

An Acad Bras Cienc (2023) 95(4): e20190120 DOI 10.1590/0001-3765202320190120

Anais da Academia Brasileira de Ciências | Annals of the Brazilian Academy of Sciences Printed ISSN 0001-3765 | Online ISSN 1678-2690 www.scielo.br/aabc | www.fb.com/aabcjournal

CROP SCIENCE

Cross preferences and genetic diversity of Psidium interspecific hybrids through morphoagronomic traits and resistance to *Meloidogyne enterolobii*

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Abstract: The introgression of M. enterolobii resistance-related genes in guava breeding programs can be compromised by incompatibility among Psidium species. This study aimed to evaluate the female parent preference and genetic diversity of Psidium interspecific hybrids using morphoagronomic traits and resistance to M. enterolobii. There were evaluated cross successes and germination from crosses between accesses of P. cattleyanum, P. guineense and P. guajava and the genetic diversity by Ward-MLM method of hybrids according to descriptors developed for the genus. Crosses were more successful when P. cattleyanum was the female parent. Germination was more successful in crosses involving P. cattleyanum and P. guajava. Four groups were formed. The group IV clustered the most resistant genotypes, composed by genotypes of *P. cattleyanum* x P. guineense, while the group II was the most susceptible. The groups I and III grouped some genotypes of P. cattlevanum x P. quajava with low levels of susceptibility. There are preferences of female parent species among crosses. Some individuals of groups I and III can be used as source of resistance genes for the breeding program, due the presence of favorable alleles inherited from guava parent. The high susceptibility leads to reduction in root development.

Key words: Genetic diversity, guava decline, interspecific hybridization, root-knot nematode, ward-MLM.

INTRODUCTION

The guava (*Psidium guajava* L.) is an important crop for Brazilian pomology, highlighting the states of São Paulo and Pernambuco (IBGE 2016). The Brazilian guava production count with a wide diversity of varieties, being some of them dedicated to *in natura* fruit consume (Ogawa, Pedro Sato, Kumagai and Sassaoka varieties) and to industry process fruits (Paluma, Rica and Século XXI) varieties (Pereira & Kavati 2011).

However, some concerns come up as the disease that becoming widespread worldwide,

the guava decline. It is a synergic association between the nematode *Meloidogyne enterolobii* Yang and Eisenbank, 1983 (syn. *Meloidogyne mayanguensis* Rammah & Hirschmann 1988), known as root-knot nematode, and the fungus *Fusarium solani*, promoting symptoms as galls on plant root, tanning and yellowing on leaves, defoliation of whole aerial part and, consequently, the plant death (Gomes et al. 2014, Sousa et al. 2017). The impact promoted by guava decline in Brazil has been threatening all production chain of guava in many poles, estimating a loss of million dollars (Gomes et al. 2011). In India, Jain et al. (2007) estimated that crop losses due plants parasitic nematodes were about of \$3.5 billion. Searching for efficient methods of control against this pathogen, several strategies were employed (Almeida et al. 2011, 2012, Robaina et al. 2015), however, the introduction of resistant plants of guava in orchards is the most efficient control against the nematode.

Different sources of resistance genes can promote a durable resistance in cultivars against any pathogen and some species of the same genus Psidium, commonly known as "araçá", carry these M. enterolobii resistancerelated genes (Miranda et al. 2010). The species P. guineense Swarts and P. cattleyanum Sabine, although have less economic importance than guava, have genotypes that could potentially be used in guava breeding programs as source of resistance genes through interspecific hybridization for introgression on target population (Costa et al. 2017; Gomes et al. 2017). Occasionally, species of the same genus can present cross-incompatibility in several crop species (Lu et al. 2014, Martins et al. 2015, Maune et al. 2018). This phenom limits the number of samples of a segregant population, which can lead to poor estimation of genetic divergence among individuals or groups of individuals. In *Psidium* spp., genetic diversity study was employed by Santos et al. (2011) to determinate an effective discrimination between "araçá" and guava accessions evaluated for some morphoagronomic traits. In breeding programs aiming the obtention of inbred lines, determinate the contrasting genotypes is useful to characterize heterotic groups and to explore the maximum of heterosis (Yuwono et al. 2017).

Therefore, multivariate methods are more efficient in quantifying dissimilarity between individuals, because it allows to consider many traits at the same time, even from different natures (Gonçalves et al. 2008, Campos et al. 2013). In this way, the Ward - Modified Location Model (MLM) procedure is a great strategy to quantify variability using both quantitative and gualitative variables (Franco et al. 1998). Some diversity studies have been developed through the Ward-MLM method. In cultivated species, divergence in lim'e bean accesses originated from different domestication centers was identified (Oliveira-Silva et al. 2017), while in peanut populations, groups with moderate variability were observed (Nunes da Luz et al. 2014). In cases of fruit trees, genetic divergence was verified in passion fruit progenies using quantitative and qualitative variables (Silva et al. 2014) and in different species of the genus Passiflora, the method allowed a clear differentiation of 11 species (Paiva et al. 2014).

Thus, the objective of this study was to determinate the cross preference among three *Psidium* species, as well as to characterize and estimate the genetic diversity of first generation of interspecific hybrids (H₁) of *Psidium* based on quantitative and qualitative traits related to morphoagronomic and response to inoculation with *M. enterolobii*. There were used seedlings to characterize genotypes due to the easiness of quantifying nematodes in the roots. This is the first study in which the Ward-MLM method is applied to discriminate Psidium genotypes according to *M. enterolobii* resistance and other morpho-agronomic traits in early stages.

MATERIALS AND METHODS

Experiment structure

The experiment was installed in a greenhouse at Universidade Estadual do Norte Fluminense Darcy Ribeiro - UENF in April of 2015, using interspecific hybrids between resistant genes donator specie (*P. cattleyanum*) and resistant genes receptor species (*P. guajava* and *P. guineense*) (Miranda et al. 2012) to compound a segregate population.

The crosses were carried out stepwise, according to the floral synchronism between parents, therefore all following procedures of evaluation were carried out stepwise. The accesses used as parents were obtained from germplasm bank of UENF (Table I) and crossed randomly, according to recommendation of Alves and Freitas (2007). A total of 412 crosses between *P. cattleyanum* vs. *P. guineense* and 284 crosses between *P. cattleyanum* vs. *P. guajava* were carried out at late 2014. There were evaluated, at the total, 885 genotypes of the five families for diversity study.

Therefore, the seeds collected from crosses were planted in trays and kept in a nebulization chamber until germination, which occurred between 22 and 30 days. The H₁ generation seedlings were conducted under greenhouse conditions, transplanted to 5 L plastic pots that received one seedling each, with properly fertilized substrate, and maintained in greenhouse.

Inoculation procedure

There were used tomato roots as inoculum source, planted on sterile soil being inoculated with *M. enterolobii* isolates previously collected from guava orchards in São João da Barra, RJ. After the development of the sixth pair of Psidium plant leaves, each pot received a 5 mL nematode egg concentration inoculated around the plant. The nematode egg concentration inoculated had an initial population (Pi) with 1,000 eggs of *M. enterolobii*. The inoculation procedure and the period of 135 to 180 days, consisting of the inoculation until evaluation, follows the methodology proposed by Burla et al. (2010).

 Table I. Families of interspecific hybrids of Psidium discriminating the female and male species, and data about

 the cross performance.

Families*	Parents	NC**	CS (%)	NF	NS	G (%)
CV1 X CV11	P. cattleyanum x P. guineense	87	24.13	21	410	47.60
CV11 X CV8	P. guineense x P. cattleyanum	100	1.00	1	21	4.70
CV11 X UENF-20181	P. guineense x P. cattleyanum	200	0.50	1	297	66.70
CV6 X 13.10 I	P. cattleyanum x P. guajava	30	10.00	3	16	43.80
CV6 X 9.7 I	P. cattleyanum x P. guajava	20	5.00	1	77	3.90
CV8 X CV11	P. cattleyanum x P. guineense	25	40.00	10	298	67.10
UENF-20181 X 9.7 I	P. cattleyanum x P. guajava	20	10.00	2	8	0.00
UENF-20181 X 9.9 I	P. cattleyanum x P. guajava	25	4.00	1	8	12.50
UENF-20175 X 9.7 I	P. cattleyanum x P. guajava	40	5.00	2	5	80.00
9.9 I X P52	P. guajava x P. cattleyanum	50	2.00	1	117	76.90
13.4 II X P51	P. guajava x P. cattleyanum	34	6.00	2	394	87.60
13.4 II X UENF-20176	P. guajava x P. cattleyanum	20	5.00	1	335	76.70
9.5 II X UENF-20181	P. guajava x P. cattleyanum	25	4.00	1	68	92.60
13.2 II X CV4	P. guajava x P. cattleyanum 20		5.00	1	101	86.10
Total		696	6.60	46	1,858	67.40

*The prefix CV means that patents are accesses of *Psidium* sp. placed in greenhouse, the prefix UENF means that parents are accesses of *Psidium* sp. placed at field and the other parents are genotypes of guava under field experimentation. **Number of crosses (NC); Crosses success rate (CS); Number of fruits (NF); Number of seeds (NS); Germination rate (G).

Qualitative variables

We evaluated the plants according fourteen qualitative morphological descriptors from each plant, being four binary and ten multicategory, based on International Union for the Protection of New Varieties of Plants (UPOV) document as described at the Table II. These characteristics were all evaluated when the progenies showed the sixth pair of leaves, just before the inoculation procedure.

Quantitative variables

Six guantitative variables measured from each plant were evaluated in segregate population. Plant height was obtained using a measuring tape since soil to the tip of plant. Number of leaves was obtained by counting leaves. Fresh mass of aerial part and root was obtained using an electronic balance model Shimadzu BL73200H. Fresh volume of root was measured using cylinder graduated by the difference between column water after root submersion. The variables associated to roots were processed according to the method proposed by Miranda et al. (2010) for screening guava plants to M. enterolobii resistance, where the root plant was halved allowing the plants to continue their development, while the other part of root was used as sample for analysis. These sample were used to estimate the reproduction factor (RF) following the methodology proposed by Oostenbrink (1966), where $RF = P_i/P_r$, being P_ithe initial population and P_e final population, and RF > 1 means that genotype is susceptible for nematode infection, RF < 1 means that genotype is resistant for nematode infection and RF = 0 means that genotype is immune for nematode infection.

Genetic diversity analysis

The clustering method approached to estimate the genetic divergence was the Ward-MLM

Table II. Descriptors used to evaluate morphologictraits from interspecific hybrids of *Psidium*.

Descriptor	Characteristic	Code
	Absent or very sparse	1
	Sparse	3
Pubescence on lower side	Medium	5
of young leaf	Dense	7
	Very Dense	9
	Absent	1
Twisting	Present	9
Pubescence on lower side	Absent	1
of fully developed leaf	Present	9
	Round	1
	Ovate	2
	Obovate	3
Shape of leaf	Trullate	4
	Obtrullate	5
	Oblong	6
	Obtuse	1
Shape of base	Rounded	2
Sliape of base	Cordate	3
		1
	Attenuate	
Channa a Chin	Apiculate	2
Shape of tip	Acute	3
	Obtuse	4
	Rounded	5
Relief of surface of upper	Smooth	3
side	Medium	5
	Wrinkled	7
	Weak	3
Curvature in cross section	Medium	5
	Strong	7
Curvature of midrib	Absent	1
	Present	9
Degree of curvature of	Weak	3
midrib	Medium	5
	Strong	7
Undulation of margin	Absent	1
	Present	9
Dograp of undulation of	Weak	3
Degree of undulation of margin	Medium	5
marsin	Strong	7
	Grey green	1
Croop salar	Yellow green	2
Green color	Green	3
	Dark green	4
	Cream	1
Color of midrib on lower	Yellow	2
side	Reddish	3

technique proposed by Franco et al. (1998), where the Gower's algorithm (Gower 1971) was used to estimate the distance between access contemplating both quantitative and qualitative variables. This method was processed by software SAS, through the commands CLUSTER and IML (SAS institute 2011). The optimal number of groups was defined based on pseudo-f and pseudo-t² criteria, associated to likelihood profile and likelihood test ratio. The MLM procedure estimated the mean vector of quantitative traits for each subpopulation. The distance between groups and the correlation of variables with canonical variables was obtained by CANDISC procedure. The distance, proposed by Matusita (1956), adapted by Krzanowski (1983) and Franco et al. (1998) for distribution of variables, defined the dissimilarity of groups based on generalized Mahalanobis distance, where each plant of the group was considered as replication.

RESULTS

Interspecific crosses

The results obtained from the crosses (Table I) demonstrate that the highest cross success rates between *P. cattleyanum* x *P. guineense* were observed in crosses CV8 x CV11 and CV1 x CV11 (40% and 24.13%, respectively). For crosses between *P. cattleyanum* x *P. guajava*, the progenies with highest cross success rates were CV6 x 13.10I and UENF-20181 x 9.9I (10% each).

There were remained 1,858 seeds from the crosses, however, the germination rate above 70% were only observed for the crosses *P. guajava* x *P. cattleyanum*, highlighting the crosses 9.5II x UENF-20181, 13.4II x P51, 13.2II x CV4, 9.9I x P52 and 13.4II x UENF-20176, with germination rate of 92.6%, 87.6%, 86.1%, 76.9% and 76.7%, respectively, as well as the cross *P. cattleyanum* (UENF-20175) x *P. guajava* (9.7I) which presented

germination rate of 80%. Among 70% and 40% of germination rate, the crosses *P. cattleyanum* (CV8) x *P. guineense* (CV11), *P. guineense* (CV11) x *P. cattleyanum* (UENF-20181), *P. cattleyanum* (CV1) x *P. guineense* (CV11) and *P. cattleyanum* (CV6) x *P. guajava* (13.101) were observed in this group, showing the rates 67.1%, 66.7%, 47.6% and 43.8%, respectively.

To follow up the study, only 885 seeds had emerging success and vigor enough to reach at the six pairs of leaves admitted here for the characterization and diversity analysis.

General classification of population

The 885 genotypes were mostly classified as absent or very sparse pubescence in lower side of young leaves (40.11%), followed by medium, sparse, dense and very dense (27.23%, 24.75%, 6.33% and 1.58%, respectively) (Figure 1).

The descriptor twisting of leaves showed most of genotypes with absence (96.95%), as well observed for the mostly genotypes when evaluated pubescence in downside of mature leaves (98.87%). For developed leaves the genotypes were classified mostly as oblong shape (68.81%), followed by oblanceolate (30.85%), ovulate (0.23%) and obovate (0.11%). For shape of base all genotypes were classified as obtuse (100%) and for tip of leaves the genotypes were classified as obtuse (69.83%) and as acute (30.17%).

The curvature in cross section has the most of genotypes classified as medium (50.40%) followed by weak (29.60%) and strong (20.00%). The descriptor degree of curvature of midrib, that was observed in 69.49% of genotypes, had weak (49.27%), medium (40.81%) and strong (9.92%), while the other 30.51% did not presented this characteristic. In turn, the degree of undulation of margin was observed in only 69.83% and the most of these genotypes was classified as weak (58.74%), followed by medium (33.66%) and strong (7.60%).

The descriptor green color classified the genotypes mostly as dark green leaves (58.54%), followed by green (41.46%) and the color of midrib on lower side were yellow for the most genotypes (74.69%), followed by cream (13.01%) and red (12.09%).

Clustering

According to pseudo-F and pseudo-t² criteria, the 885 genotypes were clustered in three and two groups, respectively. The logarithmic function of probability based at highest increment on loglikelihood's value determined the formation of four groups as optimal number of groups, as illustrate the figure 2.

Clusters characterization

The group I was composed by 44.3% of genotypes of progeny CV11 x UENF-20181, 26.0% of progeny CV8 x CV11, 15.9% of progeny CV1 x CV11, 7.8% of progeny 13.4II x UENF-20176 and 6% of progeny 13.4II x P51, a total of three hundred thirty-four genotypes, being this the only one group that allocated genotypes of all crosses. The group II was composed by 92.2% of genotypes of progeny 13.411 x P51, 6.4% of progeny CV11 x UENF-20181 and 1.4% of progeny CV8 x CV11, a total of 281 genotypes. The group III was composed by 59.2% of genotypes of progeny CV11 x UENF-20181, 35.4% of progeny 13.4II x UENF-20176 and 5.4% of progeny 13.4II x P51, a total of 147 genotypes. All group IV was composed by genotypes of progeny CV1 x CV11, a total of 123 genotypes.

In group I, the genotypes were represented mostly with absence of pubescence on lower side of young leaves (52.99%), absence of twisting (96.71%), absence of pubescence on lower side of developed leaves (99.40%), oblong shape of leaves (55.99%), obtuse shape of base (100.00%), obtuse shape of tip (58.08%), smooth surface of upper side (39.22%), medium curvature in cross section (45.51%), presence of curvature of midrib (57.49%), weak degree of curvature of midrib (59.90%) when observed, presence of undulation of margin (58.08%), weak degree of undulation of margin (68.04%) when observed, dark green color of leaves (62.28%) and yellow color of midrib on lower side (68.26%) (Figure 3).

The second group had the most of genotypes with medium pubescence on lower side of young leaves (39.86%), absence of twisting (98.93%), absence of pubescence on lower side of developed leaves (97.51%), oblong shape (98.58%), obtuse shape of base (100.00%), obtuse shape of tip (98.58%), wrinkled surface of upper side (71.53%), medium curvature in cross section (51.96%), presence of curvature of midrib (98.58%), weak degree of curvature of midrib (54.51%) when observed, presence of undulation of margin (98.58%), weak degree of undulation of margin (60.65%) when observed, dark green color of leaves (61.21%) and yellow color of midrib on lower side (80.43%).

The third group was represented by genotypes with medium pubescence on lower side of young leaves (39.46%), absence of twisting (91.16%), absence of pubescence on lower side of developed leaves (100.00%), oblong shape (98.64%), obtuse shape of base (100.00%), obtuse shape of tip (100.00%), wrinkled surface of upper side (56.46%), medium curvature in cross section (50.43%), presence of curvature of midrib (99.32%), medium degree of curvature of midrib (54.11%) when observed, presence of undulation of margin (99.32%), medium degree of undulation of margin (47.26%) when observed, dark green color of leaves (61.22%) and yellow color of midrib on lower side (68.71%).

The fourth group clustered genotypes with characteristics of absence of pubescence on lower side of young leaves (96.75%), absence of twisting (100.00%), absence of pubescence on lower side of developed leaves (99.19%), obtrullate shape (100.00%), obtuse shape of base (100.00%), acute shape of tip (100.00%), smooth surface of upper side (90.24%), medium curvature in cross section (60.16%), absence of curvature of midrib (100.00%), absence of undulation of margin (99.19%), weak degree of undulation of margin (100.00%) when observed, dark green color of leaves (58.54%) and yellow color of midrib on lower side (86.18%).

Descriptive statistical

The group II present the highest mean of reproduction factor, 130.00 and 42.73 standard deviation, while the group IV present the lowest mean 1.11 and standard deviation 1.08, the group I and III present mean 4.28 and 6.13, respectively, and standard deviation 9.07 and 9.80, respectively (Figure 4).

For number of leaves, the highest mean belongs to group I with 42.38 and standard deviation 19.20, followed by group IV with mean 40.27 and standard deviation 11.65. The groups II and III presented mean 36.34 and 28.39, respectively, and standard deviation 11.61 and 11.21, respectively.

The height of plant presents the highest mean for group III, followed by groups IV, II and I, with values 43.71, 37.31, 36.68 and 29.48 cm, respectively, and the standard deviation of these groups was 9.85, 7.14, 10.92 and 7.76 cm, respectively.

For fresh mass of aerial part, the highest mean was registered by group IV with mean 27.44 g and standard deviation 9.48 g, the second highest mean belongs to groups I and III, both present mean 17.99 and standard deviation 9.10 and 9.44 g, respectively. The lowest mean was registered by group II with 17.90 g of mean and 8.55 g of standard deviation.

The highest mean of fresh mass of root belongs to group IV, with 33.85 g, presenting

9.95 g of standard deviation. The groups I, III and II presenting mean 7.49, 6.29 and 5.28 g, respectively, with standard deviation 4.86, 2.72 and 2.38 g, respectively.

Similar rank was observed for fresh volume of root, where the group IV presented mean of 27.60 mL and standard deviation 10.95 mL, followed by group I with 6.27 mL of mean and 4.75 mL of standard deviation, group III with 5.16 mL of mean and 2.68 mL of standard deviation, and group II with 4.09 mL of mean and 2.36 mL of standard deviation.

Canonical correlation

The procedure CANDISC (Table III) estimated a positive and strong correlation of RF with canonical variable 1 (0.841), while root fresh mass and root fresh volume showed negative and strong correlation with the same canonical variable (-0.755 and -0.717, respectively). For the correlation of the canonical variable 2 and reproduction factor, root fresh mass and root fresh volume, the results were all positives (0.538, 0.601 and 0.548, respectively) and were not so strong as canonical variable 1. Therefore, the second canonical variable do not represent the correlation between these characteristics as well as the first canonical variable. The characteristics number of leaves, plant height and fresh mass of aerial part showed low values

Table III. Two first canonical variables correlated withsix quantitative variables measured in interspecifichybrids of *Psidium*.

Variables*	Canonical Variables			
Variables*	Can1	Can2		
RF	0.841	0.538		
NL	-0.099	0.000		
PH	0.045	0.172		
MA	-0.275	0.255		
RM	-0.755	0.601		
RV	-0.717	0.548		

*RF (Reproduction Factor), NL (Number of leaves), PH (Plant Height), MA (Mass of Aerial Part), RM (Root Mass) and RV (Root Volume). of coefficients of correlation with both canonical variables, suggesting no correlation between these characteristics with the others.

Distance between groups

According to Ward-MLM method, the estimation of distance between groups (Table IV), based on generalized distance of Mahalanobis, showed that the group II presented highest distance with group IV (70.95), while the most similar groups were I and III (5.77). The groups I and II presented values of distance 26.83, while the groups I and IV presented value of distance 40.68, otherwise, the groups II and III presented value of distance 26.26, and the groups III and IV presented value of distance 45.06 (Figure 5).

DISCUSSION

Interspecific crosses of Psidium species

The results elucidated that interspecific crosses among *Psidium* species were not homogenous and some crosses have advantages when carried out in one direction. This was noted when analyzed the cross-success rates, as observed when the *P. cattleyanum* was the female parent, where rates were under than 10%. Furthermore, the highest cross success rates were observed in crosses among *P. cattleyanum* and *P. guineense*. Studies carried out for interspecific hybridization in *Capsicum* showed difficulties caused by pre-fertilization barriers, such as possibility of crosses only in one direction, as example of crosses between *C. baccatum* and *C. frutescens*,

 Table IV. Mahalanobis distance among four groups

 formed by Ward-MLM method from interspecific

 hybrids of *Psidium*.

Group	1	2	3	4
1	-	26.83	5.77	40.68
2		-	26.26	70.95
3			-	45.06
4				-

showing non-germination of pollen grains or stopping the pollen tube growing (Martins et al. 2015). Many seeds obtained by crosses suffered with lack of vigor, or even germinated. These phenomena are usually common in interspecific hybridization (Parton et al. 2001, Martins et al. 2015, Okamoto and Ureshino 2015). The reduction in the number of progenies analyzed in the study (from 1,858 seeds to 885) could be affected by a post-fertilization barrier in interspecific hybridization among *Psidium* species, which can be confirmed by cytological studies.

Some genotypes suffered from lack of vigor, causing losses of genotypes mostly registered in progenies of *P. cattleyanum* x *P. guajava*. The same lack of vigor was reported by Costa et al. (2012), that had unsuccess in interspecific hybridization among guava and *P. cattleyanum*, however, these authors had success in cross among *P. guajava* and *P. guineense*. The high number of different parents used, as well as crosses made in this study, was the differential to acquisition of satisfactory number of viable seeds among *P. cattleyanum* and *P. guajava*.

Besides that, the basic chromosome number of Psidium species is x = 11, however, P. guajava is the only diploid specie in this genus (2n = 2x)= 22), while *P. quineense* and *P. cattleyanum* are admitted as polypoid (2n = 4x = 44) (Costa et al. 2008, Souza et al. 2015). The difficult to cross P. quajava and P. cattleyanum presented here could be an effect of the ploidy level differences, reducing the viability of crosses due the called triploid block (Marks 1966). It's expected that progenies of cross P. cattleyanum (2n = 44) x *P. quajava* (2n = 22) might present karyotype with 2n = 3x = 33 chromosomes, generating rarely gametes with up to 22 chromosomes, or usually sterile, due to unbalanced chromosome numbers caused by a failure to separate oddnumbered chromosome sets during meiosis (Köhler et al. 2010). The next generation (H_a) should have some genotypes with chromosomal complement tetraploid. However, if considering backcross with *P. guajava*, triploid hybrids should be expected in the first generation of backcross.

Cluster characterization

The number of groups formed using the Pseudo-F and Pseudo-t² criteria, associated to likelihood function, grouped the evaluated genotypes in the best way possible. These cluster number optimization criteria are preferable for non-accept a subjective clustering, however, the different number of accesses, descriptors and traits evaluated, can result in different number of groups (Gonçalves et al. 2009). Campos et al. (2013), evaluating the diversity of 138 guava access based on morphological, agronomical and physic-chemical descriptors, used this methodology and obtained eight groups in their analysis.

Considering the study of Santos et al. (2008) and comparing the data, the standard of segregation of genotypes in group I showed average similarities to guava in characteristics of curvature in shape of leaves, cross section and curvature of midrib. In group II, the similarities of genotypes with guava were observed in curvature in cross section, curvature of midrib and undulation of margin. The group III showed similarities in curvature of midrib, undulation of margin and degree of undulation of margin. For group IV, all genotypes showed standard segregation like P. cattleyanum. The group IV obtained in this study also clustered the genotypes with the lowest RF, followed by group I and III. Although Souza et al. (2014) clustered mostly genotypes resistant to M. enterolobii in one group, being these genotypes a *P. cattleyanum* access, the present study is the first one to estimate the diversity among interspecific hybrids of Psidium. Miranda et al. (2012) suggested that identifying resistant genotypes for *M. enterolobii* in accesses of *Psidium spp.* could open a novel path for strategies in guava breeding programs.

Canonical Variables

The estimative of eigenvalues of the first two canonical variables obtained in this study represented 95.21% of total variation. It's a value above the suggested by Gonçalves et al. (2009), which recommend that estimative above 80% is satisfactory for interpretation of variability among access. The correlation estimated about the six quantitative variables, showed that the RF of nematode have a high negative correlation with root variables on the first canonical variable. According Gomes et al. (2014) one of primary symptoms caused by M. enterolobii is the necrosis in all kind of root tissues, which, consequently, leads to losses of fine roots of absortion, by this way, infected genotypes have low mass and volume of roots compared to resistant genotypes, suggesting that the low mass and volume of roots are associated with the damage caused by *M. enterolobii*. However, carry out indirect selection through fresh mass and volume of roots might not provide an effective accuracy for screening resistant genotypes to *M*. enterolobii.

Distance between groups

The distance of groups I and III, composed by *P. cattleyanum* x *P. guajava* progenies, from the group IV, composed by *P. cattleyanum* x *P. guineense* progenies, where genotypes showed low mean of RF, was relatively high and it can be exploited for the ongoing guava breeding program. However, the genotypes obtained in group I and III could be preferable for backcross because they have more probabilities to drag agronomic favorable genes in linkage to resistant genes rather than genotypes from

group IV, that have no or few related genes to *P. guajava*. The backcross method is efficient for introgression of desired genes in populations already improved, such as commercial varieties and hybrids (Frisch & Melchinger 2005). Directing crosses through divergent clusters may avoid the narrowing of genetic base of a breeding program by expanding the genetic variability, providing options for the development of new varieties and hybrids (Govindaraj et al. 2015). Silva et al. (2014) had well used the Ward-MLM as strategy to define heterotic groups, and then advance the generations of passion fruit in a program of recurrent selection.

CONCLUSIONS

There are some preferences among crosses of *Psidium* sp. The crosses aiming the introgression of resistant genes had great results in *P. cattleyanum* x *P. guineense*, in comparison to *P. cattleyanum* x *P. guajava* families, however the germination rate was higher in *P. gujava* x *P. cattleyanum* families. Therefore, only with high number of cross-pollinations between *P. guajava* and *P. cattleyanum* there will be possible to reach a satisfactory number of interspecific hybrid seeds.

The Ward-MLM method was efficient to cluster genotypes from similar crosses. The group I and III could cluster several resistant genotypes, being some of them originated from crosses between *P. cattleyanum* and *P. guajava*. The potential of these clusters for use in the guava breeding program on going is based on the presence of *P. guajava* as parent where some agronomic traits are already present.

Among quantitative traits evaluated, the first two canonical variables could demonstrate that the susceptibility to *M. enterolobii* is associated to the loss of fresh mass of roots and fresh volume of roots. However, it is still interesting select genotypes through reproduction factor of *M. enterolobii* rather than roots-related variables measured in this study, due the high information promoted by reproduction factor according to the resistance of the genotype.

Acknowledgments

The authors wish would acknowledge. the Nematology Laboratory of Universidade Estadual do Norte Fluminense Darcy Ribeiro by support with instruments, the Fundação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) by the scholarship for OFA, and the Conselho Nacional de Desenvolvimento Científico (CNPq) and the Rio de Janeiro Search Foundation (FAPERJ) by financial support.

REFERENCES

ALMEIDA AM, GOMES VM & SOUZA RM. 2011. Greenhouse and field assessment of rhizobacteria to control guava decline. Bragantia 70(4): 837-842.

ALMEIDA AM, SOUZA RM, GOMES RM, MIRANDA GB. 2012. Greenhouse and field assessment of different organic compounds against guava-parasitic *Meloidogyne enterolobii*. Bragantia 71(1): 67-74.

ALVES JE & FREITAS BM. 2007. Requerimentos de polinização da goiabeira. Ciênc Rural 37(5): 1281-1286.

BURLA RS, SOUZA RM, GOMES VM & CORRÊA FM. 2010. Assessment of inoculum level, evaluation time and variables for screening of *Psidium* spp. for resistance to *Meloidogyne mayanguensis*. Nematol Bras 34(2): 82-90.

CAMPOS BM, VIANA AP, QUINTAL SSR, GONÇALVES LSA & PESSANHA PGO. 2013. Quantification of the genetic divergence among guava accessions using Ward-MLM strategy. Rev Bras Frut 35(2): 87-94.

COSTA IR, DORNELAS MC & FORNI-MARTINS ER. 2008. Nuclear genome size variation in fleshy-fruited Neotropical Myrtaceae. Plant Syst Evol 276: 209-217.

COSTA SR, SANTOS CAF & CASTRO JMC. 2012. Assessing *Psidium* guajava x *P. guineense* hybrids tolerance to *Meloidogyne* enterolobii. Acta Hort 959: 59-65.

COSTA SR, SANTOS CAF & CASTRO JMC. 2017. Inheritance of resistance to *Meloidogyne enterolobii* in *Psidium guajava* x *P. guineense* hybrid. Eur J Plant Pathol 148(2): 405-411. FRANCO J, CROSSA J, VILLASEÑOR J, TABA S & EBERHART SA. 1998. Classifying genetic resources by categorical and continuous variables. Crop Sci 38: 1688-1696.

FRISCH M & MELCHINGER AE. 2005. Selection theory for marker-assisted backcrossing. Genetics 170: 909-917.

GOMES VM, RIBEIRO RM, VIANA AP, SOUZA RM, SANTOS EA, RODRIGUES DL & ALMEIDA OF. 2017. Inheritance of resistance to *Meloidogyne enterolobii* and individual selection in segregating populations of *Psidium* spp. Eur J Plant Pathol 148(3): 699-708.

GOMES VM, SOUZA RM, ALMEIDA AM & DOLINSKI C. 2014. Relationship between *M. enterolobii* and *F. solani:* spatial and temporal dynamics in the occurrence of guava decline. Nematoda 1: e010144.

GOMES VM, SOUZA R, MUSSI-DIAS V, SILVEIRA SF & DOLINSKI C. 2011. Guava decline: a complex disease involving *Meloidogyne mayaguensis* and *Fusarium solani*. J Phytopathol 159: 45-50.

GONÇALVES LSA, RODRIGUES R, AMARAL JÚNIOR AT, KARASAWA M & SUDRÉ CP. 2008. Comparison of multivariate statistical algorithms to cluster tomato heirloom accessions. Genet Mol Res 7(4): 1289-1297.

GONÇALVES LSA, RODRIGUES R, AMARAL JÚNIOR AT, KARASAWA M & SUDRÉ CP. 2009. Heirloom tomato gene bank: assessing genetic divergence based on morphological, agronomic and molecular data using a Ward-modified location model. Genet Mol Res 8(1): 364-374.

GOVINDARAJ M, VETRIVENTHAN M & SRINIVASAN M. 2015. Importance of genetic diversity assessment in crop plants and its recent advances: an overview of its analytical perspectives. Genet Res Int 2015: 431-487.

GOWER JC. 1971. A general coefficient of similarity and some of its properties. Biometrics 27(4): 857-871.

IBGE. 2016. Instituto Brasileiro de Geografia e de Estatística. Sistema IBGE de Recuperação Automática – SIDRA. Available in: <http://www.sidra.ibge.gov.br/bda/ tabela/protabl.asp?c=1613&z=p&o=28&i=P>. Access in: August 24th2017.

JAIN RK, MATHUR KN & SINGH RV. 2007. Estimation of losses due to plant parasitic nematodes on different crops in India. Indian J Nematol 37: 219-220.

KÖHLER C, SHEID OM & ERILOVA A. 2010. The impact of the triploid block on the origin and evolution of polyploid plants. Trends Gent 26(3): 142-148.

KRZANOWSKI WJ. 1983. Distance between populations using mixed continuous and categorical variables. Biometrika 70(1): 235-243. MARKS GE. 1966. The origin and significance of intraspecific polyploidy: experimental evidence from *Solanum* chacoense. Evolution 20(4): 552-557.

MARTINS KC, PEREIRA TNS, SOUZA SAM, RODRIGUES R & AMARAL JÚNIOR AT. 2015. Crossability and evaluation of incompatibility barriers in crosses between Capsicum species. Crop Breed Appl Biotechnol 15: 139-145.

MATUSITA K. 1956. Decision rules based on the distance for problem of fit, two samples, and estimation. Ann of Math Stat 8: 67-77.

MIRANDA GB, SOUZA RM, GOMES VM, FERREIRA TF & ALMEIDA AM. 2012. Assessment of *Psidium* spp. accessions for resistance to *Meloidogyne enterolobii*. Bragantia 71(1): 52-59.

MIRANDA GB, SOUZA RM & VIANA AP. 2010. Assessment of methods and criteria for screening *Psidium* spp. for resistance to *Meloidogyne enterolobii*. Nematol Bras 34(4): 211-219.

NUNES DA LUZ L, CAVALCANTI DOS SANTOS R, MELO FILHO PA & AZEREDO GONÇALVES LS. 2014. Combined selection and multivariate analysis in early generations of intraspecific progenies of peanuts. Chil J Agric Res 74(1): 16-22.

OKAMOTO A & URESHINO K. 2015. Pre- and post-fertilization barriers in interspecific hybridization between evergreen azalea species and *Rhododendron uwaense* H. Hara & T. Yamanaka. Hort J 84(4): 355-364.

OLIVEIRA-SILVA RN, LOBO-BURLE M, GOMES-PÁDUA J, ALMEIDA-LOPES AC, FERREIRA-GOMES RL & MARTÍNEZ-CASTILLO J. 2017. Phenotypic diversity in lima bean landraces cultivated in Brazil, using the Ward-MLM strategy. Chil J Agric Res 77(1): 35-40.

OOSTENBRINK M. 1966. Major characteristics of the relation between nematodes and plants. Meded Land bouwhogesch 66: 1-46.

PAIVA CL, VIANA AP, SANTOS EA, SILVA RNO & OLIVEIRA EJ. 2014. Genetic diversity of the genus *Passiflora* using the Ward-MLM strategy. Rev Bras Frutic 36(2): 381-390.

PARTON E, VERVAEKE I, DEROOSE R & DE PROFT MP. 2001. Interspecific and intergeneric fertilization barriers in Bromeliaceae. Acta Hort 552: 43-53.

PEREIRA FM & KAVATI R. 2011. Contribution of the brazilian scientific research in the development of some fruit trees of subtropical climate. Rev Bras Frutic v.e.92-180.

ROBAINA RR, CAMPOS GS, MARINHO CS, SOUZA RM & BREMENKAMP CA. 2015. Grafting guava on cattley guava resistant to *Meloidogyne enterolobii*. Ciênc Rural 45(9): 1579-1584.

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CROSS PREFERENCES AND DIVERSITY OF PSIDIUM HYBRIDS

SANTOS CAF, CASTRO JMC, SOUZA FF, VILARINHO AA, FERREIRA FR, PÁDUA JG, BORGES RME, BARBIERI RL, SOUZA AGC & RODRIGUES MA. 2008. Preliminary characterization of Psidium germplasm in different Brazilian ecogeographic regions. Pesq Agropec Bras 43(3): 437-440.

SANTOS CAF, CORRÊA LC & COSTA SR. 2011. Genetic divergence among Psidium accessions based on biochemical and agronomic variables. Crop Breed Appl Biotecnol 11: 149-156.

SAS INSTITUTE. 2001. SAS's user's guide: statistics. SAS Institute. Available in: <http://documentation.sas.com/ doc/en/pgmsascdc/9.4_3.4/statug/titlepage.htm>. Access in: April 06th 2023.

SILVA FHL, VIANA AP, FERREIRA RT, FREITAS JCO, SANTOS JO & RODRIGUES DL. 2014. Measurement of genetic diversity in progenies of sour passion fruit by Ward-MLM methodology: a strategy for heterotic group formation. Ciênc Agrotec 38(3): 240-246.

SOUSA AD, PEDROSA EMR, SILVA CUC, CASTRO JMC & RIBEIRO JM. 2017. Penetration, development, and reproduction of *Meloidogyne enterolobii* on *Psidium* species and induced cellular responses in the roots. Rev Bras Frutic 39(2): e-453.

SOUZA AG, RESENDE LV, LIMA IP, MARTIN LSS & TECHIO VH. 2015. Chromosome number and nuclear DNA amount in *Psidium* spp. resistant and susceptible to *Meloidogyne enterolobii* and its relation with compatibility between rootstocks and commercial varieties of guava tree. Plant Syst Evol 301(1): 231-237.

SOUZA AG, RESENDE LV, LIMA IP, SANTOS RM & CHALFUN NNJ. 2014. Genetic variability of araça and guava accessions susceptible and resistant *Meloidogyne enterolobii*. Ciênc Rural 44(5): 822-829.

YUWONO PD, MURTI RH & BASUNANDA P. 2017. Heterosis and specific combining ability in sweet corn and its correlation with genetic similarity of inbred lines. J Agr Sci 9(3): 245-253.

How to cite

ALMEIDA OF, VIANA AP, GOMES VM, SOUZA RM, SANTOS PR, AMARAL JÚNIOR AT & RODRIGUES DL. 2023. Cross preferences and genetic diversity of Psidium interspecific hybrids through morphoagronomic traits and resistance to *Meloidogyne enterolobii*. An Acad Bras Cienc 95: e20190120. DOI 10.1590/0001-3765202320190120.

Manuscript received on February 2, 2019; accepted for publication on August 1, 2019

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Odimar Almeida performed the measurements and data analysis. Alexandre Pio Viana conceived the idea. Vicente Gomes carried out the crosses and planned the experiments. Ricardo de Souza provided the lab facilities to process samples. Paulo Santos, Antônio Amaral Júnior and Daniele Rodrigues supported the analysis concepts.

