

## HEPARIN-ANTIVENOM ASSOCIATION: DIFFERENTIAL NEUTRALIZATION EFFECTIVENESS IN *Bothrops atrox* AND *Bothrops erythromelas* ENVENOMING

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### SUMMARY

Heparin, in some regions of Brazil has been used in the treatment of bothropic accidents, but the data found in the literature are inconclusive about its effectiveness. The venoms of *Bothrops atrox* and of *B. erythromelas* were characterized according to their biological activities. The capacity of heparin in neutralizing these activities was tested with doses of 3 and 6 IU in isolated form and associated to Antithrombotic Serum (ABS). It was verified that heparin, in doses of 3 and 6 IU, was not effective in neutralizing the defibrinating and edema-forming activities of *B. atrox* venom and the hemorrhagic and coagulant actions of both venoms. Heparin diminished the effectiveness of the ABS in the neutralization of the hemorrhagic and edema-forming activities of the *B. atrox* venom. However, heparin in the 6 IU dose was capable of neutralize the edema-forming of the *B. erythromelas* and increase the effectiveness of the ABS. Heparin also neutralized the phospholipase A<sub>2</sub> activity of *B. atrox* (14.3%) and *B. erythromelas* (28.0%) venoms. For *B. erythromelas* venom, the associated treatment, heparin and ABS, was more effective in the neutralization of its lethal activity.

**KEYWORDS:** Heparin; Neutralization; *Bothrops erythromelas*; *Bothrops atrox*; Venom.

### INTRODUCTION

In Brazil, snake venoms accidents represent an important public health problem. Around 90% of which are caused by snakes of the *Bothrops* genus. The species *Bothrops jararaca* (South and Southeast), *Bothrops moojeni* (Center-West), *Bothrops erythromelas* (Northeast) and *Bothrops atrox* (North) which are responsible for most of these accidents, with humans, are distributed in different regions of Brazil<sup>5</sup>.

Despite the interspecies and intra-species variations existent in the chemical composition of venom, generally, the signs and symptoms presented by patients that have had accidents with *Bothrops* genus are: (a) local - pain, edema, equimosis, hemorrhage, myonecrosis; (b) systemics - blood incoagulability, hemorrhages being it possible to also occur shock and acute kidney failure. In severe cases of envenoming, these local effects may lead to permanent tissue loss, disability or amputation<sup>18,25</sup>.

The effective therapeutic treatment for ophidian accidents is serotherapy. Antivenom serotherapy, however, although efficient for neutralization of systemic manifestations of bothropic venom, contribute little to the improvement of the local tissue damage<sup>6</sup>. Experiments on mice, show that antivenoms are effective in neutralizing necrosis and local hemorrhage, only when injected at the same time or immediately after poison inoculation<sup>15,26</sup>. Therefore, new therapies parallel antivenom

injection, would be fundamental in minimizing local effects induced by snakes venoms, specially from *Bothrops* genus.

The use of heparin as a treatment in snake venom accidents, was first proposed in the late 40's by AHUJA *et al.*<sup>2</sup> and AHUJA & SINGH<sup>1</sup>.

Heparin is a family of glycosaminoglycans, highly sulfated, with molecular weight that varies between 5,000 and 40,000, capable of interacting with the antithrombin III, producing anticoagulant effect. The clinical use of heparin has been secured as anticoagulant and antithrombotic, but, recently, a possible anti-inflammatory action has been suggested<sup>21</sup>.

Due to its strongly polyanionic nature, heparin can interact with many molecules that have cationic sites<sup>33</sup>. Among the substances capable of interacting with heparin, are included proteins from the extracellular matrix (fibronectin, laminar, vitronectin), proteins involved in lipid metabolism, components of the complement system, serine protease inhibitors, viral proteins, and enzymes<sup>21,22,33</sup>. Within the enzymes, heparin can interact with phospholipase A<sub>2</sub>, present in many snake venoms, affecting<sup>8,9,12</sup>, or not<sup>7,19</sup> the enzymatic activity of these proteins.

TINOCO<sup>30</sup> in a clinical and experimental study with dogs, suggested the use of heparin in substitution of specific serum therapy in the case of poisoning by *Bothrops jararaca*.

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In 1975, NAHAS *et al.*<sup>27</sup> demonstrated that heparin was not capable of neutralizing the thrombin-like activity of bothropic venoms and in prevention of the desfibrinating syndrome, induced by *Bothrops jararaca* venom. Nevertheless, they observed that heparin was capable of neutralizing the venom that acts only on factor X. Those venoms that act directly on the fibrinogen and on factor X, did not have their coagulant activity blocked by heparin.

Heparin neutralized the myotoxic activity of bothropstoxin enzyme of *Bothrops jararacussu* venom. This protein belongs to phospholipase A<sub>2</sub> group that do not have neurotoxic activity<sup>23,24</sup>. Heparin inhibited *in vitro* the citotoxic activity of *Bothrops asper* venom and also neutralized *in vivo*<sup>22</sup> its myotoxic activity.

The present experiments were undertaken to verify the action of commercial heparin (Liquemine-Roche) within the main biological activities (hemorrhagic, *in vitro* and *in vivo* coagulant, edematogenic, phospholipasic and lethality) of the *Bothrops atrox* and *Bothrops erythromelas* venoms, that are responsible for most accidents in the North and Northeast of Brazil, respectively.

## MATERIAL AND METHODS

**Venoms:** Lyophilized venom of *Bothrops atrox* was obtained at the Centro de Animais Peçonhentos do Instituto de Medicina Tropical do Amazonas, Manaus (AM), Brazil. The freezing dried venom of *Bothrops erythromelas* was kindly given by Prof. Dr. Augusto S. Abe of the Departamento de Zoologia, Universidade Estadual de São Paulo, Rio Claro (SP), Brazil. The venoms were kept at -20 °C and diluted at the moment of the use.

**Animals:** Outbred Swiss mice (18 - 22 g), females proceeding from the Instituto Nacional de Pesquisas da Amazônia (INPA - Manaus, Amazonas, Brazil) were used in the experiments. The animals were kept in plastic cages receiving water and food *ad libitum* under controlled temperature conditions.

**Bovine Fibrinogen:** Samples of lyophilized bovine fibrinogen were donated by the Seção de Hematologia do Instituto Butantan (São Paulo, SP, Brazil).

**Heparin (HEP):** Heparin used in the experiments was purchased from Roche (Liquemine®).

**Antithrotoppic Serum (ABS):** The antithrotoppic serum used in the neutralization experiments was produced by Instituto Butantan (São Paulo, SP, Brazil) -Lot 9406081.

### Biological activities of venoms

**Lethal Dose 50% (LD<sub>50</sub>):** For determination of the LD<sub>50</sub>, different doses of venoms were injected by intraperitoneal route (i.p). Four mice were used for each venom dose. Deaths were recorded after 24 and 48 hours. The LD<sub>50</sub> were calculated by the probit analysis<sup>11</sup>.

**Minimum Hemorrhagic Dose (MHD):** Hemorrhage was quantified by the method of GUTIÉRREZ *et al.*<sup>16</sup>. Four mice were used for each venom dose. The venom doses were injected by intradermic route. Mice

were killed by ether inhalation 2 hr after injection, and the areas of hemorrhage measured on the internal surface of the dissected skin. The minimum hemorrhagic dose was defined as the amount of venom which induced a hemorrhagic area of 10 mm diameter.

**Minimum Coagulant Dose:** The plasma and fibrinogen minimum coagulant doses were estimated as previously described<sup>29</sup>. The minimum coagulant dose of a venom is defined as the minimum amount of venom resulting in clot formation within 60 sec at 37 °C. The tests were done in triplicate.

**Minimum Defibrinagenating Dose (MDD):** For defibrinating activity the method described by GENÉ *et al.*<sup>13</sup> was used. Briefly, five doses for each venom were used and, for each dose, four mice. The venom doses were inoculated by intravenous route. One hour after the injection of the venom, blood samples (0.1 ml) were collected from the animals. Blood was placed in tubes and kept at 37 °C until clotting occurred.

**Minimum Edema-forming Dose (MED):** The minimum edema-forming doses were determined according to the procedures of YAMAKAWA *et al.*<sup>32</sup> with some modifications. Briefly, hind paw edema was produced by a subplantar injection of 50µl venom solution or an equal volume of saline into the right and left hind-paw of mice, respectively. The paws (right and left) were measured with a calliper (Mitutoyo) at 0, 30, 60, 120, 180, 240, 300, 360 minutes. The minimum edema-forming dose was defined as the amount of venom that induced an edema of 30%.

**Determination of the Heparin Doses:** The heparin doses were obtained with basis on that used in human therapeutics (60 kg of body mass) and calculated for mice with 20 grams of body mass.

**Anticoagulant Effect of the Heparin:** The doses of heparin obtained by the previous method were injected in mice, by intravenous route and the effect of each dose on the blood coagulation was observed. For each dose, four mice were used.

### Effects of Heparin on the biological activities of Venoms

**Group Schemes for the neutralization experiments:** For all the neutralization experiments (hemorrhagic, edematogenic, desfibrinating and lethality activities) six groups of four animals each were used, for each venom, and received the following treatment: group I (GI) - venom in the corresponding doses for each activity (venom control); Group II (GII) - venom and heparin in a 3 IU dose; Group III (GIII) - venom and heparin in a 6 IU dose; Group IV (GIV) - venom and Antithrotoppic Serum (ABS); Group V (GV) - venom, ABS and 3 IU heparin; Group VI (GVI) - venom, ABS and 6 IU heparin. The doses of heparin were injected by intravenous route (e.v). A 100µl ABS dose was used for each animal and injected (e.v) in all experiments. Heparin and ABS were injected immediately after injection of the venom (zero time).

• **Hemorrhagic:** The challenge dose used, for each venom, in the neutralization of the hemorrhagic activity corresponded to three times the Minimum Hemorrhagic Dose and was injected by intradermic route at zero time. The animals were sacrificed two hours later, with ethylic ether and the hemorrhagic halos were measured.

- **Edematogenic:** The challenge dose was equivalent to three times the Minimum Edematogenic Dose of the corresponding venom and was injected in the right paw. Saline, used as control, was injected in the left paw, as described previously. The paws were measured at times 0, 30, 60, 120, 180, 240, 300 and 360 minutes after injection of the venom and saline.
- **Desfibrinating:** The dose used, for each venom, corresponded to three times the Minimum Desfibrinating Dose and was injected by i.v. route, at zero time. Blood samples were collected one hour after the injection, kept at 37 °C and the time of coagulation, for each blood sample, was observed for one hour.
- **Lethality:** A 250 µg challenge dose was used for each venom injected by i.v. route, at zero time. The time of death and the reactions presented by the animals were observed and noted. The animals were submitted to necropsy and the organs (liver, lungs, kidneys, brain and heart) kept in a 10% formaldehyde solution. Histopathological analysis was performed by classical techniques.
- **Coagulant Activity:**
  - *On the fibrinogen.* The dose of venom used was equivalent to two times the minimum coagulant dose on the fibrinogen (MCD-F). The venom (2 MCD-F) was incubated at 37 °C, along with the different doses of heparin (3, 6, 12, 24, 48 IU) for 30 minutes. After incubation, 100µl of each venom solution and HEP were added to 400µl of Bovine fibrinogen (2g/L) and the time of coagulation was observed.
  - *On the plasma.* The dose of venom used was equivalent to two times the minimum coagulant dose on human plasma, used as substrate, following the methodology mentioned above.
- **Phospholipase A<sub>2</sub> Activity (Indirect Hemolytic Assay):** The method described by GUTIÉRREZ *et al.*<sup>17</sup> was used to evaluate this activity. 100µl samples of venom solution (10mg/ml) were incubated with 100µl of heparin, ABS and saline, at 37 °C, for 30 minutes. After incubation, 10µl of each mixture was added to wells, previously covered with agarose containing sheep erythrocytes and egg yolk. The hemolysis halos were measured 24 hours after sample application.
- **Statistical Analysis.** The T Student test was applied, using the Statgraphics Plus statistical program to verify the statistical difference between groups. The value of p=0.05 was defined.

## RESULTS AND DISCUSSION

**Characterization of Biological Activities of the Venoms:** The values obtained for the minimum doses of venoms activities are showed in Table 1. These values were similar to the ones described by other authors<sup>3,10,28</sup>. The venom of the *Bothrops erythromelas* has no coagulant activity on bovine fibrinogen. This venom, however, was more toxic, for the lethal and coagulant activities on the plasma, than the venom of the *B. atrox* and less active, for the edematogenic, hemorrhagic and desfibrinating actions. Although FERREIRA *et al.*<sup>10</sup> showed that hemorrhagic and edematogenic actions are related to the lethality of bothropic venom, this fact was not observed in this experiments.

**Determination of Experimental Doses of Heparin:** The doses of heparin used on mice corresponding to the ones injected, in human therapeutics, for individuals weighing 60kg. From the blood samples collected from the animals that received different doses of heparin (3, 6, 12, 24 and 48 IU) it was observed that the blood coagulation time remained normal (<5 minutes) for the animals that received 3 and 6 IU heparin doses. The time of blood coagulation was altered, occurring the formation of microclots and coagulation after 30 minutes, for the animals that received the 12 UI dose; and uncoagulative (> 60 minutes) for the animals that received the 24 and 48 IU doses. Based on these results, 3 and 6 IU heparin doses were chosen for the neutralization tests, since the higher doses would induce alterations in the blood coagulation.

**Study of Neutralizing Effect of Heparin on the Coagulant Activity:** Heparin was capable of neutralizing the thrombin time coagulant activity of the *B. atrox* (Table 2) only in the doses over 12 IU (equal to 36,000 IU in human therapeutics).

**Table 1**

Establishment of the minimum doses for the venom of *Bothrops atrox* and *Bothrops erythromelas*

Biological Activities	<i>Bothrops atrox</i>	<i>Bothrops erythromelas</i>
Lethality (LD <sub>50</sub> )	5.6 mg/kg (4.4 – 6.6 mg/kg)	3.0 mg/kg (2.5 – 3.5 mg/kg)
Coagulant on Fibrinogen (MCD-F)	77.5 mg/l	*
Coagulant on Plasma (MCD-P)	150.0 mg/l	52.5 mg/l
Edematogenic (MED)	1.2 µg/animal	1.4 µg/animal
Hemorrhagic (MHD)	2.9 µg/animal	5.9 µg/animal
Desfibrinating (MDD)	10.0 µg/animal	45.0 µg/animal

\* *B. erythromelas* venom did not present coagulant activity on the bovine fibrinogen.

Heparin did not present, in the tested doses, inhibitory effect of the coagulant action of *B. atrox* venom on the human plasma (Table 2). However, in the 12, 24 and 48 IU doses, heparin was capable of partially neutralize the coagulant action of *B. erythromelas* venom on the plasma (Table 2).

NAHAS *et al.*<sup>27</sup> demonstrated that heparin neutralizes only the coagulant activity of venoms that act selectively on the factor X of the blood coagulation system. This is the reason that heparin did not inhibit the coagulant activity of *B. atrox* venom, since this venom acts on factor X, on the prothrombin, and on the fibrinogen (thrombin-like action). The *B. erythromelas* venom which acts on the factor X and does not have thrombin-like activity, explains the partial neutralization by heparin of the coagulant activity on the human plasma.

**Neutralizing Effect of Heparin on the Desfibrinating Activity:** The

**Table 2**  
Neutralization of the coagulant activity of the venoms

Heparin (IU)	Bovine Fibrinogen (min.)		Human Plasma (sec.)	
	<i>B. atrox</i>	<i>B. erythromelas</i>	<i>B. atrox</i>	<i>B. erythromelas</i>
-	<2	*	51	62.5
3	<2	*	48	62
6	<2	*	54	63
12	>60	*	55	91
24	>60	*	63	127
48	>60	*	44	146

\* *B. erythromelas* venom did not present coagulant activity on the bovine fibrinogen.

results presented in Table 3 demonstrate that heparin was not capable of neutralizing the desfibrinating action (MDD), *in vivo*, of the *B. atrox* venom in the doses tested. On the other hand, the antithrombotic serum (ABS) was capable of totally neutralizing this activity, and its action was not affected when associated with heparin (Table 3).

**Table 3**  
Neutralization of the desfibrinating activity of *Bothrops atrox* venom by heparin

Heparin (IU)	Venom	Antithrombotic	Time of Coagulation (min)
-	+	-	> 60
3	+	-	> 60
6	+	-	>60
-	+	+	< 10
3	+	+	< 10
6	+	+	< 10

Due to the proximity of the Lethal Dose 50% and the Minimum Desfibrinating Dose (MDD) of *B. erythromelas* venom (see Table 1), it was not possible to study the neutralizing effect of the heparin, since the animals died before one hour after the injection of three times the MDD.

**Neutralizing Effect of Heparin on the Hemorrhagic Activity:** Heparin did not neutralize the hemorrhagic activity of *B. atrox* venom

as shown in Table 4. There was a significant dose-dependent increase of the hemorrhagic halo produced by the venom, compared to the control, when the animals received heparin (3 and 6 IU doses) at the same time (Table 4).

The antithrombotic serum (ABS) was capable of neutralizing 38% of the hemorrhagic activity of *B. atrox* venom. The antivenom's action was totally neutralized when the animals were treated with ABS and heparin in the 3 and 6 IU doses, being observed, in these groups, significant increases of the hemorrhage compared to the group that received antithrombotic serum, alone. MELO *et al.*<sup>24</sup> reported that polyanions with high molecular weight, as heparin family, are not effective in neutralizing the hemorrhagic activity of *B. jararacussu* venom. LOMONTE *et al.*<sup>22</sup> demonstrated through histological sections that hemorrhagic activity of *B. asper* venom was potencialized by addition of heparin standard solution, which has high molecular weight polyanions capable of interacting with the antithrombin III, promoting the anticoagulant effect. In addition, when low weight heparin was used, the increase in the hemorrhagic activity was not observed<sup>22</sup>.

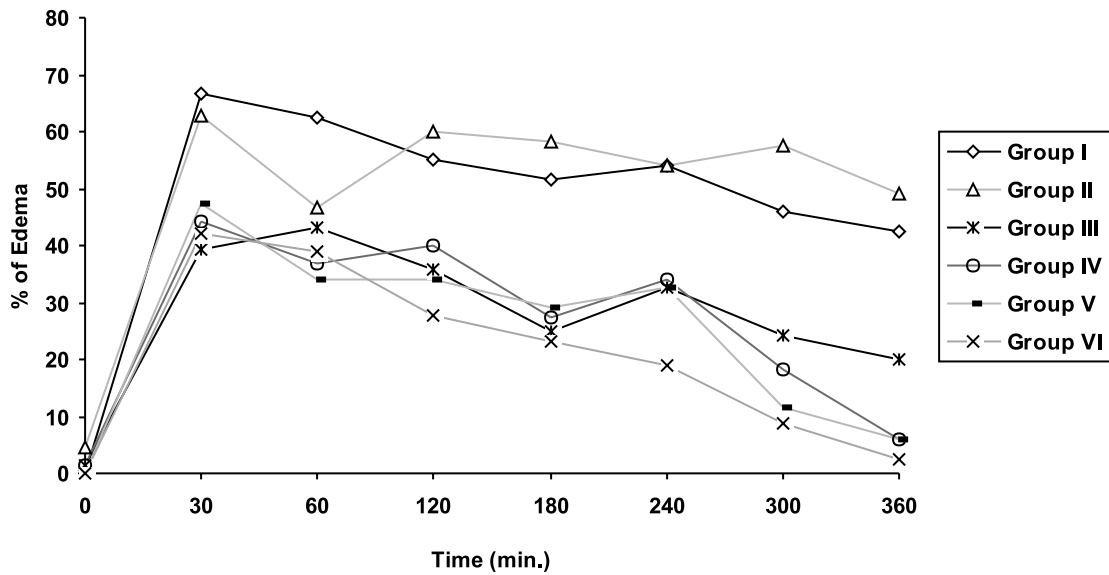
There was no significant difference between the hemorrhage produced by *B. erythromelas* venom in the control and experimental groups that received heparin (3 and 6 IU doses). The bothropic antivenom, on the other hand, was capable of neutralizing 57% of the hemorrhagic activity, without having its action harmed by the heparin in the studied doses - 3 and 6 IU.

**Neutralizing Effect of Heparin on the Edematogenic Activity:** Figure 1 shows the action of heparin, in the 3 and 6 IU doses, on the kinetics of edema formation produced by the venom of *B. atrox* in the dose corresponding to three times the Minimum Edema-forming Dose (MED). It was observed that the antithrombotic serum reduced, in a significant way, the edema from 180 minutes on, obtaining its maximum effect of neutralization (44.6%), in 360 minutes. GUTIÉRREZ & LOMONTE<sup>18</sup> demonstrated that the antivenoms are not capable of totally neutralizing the edematogenic activity. Heparin, by itself, did not induce a significant reaction in the edema levels. Interestingly, the presence of heparin decreased the anti-edematogenic activity of the antiserum. The results presented for the edematogenic action of *B. atrox* venom show that heparin was not effective in neutralizing this activity.

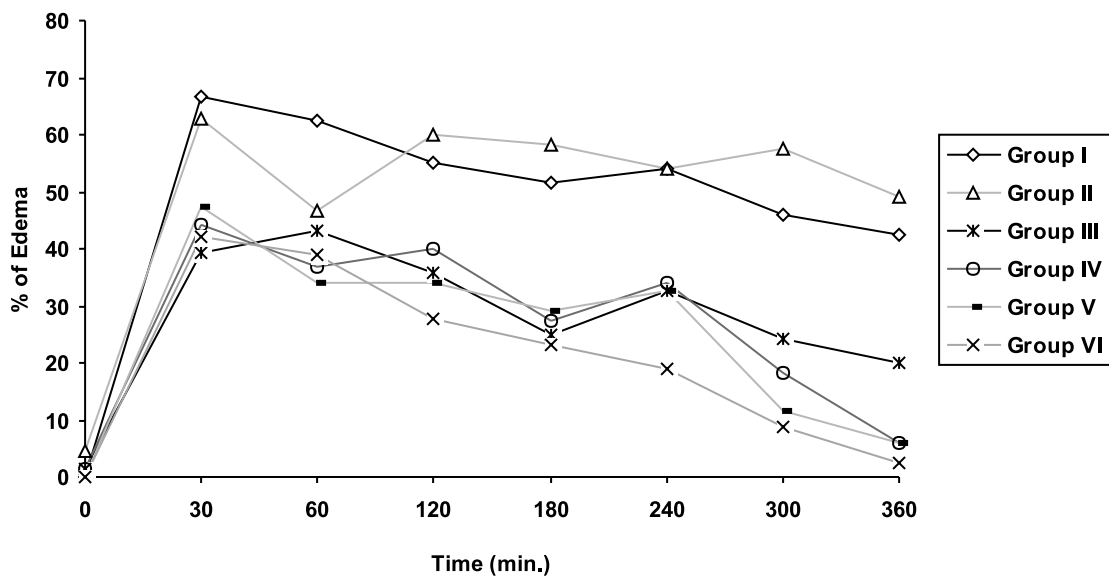
Effects of heparin, in the 3 and 6 IU doses, on the edematogenic activity of *B. erythromelas* venom are expressed in Figure 2. It was observed that the bothropic antivenom promoted an 85.8% reduction of the edema

**Table 4**  
Neutralization of the hemorrhagic activity of the venom of the *Bothrops atrox* and *Bothrops erythromelas*

Groups	Hemorrhagic area (mm <sup>2</sup> )	Protection (%)	Hemorrhagic area (mm <sup>2</sup> )	Protection (%)
	<i>Bothrops atrox</i>		<i>Bothrops erythromelas</i>	
Control (venom)	128.2±14.5	-	310.3±41.2	-
Heparin 3UI	169.2±14.1	-	399.2±56.5	-
Heparin 6 UI	207.3±22.3	-	401.3±109.9	-
Antithrombotic	78.6±22.2	38.7	130.3±37.2	57.9
Antithrombotic + heparin 3 UI	132.3±18.9	-	159.6±14.1	48.5
Antithrombotic + heparin 6 UI	138.0±6.1	-	157.3±30.7	49.3



**Fig. 1** - The effect of heparin (3 and 6 IU) in neutralizing the edematogenic activity of *Bothrops atrox* venom: Group I - the mice were injected with a dose of 3.6 µg/animal (3DME) of the *Bothrops atrox* venom; Group II - received the *Bothrops atrox* venom in a dose of 3.6 µg/animal and 3 IU of heparin; Group III - venom in a dose of 3.6 µg/animal and 6 IU of heparin; Group IV - venom in a dose of 3.6 µg/animal and Antithbotropic serum; Group V - venom in a dose of 3.6 µg/animal, Antithbotropic serum and 3 IU of heparin; Group VI - venom in a dose of 3.6 µg/animal, Antithbotropic serum and 6 IU of heparin.



**Fig. 2** - The effect of heparin (3 and 6 IU) in neutralizing the edematogenic activity of *Bothrops erythromelas* venom: Group I - the mice were injected with a dose of 4.2 µg/animal (3DME) of *Bothrops erythromelas* venom; Group II - received *Bothrops erythromelas* venom in a dose of 4.2 µg/animal and 3 IU of heparin; Group III - venom in a dose of 4.2 µg/animal and 6 IU of heparin; Group IV - venom in a dose of 4.2 µg/animal and Antithbotropic serum; Group V - venom in a dose of 4.2 µg/animal, Antithbotropic serum and 3 IU of heparin; Group VI - venom in a dose of 4.2 µg/animal, Antithbotropic serum and 6 IU of Heparin.

**Table 5**  
Neutralization of the phospholipase A<sub>2</sub> activity of the venom *Bothrops atrox* and *Bothrops erythromelas*

<i>Bothrops atrox</i>	<i>Bothrops erythromelas</i>	Heparin (IU)	Antithrotophic	Hemolysis halo (mm <sup>2</sup> )	Neutralization (%)
+	-	-	-	2.1	-
+	-	+	-	1.8	14.3
+	-	-	+	1.6	23.8
-	+	-	-	2.5	-
-	+	+	-	1.8	28.0
-	+	-	-	2.1	16.0

produced by this venom, in the sixth hour after injection. Heparin caused a significant anti-edematogenic effect only in the 6 IU dose. It is interesting to note that, up to 180 minutes, the effect of the heparin does not significantly differ from the neutralizing effect produced by the antithrotophic, having it promoted maximum neutralizing effect (52.9%) in 360 minutes. When associated to the antithrotophic venom, heparin induced according to the dose the following results: (a) with 3 IU dose, the neutralization did not differ from that produced by the antivenom, alone; (b) with 6 IU dose, the action of the antithrotophic improved, at the sixth hour (360 minutes), reducing the edema formation to 94.1%.

The neutralization of local action of *B. atrox* venom by the antithrotophic and by the heparin was less effective than *B. erythromelas* venom (Figures 1 and 2). This phenomenon can be explained considering the difference of the phospholipasic and proteolytic activities of both venoms. The indirect hemolytic activity (by phospholipase A<sub>2</sub>) of the venom is presented in Table 5, showing that the venom of *B. erythromelas* has a higher phospholipasic A<sub>2</sub> activity. *B. atrox* venom is known as one of the most proteolytic bothropic venoms<sup>3,10,28</sup>. Furthermore, it is more hemorrhagic than *B. erythromelas* venom. Therefore, it is possible to suggest that the edematogenic activity of *B. atrox* venom may be related with the proteolytic and hemorrhagic activities. On the other hand, the edematogenic activity of *B. erythromelas* venom may be due to a potent phospholipasic activity. The observation of necrosis, flicrens and hemorrhage on the paws of the animals that received the venom of the *Bothrops atrox* corroborates this hypothesis. Animals that received *Bothrops erythromelas* venom, besides edema, presented only a discreet local hemorrhagic action, without signs of macroscopic tissue destruction. According to this hypothesis, the intense tissue destruction produced by the *Bothrops atrox* venom (microvasculature, conjunctive tissue), along with the overcoming of the microtrombs<sup>18</sup> hinders the access of antibodies and heparin to the site of venom inoculation.

Finally, it remains to explain how heparin contributes to reduce the levels of edema produced by *B. erythromelas* venom. It is known that heparin is constituted of radical sulfates, which make it possible to interact electrostatically with diverse proteic locations of a basic character, including phospholipase A<sub>2</sub>, which is present in the *Bothrops atrox*, *B. erythromelas* and in diverse venom of other kinds and species of snakes<sup>21</sup>. Heparin was capable of neutralizing the phospholipasic activity of the *Bothrops atrox* and *Bothrops erythromelas* venom on an average of 14.3% and 28.0% respectively (Table 5). Given that fact, the *Bothrops erythromelas* venom produces less local tissue destruction allowing heparin access to the inoculation site where, by electrostatic interaction, it would reduce the phospholipasic activity of *Bothrops erythromelas* venom, with a consequent reduction of edema.

**Study of Neutralizing Effect of Heparin on Lethality:** Heparin (dose 3 IU), prolonged Time of Death of the animals which received the *Bothrops atrox* and *Bothrops erythromelas* venom, but did not keep the animals alive (Table 6). The animals that received the venom with heparin presented macroscopic lesions of greater intensity. It is interesting to point out that there was a difference in the macroscopic lesions provoked by the venom; those that received the *Bothrops atrox* venom died with rhinorrhagia, not present in the control animals and in those which received the *Bothrops erythromelas* venom.

**Table 6**  
Effect of the Heparin (3 IU) on time of death

Groups	Time of Death (min)	
	<i>Bothrops atrox</i>	<i>Bothrops erythromelas</i>
Control (Venom)	9.0 ± 2.6	5.3±0.3
Heparin 3 UI	26.7 ± 18.4	11.2±1.9
Antithrotophic	-	*
Antithrotophic + Heparin 3UI	-	**

(-) did not die; (\*) protection of 58%; (\*\*) protection of 92%

Necropsy revealed that envenomed mice treated with heparin presented, when compared with control animals: (a) more intense intracranial hemorrhaging; (b) macroscopic signs of intense pulmonary hemorrhaging; (c) a higher increase in the weights of lungs and brain. However, the increase was not statistically significant. On the other hand, the hystological sections of the animals injected with *Bothrops atrox* venom confirmed the intensity of lesions observed macroscopically, as well as confirmed that heparin potentialized the hemorrhagic activity, even in small doses (3IU) (personal communication of Dr. Luiz Carlos de Lima Ferreira – Laboratório de Patologia, Instituto de Medicina Tropical do Amazonas, Manaus, Amazonas, Brazil).

Many authors reported intracranial hemorrhaging produced by ophidian accidents<sup>4,14,31</sup>. KOUYOUMDJIAN<sup>20</sup>, showed six cases of patients with serious bothropic poisoning experiencing intracranial hemorrhaging, where four led to death, one survived with sequel and one without it. This indicates that the poisoning produced in Time of Death experiment was serious. Comparing the groups by intensity of the hemorrhagic phenomena, it suggests that heparin can increase the gravity of the accidents.

The antithrotophic serum was more effective in neutralizing the lethality of the *Bothrops atrox* venom (100% protection), but promoted

only 58% protection against the lethality of the *Bothrops erythromelas* venom. Heparin increased the efficacy of the bothropic antivenom for *Bothrops erythromelas* when associated with this antivenom (92% protection). A possible explanation could be that heparin interacted with diverse fractions of the venom making the efficacy of the antibothropic better, considering that heparin does not interfere with the ability of the antibothropic in neutralizing the hemorrhagic activity of *Bothrops erythromelas* venom (Table 4).

The differences observed in the neutralization of the activities of both venom calls into question the efficacy of the antibothropic serum supplied by the Ministry of Health. The Antibothropic Serum (ABS) was less effective in neutralizing the lethal activity of *Bothrops erythromelas* venom. This snake is responsible for most of the ophidian accidents in Northeast of Brazil and its venom is not included in the production of this immunobiologic.

## RESUMO

### Associação de heparina e antiveneno: eficácia da neutralização dos venenos de *Bothrops atrox* e *Bothrops erythromelas*

A heparina tem sido utilizada no tratamento dos acidentes botrópicos em algumas regiões do Brasil, porém, os dados encontrados na literatura são inconclusivos sobre sua eficácia. Os venenos de *Bothrops atrox* e de *B. erythromelas* foram caracterizados segundo suas atividades biológicas. A capacidade da heparina em neutralizar estas atividades foi testada com as doses de 3 e 6UI de forma isolada e associada ao soro Antibotrópico (SAB). Verificou-se que a heparina nas doses de 3 e 6UI, não foi eficaz em neutralizar a atividade desfibrinante e edematogênica do veneno de *B. atrox* e as ações hemorrágica e coagulante dos dois venenos. A heparina diminuiu a eficácia do soro SAB na neutralização das atividades hemorrágica e edematogênica do veneno de *B. atrox*. Contudo, a heparina na dose de 6UI foi capaz de neutralizar a atividade edematogênica do veneno de *B. erythromelas* e aumentar a eficácia do soro SAB. A heparina neutralizou ainda a atividade fosfolipásica A<sub>2</sub> dos venenos de *B. atrox* (14,3%) e de *B. erythromelas* (28,0%). Para o veneno de *B. erythromelas*, o tratamento associado de heparina e soro SAB, foi mais eficaz na neutralização da atividade letal.

## REFERENCES

- AHUJA, M.L. & SINGH, G. - Snake bite in India. **Indian J. med. Res.**, **42**: 661-686, 1954.
- AHUJA, M.L.; VEERARAGHAVAN, N. & MENON, I.G.K. - Action of heparin on the venom of *Echis carinatus*. **Nature (Lond.)**, **158**: 878, 1946.
- ASSAKURA M.T.; FURTADO, M.F. & MANDELBAUMM, F.R. - Biochemical and biological differentiation of the venoms of Lanceheads vipers (*B. atrox*, *B. asper*, *B. marajoensis* and *B. moojeni*). **Comp. Biochem. Physiol.**, **102b**: 727-732, 1992.
- BURKE, C.W. - The anterior pituitary, snake bite and Sheehan's syndrome. **Quart. J. Med.**, **75**: 331-333, 1990.
- CARDOSO, J.L.C. - Bothropic accidents. **Mem. Inst. Butantan**, **52** (suppl.): 43-44, 1990.
- CARDOSO, J.L.C.; FAN, H.W.; FRANÇA, F.O.S. *et al.* - Randomized comparative trial of three antivenoms in the treatment of envenoming by lance-headed vipers (*Bothrops jararaca*) in São Paulo, Brazil. **Quart. J. Med.**, **86**: 315-325, 1993.
- CONDREA, E. & DE VRIES, A. - Hemolysis and splitting of human erythrocyte phospholipids by snake venoms. **Biochim. Biophys. Acta**, **84**: 60-73, 1964.
- DICCIANNI, M.B.; MISTRY, M.J.; HUG, K. & HARMONY, J.A.K. - Inhibition of phospholipase A<sub>2</sub> by heparin. **Biochim. Biophys. Acta**, **1046**: 242-248, 1990.
- DICCIANNI, M.B.; LILLY STAUDERMAN, M.; MCLEAN, L.R.; BALASUBRAMANIAM, A. & HARMONY, J.A.K. - Heparin prevents the binding of phospholipase A<sub>2</sub> to phospholipid micelles: importance of the amino-terminus. **Biochemistry**, **30**: 9090-9097, 1991.
- FERREIRA, M.L.; MOURA-DA-SILVA, A.M.; FRANÇA, F.O.S.; CARDOSO, J.L.C. & MOTA, I. - Toxic activities of venoms from nine *Bothrops* species and their correlation with lethality and necrosis. **Toxicol.**, **30**: 1603-1608, 1992.
- FINNEY, D.J. - **Probit analysis**. 3. ed. Cambridge, Cambridge University Press, 1971.
- FRANSON, R.; PATRIARCA, P. & ELSBACH, P. - Phospholipid metabolism by phagocytic cells. Phospholipases A<sub>2</sub> associated with rabbit polymorphonuclear leukocyte granules. **J. Lipid Res.**, **15**: 380-388, 1974.
- GENÉ, J.A.; ROY, A.; ROJAS, G.; GUTIÉRREZ, J.M. & CERDAS, L. - Comparative study on coagulant, desfibrinating, fibrinolytic and fibrogenolytic activities of Costa Rican crotaline snake venoms and their neutralization by a polyvalent antivenom. **Toxicol.**, **27**: 841-848, 1989.
- GUPTA, P.K. - Bilateral thalamic hematoma following snake bite [letter]. **J. Ass. Phycns. Índia**, **40**: 549-550, 1992.
- GUTIÉRREZ, J.M.; CHAVES, F.; BOLAÑOS, R. *et al.* - Neutralización de los efectos locales del veneno de *Bothrops asper* por un antiveneno polivalente. **Toxicol.**, **19**: 493-500, 1981.
- GUTIÉRREZ, J.M.; GENÉ, J.A.; ROJAS, G. & CERDAS, L. - Neutralization of proteolytic and hemorrhagic activities of Costa Rican snake venoms by a polyvalent antivenom. **Toxicol.**, **23**: 887-893, 1985.
- GUTIÉRREZ, J.M.; AVILA, C.; ROJAS, G. & CERDAS, L. - An alternative *in vitro* method for testing the potency of the polyvalent antivenom produced in Costa Rica. **Toxicol.**, **26**: 411-413, 1988.
- GUTIÉRREZ, J.M. & LOMONTE, B. - Local tissue damage induced by *Bothrops* snakes. **Mem. Inst. Butantan**, **51**: 211-223, 1989.
- HORIGOME, K.; HAYAKAWA, M.; INOUE, K. & NOJIMA, S. - Purification and characterization of phospholipase A<sub>2</sub> released from rat platelets. **J. Biochem.**, **101**: 625-631, 1987.
- KOUYOUMDJIAN, J.A. - Intracranial haemorrhage after snake bite. **Mem. Inst. Butantan**, **52**(supl.): 45, 1990.
- LANE, D.A. & ADAMS, L. - Non-anticoagulant uses of heparin. **New Engl. J. Med.**, **329**: 129-130, 1993.
- LOMONTE, B. - Tissue damage and inflammation induced by snake venoms. Sweden, University of Göteborg, 1994.
- MELO, P.A. & SUAREZ-KURTZ, G. - Release of sarcoplasmic enzymes from skeletal muscle by *Bothrops jararacussu* venom: antagonism by heparin and by the serum of South American marsupials. **Toxicol.**, **26**: 87-95, 1988.
- MELO, P.A.; HOMSI-BRANDEBURGO, M.I.; GIGLIO, J.R. & SUAREZ-KURTZ, G. - Antagonism of the myotoxic effect of *Bothrops jararacussu* venom and Bothropstoxin by polyanions. **Toxicol.**, **31**: 285-291, 1993.
- MINISTÉRIO DA SAÚDE & FUNDAÇÃO NACIONAL DE SAÚDE - Manual de diagnóstico e tratamento de acidentes por animais peçonhentos. Brasília, 1998.
- MORAIS, J.F.; FREITAS, M.C.W.; YAMAGUCHI, I.K.; DOS-SANTOS, M.C. & DIAS DA SILVA, W. - Snake antivenoms from hyperimmunized horses: comparison of the antivenom activity and biological properties of their whole IgG and F(ab')<sub>2</sub> fragments. **Toxicol.**, **32**: 725-734, 1994.

27. NAHAS, L.; KAMIGUTI, A.S.; RZEPPA, H.W.; SANO, I.S. & MATSUNAGA, S. - Effect of heparin on the coagulant action of snake venoms. **Toxicon**, **13**: 457-463, 1975.
28. SANCHEZ, E.F.; FREITAS, T.V.; FERREIRA-ALVES, D.L. *et al.* - Biological activities of venoms from South American snakes. **Toxicon**, **30**: 95-103, 1992.
29. THEAKSTON, R.D.G. & REID, H.A. - Development of simple standard assay procedures for the characterization of snake venoms. **Bull. Wld. Hlth. Org.**, **61**: 949-956, 1983.
30. TINOCO, R.C. - Picada de cobra (uso de heparina como substituto de soro antiofídico). **J. bras. Med.**, **23**: 109-114, 1972.
31. TRAVETT, A.J.; NWOKOLO, N.C.; KEVAU, I.H. & SEATON, R.A. - Cerebrovascular accident after Taipan bite [letter]. **Med. J. Aust.**, **160**: 94, 1994.
32. YAMAKAWA, M.; NOZAKI, M. & HAKAMA, Z. - Fractionation of sakishima-habu (*Trimeresurus elegans*) venom and lethal haemorrhagic and edema-forming activities of the fraction. **Animal Plant & Microbial Toxins**, **1**: 98-111, 1976.
33. ZHOU, F.; HÖÖK, T.; THOMPSON, J.A. & HÖÖK, M. - Heparin protein interactions. In: LANE, D.A.; BJÖRK, I. & LINDADAHN, U., ed. **Heparin and related polysaccharides**. New York, Plenum Press, 1992. p.141-153.

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