

SPATIAL AND TEMPORAL VARIABILITY OF PLANT-PARASITIC NEMATODES POPULATION IN SUGARCANE ⁽¹⁾

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ABSTRACT

Nematodes are important pests of sugarcane crop in Brazil. However, no studies have been reported on the spatial and temporal variability of nematodes population in sugarcane fields and such studies are indispensable for the development of sampling plans, aimed at their application in integrated management programs. This work was carried out in two commercial fields of approximately 1 ha, both of them infested by *Meloidogyne javanica* and *Pratylenchus zaei*. In each area, samples represented by about 50 g of roots, were collected every two months, within a grid measuring 10.5 × 10 m (experiment 1) and 9.8 × 10 m (experiment 2). The highest nematodes populations were obtained during the rainy season, when high temperatures and moistures were favorable to root development. For this reason the rainy season was considered the best time to collect samples to identify areas with nematodes problem. Among the 52 calculated semivariograms (28 for *M. javanica* or *P. zaei* populations in area 1 and 24 for *M. javanica* or *P. zaei* populations in area 2), 22 could not be fitted to any model and presented a pure nugget effect. The spherical model showed the best fit to the semivariograms of data observed in the other 30 conditions. In those cases, values of range in semivariogram varied from 22 to 77 m, with 47 m on average and suggesting that, to obtain a reliable estimate for the population of these plant parasites in a given area, samples should be collected 47 m apart, which is equivalent to 1.5 sampling points per hectare.

Key words: *Saccharum*, *Meloidogyne*, *Pratylenchus*, semivariogram, kriging.

RESUMO

VARIABILIDADE ESPACIAL E TEMPORAL DE POPULAÇÕES DE NEMATÓIDES PARASITOS DE PLANTAS EM CANA-DE-AÇÚCAR

Nematóides são importantes parasitos da cana-de-açúcar no Brasil. Apesar disso, não há estudos sobre a variabilidade espacial e temporal de suas populações em canaviais e tais estudos são indispensáveis para o desenvolvimento de planos de amostragem, visando à aplicação em programas de manejo integrado. Por essa razão, realizou-se este trabalho, em duas áreas comerciais, de cerca de 1 ha cada uma, ambas infestadas por *Meloidogyne javanica* e *Pratylenchus zaei*. Em cada área, amostras representadas por cerca de 50 g de raízes foram coletadas aproximadamente a cada dois meses, em *grid* de 10,5 × 10 m (Ensaio 1) e 9,8 × 10 m (Ensaio 2). As populações de nematóides mais elevadas foram obtidas na estação chuvosa, quando as altas temperaturas e umidade favorecem o desenvolvimento das raízes das plantas. Assim, esta estação foi considerada a melhor para coleta de amostras visando identificar áreas com problemas nematológicos. Entre os 52 semivariogramas calculados (28 para populações de *M. javanica* ou *P. zaei* na área 1 e 24 para populações de *M. javanica* ou *P. zaei* na área 2), 22 não foram ajustados a qualquer modelo e tiveram efeito pepita puro. O modelo esférico foi o que mais se ajustou aos semivariogramas dos dados observados nas outras 30 situações. Nesses casos, o alcance variou de 22 a 77 m, sendo em média de 47 m, sugerindo que para uma estimativa confiável da população desses fitoparasitos em determinada área, amostras deveriam ser coletadas à distância de 47 m umas das outras, o que equivale a 1,5 pontos de amostragem por hectare.

Palavras-chave: *Saccharum*, *Meloidogyne*, *Pratylenchus*, semivariograma, krigagem.

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1. INTRODUCTION

Among several species of nematodes found attacking sugarcane in Brazil, three have been rated as major economic problems: *Meloidogyne javanica*, *Meloidogyne incognita* and *Pratylenchus zeae* (NOVARETTI and TÉRAN, 1983, MOURA et al., 1990). On the average, these species cause losses productivity in the first harvest of the highly susceptible varieties that range from 20 to 40%; the productivity and the longevity may also be reduced for the subsequent sugarcane ratoons (DINARDO-MIRANDA, 2006).

A number of studies dealing with the efficacy of different nematode control methods in sugarcane fields were developed in Brazil along the last three decades. In opposition, studies related to the spatial and temporal distribution of nematode parasites of sugarcane are quite uncommon and they are indispensable for the development of sampling plans with the purpose of applying in integrated management programs (TAYLOR, 1984).

CARNEIRO et al. (1980) verified that the population levels of *P. zeae*, *Hoplolaimus galeatus*, *Helicotylenchus dihystra* and *Paratrichodorus porosus* occurring in a sugarcane field at Piracicaba, São Paulo State, increased progressively from November on, due to frequent rainfall and increased soil moisture, decreasing again by late March or early April. In relation to spatial distribution, DINARDO-MIRANDA and FRACASSO (2009) used geostatistical techniques to analyze populations data of *M. javanica*, *P. zeae* and *Pratylenchus brachyurus*, occurring in individual or mixed infestations in six sugarcane fields in São Paulo State. The samples were collected during the rainy season. The nematode aggregation area was estimated in 2,110 m² but, In several instances, the semivariograms could not be fitted to any of the tested models and were assumed to represent pure nugget effects.

The objective of this study was to characterize the spatial and temporal variability distribution of *M. javanica* and *P. zeae* in two Brazilian sugarcane fields based on statistical analyses of population data and with selected geostatistical procedures.

2. MATERIAL AND METHODS

Two experiments were conducted in areas located in the northern region of the State of São Paulo. The experiment 1 was carried out in clay; soil area (62.0% clay, 13.1% silt; 16.9% fine sand; 8.0% coarse sand; 24.9% total sand) planted with the sugar cane variety SP91-1049 on 2/19/05 and harvested for the first time on 6/13/06. The experiment 2 was carried out in sandy soil area (5.2% clay; 3.3% silt; 46.5% fine sand; 45.0% coarse sand; 91.5% total sand), planted with the sugar cane

variety RB928064 on 3/29/05 and harvest for the first time on 11/25/06.

Each experimental area was a field of 1 ha, where the samples were collected within a rectangular grid measuring 10.5 × 10 m (experiment 1) and 9.8 × 10 m (experiment 2), totalizing 100 sampling point in each sampling date, in each experiment. The samplings were done every two months, approximately, beginning two months after planting and ending on August 2007, in experiment 1 and on June 2007, in experiment 2. Fourteen and twelve samplings were done on experiment 1 and 2, respectively.

In each sampling point, 50 g sugarcane roots were collected to determine the nematode population. Nematodes were extracted from roots by a combination of sieving and centrifugal flotation methods, in sucrose solution, according to COOLEN and D'HERDE (1972).

Data were initially analyzed by descriptive statistics and mean, standard deviation, coefficient of variation, maximum value, minimum value, skewness and kurtosis were obtained using the Statistical Analysis System (SAS INSTITUTE, 1995). In order to decrease the coefficient of variation, the populations data were transformed to $\log(x+1)$, and basic statistics (mean, standard deviation, coefficient of variation and others) were calculated again.

In order to verify the hypothesis of normality of the data, the SHAPIRO and WILK (1965) test was carried out using the Statistical Analysis System (SAS INSTITUTE, 1995).

After this, geostatistical analyses of the transformed data were run using semivariograms and kriging interpolation to construct maps, as described by VIEIRA et al. (1983). The semivariogram analyses were conducted using the GEOSTAT software (VIEIRA et al., 2002). Based on the models fitted to the semivariograms, the jack knifing test was used to verify whether the estimates of semivariogram parameters were adequate and to estimate the number of neighbors that should be used in the kriging interpolation (VIEIRA, 2000). Once the parameters for the model were validated and the adequate numbers of neighbors was determined, values were interpolated for the locations where they were not measured, by the kriging method, using the GEOSTAT software (VIEIRA et al., 2002). The kriging-estimated values were used in the Surfer software (GOLDEN SOFTWARE, 1999) to construct the maps.

3. RESULTS AND DISCUSSION

Among the nematode species important for sugarcane, *M. javanica* and *P. zeae* were found in both experimental areas.

Great differences were observed in the experiments between the maximum and minimum populations, for both species under study, considering either the original data or data transformed to $\log(x+1)$. However, when transformed data were considered, the coefficients of variation, skewness and kurtosis were smaller as compared to the original data. For this reason, transformed data were used to conduct all the analysis. The coefficients of variation for *P. zae* were between 8.2 and 28.1 in experiment 1, and between 10.8 and 41.2% in experiment 2. These values are smaller than the observed for *M. javanica*, that reached 575.0 and 319.0% in experiments 1 and 2, respectively (Tables 1 and 2). Higher variability was also observed by DINARDO-MIRANDA and FRACASSO (2009) in *Meloidogyne* populations, as compared to *Pratylenchus*. This is probably related to the biology of nematodes that belong to that genus. According to FERRIS et al. (1990), because *Meloidogyne* is a sedentary endoparasite, females lay all

their eggs in the same place, in masses, resulting in a highly-aggregated spatial pattern. On the other hand, nematodes of the genus *Pratylenchus* are migratory and therefore move through the plant and the soil, laying their eggs individually, resulting in a more uniform distribution in the area.

In both experiments, the nematode populations increased rapidly at the beginning of crop development, with a population peak for both species occurring in June 2005 in experiment 1 and in June 2005 and October 2005, respectively, for *P. zae* and *M. javanica* in experiment 2 (Figures 1 and 2). Such rapid population growth can be attributed to the high soil moisture conditions as a resulted from rainfall (Figure 3).

For brazilian conditions, soil moisture plays a preponderant role on nematode population dynamics; Many factors may directly affect the spatial and temporal distribution of these parasites, but soil temperature and

Table 1. Statistical parameters of transformed by $\log(x+1)$ data of *M. javanica* (Mj) and *P. zae* (Pz) populations in SP91-1049 sugarcane variety, in experiment 1

Sampling date		Mean (n° /50g roots)	Minimum value (n° /50g roots)	Maximum value (n° /50g roots)	Standart deviation	Coefficient of variation	Skewness	Kurtosis	Normality*
4/15/05	Mj	0.08	0	3.20	0.48	575.0	5.70	31.60	n
	Pz	3.56	2.00	4.40	0.46	12.8	-1.11	1.68	n
6/23/05	Mj	1.47	0	4.45	1.31	88.7	0.03	-1.32	n
	Pz	3.80	2.60	4.40	0.32	8.4	-0.68	0.96	n
8/26/05	Mj	0.58	0	3.97	1.16	200.6	1.66	1.14	n
	Pz	3.72	2.30	4.31	0.33	8.9	-0.80	2.59	n
10/24/05	Mj	0.23	0	3.38	0.74	362.2	3.13	8.50	n
	Pz	3.47	2.60	4.16	0.36	10.3	-0.29	-0.59	n
12/22/05	Mj	1.22	0	4.04	1.43	116.0	0.47	-1.46	n
	Pz	3.32	2.00	4.80	0.51	15.2	-0.28	-0.31	n
2/22/06	Mj	0.69	0	4.40	1.27	184.1	1.45	0.47	n
	Pz	3.27	2.00	4.25	0.44	13.6	-0.25	-0.13	N
4/28/06	Mj	0.74	0	3.85	1.23	166.2	1.20	-0.20	n
	Pz	3.57	2.00	4.58	0.57	16.0	-0.50	-0.25	n
6/27/06	Mj	0.79	0	3.85	1.31	166.6	-0.35	1.19	n
	Pz	3.29	0	4.56	0.75	22.8	9.18	-2.46	n
8/25/06	Mj	0.88	0	3.71	1.28	145.4	-0.78	0.94	n
	Pz	2.65	0	3.81	0.70	26.5	6.16	-20.40	n
10/26/06	Mj	0.34	0	3.11	0.82	241.5	2.58	2.08	n
	Pz	3.48	0	4.61	0.61	17.4	10.16	-2.02	n
12/28/06	Mj	1.15	0	3.98	1.40	121.3	-1.52	0.50	n
	Pz	3.98	2.90	4.65	0.33	8.2	0.97	-0.55	n
2/27/07	Mj	0.85	0	4.00	1.37	160.3	-0.73	1.05	n
	Pz	3.65	2.30	4.60	0.46	12.4	0.32	-0.50	n
4/25/07	Mj	0.85	0	3.86	1.32	154.1	-0.74	1.01	n
	Pz	4.01	2.70	4.76	0.37	9.2	0.90	-0.64	n
8/24/07	Mj	0.08	0	2.30	0.41	493.9	21.58	4.80	n
	Pz	2.79	0	3.88	0.78	28.1	5.83	-2.12	n

* N = normal distribution and n = not normal distribution, according to SHAPIRO and WILK (1965).

moisture are considered the most important abiotic factors (McSORLEY, 1998).

Soil temperature interferes with nematode survival, life cycle duration, and reproduction (NOE, 1991). For *M. incognita*, *M. javanica*, *P. zaeae*, and *P. brachyurus* the optimal range for development is between 25 and 30 °C (TAYLOR and SASSER, 1978). Consequently, in Brazilian sugarcane plantations soil temperature does not have a significant influence on nematode populations, because it is generally near the ideal range, with little seasonal variation, rarely reaching extreme values, lower than 15 °C or higher than 35 °C, which would be limiting for the development of these parasites.

On the other hand, the effect of soil moisture is very significant, either directly, on the activity and mobility of these parasites (SULTAN and FERRIS, 1991) or indirectly, by interfering with root development, which is the nematodes feeding site.

According to VASCONCELLOS and DINARDO-MIRANDA (2006), sugarcane begins to emit roots right after planting, and this process is maintained so long as soil moisture

is available. Under drought conditions, the root system becomes smaller, with the initial death of superficial roots, followed by deeper roots in extended drought periods. New roots are emitted only when soil moisture increases as a result of rains or irrigation. Therefore, in the present experiments, regular and abundant rains in the months that followed planting favored root growth, the feeding site of nematodes, indirectly benefiting their populations.

Because in 2005 regular rains occurred throughout the year, except in August, the nematode populations remained high in both experiment areas during that period. Significant reductions in nematode populations were observed only in the months ranging from June to October 2006, when the volume of rains was very small. Starting in November 2006, with the beginning of the rainy season, populations again increased, reaching another peak between the months of December and April (Figures 1 to 3).

These data are in agreement with those obtained by CARNEIRO et al. (1980), who also observed that nematode populations increased beginning in November with the rains and soil moisture, and decreased starting in

Table 2. Statistical parameters of transformed by log(x+1) data of *M. javanica* (Mj) and *P. zaeae* (Pz) populations in RB928064 sugarcane variety, in experiment 2

Sampling date		Mean (n° /50g roots)	Minimum value (n° /50g roots)	Maximum value (n° /50g roots)	Standart deviation	Coefficient of variation	Skewness	Kurtosis	Normality*
6/1. st /05	Mj	0.48	0	3.53	0.99	205.1	1.74	1.45	n
	Pz	3.57	2.30	4.53	0.41	11.6	-0.47	0.21	n
8/4/05	Mj	1.41	0	4.73	1.50	107.0	0.31	-1.55	n
	Pz	3.21	2.00	4.08	0.36	11.3	-0.46	0.46	n
10/6/05	Mj	1.19	0	4.90	1.51	128.3	1.72	-1.04	n
	Pz	3.23	0	4.12	0.71	22.1	-2.83	10.71	n
12/16/05	Mj	1.51	0	4.44	1.56	103.6	0.19	-1.76	n
	Pz	3.09	0	4.23	0.58	18.9	-1.72	6.77	n
2/10/06	Mj	1.08	0	3.75	1.34	124.4	0.58	-1.41	n
	Pz	3.25	0	4.21	0.67	20.6	-2.37	9.55	n
4/5/06	Mj	0.97	0	3.72	1.28	132.7	0.72	-1.19	n
	Pz	3.42	0	4.45	0.59	17.3	-2.08	10.13	n
6/21/06	Mj	0.71	0	3.57	1.13	159.1	1.07	-0.52	n
	Pz	2.81	0	4.37	0.79	28.3	-2.12	5.75	n
7/17/06	Mj	0.44	0	3.58	0.93	232.5	1.79	1.68	n
	Pz	2.45	0	3.85	1.01	41.2	-1.54	1.58	n
11/25/06	Mj	0.36	2.00	4.14	0.90	2143.2	2.26	3.97	n
	Pz	2.96	0	4.00	0.60	20.2	-1.51	4.68	n
2/14/07	Mj	0.28	0	3.36	0.84	288.9	2.71	5.85	n
	Pz	2.93	0	4.17	0.81	27.5	-2.28	6.63	n
4/20/07	Mj	0.54	0	3.70	1.11	205.5	1.66	1.00	n
	Pz	3.47	2.00	4.57	0.51	14.5	-0.73	0.34	n
6/15/07	Mj	0.20	0	3.14	0.67	319.0	3.16	8.90	n
	Pz	3.39	2.30	4.07	0.37	10.8	-0.43	-0.18	n

* N = normal distribution and n = not normal distribution, according to SHAPIRO and WILK (1965).

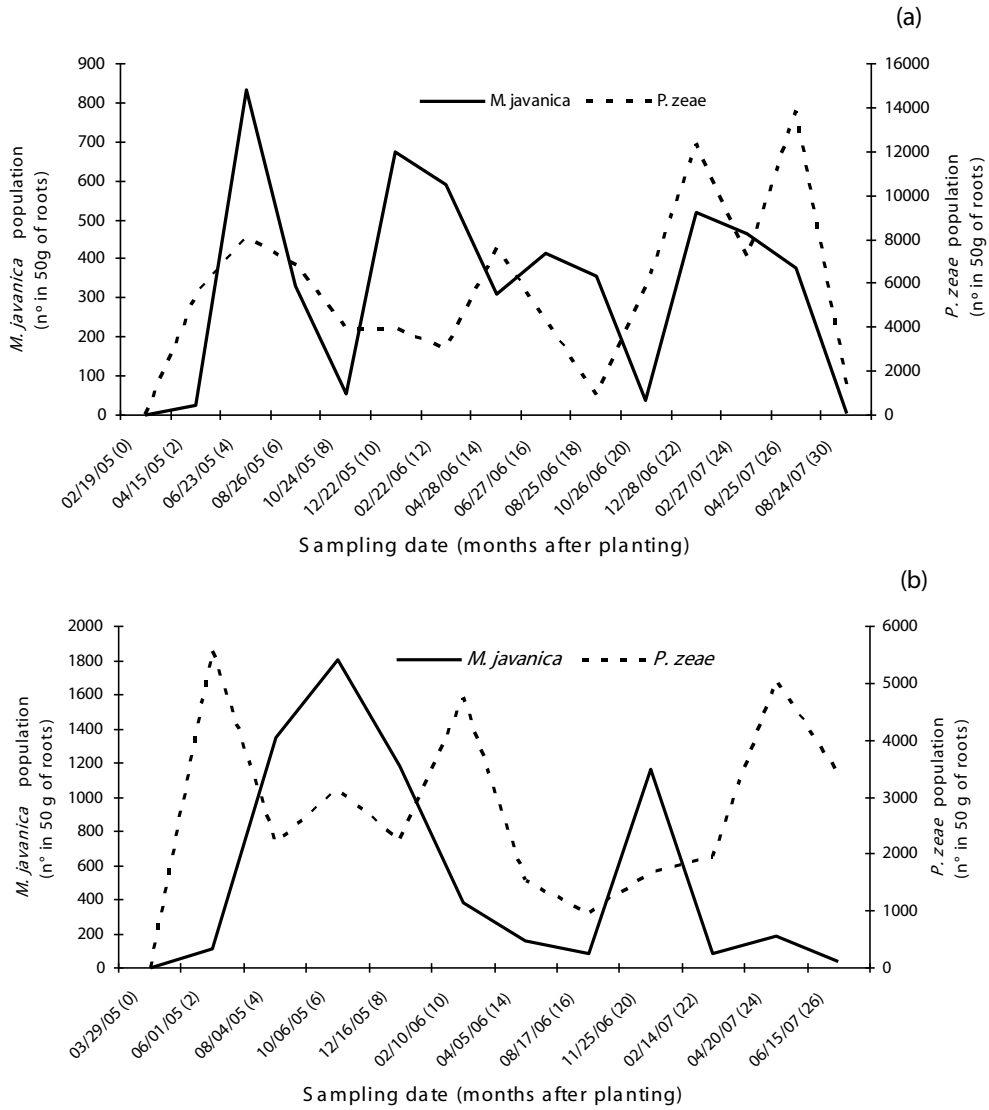


Figure 1. Population dynamics of *M. javanica* and *P. zeae* in roots of SP91-1049 (a, Experiment 1) and RB928054 (b, Experiment 2) sugarcane varieties.

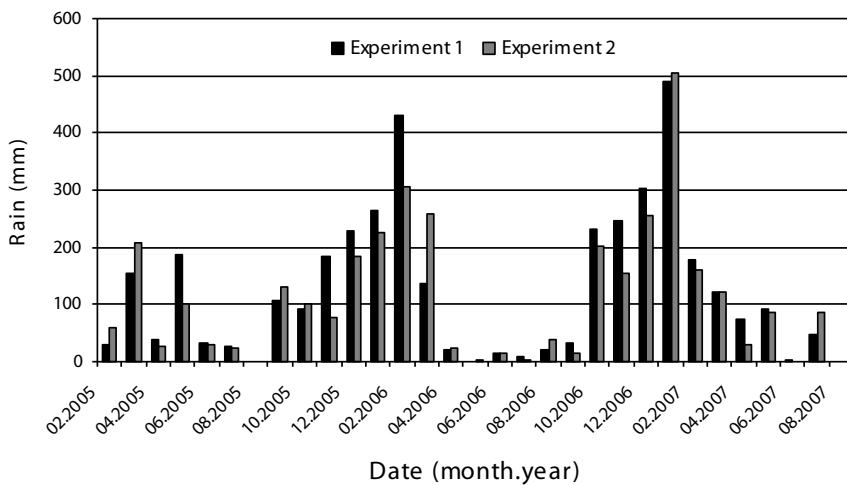


Figure 2. Rain fall during experimental period.

April, when the rains and consequently soil moisture decreasing.

Although soil moisture significantly interfered with *M. javanica* and *P. zae* population dynamics, the competition between both species also played an important role on population fluctuation, in certain seasons making one population of one species to increase while the population of the other decreased. This fact occurred in experiment 2 and between December and April 2006 in experiment 1. According to EISENBACK (1985), this illustrates the complexity of the relations between the various species of nematodes.

According to Shapiro and Wilk's test (SHAPIRO and WILK, 1965), only the *P. zae* population in the sampling made on 2/22/06 in experiment 1 had normal distribution (Tables 1 and 2). Despite the lack of normality in most data series, the analysis of transformed data showed that skewness and kurtosis values were near zero,

indicating they are nearly normal, and for this reason were considered adequate for geostatistical analysis. According to CRESSIE (1991), normality of data is not a requirement in geostatistics; it is simply convenient that distribution will not have very long tails, which could compromise the analyses.

Among the 52 calculated semivariograms, those corresponding to the *M. javanica* populations on 4/15/05, 10/24/05, 12/22/05, 2/22/06, 4/28/06, 8/24/06 and 10/26/06 in experiment 1, on 4/5/06, 6/21/06, 8/17/06, 11/25/06, 4/20/07, and 6/15/07 in experiment 2, and to the *P. zae* populations on 10/24/05, 6/27/06, 8/25/06, 10/26/06, 2/27/07, and 8/24/07 in experiment 1, and 8/4/05, 10/6/05, 6/21/06, and 8/17/06 in experiment 2 did not fit any model, showing a pure nugget effect (Tables 3 and 4). This means that the spatial distribution of nematodes in the above mentioned cases occurred simply at random, within the sampling distance used in the study, that

Table 3. Parameters of fitted semivariogram and jack kinifing, coefficient of determination (r^2), $C_0/(C_0+C_1)$ of *M. javanica* (Mj) and *P. zae* (Pz) populations, in experiment 1

Sampling date		Semivariogram parameters			Jack kinifing parameters (reduced errors)		r^2	$C_0/(C_0+C_1)$	Area (m^2) ¹
		C_0	C_1	a (m)	mean	Variance			
4/15/05	Mj				Pure nugget effect				
	Pz	0.13	0.08	40	-0.016	1.049	0.99	0.62	5026
6/23/05	Mj	1.25	0.50	38	0.0005	0.976	0.30	0.71	4536
	Pz	0.07	0.035	50	0.0030	1.075	0.99	0.67	7790
8/26/05	Mj	0.95	0.33	22	-0.0006	0.967	0.17	0.74	1521
	Pz	0.065	0.045	28	0.0007	0.990	0.56	0.59	2463
10/24/05	Mj				Pure nugget effect				
	Pz				Pure nugget effect				
12/22/05	Mj				Pure nugget effect				
	Pz	0.17	0.09	20	-0.0138	1.005	0.04	0.65	1257
2/22/06	Mj				Pure nugget effect				
	Pz	0.14	0.065	28	0.0002	1.011	0.62	0.68	2463
4/28/06	Mj				Pure nugget effect				
	Pz	0.20	0.125	35	0.014	1.034	0.69	0.62	3848
6/27/06	Mj	1.51	0.25	30	0.0038	1.013	0.99	0.86	2827
	Pz				Pure nugget effect				
8/25/06	Mj	0.9	0.8	42	0.0085	1.017	0.74	0.53	5542
	Pz				Pure nugget effect				
10/26/06	Mj				Pure nugget effect				
	Pz				Pure nugget effect				
12/28/06	Mj	1.0	1.1	65	-0.0026	1.150	0.01	0.48	13273
	Pz	0.075	0.035	55	0.0083	1.065	0.99	0.68	9503
2/27/07	Mj	0.8	1.3	75	-0.0007	1.099	0.82	0.38	17672
	Pz				Pure nugget effect				
4/25/07	Mj	1.25	0.61	80	-0.0390	1.081	0.43	0.67	20106
	Pz	0.08	0.05	25	0.0078	1.378	0.07	0.62	1964
8/24/07	Mj				Pure nugget effect				
	Pz	0.45	0.20	30	-0.0035	0.9877	0.98	0.69	2827

(¹) Area calculated by $\frac{C_0}{r^2}$, where $\frac{C_0}{r^2}=3.1416$ and $r = a$.

is, the distance between sampling points was too large to allow the detection of spatial dependence between them.

DINARDO-MIRANDA and FRACASSO (2009) also calculated semivariograms for ten nematode populations in sugarcane and were not able to fit four of them to any model.

For the other populations the spherical model provided the best fit to the semivariograms of the observed data (Tables 3 and 4, Figures 3 and 4). Although observed r^2 values were low, the parameters estimated for the spherical model (C_0 , C_1 , a) were endorsed by the jack knifing test, since the mean values for the reduced errors were near zero and the values for the variance of reduced errors were near 1 (Tables 3 and 4).

The portion of variability attributed to spatial dependence, given by the $C_0/(C_0 + C_1)$ ratio, varied from 0.39 to 0.86 in experiment 1 (Table 3) and from 0.12 to 0.73 in experiment 2 (Table 4). According to CAMBARDELLA et al. (1994), values below 0.25, between 0.25 and 0.75, and above 0.75 indicate strong, moderate, and weak spatial dependence between data, respectively. Consequently,

all variables showed moderate spatial dependence, except *M. javanica* on 8/4/05 in experiment 2, which showed strong spatial dependence, and *M. javanica* on 6/27/06 in experiment 1, with weak spatial dependence. The moderate degree of spatial dependence, found in most situations, indicates that the nematode distribution in these fields is aggregated and it is in agreement with results from other studies on the spatial distribution of nematodes, conducted in Brazil, such as those by FARIAS et al. (2002), who worked with *Rotylenchulus reniformis* in cotton and found a $C_0/(C_0 + C_1)$ ratio value of 0.27, and TORRES et al. (2006), who observed ratio ranging from 0.05 to 0.67, working with *R. reniformis* and Dorylaimidae and Aphelenchidae nematodes in melon. DINARDO-MIRANDA and FRACASSO (2009), who worked with *M. javanica*, *P. zaeae*, and mixed populations of *P. zaeae* and *P. brachyurus* in sugarcane, determined values for that ratio ranging from 0.29 to 0.68.

The range (a), which represents the distance up to which there is spatial dependence between samples, varied from 20 to 75 m in experiment 1 and from 30 to 95 m in experiment 2 (Tables 3 and 4), indicating that aggregation areas of *M. javanica* and *P. zaeae* in experiment

Table 4. Parameters of fitted semivariogram and jack kinifing, coefficient of determination (r^2), $C_0/(C_0+C_1)$ of *M. javanica* (Mj) and *P. zaeae* (Pz) populations, in experiment 2

Sampling date		Semivariogram parameters			Jack kinifing parameters (reduced errors)		r^2	$C_0/(C_0 + C_1)$	Area (m ²) ¹
		C_0	C_1	a (m)	mean	Variance			
6/1. st /05	Mj	0.40	0.65	35	0.0014	0.9266	0.5371	0.38	3848
	Pz	0.06	0.135	95	-0.0038	1.113	0.8424	0.31	13273
8/4/05	Mj	0.30	2.30	55	-0.0051	1.2240	0.8837	0.12	9503
	Pz				Pure nugget effect				
10/6/05	Mj	1.20	1.30	55	0.0011	0.9875	0.8779	0.48	9503
	Pz				Pure nugget effect				
12/16/05	Mj	1.75	0.75	45	0.0011	1.023	0.6122	0.70	6361
	Pz	0.15	0.20	35	-0.0025	1.169	0.1329	0.43	3848
2/10/06	Mj	0.70	1.20	30	-0.0088	1.122	0.5433	0.36	2827
	Pz	0.26	0.24	65	-0.0030	1.003	0.3696	0.52	13273
4/5/06	Mj				Pure nugget effect				
	Pz	0.16	0.23	55	0.0038	0.8355	0.4873	0.41	9503
6/21/06	Mj				Pure nugget effect				
	Pz				Pure nugget effect				
7/17/06	Mj				Pure nugget effect				
	Pz				Pure nugget effect				
11/25/06	Mj				Pure nugget effect				
	Pz	0.28	0.10	38	-0.0216	1.297	0.3889	0.73	4536
2/14/07	Mj	0.52	0.23	77	-0.0048	1.079	0.7899	0.69	18627
	Pz	0.42	0.30	77	-0.0013	0.9651	0.5640	0.58	18627
4/20/07	Mj				Pure nugget effect				
	Pz	0.13	0.16	60	0.0040	1.004	0.5688	0.45	11310
6/15/07	Mj				Pure nugget effect				
		0.09	0.052	55	-0.0090	1.066	0.1357	0.63	9503

(¹) Area calculated by $\frac{4}{\pi}r^2$, where $\frac{4}{\pi}=3.1416$ and $r = a$.

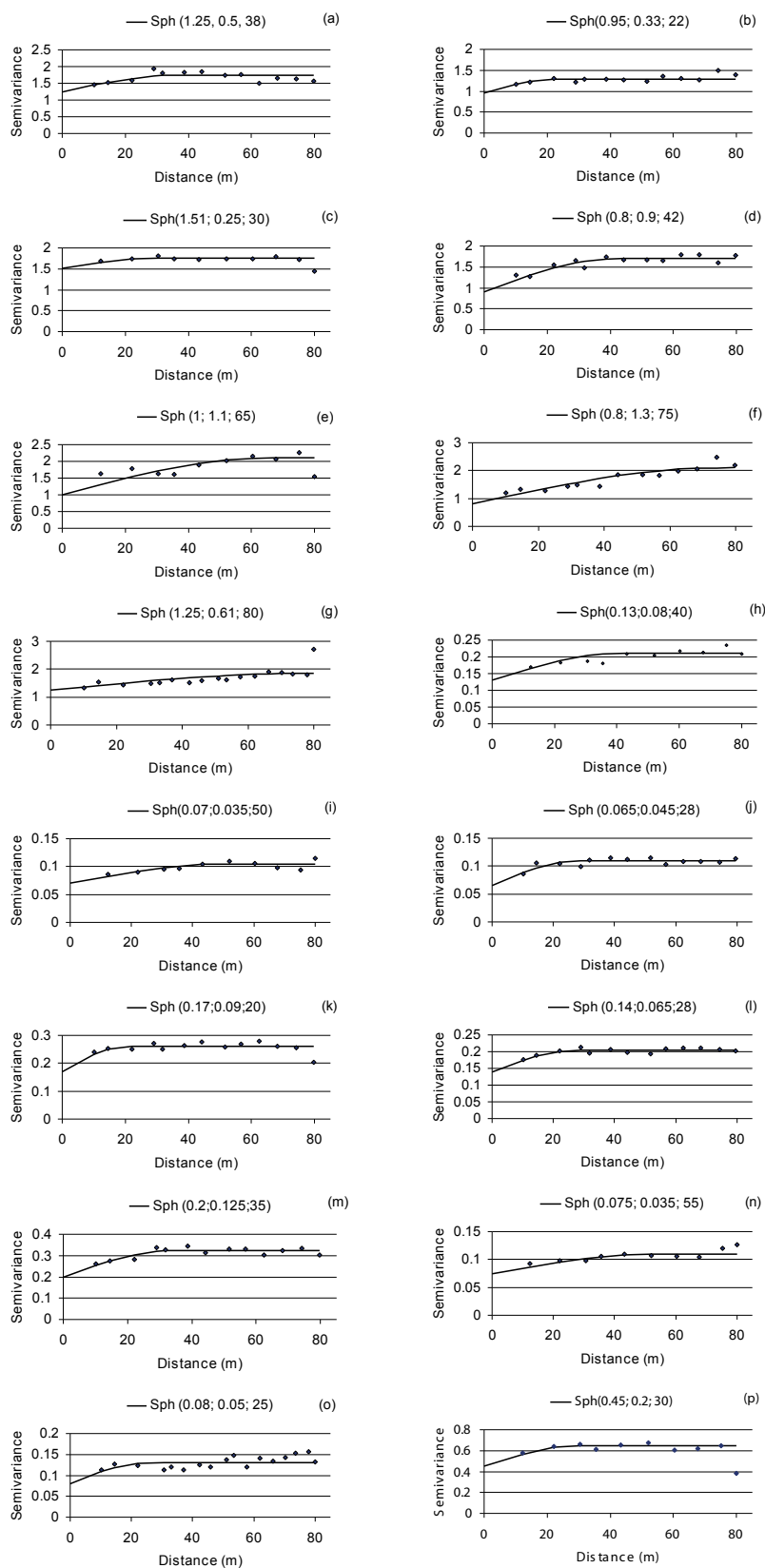


Figure 3. Semivariogram for populations of *M. javanica* (a, on 6/23/05; b, on 8/26/05; c, on 6/27/06; d, on 8/25/06; e, on 12/28/06; f, on 2/27/07 and g, on 4/25/07) and *P. zeae* (h, on 4/15/05; i, on 6/23/05; j, on 8/26/05; k, on 12/22/05; l, on 2/22/06; m, on 4/28/06; n, on 12/28/06; o, on 4/25/07 and p, on 8/24/07), in experiment 1. Numbers in parenthesis are nugget effect value (C_0), C_1 and range (a) of spherical model (Sph).

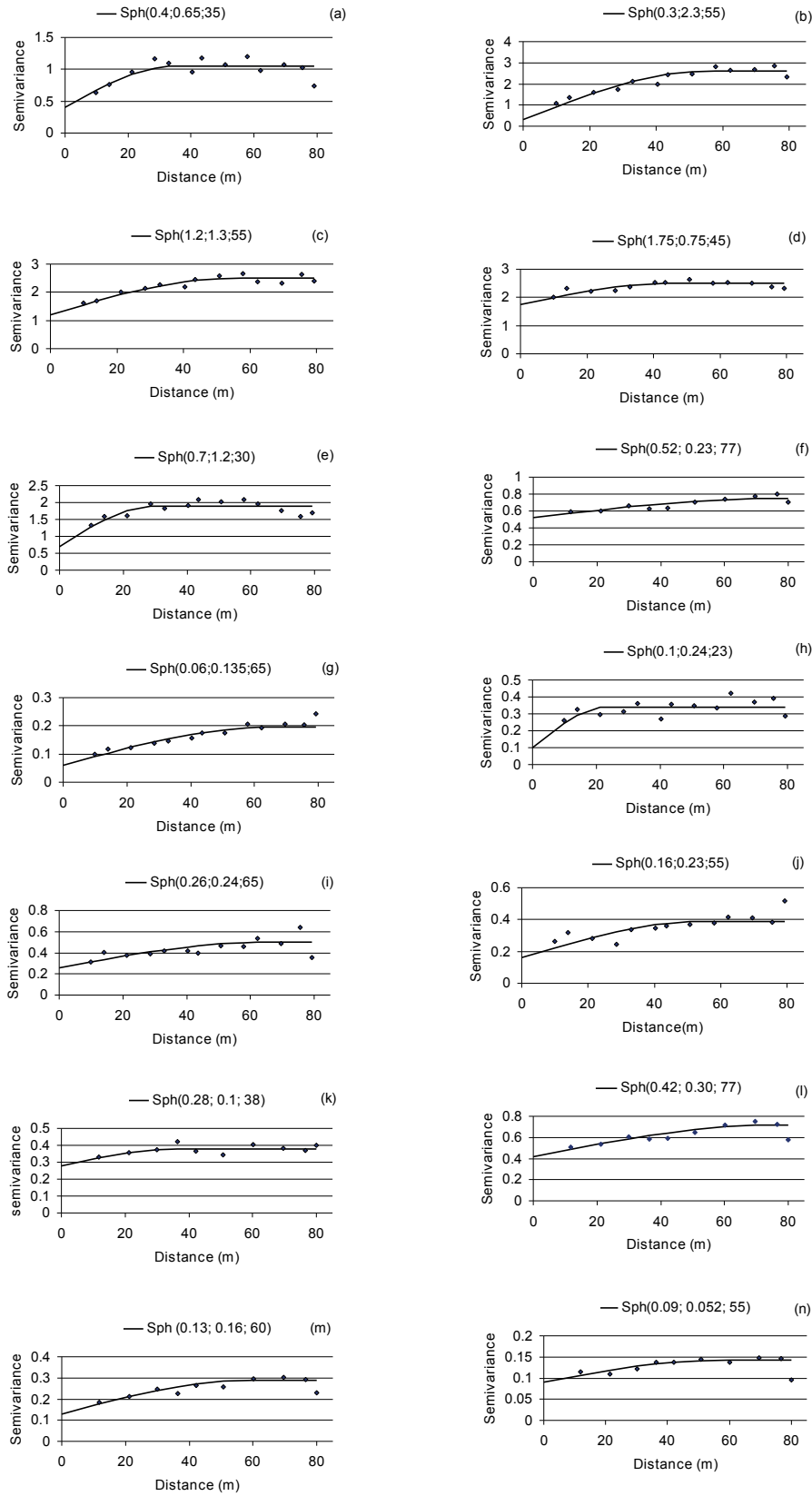


Figure 4. Semivariogram for populations of *M. javanica* (a, on 6/1st/05; b, on 8/4/05; c, on 10/6/05; d, on 12/16/05; e, on 2/10/06 and f, on 2/14/07) and *P. zeae* (g, on 6/1st/05; h, on 12/16/05; i, on 2/10/06; j, on 4/5/06; k, on 11/25/06; l, on 2/14/07; m, on 4/20/07 and n, on 6/15/07), in experiment 2. Numbers in parenthesis are nugget effect value (C_0), C_1 and range (a) of spherical model (Sph).

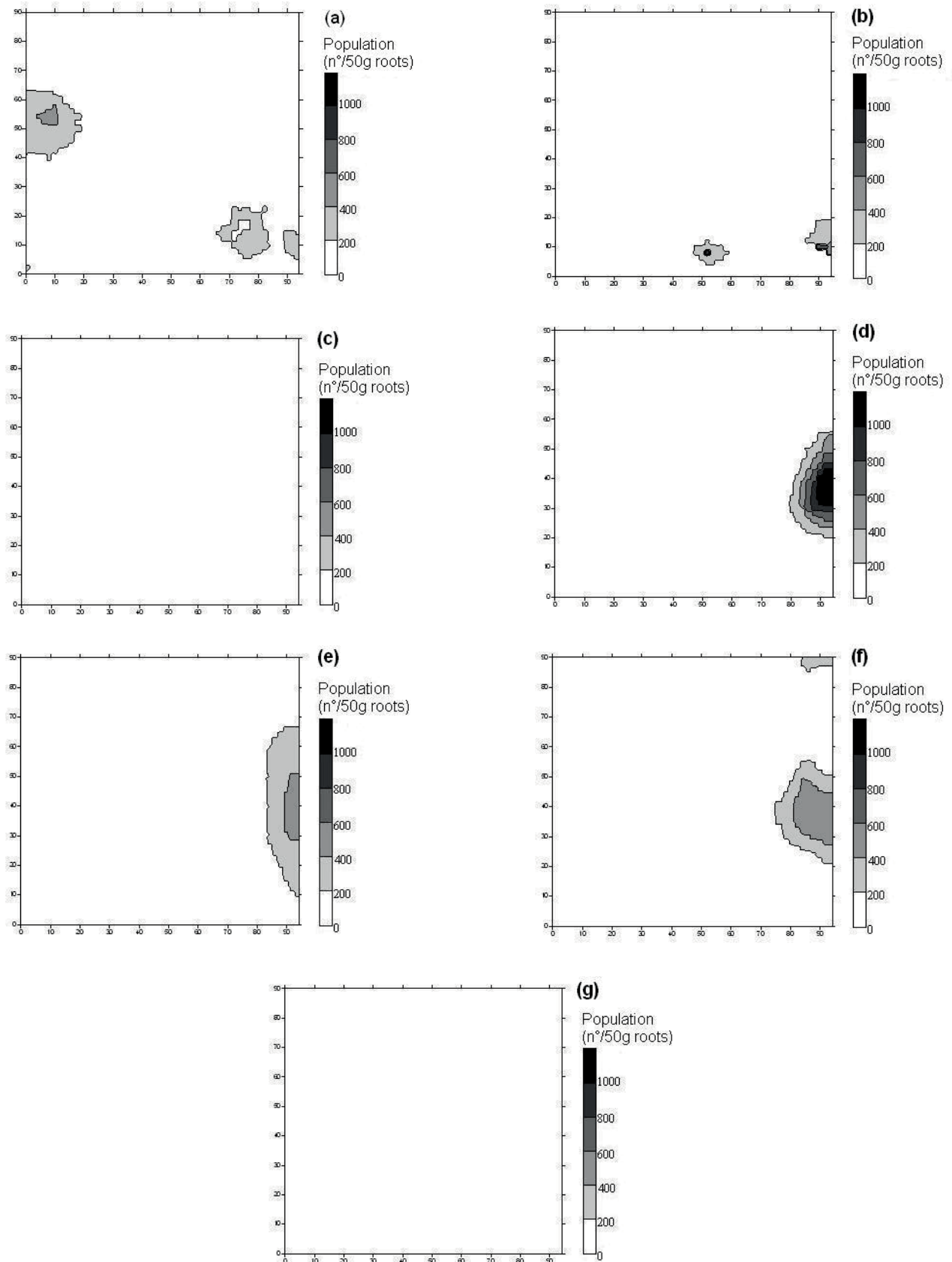


Figure 5. Maps of spatial distribution of *M. javanica* (a, on 6/23/05; b, on 8/26/05; c, on 6/27/06; d, on 8/25/06; e, on 12/28/06; f, on 2/27/07 and g, on 4/25/07), in experiment 1.

1 are smaller than those in experiment 2. The soil texture could be contributed to this result; According to VAN GUNDY (1985), nematodes migration decreases when the clay and silt fractions in the soil increase. However, further studies are needed to confirm this phenomenon.

Just like in the present study, other studies have also revealed that the spherical model best describes the spatial distribution of plant-parasite nematodes, with similar range values as those herein determined. In Brazil, FARIAS et al. (2002) worked with *R. reniformis* in cotton and determined a mean range of 15 m, and DINARDO-MIRANDA and FRACASSO (2008), in a study involving *M. javanica*, *P. zaeae*, and a mixture of *P. zaeae* and *P. brachyurus* in sugarcane, verified that range varied from 18 to 35 m. Working with sugarcane in Martinica, ROSSI et al. (1996) found range of 66.9 m to *P. zaeae*, but exponential model was used to fit the semivariogram of this specie.

The maps constructed based on the data are presented in Figures 5 to 8. In experiment 1, high values of *P. zaeae* population were found in several regions in the field, except in a small part on the right side of experimental area, where the highest populations of *M. javanica* were registered, indicating that competition between the species could be occurred. In experiment 2, the highest populations of *M. javanica* were in the upper side of the field, whereas the highest populations of *P. zaeae* were registered in the left side of the field. It is also possible to note that, in experiment 2, the maps for April 2006 (first year cane) and in April 2007 (ratoon) are very similar, indicating that few changes occurred in the area in relation to *P. zaeae* population, during the considered period.

Since population surveys in an integrated management program are intended to identify areas with nematode problems, the surveys should be carried out when populations are higher, that is, during the

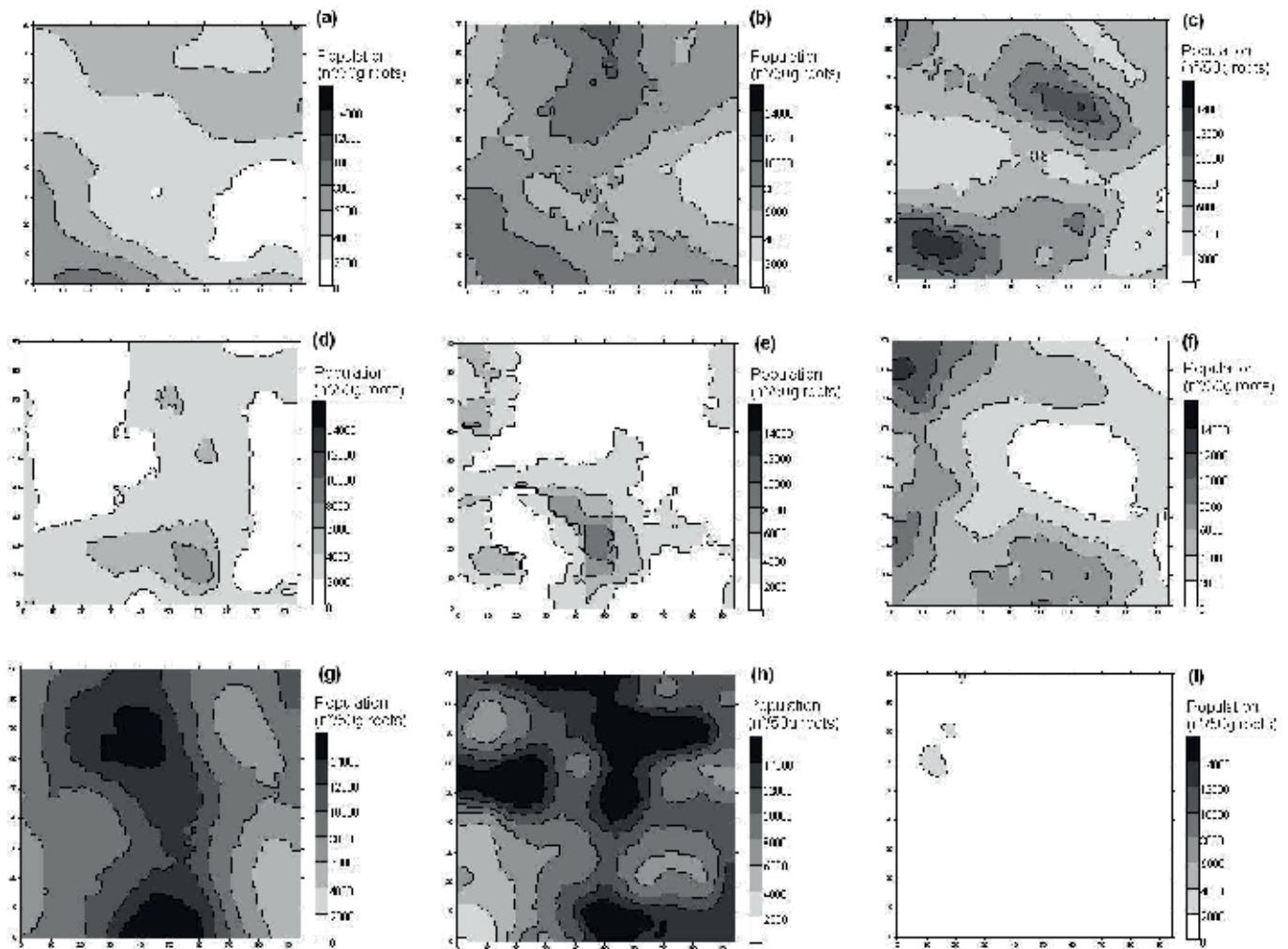


Figure 6. Maps of spatial distribution of *P. zaeae* (a, on 4/15/05; b, on 6/23/05; c, on 8/26/05; d, on 12/22/05; e, on 2/22/06; f, on 4/28/06; g, on 12/28/06; h, on 4/25/07 and i, on 8/24/07), in experiment 1.

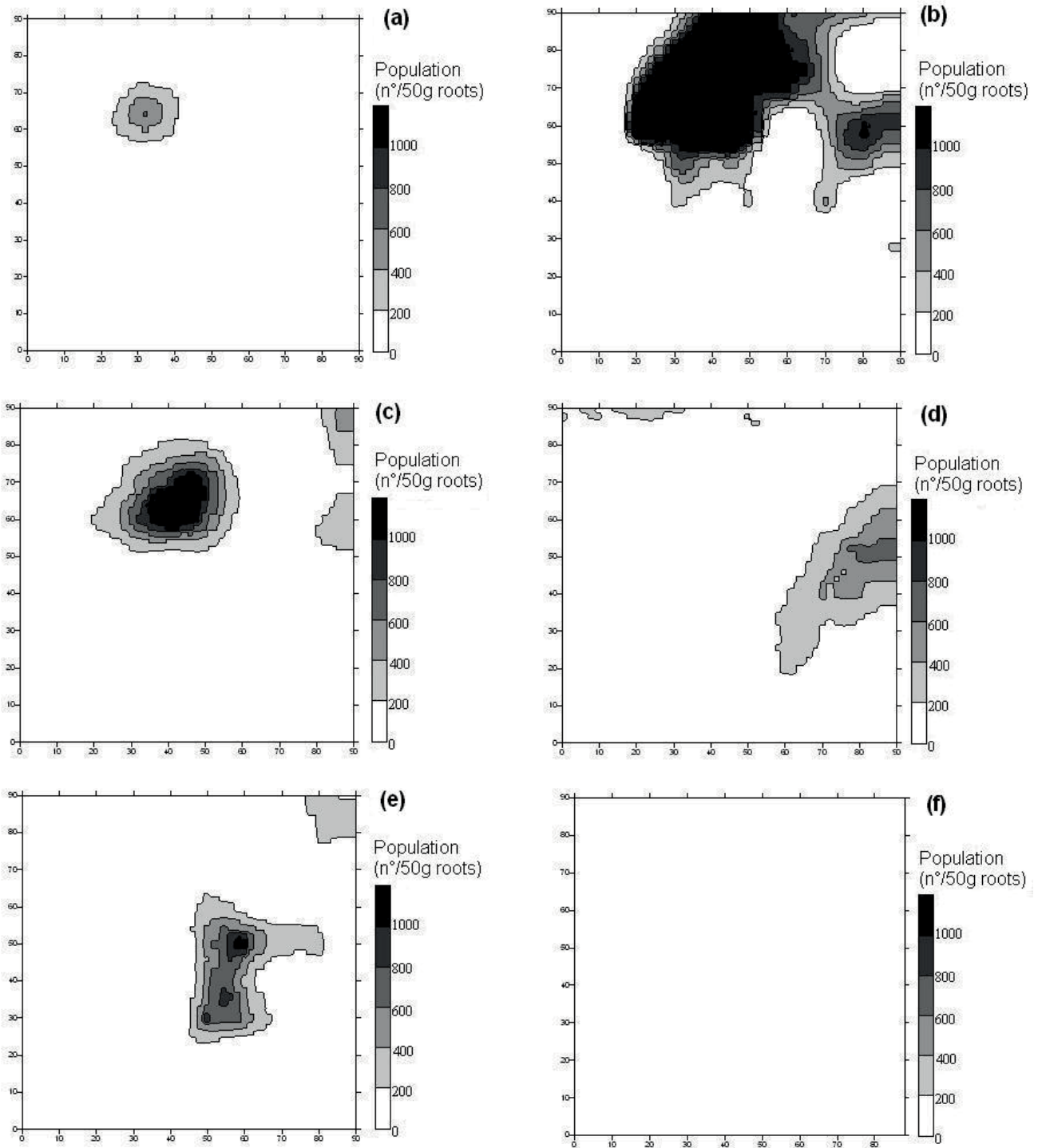


Figure 7. Maps of spatial distribution of *M. javanica* (a, on 6/1.st/05; b, on 8/4/05; c, on 10/6/05; d, on 12/16/05; e, on 2/10/06 and f, on 2/14/07), in experiment 2.

rainy period of the year. Therefore, in the conditions of our experiments, the best periods to perform samplings were between February 2005 and April 2006 and between October 2006 and April 2007. In those periods, the mean range for both nematode species in both experiments was 47 m, therefore revealing that samples separated from one another by smaller distances would

not be statistically independent. This information allowed to estimate the nematode aggregation area at about 7.000 m² ($A = \pi r^2$, where $r = a$) and suggests that samples would be collected 47 m apart in order to obtain a reliable estimate of the nematode population in a given area. This value is higher than the value found by DINARDO-MIRANDA and FRACASSO (2009), who

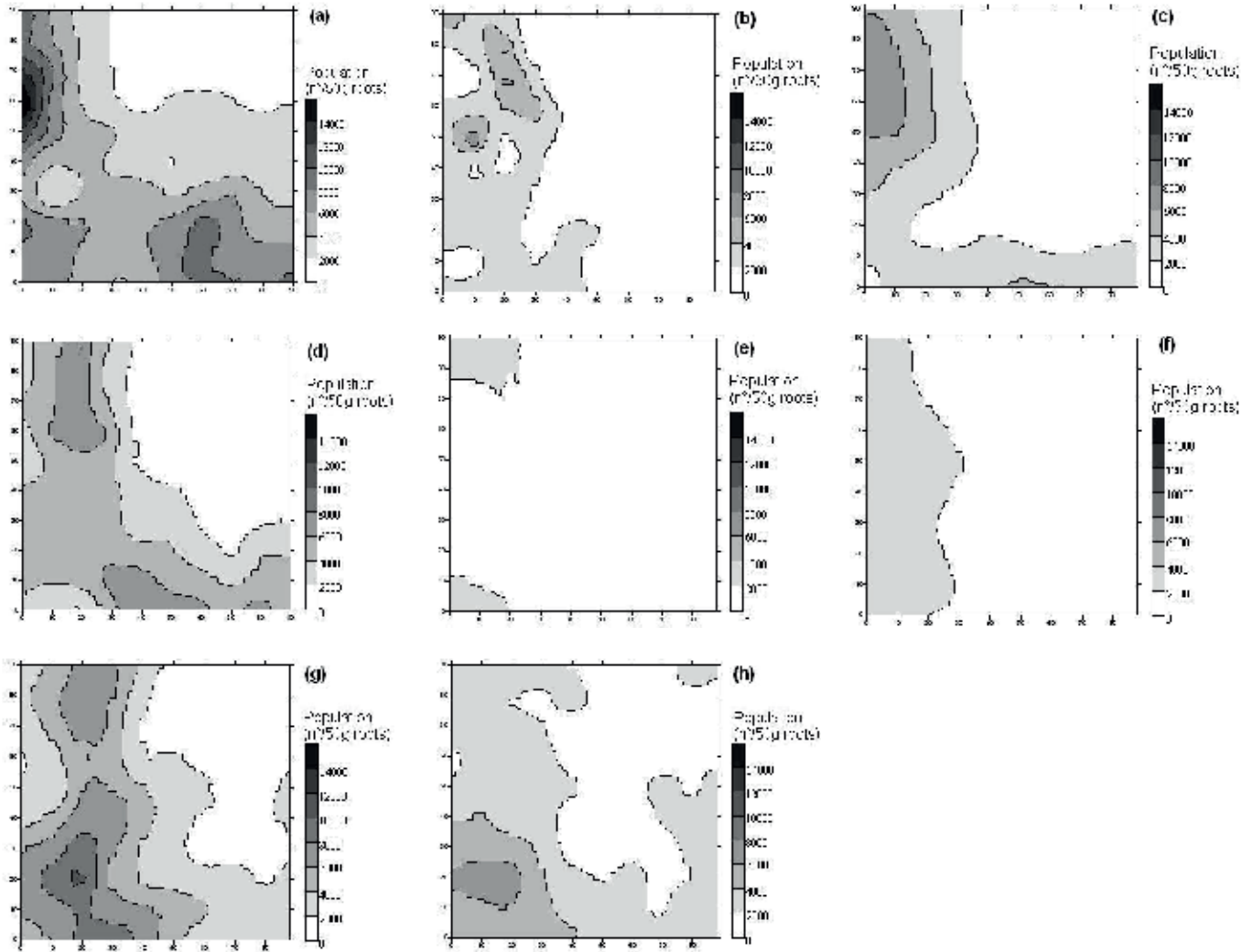


Figure 8. Maps of spatial distribution of *P. zeae* (a, on 6/1st/05; b on 12/16/05; c, on 2/10/06; d, on 4/5/06; e, on 11/25/06; f, on 2/14/07; g, on 4/20/07 and h, on 6/15/07), in experiment 2.

determined that sampling points should be adopted 27 m apart, on average, to estimate nematode populations, in a study in which six areas were sampled. It must be pointed out, however, that the commercial fields of sugarcane in Brazil involve a great number of varieties, grown in a wide variety of production environments; all these factors interfere with the spatial distribution of nematodes and explain the differences observed in the present work when compared with the study of DINARDO-MIRANDA and FRACASSO (2009).

Hence, the data in the present work suggest that in order to obtain a reliable estimate of nematode populations, samplings should be taken in the rainy period of the year and should be collected 47 m apart, meaning at least 1.5 sample collection points per hectare.

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