potential of orally consuming GB juice to alter the sun damage caused to the skin by ultraviolet (UV) radiation. This evaluation was conducted in rats. The results showed that juice consumption at 5% was able to promote oedema reduction due to sunburn. In addition, the juice was able to protect against immunosuppression promoted by UV radiation.

Zhao and Bojanowski (2015) described the dermatological uses of *Lycium barbarum*. These authors found that, although this fruit is a strong candidate for dermatological use, few publications report its use. Some previously reported effects include antioxidant, anti-inflammatory and immunomodulatory activities (Zhao, Bojanowski, 2015). However, no publications exist in the literature about a nanostructured system containing this extract.

Thus, this study assessed the incorporation of a GB extract into a structured cosmetic system aiming to optimise its transport through the skin. Highly organised systems are more suitable than conventional cosmetic systems, since, due to the highly structural organisation of the formulation, they have greater stability (Chiari *et al.*, 2012a). Here, we report the design and characterisation of structured liquid crystalline cosmetic formulation incorporated with GB extract with antioxidant activity aimed at the future topical treatment or prevention of skin disorders associated with ageing.

MATERIAL AND METHODS

Goji berry extract preparation

Dehydrated fruits of goji berry were purchased in the market of Leme city, São Paulo, Brazil in November 2015.

After being selected, 130 g of fruits was weighed and macerated in 96% (v/v) ethanol solution for 2 days at room temperature. After this period, the extract was separated and another 200 mL of the same solvent was added to the fruits for a new extraction period (remaceration). This cycle was maintained for 14 days. The total volume of solvent used was 1000 mL.

The obtained extract was stored in an amber glass bottle at 5 ± 2 °C. At the end of the 14 days, the resulting filtrates were collected and evaporated with the aid of a rotavaporator at 60 ± 2 °C until complete elimination of the organic solvent (ethanol), thereby obtaining the goji berry soft extract (Cui *et al.*, 2011; Pai *et al.*, 2013).

The goji berry soft extract was packed in an amber glass vial and stored at -5 ± 2 °C until use.

Measurement of the antioxidant activity

The assessment of the antioxidant activity of GB extract was performed using the 1,1-diphenyl-2-picrylhydrazyl radical (DPPH) assay, according to methodology described by Mensor *et al.* (2001) and Falcão *et al.* (2006), modified by Chiari *et al.* (2012b).

One millilitre of the extract aqueous solution at various concentrations was added to 2.5 mL of DPPH methanolic solution (0.004%). The solutions were kept out of the light and after 30 minutes the absorbance of the solutions was determined at 515 nm.

Control solutions containing only 1 mL of water and 2.5 mL of the 0.004% methanolic solution of DPPH were used. The mean absorbance of these samples was used as the maximum absorbance, which was used to calculate the inhibition percentage (% DPPH inhibition) of the DPPH radical (Molyneux, 2004).

$$\%DPPH\ inhibition = \frac{(A_{max} - A_{test})}{A_{max}} \times 100,$$

A_{max} is the absorbance of DPPH at 515 nm in the absence of sample (control).

A_{test} is the absorbance of DPPH at 515 nm in the presence of sample.

The DPPH assay was performed in triplicate.

Design of the cosmetic formulation

For the formulation design, an extensive search was made in the literature regarding raw materials that favour the permeation of active principles (permeation promoters) in order to ensure the penetration of the GB extract through the stratum corneum.

After designing the composition of the formulations (Table I), they were prepared by the conventional technique of emulsions preparation. For this, the aqueous and oily phases were separately heated to 75 °C for further mixing and vigorous stirring until cooling. In the formulation containing GB extract, the soft extract was solubilised in 10% of the water (discounted from the total water content of the formulation). Right after preparation and cooling, the extract solution was mixed with vigorous agitation (Cefali *et al.*, 2015).

Measurement of the formulation antioxidant activity

This assay was performed with the same methodology as for the assessment of the antioxidant activity of the GB extract, but using as samples the formulation containing the extract. Thus, 0.5 g of the emulsion containing the GB soft extract was weighed and solubilised in 0.5 mL of ethanol. To the DPPH solution, 0.3% Tween-20 was added to facilitate dispersion of the formulation and to prevent turbidity. After the addition of 2.5 mL of the DPPH solution, a spectrophotometer reading was performed at 515 nm after 30 minutes of the

reaction. As a negative control, the base formulation (not containing the GB extract) was used, to assess interference from any raw material in the emulsion. Thus, the Amax used in the calculation of % of DPPH inhibition was the absorbance of the reaction with the DPPH solution and base formulation solubilised in ethanol (0.5 g of base formulation solubilised in 0.5 mL of ethanol) (Cefali *et al.*, 2015).

Analysis of the preliminary stability of the cosmetic formulation

The organoleptic characteristics (colour, odour and appearance), in addition to the pH values and relative density of the formulation, were monitored during a period of 30 days, keeping the samples at room temperature (24 ± 2 °C), at 38 ± 2 °C and at 4 ± 2 °C, to assess formulation stability (Brazil, 2004; Isaac *et al.*, 2008).

The pH was measured using a digital pH meter. For this, dispersions (10% in deionised water) of the formulation were prepared (Davis, 1977).

The relative density was measured with the aid of a pycnometer. For this, the mass of the empty pycnometer, of the pycnometer with water and of the pycnometer with the formulation was determined. The mass of the product contained in the pycnometer volume was divided by the mass of the water contained in the same volume (Brazil, 2004; Isaac *et al.*, 2008).

These determinations were performed 24 hours after the preparation of the emulsion (day 0) and after 7, 15 and 30 days (Brasil, 2004).

TABLE I - Percent composition (w/w) of test emulsions 1, 2 and 3

Raw material (INCI name)	Test formulation 1	Test formulation 2	Test formulation 3
Cetyl alcohol	5.00	3.00	3.00
Grape seed oil	-	-	3.00
Shea butter	2.00	2.00	-
Emulsifyin wax NF (Polawax NF®)	8.00	5.00	5.00
Cetearyl alcohol	0.50	0.50	0.50
Phenoxyethanol (and) Methylparaben (and) Ethylparaben (and) Propylparaben (and) Butylparaben (and) Isobutylparaben (Phenonip®)	0.50	0.50	0.50
BHT	0.05	0.05	0.05
Disodium EDTA	0.01	0.01	0.01
Glycerin	5.00	5.00	5.00
Ammonium Acryloyldimethyltaurate/VP Copolymer (Aristoflex® AVC)	1.00	1.00	1.00
Water	q.s.p. 100.00g	q.s.p. 100.00g	q.s.p. 100.00g

Rheological characterisation of the formulations

For the characterisation of the formulations (containing or not the GB extract) regarding the flow behaviour and spreadability, rheology was employed. For this, an AR2000ex rheometer was used. The assays were conducted with cone-plate type sensor (2° angulation and 52 mm gap) and data were analysed by Rheology Advantage Data Analysis software. The assays were conducted in duplicate.

The rheological behaviour was evaluated by determining:

- The flow property of the samples, with shear stress of 0-100 Pa for a period of 120 seconds for the ascending curve and shear stress of 100-0 Pa for 120 seconds for the descending curve;
- A stress sweep by subjecting samples to a constant frequency of 1 Hz using variable shear stresses of 0.1 to 10 Pa.
- A frequency sweep by subjecting samples to a constant shear stress of 1 Pa, using variable frequencies from 0.1 to 50 Hz.
- A creep and recovery assay, subjecting samples to a shear stress of 1 Pa for 200 seconds. Then, the stress was removed and the behaviour of the sample was followed for another 200 seconds (Isaac *et al.*, 2013b).

All the tests were performed at a temperature of 25 ± 1 °C, with the formulations also kept at room temperature. The graphs were plotted in the software Origin 7.0.

Assessment of the formulations using small angle X-ray scattering (SAXS)

The nanostructures of the phases that compose the base formulation and the formulation containing the GB extract were assessed using small angle X-ray scattering (SAXS) measurements.

Data collection was performed at the National Synchrotron Light Laboratory (Laboratório Nacional de Luz Síncrotron LNLS in Campinas), using the synchrotron light line for SAXS, equipped with a monochromator ($\lambda = 1.608\text{\AA}$) Si (1 1 1) that produces a horizontally focused beam of light. A vertical X-ray detector and a multichannel analyser were used to capture the intensity of the radiation scattered by the samples, $I(q)$, as a function of the scattering vector modulus q , $q = (4\pi/\lambda) \sin(\varepsilon/2)$, with ε as the scattering angle. The dispersion of the rays produced by the slits was subtracted from the total scattering intensity. Both intensities are described in relative units,

but for a quantitative comparison, they were normalised to the same experimental conditions (Manaia *et al.*, 2012).

Evaluation of formulations using polarised light microscopy

Samples of the formulation containing or not the GB extract were arranged under slides and coverslips, and were then observed under a polarised light microscope (Carl Zeiss Axioskop-40). The magnification used was 400x (Manaia *et al.*, 2012).

Analysis of the bioadhesion force

The bioadhesion strength of the formulations was analysed using a TAXT plus texture analyser (Stable Micro Systems®). Samples of the formulation containing or not the GB extract were analysed and a commercial anti-ageing formulation (Vitanol-A®) as the standard. This standard was chosen due to its similar characteristics to the formulation investigated in the study, i.e. it is an anti-ageing emulsion intended for facial skin care (Oshiro-Jr *et al.*, 2015; Oshiro *et al.*, 2018).

Dissected porcine ear skin was obtained from a slaughterhouse and dermatomed was used as a biological substrate at a thickness of 500 μm . Prior to the test, hair was removed with a pair of scissors and then adhered to the probe.

The sample to be assessed was previously packed in a cylindrical and shallow plastic bottle (2 cm in diameter and 1 cm high) that was placed under the probe. Thereafter, the lowering of the probe was run at a rate of 1 mm/s until the pig ear skin, adhered to the probe, came into contact with the surface of the formulation. Instantly, a downward force of 0.05 N was applied for a set time (30 s) to ensure intimate contact between the sample and the substrate. After this time, the probe was removed at a speed of 1 mm/s and, the force required to remove the substrate from the formulation surface was determined as the resultant force *versus* time. Assays were repeated five times. Data were compared using one-way ANOVA and Tukey's multiple comparison test ($p < 0.05$).

RESULTS AND DISCUSSION

Goji berry extract preparation

The GB soft extract was obtained by the maceration method, with the extractive solvent renewed every two days the (re-maceration) (Sajjadi *et al.*, 2016; Das Dores *et al.*, 2017). This practice aimed to avoid solvent

saturation, allowing the maximum extraction rate of the components of interest from the dehydrated fruits. Maceration, according to Timaco (2012), is capable of promoting the extraction of important substances such as phenolic compounds, polysaccharides and mineral elements.

After extract rotavaporation, 90 mL of soft GB extract with a dark yellow colour was obtained.

Measurement of the antioxidant activity

In order to assess the antioxidant activity, the GB soft extract in various concentrations was added to a 0.004% solution of DPPH, obtaining a colour variation that was measured spectrophotometrically, ranging from purple to yellow. After reading in a spectrophotometer at 515 nm, the results were plotted and are shown in Figure 1.

As seen in Figure 1, the antioxidant activity of the GB soft extract was proportional to its concentration. This was represented by the equation $y = -93.185x^2 + 185.88x + 0.1671$, which describes, through polynomial regression, the relationship between the extract concentration and the percentage of free radical inhibition. By means of this equation, the IC_{50} value of the GB soft extract was calculated, which in this case corresponded to 0.32% of the extract in solution. The IC_{50} is the concentration capable of inhibiting 50% of the free radicals present in the evaluated solution (Chiari *et al.*, 2012b).

This result aided in determining the concentration of GB soft extract to be added to the base formulation. Considering that 1% of the extract promotes approximately 100% inhibition of the DPPH radical, this concentration was multiplied by 2 for addition to the base emulsion, thus 2% was used (an approximately concentration). The reason for doubling the concentration capable of inhibiting 100% of free radicals is the fact that the skin acts as a barrier to the penetration of the active compounds. Thus, using a higher concentration than that required is a useful way to increase the available concentration in the epidermis and dermis, since the penetration rate will never be equal to 100% (Idson, 1971). Increasing the drug concentration in topical formulations to improve efficacy has been used in recent studies (Anand, Blay, 2011).

Design of the cosmetic formulation

For the design of a structured cosmetic formulation, raw materials that favour the penetration of the active substance through the skin were chosen; these raw materials are known as permeation enhancers. The aim

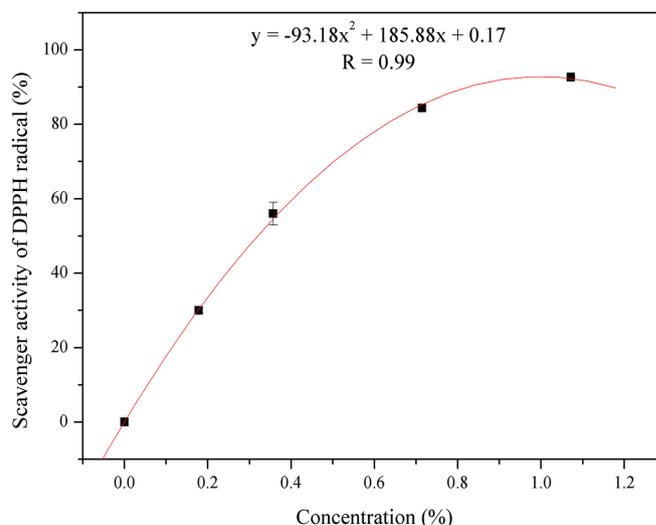


FIGURE 1 - Inhibition percentage of the DPPH radical *versus* concentration of the GB soft extract.

was to ensure that the antioxidant GB soft extract could be able, if applied to the skin, to reach both the epidermis and the dermis. According to Martins and Veiga (2002), permeation enhancers are compounds that can permeate or interact with stratum corneum constituents, when incorporated into a transdermal formulation, but do not possess pharmacological activity, but merely reduce skin resistance to drug diffusion. In this case, the surfactants and humectants used in the formulation can be cited.

A base formulation (test formulation 1) was proposed and, from this, two other formulations (test formulations 2 and 3) were prepared by altering the raw materials and their respective concentrations (Table I). The test formulations were macroscopically analysed for viscosity and spreadability. Emulsions 1 and 2 did not disperse the self-emulsifying wax completely, leaving them with a rough appearance and the presence of lumps. Considering this, test formulation 3 was considered most suitable due to its lower viscosity, a characteristic considered excessive in the other formulations, as well as a more pleasant sensorial aspect.

Measurement of the antioxidant activity of the formulation

The antioxidant activity of the formulation containing the GB soft extract was evaluated. The inhibition percentage of DPPH radicals obtained using 0.5 g of the formulation in the reaction was $20.16 \pm 10.34\%$. Considering that 0.5 g of the formulation were dispersed in 3.5 mL for the reaction with the DPPH radical, a final concentration of 0.14 g of emulsion/mL was obtained during the reaction. Thus, each 0.14 g of

emulsion containing the GB soft extract per millilitre was capable of inhibiting approximately 20% of the free radicals in the reaction.

Cefali *et al.* (2015) evaluated a phytocosmetic containing lycopene; the percentage inhibition of DPPH radicals was 19.51% (0.1 g/mL). The authors reported the antioxidant effect to avoid skin ageing, considering daily use and accumulation of the product on the skin. Also, it should be emphasised that daily application of the product on the face would probably exceed a mass of 0.14 g, reinforcing the anti-ageing potential of the formulation.

Analysis of the preliminary stability of the cosmetic formulation

Assessment of the organoleptic characteristics

The assessment of the organoleptic characteristics, i.e. colour, odour and appearance, is important to follow the changes in these characteristics over time, which is an indicative of stability of the formulation (Isaac *et al.*, 2008).

During the 30-day stability evaluation, macroscopic analyses were performed, i.e. the samples were visually observed for their appearance, relating them to the reference sample, understood as the formulation at the time of preparation. The sample maintained at room temperature could also be considered as a control, since there was no acceleration of instabilities due to temperature and/or light radiation. The odour and colour of the formulations were also evaluated.

On the day of preparation, the formulations without GB extract and with GB extract had similar but not identical characteristics. The formulation without GB extract was white in colour, with a characteristic odour and homogeneous emulsion appearance. The formulation with GB soft extract presented a slightly yellowish colour, with the characteristic odour of the extract and the appearance of a homogeneous emulsion.

After 30 days, it was possible to observe that the formulations with and without the GB extract were stable, as no changes were observed in its appearance (no gas formation or phase separation), odour or colour. The same behaviour was observed after storage at room temperature, 38 °C and 4 °C for a period of 30 days.

Determination of pH

The pH values of the formulations containing or not the GB soft extract were monitored for 30 days are shown in Tables II and III.

Analysing the data presented in Tables II and III, it can be observed that the emulsions did not present significant differences in pH with the passage of time. According to Friedrich *et al.* (2007), this shows that even with changes in temperature and luminosity, there was no degradation of the fatty compounds. The pH values remained within the recommended range for cosmetic products, varying approximately between 5 and 6.5. Vieira *et al.* (2009) considered that pH variations greater than 15 should be investigated. In the formulations investigated here, with or without the presence of the GB soft extract, the maximum variation between t0 and t30 was 9.24%.

The determination of the pH of cosmetic formulations is of extreme importance as it should be a value compatible with that of the region of the body in which it will be applied. According to Leonardi *et al.* (2002), the skin has a slightly acidic pH, ranging from 4.8 to 5.8, which favours the bactericidal and fungicidal protection of the skin surface. It is noteworthy that skin secretions have an important buffering capacity, since the pH of the skin can be altered as a result of the use of topical products.

It was determined that the pH values obtained in this study were satisfactory since they presented values between 5.19 and 6.22, indicating slightly acidic characteristics. The formulation with GB was slightly more acidic than the base formulation, which suggests that the extract also has acidic characteristics.

TABLE II - pH values determined during 30 days in the formulation without the GB soft extract (mean \pm standard deviation). The percentage of variation comparing t0 and t30 is shown

Time (days)	24 \pm 2 °C	38 \pm 2 °C	4 \pm 2 °C
0	5.34 \pm 0.27	5.34 \pm 0.27	5.34 \pm 0.27
7	6.22 \pm 0.12	6.12 \pm 0.11	6.16 \pm 0.26
15	5.83 \pm 0.09	6.01 \pm 0.10	5.23 \pm 0.16
30	5.80 \pm 0.10	5.70 \pm 0.17	5.83 \pm 0.15
%variation (comparing t0 and t30)	8.61	6.74	9.24

TABLE III - pH values determined during 30 days in the formulation with GB soft extract (mean \pm standard deviation). The percentage of variation comparing t0 and t30 is shown

Time (days)	24 \pm 2 °C	38 \pm 2 °C	4 \pm 2 °C
0	5.84 \pm 0.05	5.84 \pm 0.05	5.84 \pm 0.05
7	5.64 \pm 0.47	5.20 \pm 0.14	5.25 \pm 0.08
15	5.72 \pm 0.14	5.42 \pm 0.18	5.23 \pm 0.14
30	5.63 \pm 0.15	5.40 \pm 0.26	5.67 \pm 0.40
%variation (comparing t0 and t30)	3.48	7.48	2.91

Determination of the relative density

Physicochemical tests are important for the investigation of structural alterations of the formulation that are not visually perceptible, i.e. the relative density. The values of the formulations assessed over 30 days of analysis are shown in Tables IV and V.

The emulsions with and without GB extract did not show relative density variations during the 30-day stability study, when the samples were stored at room temperature, 38 \pm 2 °C and 4 \pm 2 °C, with a relative density value close to 1.0 g/cm³. Density values are important for semi-solid products according to the Cosmetic Products Stability Guide, as they may indicate the incorporation of air or the loss of volatile ingredients. This also happens with water, which can cause an increase in the viscosity of the

formulation. However, this apparently did not occur in this study.

Rheological characterisation of the formulations

The rheological characterisation was performed for both the base formulation and the formulation containing the GB soft extract. Besides characterising the formulations, this allowed us to evaluate if the addition of the GB soft extract induced significant changes to the formulation's structure.

The first assay performed was the flow analysis of the formulations (Figure 2). Both exhibited Herschel-Bulkley behaviour, since there was a yield value, followed by a decrease in viscosity with an increase in shear.

TABLE IV - Relative density values (g/cm³) measured periodically, for 30 days, for the base formulation (mean \pm standard deviation). The percentage of variation comparing t0 and t30 is shown

Time (days)	24 \pm 2 °C	38 \pm 2 °C	4 \pm 2 °C
0	0.977 \pm 0.006	0.977 \pm 0.006	0.977 \pm 0.006
7	0.977 \pm 0.012	0.987 \pm 0.006	0.983 \pm 0.012
15	0.987 \pm 0.006	0.983 \pm 0.012	0.987 \pm 0.006
30	0.987 \pm 0.006	0.987 \pm 0.006	0.987 \pm 0.006
%variation (comparing t0 and t30)	0.989	0.989	0.989

TABLE V - Relative density values (g/cm³) measured periodically, for 30 days, for the formulation containing GB extract (mean \pm standard deviation). The percentage of variation comparing t0 and t30 is shown

Time (days)	24 \pm 2 °C	38 \pm 2 °C	4 \pm 2 °C
0	0.967 \pm 0.006	0.967 \pm 0.006	0.967 \pm 0.006
7	0.977 \pm 0.006	0.983 \pm 0.006	0.967 \pm 0.006
15	0.973 \pm 0.012	0.973 \pm 0.006	0.983 \pm 0.006
30	0.987 \pm 0.006	0.987 \pm 0.006	0.977 \pm 0.006
%variation (comparing t0 and t30)	2.034	2.034	0.997

The formulations exhibited low thixotropy, which is a favourable feature, since it is related to the spreadability of the preparation (Morais *et al.*, 2005). It was also possible to demonstrate that the addition of the fluid GB extract promoted a slight reduction in the viscosity of the formulation. There was also an increase in thixotropy, favouring the spreadability of the formulation.

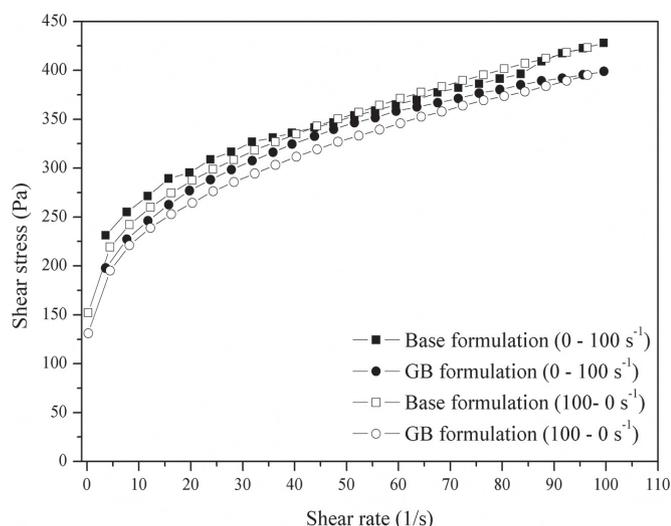


FIGURE 2 - Flow curve of the formulations containing or not the GB soft extract.

Shear stress (Figure 3) is an assay employed to verify the range of shear stress where a linear viscoelasticity region is found. For both formulations, the linear region of viscoelasticity occurred practically in the entire shear stress range evaluated, with 1 Pascal being the value determined for the continuation of the studies. A frequency sweep, on the other hand, allows for the analysis of the elastic (G') and viscous (G'') moduli (Figure 4). The GB and base formulations presented a higher elastic modulus than viscous modulus, which was considered indicative of stability for emulsified formulations (Chiari *et al.*, 2012a; Isaac *et al.*, 2013a, b). However, the addition of the GB soft extract reduced the G' and G'' values, which can be explained by the reduction in the relative fatty content of the formulation.

The results of the rheological characterisation, including the creep and recovery tests (Figure 5), showed a small reduction in viscosity and elastic modulus, interfering with the deformation (time from 0 to 200 s) and recovery of the formulation (time from 200 to 400 s).

Assessment of the formulations by SAXS

The SAXS technique is fundamental to confirm the presence of structures shown by polarised light microscopy (Figure 6). Well-defined peaks, known as

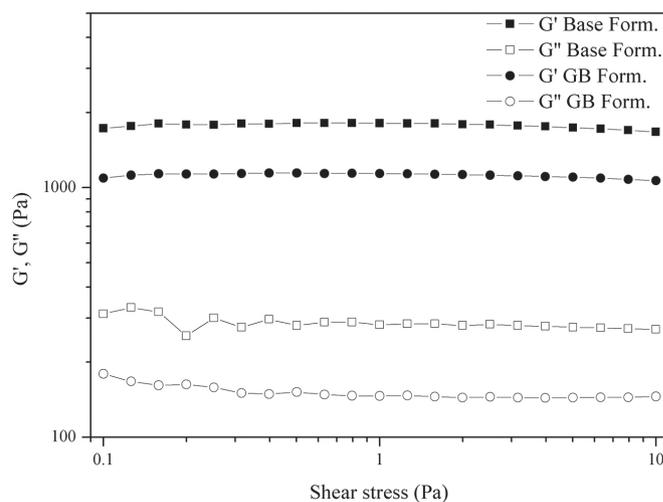


FIGURE 3 - Shear stress of the formulations containing or not the GB soft extract.

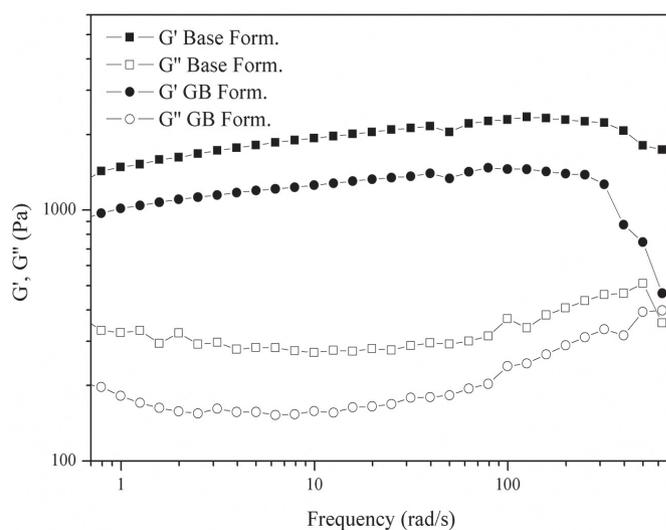


FIGURE 4 - Frequency sweep of the formulations containing or not the GB soft extract.

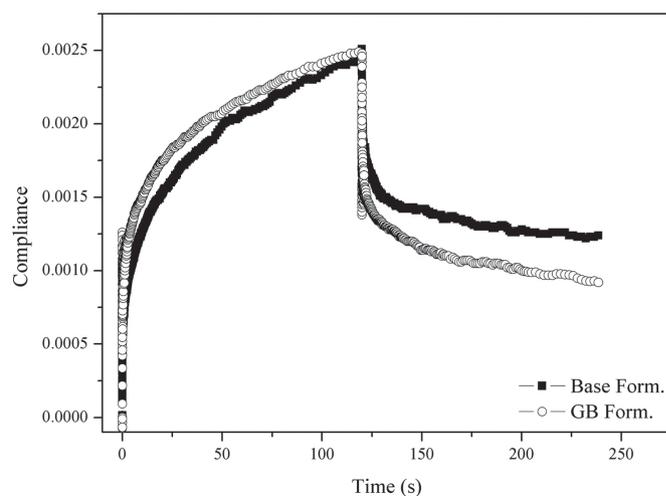


FIGURE 5 - Creep and recovery analysis of the formulations containing or not the GB soft extract.

Bragg peaks, have been found in the analysis of similar formulations, revealing the presence of highly organised structures, such as liquid crystals (Ferreira *et al.*, 2015). At medium to high concentrations of amphiphilic molecules such as surfactants, the systems are more structured than those at low concentrations. The amphiphilic molecules aggregate and form small, randomly oriented crystalline domains, with lattices exhibiting 1D, 2D or 3D long range correlations.

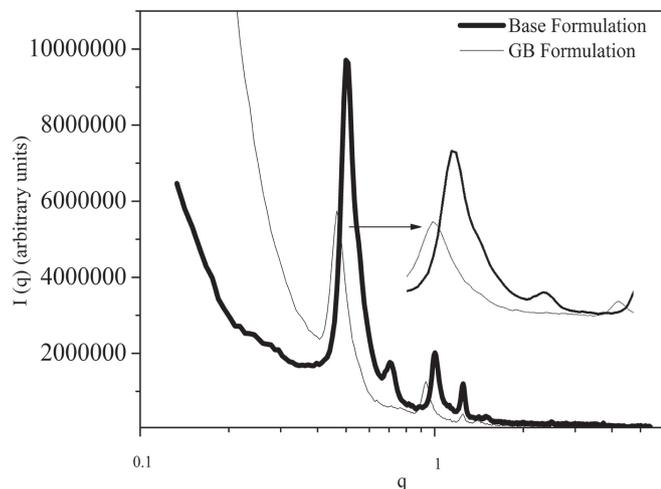


FIGURE 6 - SAXS assay results for the formulations containing or not the GB soft extract (q was determined in $1/\text{\AA}$). A zoom of the image is indicated with the arrow.

The SAXS curves for these formulations exhibited sharp maxima (or Bragg peaks) for specific values of the modulus of the scattering vector q . The scattering technique can now be called SAXD. If the SAXD patterns contain several peaks, their positions on the q axis reveal the type of crystal lattice of the structures and permit their lattice parameters to be calculated (Klug, Alexander, 1954). The relationship between the positions of these peaks on the q -axis indicates the crystalline structure type and allows structural parameters to be calculated (Klug, Alexander, 1954). The d parameter was calculated as $2\pi/q_{\text{max}}$, where q_{max} is the maximum intensity value of the scattering vector peak (Manaia *et al.*, 2015).

These analyses indicated the formation of lamellar phase liquid crystals, and thus the high structural organisation of the formulation was shown, indicating the stability of the system.

Evaluation of the formulations using polarised light microscopy

Polarised light microscopy is a simple technique that can be used to show anisotropic liquid-crystalline

structures, such as liquid crystals (Müller-Goymann, 2004). The presence of these highly organised systems (Figure 7) in emulsified formulations indicates excellent stability of the system (Chiari *et al.*, 2012a). In this case, the presence of anisotropic structures was observed, probably around the droplets of the internal phase of the emulsion.

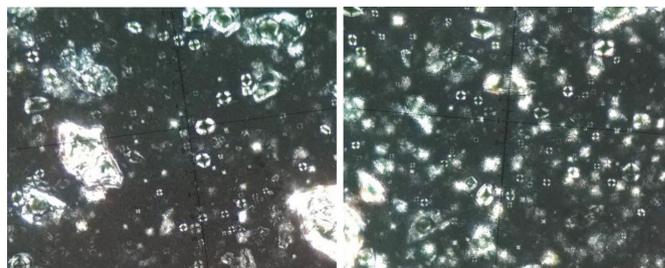


FIGURE 7 - Photomicrographs of the formulations not containing (left) and containing (right) GB soft extract.

Analysis of the bioadhesion force

In formulations for skin application, such as creams, ointments, solutions and lotions, considering a prolonged therapeutic effect is extremely difficult as external factors such as humidity, temperature and movements remove them easily. Thus, in order to evaluate the bioadhesive capacity of the formulation with a biological substrate, the adhesion strength was evaluated (Table VI). A commercial anti-ageing emulsion formulation was used as a control (Oshiro-Jr *et al.*, 2015; Oshiro *et al.*, 2018).

TABLE VI - Adhesion strength values (W_{ad}) for the formulations containing or not the GB soft extract. The results are expressed as mean and standard deviation, $n = 5$ replicates

Formulation	Adhesion strength values (W_{ad})
Base formulation	$0.073^a \pm 6.14$
Formulation with GB soft extract	$0.046^b \pm 6.59$
Vitanol-A®	$0.049^b \pm 4.28$

^{a,b} Different letters indicate statistically different values ($p < 0.05$).

The formulation without the addition of GB extract showed a higher bioadhesion value when compared to the formulation containing GB. This can be explained by the viscosity of the formulation, since formulations with higher viscosity tend to exhibit greater adhesion strength (Oshiro-Jr *et al.*, 2015). The reduction in viscosity was a result of the addition of the GB soft extract. These results corroborate the results observed in the rheology measurements. However, despite the decrease in adhesion

strength, when compared to the commercial formulation, the formulation containing the GB soft extract had a statistically equal adhesion strength, which allowed us to infer the suitability of the developed formulation. Adhesion strength tests have relevance in pharmaceutical formulations since higher values of bioadhesion may prolong the duration of action and absorption of the drug, thus improving the clinical performance of the formulation.

In conclusion, the combination of raw materials chosen in the studied formulation has the potential to provide a cosmetic with a lamellar phase. It showed adequate organoleptic characteristics and remained stable when the GB soft extract was incorporated. It also exhibited suitable bioadhesive properties on the skin, even when compared to a commercial formulation containing tretinoin. The antioxidant activity of the GB extract was also shown. Thus, this study demonstrates the suitability of applying GB extract as an anti-ageing cosmetic using this formulation.

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